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# Hyperspectral imaging for head and neck cancer detection: specular glare and variance of the tumor margin in surgical specimens

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**Abstract.** Head and neck squamous cell carcinoma (SCC) is primarily managed by surgical cancer resection. Recurrence rates after surgery can be as high as 55%, if residual cancer is present. Hyperspectral imaging (HSI) is evaluated for detection of SCC in *ex-vivo* surgical specimens. Several machine learning methods are investigated, including convolutional neural networks (CNNs) and a spectral–spatial classification framework based on support vector machines. Quantitative results demonstrate that additional data preprocessing and unsupervised segmentation can improve CNN results to achieve optimal performance. The methods are trained in two paradigms, with and without specular glare. Classifying regions that include specular glare degrade the overall results, but the combination of the CNN probability maps and unsupervised segmentation using a majority voting method produces an area under the curve value of 0.81 [0.80, 0.83]. As the wavelengths of light used in HSI can penetrate different depths into biological tissue, cancer margins may change with depth and create uncertainty in the ground truth. Through serial histological sectioning, the variance in the cancer margin with depth is investigated and paired with qualitative classification heat maps using the methods proposed for the testing group of SCC patients. The results determined that the validity of the top section alone as the ground truth may be limited to 1 to 2 mm. The study of specular glare and margin variation provided better understanding of the potential of HSI for the use in the operating room. © 2019 Society of Photo-Optical Instrumentation Engineers (SPIE) [DOI: 10.1117/1.JMI.6.3.035004]

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## 1 Introduction

Head and neck (H&N) cancer is the sixth most-common cancer worldwide, with majority of cancers of the upper aerodigestive tract, comprising the oral and nasal cavities, pharynx, and larynx, being squamous cell carcinoma (SCC).<sup>1</sup> Approximately two-thirds of the SCC patients present with advanced disease, either stage III or IV.<sup>2</sup> Surgical resection is the primary management of SCC of the aerodigestive tract, potentially with concurrent chemoradiation therapy depending on the extent of the disease.<sup>3</sup> The local recurrence rates for SCC cases depend on the successful removal of the cancer. For surgeries with negative cancer margins, the local recurrence rate is 12% to 18%, compared to local recurrence rates of up to 30% to 55%, if positive cancer margins are determined.<sup>4–6</sup> Moreover, positive cancer margins have a greatly reduced disease-free survival, with estimates ranging from 7% to 52%, compared to disease-free survival rates of 39% to 73% for negative margins.<sup>7,8</sup> Disease recurrence greatly affects the likelihood for additional surgeries,

reduced quality of life, complications from surgery, and increased mortality rates.<sup>9</sup>

Hyperspectral imaging (HSI) is a noncontact optical imaging modality capable of acquiring a single image of potentially hundreds of discrete wavelengths. Preliminary research demonstrates that HSI has the potential for providing diagnostic information for various diseases.<sup>10</sup> Preliminary studies from our group show that HSI combined with machine learning (ML) may yield diagnostic information with potential applications for surgical use in head and neck cancers.<sup>11–14</sup> Fabelo et al.<sup>15–17</sup> have demonstrated the need and utility of HSI for *in-vivo* brain cancer detection and developed a visualization system that could lead to near-real-time guidance by using a machine learning-based classification algorithm. Moreover, the use of deep learning techniques to process the same dataset was studied, demonstrating that they outperform the traditional machine learning methods.<sup>17</sup> Many applications of HSI for gastrointestinal procedures have been explored, including anatomical organ identification, anastomosis and ischemia classification, and cancer detection.<sup>18</sup> Recently, the implementation of HSI for guided

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colorectal surgery as a replacement for tactile information lost in laparoscopic surgeries has been explored in an *ex-vivo* study, and Baltussen et al.<sup>19</sup> demonstrated that HSI could distinguish between normal healthy colon wall and colorectal adenocarcinoma. Much work has been explored in head and neck HSI as well. Salmivuori et al.<sup>20</sup> detailed the assistance of HSI in ill-defined, cutaneous basal cell carcinomas of the head and neck, and the authors contend that HSI offers clinical utility in more accurately assessing margins. Farah et al.<sup>21</sup> demonstrated that narrow-band imaging (NBI) at green (400 to 430 nm) and blue (525 to 555 nm) visible light could reveal the extent of oral SCC under white light. Nonencoding portions of RNA called micro-RNA function to regulate the expression of DNA.<sup>22</sup> Moreover, Farah et al.<sup>23,24</sup> obtained micro-RNA and mRNA expression levels from both the primary tumor core and the near-tumor normal tissues; it was determined that NBI spatially correlated with tumor-like expression levels in detecting the abnormality in near-tumor normal tissues.

Machine learning methods, including support vector machines (SVMs) and convolutional neural networks (CNNs), the latter being an implementation of artificial intelligence, have demonstrated near human-level ability for image classification tasks.<sup>25,26</sup> These experiments have been conducted on gross-level HSI acquired of *ex-vivo* tissue specimens from patients undergoing surgical cancer resection. However, HSI uses wavelengths of light that can penetrate different depths into the biological tissue, so it is possible that the superficial cancer margin may change with depth and create uncertainty in ground truth, which is used to obtain evaluation metrics for these experiments.

Selected previous works from our group performing head and neck squamous cell carcinoma (HNSCC) detection in *ex-vivo* samples from the upper aerodigestive tract used only single-class tissue specimens (purely tumor or normal tissues) to demonstrate classification potential with HSI.<sup>27,28</sup> Lu et al.<sup>14</sup> incorporated tumor-involved cancer margin specimens from HNSCC, but these methods were implemented on selected regions of interest (ROIs) of normal and tumor near the margin. Therefore, the area under the curve (AUC) reported for HNSCC detection thus far from our group ranges from 0.8 to 0.95, but these results may be limited to ROIs or single-class tissues. Similarly, Manni et al.<sup>29</sup> used ROIs from tumor-margin specimens of seven patients with tongue SCC and obtained an AUC of 0.92. Weijtmans et al.<sup>30</sup> developed a deep learning architecture that separately extracts spectral and spatial features from HSI, a dual-streamed approach, also validated on seven patients with tongue SCC; the proposed model performed better with both feature streams (AUC of 0.90) compared to individual stream. A few works have investigated SCC detection at the actual cancer margin; Halicek et al.<sup>31</sup> performed SCC detection at the cancer margin, upon which the proposed method in this paper expands, but this previous work was limited by ignoring regions of specular glare. Trajanovski et al.<sup>32</sup> performed semantic segmentation with deep learning of entire *ex-vivo* tissue specimens with excellent validation performance (AUC of 0.93) on gross specimens of the cancer margin from 14 tongue SCC samples.

This study aims to investigate the ability of HSI to detect SCC in surgical specimens from the upper aerodigestive tract using several distinct machine learning pipelines. In addition, another objective of this work is to investigate the limiting factors of HSI-based SCC detection, including specular glare, noise and blur, and uncertainty in the ground truth due to changes in

superficial cancer margin with depth, all of which must be thoroughly explored to understand the potential of HSI in the operating room. A preliminary version of this work was presented at the 2019 SPIE Medical Imaging Conference.<sup>31</sup> The contributions of this journal paper include a proposed algorithm combining deep learning and unsupervised machine learning; additionally, five testing patients have been serially sectioned to reveal changes in the cancer margin. This work expands upon previous cross-validation experiments with CNN-only methods on our H&N dataset to include multiple machine learning pipelines involving CNNs and other state-of-the-art methods. The proposed methods are tested on five HSI of five SCC patients, and the accuracy of the corresponding ground truths from these five tissues are discussed in detail along with potential directions for incorporating the outcomes of this work to improve future studies with HSI for cancer detection.

## 2 Methods

### 2.1 Experimental Design

In collaboration with the Otolaryngology Department and the Department of Pathology and Laboratory Medicine at Emory University Hospital Midtown, head and neck cancer patients undergoing surgical cancer resection were recruited for our HSI studies. Written and informed consent was obtained from all patients before acquiring surgical tissue specimens for inclusion in our study, which was used for research purposes only and deidentified by a clinical research coordinator. The experimental methods and protocols were approved by the Institutional Review Board at Emory University. In previous studies, we have evaluated the efficacy of using HSI for optical biopsy of head and neck tissues.<sup>11,33</sup> Excised tissue samples of HNSCCs and normal tissues were collected from the upper aerodigestive tract sites, including tongue, larynx, pharynx, and mandible. Three tissue samples, with approximate size of  $10 \times 10 \times 3$  mm, were collected from each patient: a sample of the primary tumor specimen, a normal tissue sample, and a sample at the tumor-involved cancer margin with adjacent normal tissue; these specimens were scanned with an HSI system.<sup>14,28</sup>

In this study, we selected 26 tissue specimens from 12 patients with moderate-to-poorly differentiated primary SCC of the upper aerodigestive tract for this analysis, including primary origin sites of the larynx, pharynx, tongue, floor of mouth, alveolar ridge, buccal mucosa, and maxillary sinus. The patients were divided into two groups, cross-validation and testing groups. The first seven patients and corresponding 21 tissues samples were collected early in the course of this project that met the criteria of having the ideal distribution of tumor, tumor-normal, and normal tissues and were used for evaluating quantitative results, expanding from previously published results with this group of patients.<sup>13</sup> In addition, five tumor-involved margin tissue specimens from five patients acquired at the end of the data collection period were selected to comprise the testing group. These tissue specimens were selected because they had approximately equal amounts of cancer and normal tissues in each specimen and were verified by the collaborating pathologist as of interest to undergo serial histological sectioning through the depth of the tissue. These five patients' tissues were classified using models trained from the first seven patient cross-validation group (following a leave-one-patient-out cross-validation method), and these five patients' results are presented

qualitatively to compare with the variability in the superficial cancer margin.

## 2.2 Hyperspectral Imaging

Hyperspectral images were acquired of *ex-vivo* surgical specimens using a previously described CRI Maestro imaging system (Perkin Elmer Inc., Waltham, Massachusetts), which captures images having  $1040 \times 1392$  pixels and a spatial resolution of  $25 \mu\text{m}/\text{pixel}$ .<sup>11,28,34,35</sup> Each hypercube contains 91 spectral bands, ranging from 450 to 900 nm with a 5-nm spectral sampling interval. The HS data were normalized by a standard white–dark calibration normalization, which involved subtracting the inherent dark current from the measured spectra and dividing it by a white reference spectra for all wavelengths sampled for all pixels.<sup>11,35</sup> Figure 1 shows the RGBs, grayscale images at representative bands, and spectral signatures of the patients' HSI data. The grayscale images at selected bands highlight the choice of the cutoffs due to noise at the ends of the broadband spectrum. The HSI was used to construct RGB composite images by implementing a Gaussian function centered on each color component of the spectrum: red (625 to 700 nm), green (520 to 560 nm), and blue (450 to 490 nm).

## 2.3 Histological Imaging

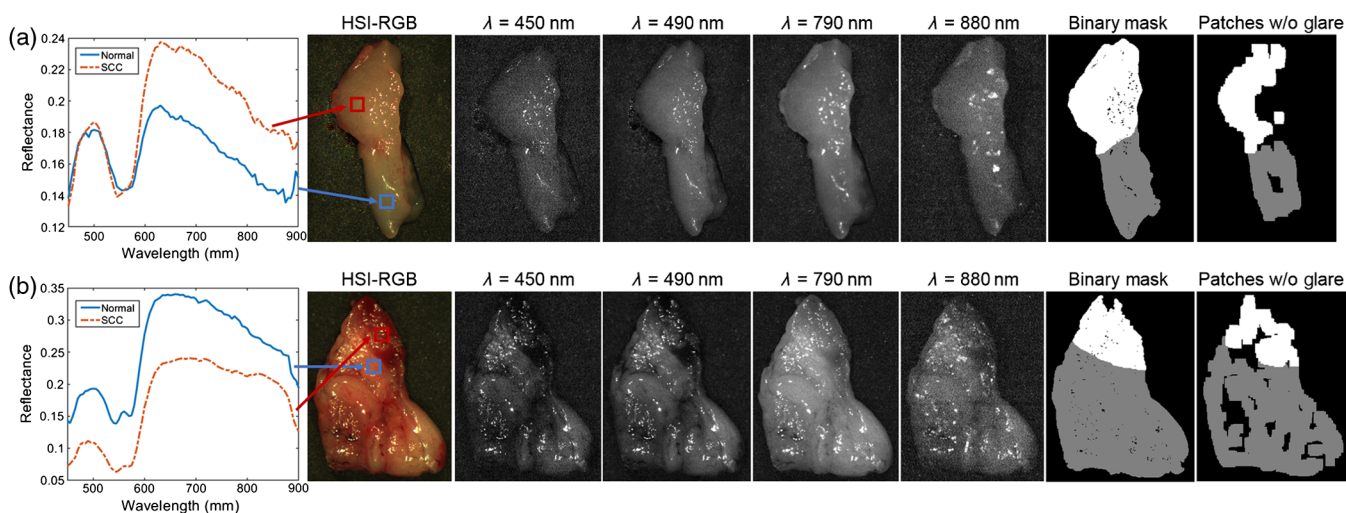
To obtain a ground-truth labeling for the tissue specimens imaged with HSI, tissues were inked after imaging to preserve orientation, fixed in formalin, paraffin embedded, sectioned from the top of the imaging surface, hematoxylin and eosin stained, and digitized using a Hamamatsu Photonics NanoZoomer at  $40\times$  objective (specimen-level pixel size,  $0.23 \mu\text{m}$ ). The *ex-vivo* tissue sections were reviewed by a board-certified pathologist with expertise in H&N pathology and the cancer margins were annotated directly on the slide using digitized histology in Aperio ImageScope (Leica Biosystems Inc., Buffalo Grove, Illinois).

There exists the possibility for substantial deformation of the histological images during processing relative to the HSI setup. Previously, the registration challenges were explored by using a deformation-based image registration pipeline of the histological image with known registration landmarks, and an error of 0.5 mm for SCC tissues was calculated.<sup>36</sup> For the cross-validation group patients in this work, the histological images were automatically registered using this deformable registration pipeline to produce the HSI ground truth. However, to calculate the error of histology ground truths for HSI in millimeters, a more accurate registration is needed than the automated approach. For the testing group patients since each had six histological slices, we manually registered each histological image using mutual landmarks to the HSI that were serially sectioned to reduce error.

## 2.4 Hyperspectral Imaging Binary Ground Truth

Specular glare is detected in each spectral band by fitting a gamma distribution to the pixel intensities of that band. Next, the top 1% of intensities are identified in this distribution, and any pixels with intensities of these bins are identified as specular glare. There were two types of specular glare observed: near-infrared (NIR) and visible, which usually were spatially independent. Visible glare occurs between 450 and 790 nm, and NIR glare occurs after 800 nm, which can be seen in the selected bands in Fig. 1. A pixel was identified as a specular glare pixel and removed if there was a glare identified using the distribution method in any spectral band from 490 to 790 nm. This range was selected because bands before and after this range were removed during preprocessing. NIR glare is observed beyond this range, so it is not relevant.

Binary masks are constructed from the histological images and used to construct the ground truth for HSI in two methods. The first method is to investigate only ideal quality pixels, which is constructed by avoiding HSI regions with a large amount of specular glare. For the ground-truth mask avoiding glare, only regions that contain sufficient area to extract  $25 \times 25$  spatial patches without any specular glare are included. The second



**Fig. 1** Two representative tissue specimens from different patients. Left: spectral signatures of SCC and normal ROIs are shown for both patients in (a) and (b). The HSI-RGB composite images and selected spectral bands are shown to highlight the noisy band cutoffs during preprocessing. Binary ground-truth masks including glare regions, generated by only removing patches centered on specular glare is shown; binary ground-truth mask excluding glare regions, generated by sufficient area to extract  $25 \times 25$  patches and avoiding specular glare (white: SCC, and normal: gray).

method is to investigate the degrading effect of specular glare on classification accuracy. For the “ground-truth mask including glare” regions, the entire tissue area is included for patch-making, but the top 1% of glare pixels are identified by fitting a gamma distribution to pixel intensities, such that no patches are centered on a glare pixel. A binary mask of the specular glare pixels was generated, and patches were produced automatically using this mask in the regions around the specular glare. Figure 1 shows an example of the two masks generated from two different patients. Both masks will be used for evaluating quantitative testing results and are referred to as “ground-truth mask excluding glare” and ground-truth mask including glare, respectively. The specular glare pixels are represented by the black pixels within the first mask in the middle. The automatic patch-making process can be seen to avoid the regions of specular glare shown in the second mask on the right of Fig. 1.

## 2.5 Effect of Sectioning Depth on Cancer Margin Ground Truth

HSI uses wavelengths of light in the visible and NIR spectrum that can penetrate different depths into biological tissue. It is possible that the cancer margin may change with depth and create uncertainty in the classification results because of the variable penetration of the HSI broadband spectrum. To investigate this, additional histological sectioning was performed on five tumor-involved margin tissue specimens (see Fig. 2).

During histological sectioning, it is a standard procedure to discard the first sections until the first good-quality section is obtained that contains the entire perimeter of the tissue specimen. The five specimens used for additional histological sectioning were retroactively enrolled for serial sectioning, so it was unknown what initial depth of tissue was discarded to obtain the first good-quality slice. In discussion with the pathology laboratory technician, who performed the histological sectioning, and through observation of specimen sectioning, it was estimated that  $\sim 100$  to  $200 \mu\text{m}$  were discarded on average during initial sectioning before obtaining the first high-quality, tissue-encompassing slide.

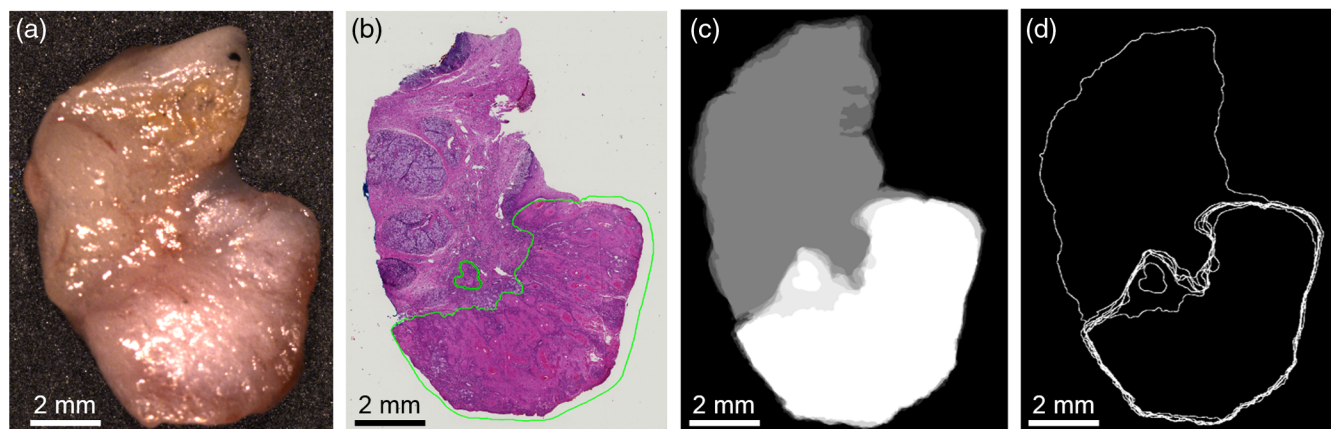
To produce serial histological sectioning further into the depth of the tissue specimens, the thickness of the remaining

paraffin-embedded tissue was estimated. The microtome used for this study produced slices at  $5 \mu\text{m}$ , so the number of additional sections was documented and the distances of additional depth could be measured. From the sectioned surface, which corresponds to the top of the tissue that was optically imaged, five more sections were obtained, up to  $300 \mu\text{m}$  further into the depth of the tissue, by discarding nonincluded slices. The extent of the additional serial sections were obtained for  $100$  to  $300 \mu\text{m}$  beyond the first good-quality slice; the exact values for the five tissue specimens’ total additional sectioning depths were  $100$ ,  $150$ ,  $200$ , and  $300 \mu\text{m}$ , depending on the unique tissue specimen. Therefore, the combined total estimate of additional sectioning is  $\sim 200$  to  $500 \mu\text{m}$  into the tissue depth from the original HS image surface. Penetration depth depends on the tissue composition and wavelength of light. This sectioning depth was investigated because it represents the effective penetration depth for biological tissue in the shorter wavelengths of HSI used, which correspond to the most relevant features for HSI classification, such as hemoglobin between  $550$  and  $600 \text{ nm}$ .<sup>37–39</sup>

## 2.6 Machine Learning Techniques

### 2.6.1 Data preprocessing

A further preprocessing chain was applied to the data mainly to reduce the noise in the spectral signatures (Fig. 3) and was compared with the standard white-reference calibration. The machine learning methods detailed below were tested with and without the following preprocessing steps. The proposed preprocessing chain is based on four steps: image calibration, operating bandwidth selection, noise filtering, and data normalization. In the first step, the previously described method to normalize the data using white reference and dark current is performed. Next, the spectral bands are truncated between  $490$  and  $790 \text{ nm}$ , so the final HS cube contains only  $61$  bands, which will be used for classification. Spectral bands outside this range were noisy because they were too close to the detectable limits of the HS camera. In the third step, a smoothing filter is applied to the data in order to reduce the spectral noise. A moving average with a span of  $10$  spectral bands was used as the smoothing filter, implemented as a sliding low-pass filter with



**Fig. 2** Representative tissue specimen that underwent serial histological sectioning to evaluate cancer margin variation with tissue depth. (a) HSI-RGB composite image; (b) first histological slice (the green outlined area is the pathologist annotation of the SCC region); (c) combined merged image of the six binary masks from the six histological images (cancer shown in white, normal in gray); (d) merged image of the SCC contours from the six histological images. The two rightmost images are used to depict the total variation of the cancer margin.

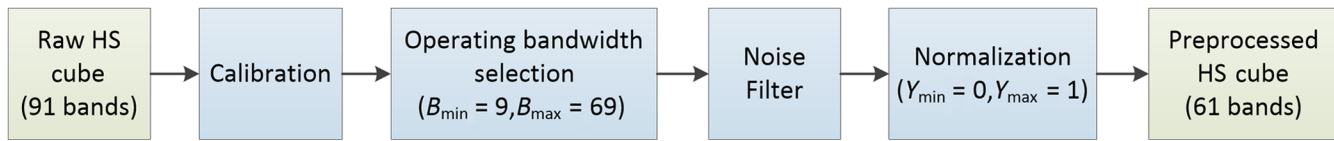


Fig. 3 Block diagram of the proposed preprocessing chain.

coefficients equal to the reciprocal of the span. Finally, each pixel's spectral signature is normalized between 0 and 1, using a function to map the minimum and maximum to these values.

### 2.6.2 Convolutional neural networks

A deep CNN was used to detect SCC in the *ex-vivo* specimens of the upper aerodigestive tract, implemented using TensorFlow in Python on a NVIDIA Titan-XP GPU.<sup>13,25,26,40</sup> In summary, an Inception-V1-style three-dimensional (3-D) CNN was designed and trained using leave-one-patient-out cross validation.<sup>26</sup> The 3-D convolutional kernels that comprised the modified 3-D inception V1 modules were of sizes  $1 \times 1 \times 1$ ,  $3 \times 3 \times 3$ , and  $5 \times 5 \times 5$ , and in total, the CNN architecture contained two inception modules, three convolutional layers, and two fully connected layers using the 3-D patch input size of  $25 \times 25 \times 91$  (height  $\times$  width  $\times$  spectral bands) for unprocessed HSI patches and  $25 \times 25 \times 61$  for preprocessed HSI patches. The total number of HSI samples in the cross-validation group was 647,000 normal HSI pixels and 877,000 cancer HSI pixels, calculated from the binary masks excluding glare, from the 21 tissue specimens from 7 patients. Each HSI pixel served as the center for an image patch (height  $\times$  width  $\times$  spectral bands). Dropout was employed to avoid overfitting in the CNN models.

In total, the CNN described was separately trained using four different scenarios using the information from two versions of the binary ground-truth masks and the two proposed processing methods, which produce different models: first, CNN trained without specular glare patches (trained separately both with and without added preprocessing of spectral data), and second, CNN trained with specular glare patches (trained separately both with and without added preprocessing of spectral data). The 95% confidence intervals (CIs) were calculated using a bootstrapping method by sampling 1000 pixels from each class with replacement from each patient and calculating the AUC of the receiver operator characteristic (ROC); the method was performed 1000 times for each patient and 2.5 and 97.5 percentiles were reported. The saved, trained models of the patients from the previous work were used to classify the HS images of the five testing group patients that underwent serial histological sectioning for surgical margin variation with depth evaluation. The probabilities of all models were averaged per patient to obtain qualitative probability heat maps, scaled from 0 to 1, where 0 represents normal class and 1 represents high probability that the tissue belongs to the cancer class.

### 2.6.3 HELICoiD algorithms

The results of the CNN classification method and the generated probability maps were compared to the results obtained by a machine learning pipeline previously developed for intraoperative detection of brain cancer using HIS.<sup>15,41–44</sup> In summary, a spatial-spectral classification algorithm, here referred to as HELICoiD [Fig. 4(a)], was implemented using a classification map obtained by an SVM classifier that is spatially

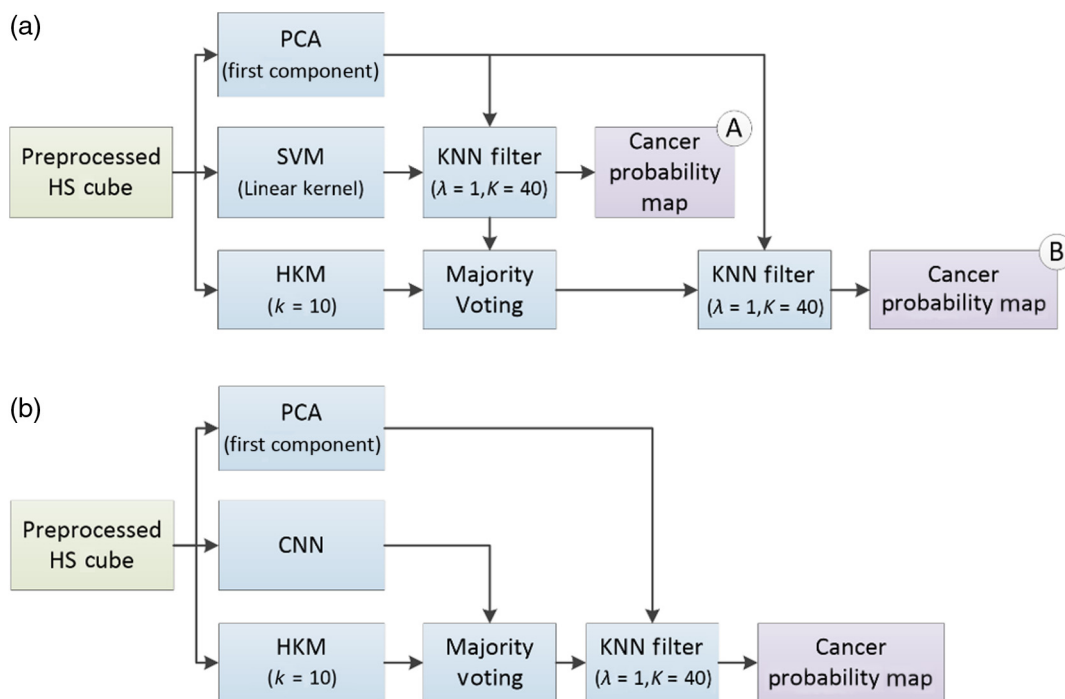
homogenized by employing a combination of a one-band representation obtained from the first principal component analysis (PCA) decomposition through a K-nearest neighbors (KNN) filtering method [Fig. 4(a), part A]. After that, the result of the KNN filtering is merged with an unsupervised segmentation map generated by a hierarchical K-means (HKM) algorithm through a majority voting (MV) method.<sup>16</sup> The result of this algorithm is a classification map that includes both the spatial and the spectral features of the HS images. In addition, for this application, the KNN filtering is applied again to the MV probabilities and the PCA one-band representation to homogenize the results [Fig. 4(a), part B].

A component of the HELICoiD algorithm was isolated and referred to as spatial SVM, which uses both the spectral and the spatial components of HSI for machine learning through a combination of PCA + SVM + KNN. This spectral-spatial implementation of SVM is performed with and without the additional preprocessing pipeline to be used as a surrogate for direct comparison to the CNN with and without preprocessing. Furthermore, a pipeline that combines the CNN architecture with the entire HELICoiD algorithm was proposed. In this case, the spatial-spectral stage of HELICoiD (PCA + SVM + KNN) is replaced by the CNN architecture trained with the preprocessed data [Fig. 4(b)]. Both HELICoiD and CNN + HELICoiD algorithms use the preprocessed HS data as input.

In summary, we present six algorithms for investigation of HSI machine learning: the CNN (with and without preprocessing), the spatial SVM (with and without preprocessing), the HELICoiD algorithm, and finally the CNN + HELICoiD algorithm. These six machine learning algorithms were tested first on the group of experiments using the binary ground-truth masks that exclude specular glare, and again the experiments were performed with the binary ground-truth masks that include specular glare. The six algorithms were compared using box plot distributions of the values and median AUC with 95% CIs, and, a paired, one-tailed *t*-test on the classification results was used to determine statistical differences with a 0.05 threshold for significance. The quantitative classification results of seven patients are reported in Table 1, obtained by using leave-one-patient-out cross validation. In addition, like the CNN method, the saved, trained models from these patients were used to classify five tissue specimens that were imaged with HSI and underwent serial histological sectioning for cancer margin variability evaluation. The probabilities of all models were averaged per patient to obtain qualitative probability heat maps, scaled from 0 to 1, where 0 represents normal class and 1 represents high probability that the tissue belongs in the cancer class.

## 3 Results

Quantitative results from the leave-one-patient-out cross validation, using both the ground-truth regions that include glare pixels and the subsampled masks that include only ideal quality regions that exclude glare, show that the CNN-based classifier group outperformed the SVM-based classifier group using the average AUC of the ROC, as shown in Table 1 and Fig. 5. The



**Fig. 4** Block diagrams of the proposed classification frameworks. (a) HELICoiD algorithm with the additional KNN filter. (b) Pipeline of the mixed algorithm.

**Table 1** Results of interpatient cross validation of SCC versus normal, obtained using the leave-one-patient-out method. Average AUCs are reported with bootstrapped 95% CI.

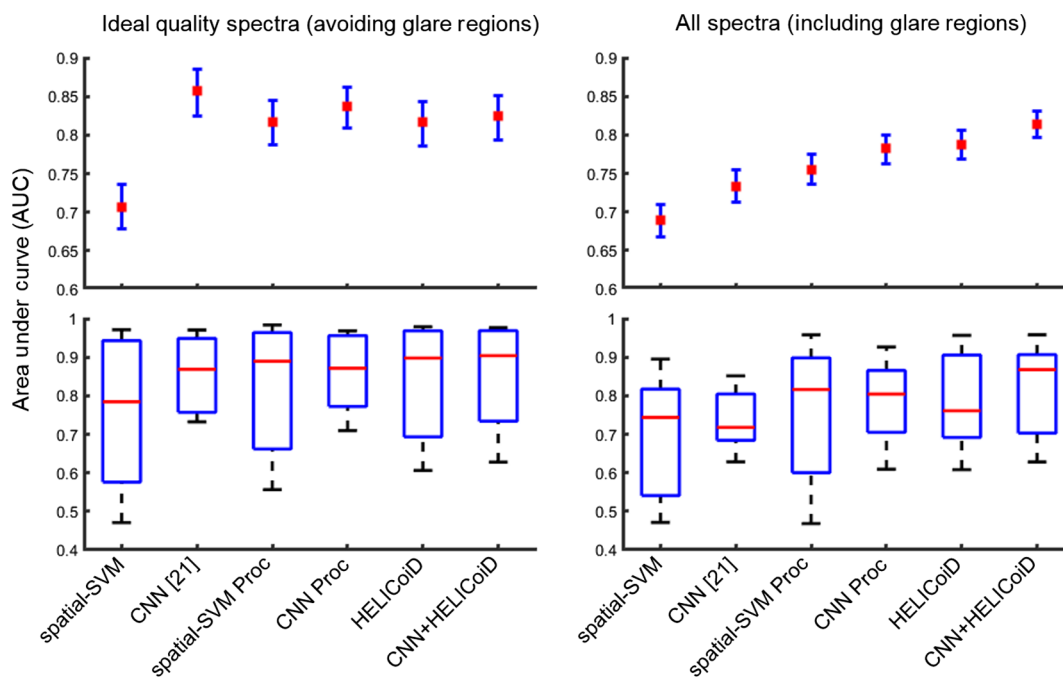
Ground truth	Classifier	Average AUC [95% CI]
Excluding glare	Spatial SVM	0.71 [0.68, 0.74]
	CNN	0.86 [0.82, 0.89]
	Spatial SVM (preprocessed HSI)	0.82 [0.80, 0.84]
	CNN (preprocessed HSI)	0.84 [0.81, 0.86]
	HELICoiD (preprocessed HSI)	0.82 [0.79, 0.84]
	CNN + HELICoiD (preprocessed HSI)	0.82 [0.79, 0.85]
Including glare	Spatial SVM	0.69 [0.67, 0.71]
	CNN	0.73 [0.71, 0.76]
	Spatial SVM (preprocessed HSI)	0.76 [0.74, 0.77]
	CNN (preprocessed HSI)	0.78 [0.76, 0.81]
	HELICoiD (preprocessed HSI)	0.79 [0.77, 0.81]
	CNN + HELICoiD (preprocessed HSI)	0.81 [0.80, 0.83]

results are reported using sevenfold cross validation to validate on all seven patients.

When classification is performed on only ideal quality pixels (obtained from the subsampled mask, see Fig. 1), the results indicate that additional preprocessing of the spectral signature

and addition of the HELICoiD method and KNN filtering do not significantly improve the results compared to only using the CNN [see Fig. 5(a)]. The average AUCs for the CNN groups are 0.86, 0.84, and 0.82 for the CNN, CNN with preprocessed input data, and CNN + HELICoiD method, respectively. The average AUCs for the SVM-based groups are 0.71, 0.82, and 0.82 for the spatial SVM without and with preprocessed input data, and HELICoiD method, respectively. The 95% CIs overlap for all groups except for spatial SVM, as shown in Table 1. In addition, all methods have a similar interquartile range and median distribution (see Fig. 5).

However, when the classification is performed over the entire HS image tissue area, which includes classification of specular glare pixels that contain more noise and variability, the CNN + HELICoiD method outperforms other methods tested with an average AUC of 0.81 for classification. This classification scheme, including specular glare pixels, represents a more realistic application of HSI. The CNN algorithm alone with or without preprocessing has an average AUC of 0.78 and 0.73, respectively, and both constitute a statistically significant decrease in performance compared the CNN + HELICoiD method ( $p = 0.03$  and  $0.04$ , respectively). The average AUC values range from 0.69 to 0.79 for the SVM-based groups, and the HELICoiD algorithm significantly outperforms the spatial SVM ( $p = 0.01$ ). The additional preprocessing pipeline, as described in Fig. 3, only offers a statistically significant increase in performance for the spatial-SVM algorithm ( $p = 0.02$ ), not the CNN ( $p = 0.1$ ) (see Table 1, for complete results with 95% CIs). In summary, from the cross-validation experiments, the best classification method was using the CNN as the input for the HELICoiD + KNN filtering method. For comparison, using the spectral-spatial SVM + PCA + KNN for input to the HELICoiD algorithm instead of the CNN component yielded slightly lower results that were not statistically significant.



**Fig. 5** Results of interpatient cross validation of SCC versus normal, obtained using the leave-one-patient-out method. Top: average AUCs reported with 95% CI. Bottom: box plots with the range in black, 75th and 25th percentile in blue, and median in red.

For generalization and application, HS images from five tissue specimens from five patients with SCC comprised the hold-out testing group and were classified using the saved models that were trained and cross validated using the seven patient cross-validation group. The testing patients were classified by all cross-validation models and averaged to obtain qualitative probability heat maps. Qualitative investigation of the five testing patients, classified with the CNN trained with preprocessed data alone and the CNN + HELICoiD + KNN filtering method is shown in Fig. 6. As shown in Figure 6, the CNN + HELICoiD + KNN technique performs better on the five testing patients, in agreement with the quantitative metrics from the cross-validation group. The SCC probability heat maps are shown with binary masks depicting the uncertainty and variation in the cancer margin with depth. Depending on the tissue, as demonstrated, the margin changes by about 1 mm depending on the sectioning depth. Therefore, our qualitative results can be interpreted within the range of uncertainty of the ground truth to provide more insight into the classification potential of machine learning methods using HSI for cancer detection.

#### 4 Discussion

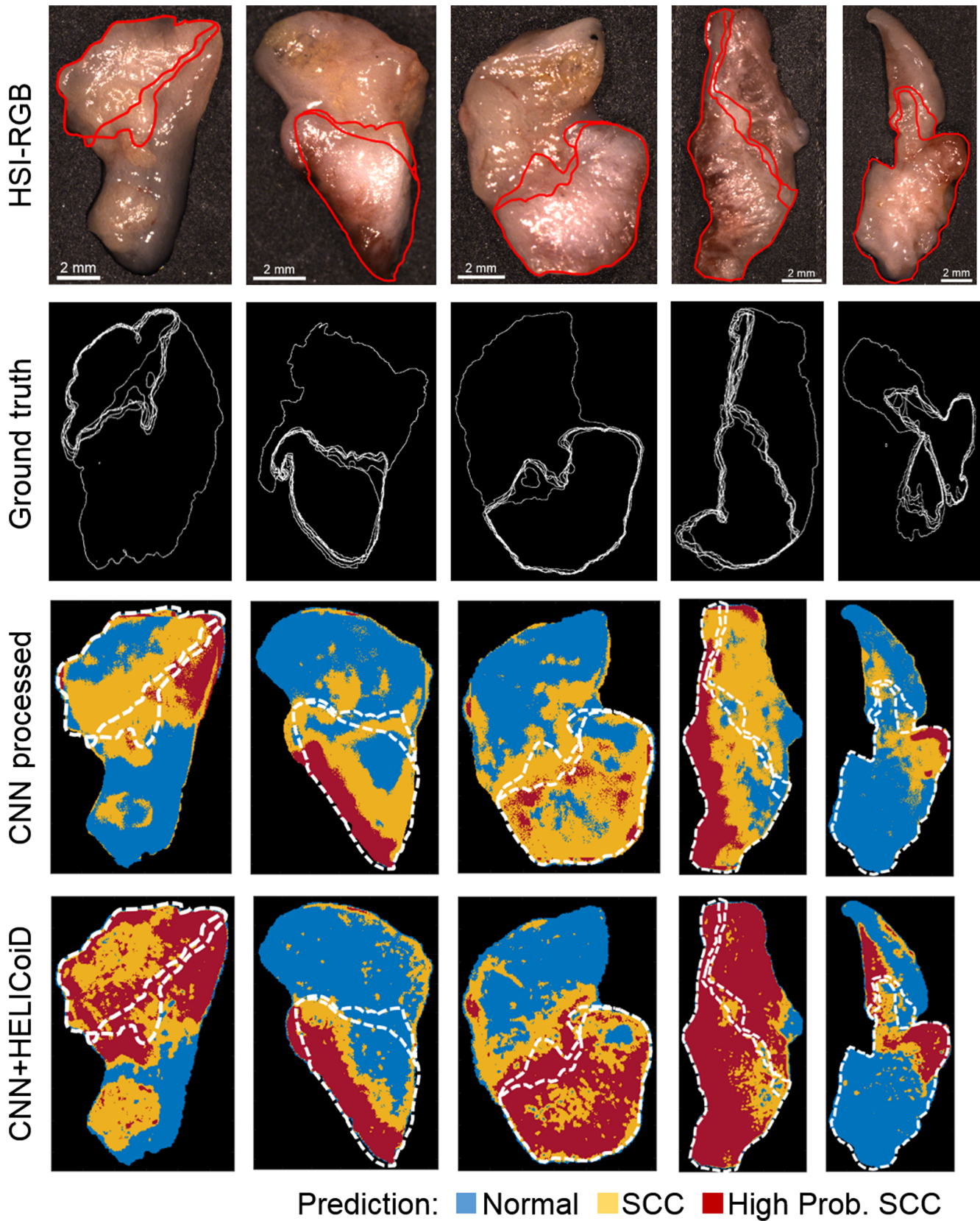
In this work, we presented and quantified the combination of two state-of-the-art machine learning-based classification methods for HSI of *ex-vivo* HNSCC surgical specimens. In summary, there are two methods for generating SCC prediction probability maps: the first uses a CNN, and the second uses a combination of SVM + PCA + KNN. After generating the predicted cancer probabilities on a pixel level, the probability map is combined with an HKM unsupervised segmentation layer through an MV algorithm that determines the class belonging to the ROI. Therefore, two distinct methods are compared using leave-one-patient-out cross validation to obtain quantitative evaluation metrics. In addition, the methods are tested on a group of HSI obtained from five SCC patients who underwent serial

histological sectioning to evaluate the variation in the cancer margin with penetration depth of the light wavelengths.

The quantitative results of this paper suggest that when working with ideal quality pixels, such as the spectral signatures generated from flat-surface-tissue surfaces with no glare, the CNN techniques and spatial-spectral machine learning algorithms will perform with no significant difference and no additional preprocessing will be necessary. The average AUCs for these methods using the preprocessed input data range from 0.82 to 0.86 with overlapping CIs. However, when the pixels classified contain noise, for example, due to sloping of the tissue edges or specular glare from completely reflected incident light, additional spectral smoothing and additional HELICoiD+KNN filtering of the classifier improve classification results of the CNN. The best performing method tested was CNN + HELICoiD + KNN with an average AUC of 0.81 [0.80, 0.83]. These tested methods outperform the traditional spectral-spatial machine learning methods employed in this study. Therefore, the HELICoiD + KNN techniques using both the CNN and SVM + PCA + KNN for cancer probability maps performed best for the seven cross-validation patients, and so both were employed on the five patients testing set. One major limitation of the approach used in this paper was the small sample size, and therefore, the proposed ML models could be prone to overfitting and lack generalization to larger testing datasets. To investigate potential overfitting in this experiment, we analyzed the training and cross-validation accuracies for the CNN trained with preprocessed HSI data. The average training accuracy was 85%, and the average cross-validation accuracy with CI was 79% [78, 81] for excluding glare and 74% [73, 76] for including glare. This result indicates that our models did not suffer substantially from overfitting.

Excluding glare, the CNN alone without preprocessing performs best. It is hypothesized that CNN and deep learning methods should be expected to outperform traditional ML techniques





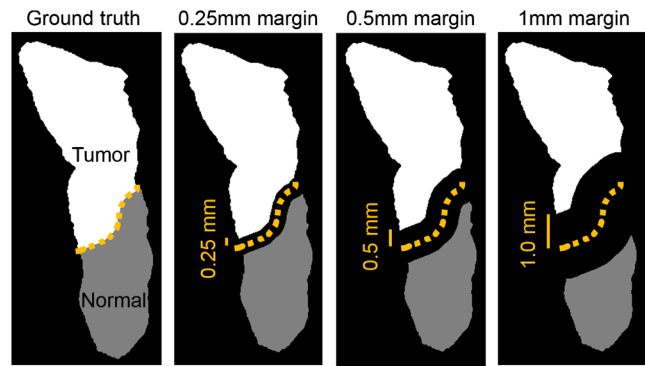
**Fig. 6** Representative results of binary cancer classification of the five testing patients. From top to bottom: HSI-RGB composite; histological ground truth showing variation in cancer margins with cancer area outlined; heat maps for cancer probability for CNN – HELICoID + KNN and HELICoID + KNN techniques. The extremes in the superficial cancer margin are overlaid on to the heat maps.

because a substantially large dataset should allow learning of variance, tolerance of noise, and removal of the need for preprocessing. Also, the CNN + HELICoiD method performs only slightly better than the original HELICoiD; this could be produced because the CNN probability maps are out-weighted by the PCA and KNN filtering that are applied afterward. After the inclusion of specular glare, the results change relative to each method, with the CNN with preprocessing outperforming the CNN without preprocessing. This outcome could be the result of the classification problem becoming more challenging compared to excluding glare, so the dataset is now too small for the CNN to perform well without preprocessing. Expanding the training dataset with more patients with specular glare and large amounts of noise may allow original CNN methods to outperform other techniques.

To test the general application of the proposed methods, HSI from five testing group patients with SCC were classified using the models that were trained and cross validated using the seven-patient group. To qualitatively investigate these results, histological images of the five *ex-vivo* tissue specimens were obtained down to about 0.3 mm to determine how the superficial cancer margin may change with depth. As shown in Fig. 6, the CNN + HELICoiD + KNN technique performs best on the five-patient SCC testing group, which was the same result obtained from the sevenfold quantitative cross-validation experiment.

The five patients' tumor-involved margin specimens were classified with the CNN trained with the fully preprocessed image patches that were extracted from the complete binary ground-truth mask including specular glare pixels. These probability maps are shown in Fig. 6, and the CNN map is also used as the input for the HELICoiD + KNN-filtering technique. In the top three rows of Fig. 6, there are regions of glare with tissue specimens that are classified incorrectly as normal with the CNN, but the CNN + HELICoiD correctly classifies these ROIs as SCC. It is also possible to observe that the CNN + HELICoiD method tends to overpredict SCC at regions of normal tissue near the cancer margin. We hypothesize that the CNN + HELICoiD technique outperforms the patch-based CNN alone because it can incorporate more local and regional spatial and spectral information to overcome the degrading effect of specular glare. Therefore, future work could involve the application of a full CNN for SCC detection on tissues with specular glare. This algorithm requires more data necessary for training as it produces labels that are end to end, a full pixel classification map for a full HSI input, and so it would require the entire HSI dataset acquired for this project.

An additional aim of this investigation was to determine the variation of the superficial cancer margin through the depth of the tissue. From the five testing patient tissue specimens, it can also be seen that the superficial cancer margin of the *ex-vivo* tissue specimens can vary from the extreme near and far margins in the range of 1 to 2 mm. These results allow interpretation of the cancer prediction probability maps with observed variation in the ground truth. However, additional possible uncertainty may exist in the histological ground truth. For example, if the angle of the sectioning plane is skewed from the tissue plane, then the ground truth could be warped, which could lead to errors that cannot be corrected by deformable registration. In a previous work, we explored a pipeline of affine and deformable demons-based registration for alignment of the histological ground truth to gross-level images of specimens, and it was



**Fig. 7** Proposed evaluation metric demonstrates millimetric, systematic cancer-margin erosion that could provide performance updates at several distances from the cancer margin to provide physicians with an easily interpretable method for understanding the results of HSI studies.

determined via needle-bored control points that target registration error was  $0.4 \pm 0.2$  mm.<sup>36</sup> The combination of these two main sources of uncertainty in the histological ground truth for the *ex-vivo* tissue specimens allows for error propagation of up to 2.5 mm in the binary mask. This would greatly affect the results of the pixel-wise evaluation metrics such as accuracy, sensitivity, and specificity.

Future work includes development and implementation of a new performance evaluation method to handle this margin uncertainty, for instance, evaluating primary tumor clearance at several millimeter increments from the ideal tumor margin. Currently, the standard for surgeons' opinions on margin adequacy is 5 mm for HNSCC, with margins between 1 and 5 mm defined as "close margins."<sup>45</sup> However, some studies suggest that outcomes may be similar for margin of 2.2 to 5 mm.<sup>46</sup> It is evident that the question of margin adequacy is still being determined, and surgeons would be interested in the performance of optical imaging methods at different margin distances. Therefore, the proposed error metric should extend to about 2 to 3 mm. Figure 7 demonstrates how the cancer margin could be systematically eroded and be used to determine accuracies at several distances from the cancer margin to better provide surgeons and physicians with a method for interpreting results for HSI studies, given the conclusions of this paper on the uncertainty of the cancer margin. The outcomes of this work suggest the use of this error metric for future studies.

Another avenue of future work involves the rethinking of the definition of normal tissues in the tissue specimens of the tumor-involved margin. In oral SCC, it has been studied that normal tissue directly adjacent to the primary SCC is molecularly distinct from normal tissues farther from the SCC, and additional resection to this extended margin may lead to increased disease-free survival and reduced local recurrence.<sup>23,24</sup> Moreover, this result was obtained by investigation that NBI at 400 to 430 nm and 525 to 555 nm reveals changes in normal tissue that correlate to significantly different levels of micro-RNA epigenetic regulation compared to primary tumor and normal tissue that are not visible under white light alone.<sup>21,23,24</sup> Therefore, it may be possible to extract micro-RNA expression levels to determine a molecular ground truth for certain tissues employed in this study, as Liu and Xu<sup>47</sup> have demonstrated that micro-RNAs can be reliably obtained from formalin-fixed, paraffin-embedded tissue samples.

## 5 Conclusion

This study investigated the effects of specular glare, noise, blurring, and tissue-edge sloping artifacts on HSI-based cancer detection. It explored the potential of HSI and machine learning for the detection of head and neck cancer. According to the experimental results, the CNN seems to be more robust against the environmental conditions of the acquired images and provides better classification results even without data preprocessing. Future work includes experimenting with deep learning methods that can incorporate more contextual information of the tissue, such as a full CNN. In addition, another objective was to evaluate the general efficacy on example test cases with uncertainty in the ground truth as the superficial cancer margin varies with penetration depth. This was tested by serially sectioning the tissue samples in the testing group to reveal the variation of the cancer margin through the depth of the tissue. This determined that the validity of the top section alone as the ground truth may be limited to 1 to 2 mm, suggesting that an alternative approach for obtaining performance metrics should be developed. All of the above factors are necessary to explore and understand the potential of HSI in the operating room. The proposed deep learning and machine learning methods employed to study these objectives require sufficiently large patient datasets for training, validation, and testing. Therefore, the preliminary results of this study encourage the inclusion of more data and further exploration into the ability of HSI for cancer detection.

### Disclosures

The authors have no relevant financial interests in this article and no potential conflicts of interest to disclose.

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## References

1. L. J. Joseph et al., “Racial disparities in squamous cell carcinoma of the oral tongue among women: a SEER data analysis,” *Oral Oncol.* **51**(6), 586–592 (2015).
2. M. Yao et al., “Current surgical treatment of squamous cell carcinoma of the head and neck,” *Oral Oncol.* **43**(3), 213–223 (2007).
3. B. Y. Kim et al., “Prognostic factors for recurrence of locally advanced differentiated thyroid cancer,” *J. Surg. Oncol.* **116**(7), 877–883 (2017).
4. T. Y. Chen, L. J. Emrich, and D. L. Driscoll, “The clinical significance of pathological findings in surgically resected margins of the primary tumor in head and neck carcinoma,” *Int. J. Radiat. Oncol. Biol. Phys.* **13**(6), 833–837 (1987).
5. T. R. Loree and E. W. Strong, “Significance of positive margins in oral cavity squamous carcinoma,” *Am. J. Surg.* **160**(4), 410–414 (1990).
6. D. N. Sutton et al., “The prognostic implications of the surgical margin in oral squamous cell carcinoma,” *Int. J. Oral Maxillofac. Surg.* **32**(1), 30–34 (2003).
7. T. M. P. Amaral et al., “Predictive factors of occult metastasis and prognosis of clinical stages I and II squamous cell carcinoma of the tongue and floor of the mouth,” *Oral Oncol.* **40**(8), 780–786 (2004).
8. P. Garzino-Demo et al., “Clinicopathological parameters and outcome of 245 patients operated for oral squamous cell carcinoma,” *J. Cranio-Maxillo-Facial Surg.* **34**(6), 344–350 (2006).
9. S. Rathod et al., “A systematic review of quality of life in head and neck cancer treated with surgery with or without adjuvant treatment,” *Oral Oncol.* **51**(10), 888–900 (2015).
10. G. Lu and B. Fei, “Medical hyperspectral imaging: a review,” *J. Biomed. Opt.* **19**(1), 010901 (2014).
11. B. Fei et al., “Label-free hyperspectral imaging and quantification methods for surgical margin assessment of tissue specimens of cancer patients,” in *39th Annu. Int. Conf. IEEE Eng. Med. and Biol. Soc.*, pp. 4041–4045 (2017).
12. M. Halicek et al., “Optical biopsy of head and neck cancer using hyperspectral imaging and convolutional neural networks,” *Proc. SPIE* **10469**, 104690X (2018).
13. M. Halicek et al., “Tumor margin classification of head and neck cancer using hyperspectral imaging and convolutional neural networks,” *Proc. SPIE* **10576**, 1057605 (2018).
14. G. Lu et al., “Detection of head and neck cancer in surgical specimens using quantitative hyperspectral imaging,” *Clin. Cancer Res.* **23**(18), 5426–5436 (2017).
15. H. Fabelo et al., “An intraoperative visualization system using hyperspectral imaging to aid in brain tumor delineation,” *Sensors* **18**(2), 430 (2018).
16. H. Fabelo et al., “Spatio-spectral classification of hyperspectral images for brain cancer detection during surgical operations,” *PLoS One* **13**(3), e0193721 (2018).
17. H. Fabelo et al., “Deep learning-based framework for in vivo identification of glioblastoma tumor using hyperspectral images of human brain,” *Sensors* **19**(4), 920 (2019).
18. S. Ortega et al., “Use of hyperspectral/multispectral imaging in gastroenterology. Shedding some different light into the dark,” *J. Clin. Med.* **8**(1), 36 (2019).
19. E. J. M. Baltussen et al., “Hyperspectral imaging for tissue classification, a way toward smart laparoscopic colorectal surgery,” *J. Biomed. Opt.* **24**(1), 016002 (2019).
20. M. Salmivuo et al., “Hyperspectral imaging system in the delineation of ill-defined basal cell carcinomas: a pilot study,” *J. Eur. Acad. Dermatol. Venereol.* **33**(1), 71–78 (2019).
21. C. S. Farah et al., “Improved surgical margin definition by narrow band imaging for resection of oral squamous cell carcinoma: a prospective gene expression profiling study,” *Head Neck* **38**(6), 832–839 (2016).
22. M. Leigh-Ann and R. M. Paul, “MicroRNA: biogenesis, function and role in cancer,” *Curr. Genomics* **11**(7), 537–561 (2010).
23. C. S. Farah, “Narrow band imaging-guided resection of oral cavity cancer decreases local recurrence and increases survival,” *Oral Dis.* **24**(1–2), 89–97 (2018).
24. C. S. Farah, S. A. Fox, and A. J. Dalley, “Integrated miRNA-mRNA spatial signature for oral squamous cell carcinoma: a prospective profiling study of narrow band imaging guided resection,” *Sci. Rep.* **8**(1), 823 (2018).

25. A. Krizhevsky, I. Sutskever, and G. E. Hinton, "ImageNet classification with deep convolutional neural networks," in *Proc. 25th Int. Conf. Neural Inf. Process. Syst.*, Vol. 1, pp. 1097–1105 (2012).
26. C. Szegedy et al., "Going deeper with convolutions," in *IEEE Conf. Comput. Vision and Pattern Recognit.* (2014).
27. M. Halicek et al., "Optical biopsy of head and neck cancer using hyperspectral imaging and convolutional neural networks," *J. Biomed. Opt.* **24**(3), 036007 (2019).
28. M. Halicek et al., "Deep convolutional neural networks for classifying head and neck cancer using hyperspectral imaging," *J. Biomed. Opt.* **22**(6), 060503 (2017).
29. F. Manni et al., "Automated tumor assessment of squamous cell carcinoma on tongue cancer patients with hyperspectral imaging," *Proc. SPIE* **10951**, 109512K (2019).
30. P. J. C. Weijtmans et al., "A dual stream network for tumor detection in hyperspectral images," in *IEEE 16th Int. Symp. Biomed. Imaging* (2019).
31. M. Halicek et al., "Cancer margin evaluation using machine learning in hyperspectral images of head and neck cancer," *Proc. SPIE* **10951**, 109511A (2019).
32. S. Trajanovski et al., "Tumor semantic segmentation in hyperspectral images using deep learning," in *2nd Int. Conf. Med. Imaging with Deep Learn.*, London, England (2019).
33. B. Fei et al., "Label-free reflectance hyperspectral imaging for tumor margin assessment: a pilot study on surgical specimens of cancer patients," *J. Biomed. Opt.* **22**(8), 086009 (2017).
34. G. Lu et al., "Hyperspectral imaging of neoplastic progression in a mouse model of oral carcinogenesis," *Proc. SPIE* **9788**, 978812 (2016).
35. G. Lu et al., "Framework for hyperspectral image processing and quantification for cancer detection during animal tumor surgery," *J. Biomed. Opt.* **20**(12), 126012 (2015).
36. M. Halicek et al., "Deformable registration of histological cancer margins to gross hyperspectral images using demons," *Proc. SPIE* **10581**, 105810N (2018).
37. L. A. Sordillo et al., "Deep optical imaging of tissue using the second and third near-infrared spectral windows," *J. Biomed. Opt.* **19**(5), 056004 (2014).
38. C. Ash et al., "Effect of wavelength and beam width on penetration in light-tissue interaction using computational methods," *Lasers Med. Sci.* **32**(8), 1909–1918 (2017).
39. H. Zhang et al., "Penetration depth of photons in biological tissues from hyperspectral imaging in shortwave infrared in transmission and reflection geometries," *J. Biomed. Opt.* **21**(12), 126006 (2016).
40. M. Abadi et al., "TensorFlow: large-scale machine learning on heterogeneous systems," 2015, <https://www.tensorflow.org>.
41. G. Florimbi et al., "Accelerating the K-nearest neighbors filtering algorithm to optimize the real-time classification of human brain tumor in hyperspectral images," *Sensors* **18**(7), 2314 (2018).
42. D. Ravi et al., "Manifold embedding and semantic segmentation for intraoperative guidance with hyperspectral brain imaging," *IEEE Trans. Med. Imaging* **36**(9), 1845–1857 (2017).
43. R. Lazcano et al., "Porting a PCA-based hyperspectral image dimensionality reduction algorithm for brain cancer detection on a manycore architecture," *J. Syst. Archit.* **77**, 101–111 (2017).
44. D. Madroñal et al., "SVM-based real-time hyperspectral image classifier on a manycore architecture," *J. Syst. Archit.* **80**, 30–40 (2017).
45. H. M. Baddour, Jr., K. R. Magliocca, and A. Y. Chen, "The importance of margins in head and neck cancer," *J. Surg. Oncol.* **113**(3), 248–255 (2016).
46. D. Zanoni et al., "A proposal to redefine close surgical margins in squamous cell carcinoma of the oral tongue," *JAMA Otolaryngol. Head Neck Surg.* **143**(6), 555–560 (2017).
47. A. Liu and X. Xu, "MicroRNA isolation from formalin-fixed, paraffin-embedded tissues," *Methods Mol. Biol.* **724**, 259–267 (2011).

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