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# The Protochordate *Ciona intestinalis* Has a Protein Like Full-Length Vertebrate Fibrinogen

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#### **Key Words**

Fibrinogen · Fibrin · Protochordate · Sea squirt

#### Abstract

In a recent review, a putative fibrinogen-like protein in the protochordate *Ciona intestinalis* was noted. Unfortunately, computer-directed splicing had omitted several exons, mistakenly generating a single long polypeptide chain. In fact, 3 consecutive genes exist, the translated versions of which are homologous to individual vertebrate fibrinogen chains. The circulating form is likely a 6-chain covalent dimer, just as occurs in vertebrates. Copyright © 2012 S. Karger AG, Basel

#### Introduction

In a recent minireview on the evolution of coagulation and inflammation [1], attention was called to a putative protein in the protochordate *Ciona intestinalis*, commonly called the sea squirt, that had many features characteristic of vertebrate fibrinogens. Although homologs of the carboxyl domains of  $\beta$  and  $\gamma$  chains of fibrinogen are widespread in the animal world [2–4], the cited protein was unique in that it also had sequences consistent with

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Accessible online at: www.karger.com/jin 'coiled coils,' as well as constellations of cysteines of the sort found in 'disulfide rings' known to bind 3 polypeptide chains symmetrically [5, 6]. The protein had initially been dismissed as just one of many fibrinogen-related domains during a search [7] of the draft genome of the C. intestinalis genome [8]. Subsequently, the highest scoring of those early candidates, found on scaffold 73, was backsearched against the GenBank nonredundant protein database [1], by which time it had been annotated by the NCBI (identification No. XP\_002122759). The protein sequence had been generated by automated computational analysis from genomic sequence NW001955408 using the NCBI gene prediction tool GNOMON; it was said to have been supported by EST evidence and was predicted to be a single polypeptide chain composed of 1,160 amino acids.

However, an attempt to model the single-chain protein made it clear that the prediction was structurally improbable, especially in its central region. Accordingly, a manual reevaluation of the DNA sequence was undertaken, the results of which revealed 5 additional exons, including 2 encoding signal peptides. In the end, a 3-gene system was identified that encodes a protein with 3 nonidentical polypeptide chains, likely a covalent dimer, that is remarkably similar to vertebrate fibrinogen.

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**Fig. 1. a** Exon distribution in 3 fibrinogen-like genes in *C. intestinalis.* Asterisks denote exons not found in NCBI entry XP\_002122759; S indicates exons encoding signal peptides. **b** Translated sequences of newly uncovered exons in *C. intestinalis* fibrinogen-related genes. In the cases of exons 6, 9, and 10, dotted residues denote identical residues in homologous sequences

#### Methods

The complete genome of the sea squirt, *C. intestinalis*, was downloaded, after which the central region of scaffold 73 of the original draft assembly [8] was cut out and translated into all 6 reading frames. Each of these was inspected with a variety of vertebrate fibrinogen chains in search of homologous segments. Extensive use was made of the search tool BLAST [9], as well as older in-house software [10]. The region was also examined meticulously for conventional splice sites and poly-A tails. Newly predicted gene products were aligned and phylogenetic trees constructed.

#### Results

Five new exons were uncovered, including 2 encoding signal sequences (fig. 1a, b). Two poly-A tails were also identified in the regions upstream of the new signal peptide-containing exons. It has been found that automated

from human fibrinogen. **c** Diagram showing putative connections between cysteines to form disulfides. Arrows near the amino terminals of A and B chains show connections to B' and A' chains in putative dimers. **d** Phylogenetic tree showing relationships of the three predicted *C. intestinalis* polypeptides with those of  $\beta$  and  $\gamma$ chains of human and lamprey fibrinogens.

splicing regimens are particularly bad at finding signal sequences [11]. Even with the manual curation, some lesser ambiguities remain with regard to the start and end of some introns, particularly in the  $\alpha$  chain where several anomalous cysteines occur. Nonetheless, it was clear that 3 genes are tightly packed within a 10-kB stretch. For the moment, the genes and their products are denoted A, B, and C rather than risk the implication that any one correlates better with the  $\alpha$ ,  $\beta$ , and  $\gamma$  chains of vertebrate fibrinogen.

#### Structural Aspects

The distribution of cysteine residues is consistent with all of the carboxyl domains of all three chains having the same disulfide bridging as occurs in the  $\beta$  chain of vertebrate fibrinogens (fig. 1c). Elsewhere, all three chains have a brace of cysteines near their amino termini that in vertebrate fibrinogen are known to form the 6-cysteine con-

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stellation known as a 'disulfide ring.' In vertebrate fibrinogen, such disulfide rings set the boundaries for coiled coils. Further, one disulfide ring from each half molecule packs against the other to form short  $\beta$  sheets at the dimer interface [6]. Additionally, the vertebrate dimer depends on the  $\beta$  chain crossing over to the other half of the molecule to form a disulfide bond with the opposing  $\alpha$  chain and vice versa. Equivalent cysteine residues occur in the putative sea squirt protein A and B chains, making it likely that the circulating protein is a 6-chained covalent dimer (fig. 1c).

Nonetheless, several anomalously positioned cysteine residues remain, including 2 in the A chain and 2 in the C chain in the region predicted to form coiled coils. It is possible that unusual splicing patterns could remove or replace some of these.

The sea squirt protein appears to be heavily glycosylated, with the A chain having 2 putative asparaginyl-sugar sites, the B chain 4, and the C chain 6, amounting to 12 all together or 24 in a 6-chained dimer. The molecular weight of the mature predicted protein would be approximately 330 kD, of which 53 kD would be carbohydrate. Residues consistent with calcium binding sites observed in vertebrate and invertebrate [12] fibrinogen-related domains appear intact.

## Functional Aspects

None of the chains in the sea squirt protein has an amino terminal extension with a possible site for cleavage by thrombin. Nor do the carboxyl terminal regions have extensions with sites that might be cross-linked by factor XIII. On the other hand, all three carboxyl-terminal domains have sequences compatible with binding sites ('holes') of the sort that in vertebrate fibrinogen bind peptide knobs. In most fibrinogen-related proteins, these 'holes' bind sugars or sialic acids [12], and the same is likely true of the sea squirt protein. In this regard, 2 of the 3 sites lack key residues involved in the binding of arginine-containing peptides. There is a single RGD sequence in the C chain; in mammals RGD sequences are involved in fibrinogen binding to platelets.

## Phylogenetic Relationships

Although two of the sea squirt polypeptide chains appear slightly more similar to vertebrate  $\gamma$  chains (fig. 1d), the distinction is slight and is offset by the disulfide arrangement of carboxyl domains of all three being characteristic of vertebrate fibrinogen  $\beta$  chains, which have an additional disulfide compared with  $\gamma$  chains.

The approximate time of the various gene duplications can be gauged from the resemblances between the three

*Ciona* sequences and the  $\beta$  and  $\gamma$  chains of various vertebrate fibrinogens. As it happens the three *Ciona* polypeptides have approximately the same degree of resemblance to each other (33.7%) as they do to vertebrate fibrinogen  $\beta$  and  $\gamma$  chains (32.7%) and that vertebrate  $\beta$  and  $\gamma$  chains have to each other (34.6%). It seems clear that the gene duplications leading to the evolution of the three *Ciona* polypeptides and the  $\beta$  and  $\gamma$  chains of vertebrate fibrinogen occurred within the same general time frame.

## Discussion

The absence of likely thrombin cleavage sites near any of the three amino termini suggests the putative fibrinogen-like protein in *C. intestinalis* is not transformed into an extracellular gel. This is consistent with longstanding observations that fibrin clots are not found in protochordates [13]. The lack of extended carboxyl terminal segments containing lysine and glutamine further implies that the protein is not a target for transglutaminase crosslinking.

However, urochordates like *C. intestinalis* do have circulating cells that are known to clump at sites of injury [13], and the protein could very well play a role in that process. In vertebrates, fibrinogen is well known to bind to platelets and various white cells quite independently of its ability to be changed into fibrin. Such interactions could be mediated either by the RGD sequence present in the C chain or by the binding of cryptic cell surface moieties, exposed by traumatic circumstances or infection, perhaps, to the molecule's well conserved 'holes'. Whatever its role, the protein is clearly derived from the same ancestral stock as vertebrate fibrinogen.

It has long been presumed that the prototypic vertebrate fibrinogen was composed of 3 identical chains, with globular carboxyl terminal regions for all three, in contrast to extant vertebrate fibrinogens which have only 2 globular domains at their termini. The conjecture was that the  $\alpha$  chain suffered a genetic dislocation early in vertebrate evolution, the result of which was the mostly unfolded carboxyl terminal region that exists today [14]. Models of how that ancient protein would have been configured have been proposed [15].

Ciona Fibrinogen-Like Protein

#### References

- 1 Doolittle RF: Coagulation in vertebrates with a focus on evolution and inflammation. J Innate Immun 2011;3:9–16.
- 2 Doolittle RF: A detailed consideration of a principal domain of vertebrate fibrinogen and its relatives. Protein Sci 1992;1:1563–1577.
- 3 Wang X, Zhao Q, Christensen BM: Identification and characterization of the fibrinogen-like domain of fibrinogen-related proteins in the mosquito, *Anopheles gambiae*, and the fruitfly, *Drosophila melanogaster*, genomes. BMC Genomics 2005;6:114.
- 4 Zhang SM, Zeng Y, Loker ES: Expression profiling and binding properties of fibrinogen-related proteins (FREPs), plasma proteins from the schistosome snail host *Biomphalaria glabrata*. Innate Immunol 2008;14 175–189.
- 5 Doolittle RF: Structural aspects of the fibrinogen to fibrin conversion. Adv Protein Chem 1973;27:1–105.

- 6 Yang Z, Kollman JM, Pandi L, Doolittle RF: Crystal structure of native chicken fibrinogen at 2.7 Å resolution. Biochemistry 2001; 40:12515–12523.
- 7 Jiang Y, Doolittle RF: The evolution of vertebrate blood coagulation as viewed from a comparison of puffer fish and sea squirt genomes. Proc Natl Acad Sci USA 2003;100: 7527–7532.
- 8 Dehal P, Satou Y, Campbell RK, et al: The draft genome of *Ciona intestinales*: insights into chordate and vertebrate origins. Science 2002;298:2157–2167.
- 9 Altschul SF, Madden T, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ: Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res 1997;25:3389–3402.
- 10 Doolittle RF: Of URFs, ORFs: A Primer on How to Analyze Derived Amino Acid Sequences. Mill Valley, University Science Books, 1987.
- 11 Nagy A, Hegyi H, Farkas K, Tordai H, Kozma E, Banyai L, Patthy, L: Identification and correction of abnormal, incomplete and mispredicted proteins in public databases. BMC Bioinformatics 2008;9:353.

- 12 Kairies N, Beisel HG, Fuentes-Prior P, Tsuda R, Muta T, Iwanaga S, Bode W, Huber R, Kawabata S: The 2.0-A crystal structure of tachylectin 5A provides evidence for the common origin of the innate immunity and the blood coagulation systems. Proc Natl Acad Sci USA 2001;98:13519–13524.
- 13 Rowley AF, Rhodes CP, Ratcliffe NA: Protochordate leucocytes: a review. Zool J Linn Soc 1984;80:283–295.
- 14 Doolittle RF, Watt KW, Cottrell BA, Strong DD, Riley M: The amino acid sequence of the α-chain of human fibrinogen. Nature 1979; 280:464–468.
- 15 Doolittle RF, Spraggon G, Everse SJ: Evolution of vertebrate fibrin formation and the process of its dissolution; in Bock GR, Goode JA (eds): Plasminogen-Related Growth Factors, CIBA Foundation Symposium 212. New York, Wiley, 1997.

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