




Bioactive Compound Produced by Endophytic Fungi Isolated From *Pelargonium sidoides* Against Selected Bacteria of Clinical Importance

Madira Coutlyne Manganyi^a, Christ-Donald K. Tchatchouang^a , Thierry Regnier^b,
Cornelius Carlos Bezuidenhout^c  and Collins Njie Ateba^{a,d} 

^aDepartment of Microbiology, North West University – Mafikeng Campus, Mmabatho, South Africa; ^bDepartment of Biotechnology and Food Technology, Tshwane University of Technology, Pretoria, South Africa; ^cUnit for Environmental Sciences and Management, North West University – Potchefstroom Campus, Potchefstroom, South Africa; ^dFood Security and Safety Niche Area, Faculty of Agriculture, Science and Technology, North-West University, Mmabatho, South Africa

ABSTRACT

Endophytic fungi have the ability to live inside the host plant tissues without causing neither symptoms of diseases/or harm. Opportunistic infections are accountable for majority of the outbreaks, thereby putting a burden on the health system. To investigate and characterize the bioactive compounds for the control of bacteria of clinical importance, extracts from endophytic fungi were isolated from indigenous South African medicinal plants. Extracts from endophytic fungi were isolated from 133 fungal strains and screened against Gram positive and negative bacteria namely *Bacillus cereus*, *Escherichia coli*, *Enterococcus faecium*, and *E. gallinarum* using disk diffusion. Furthermore, gas chromatography–mass spectrometry was performed to identify the bioactive compounds. Sixteen out of one hundred and thirty-three (12%) fungi extracts exhibited antibacterial properties against some of the selected bacteria. *E. coli* was found to be the most susceptible in contrast to *E. faecium* and *E. gallinarum* which were the most resistant. The isolate MHE 68, identified as *Alternaria* sp. displayed the greater spectrum of antibacterial activities by controlling selected clinical bacteria strains including resistant *E. faecium* and *E. gallinarum*. The chemical analysis of the extract from MHE 68 indicated that linoleic acid (9,12-octadecadienoic acid (Z,Z)) and cyclodecasiloxane could be accountable for the antibacterial activity. This is the first study conducted on the secondary metabolites produced by endophytic fungal strains isolated from the *Pelargonium sidoides* DC. possessing antibacterial properties.

ARTICLE HISTORY

Received 30 January 2019
Revised 5 June 2019
Accepted 9 June 2019

KEYWORDS

Endophytic fungi;
antibacterial activity;
secondary metabolites;
bioactive compounds

1. Introduction

Endophytic fungi play an essential part in the physiological and ecological roles [1] including growth promoter, stress tolerance, drought resistance, insect, and herbivores repliers. Antibiotic-producing fungi were the first and continue to be dominating the market [2]. Endophytes are defined as microorganisms that live/colonize within the plant tissues and cause no damage or symptoms of disease. There are regarded as being more beneficial to the plant than detrimental [3]. Current research focuses on using untapped location, medicinal plants and their endophytic fungi to discovery novel, affordable, efficacies pharmaceutical active compounds. This is in the hope of neutralizing the enormous problem of resistance [4].

Despite the knowledge about flora, fauna, and the traditional use of medicinal plant in Southern Africa, South Africa in particular remains an

untapped location for host medicinal and aromatic plants with novel microorganisms [5].

The variance of plant to fungal diversity is 1 to 6 [5], increases the probabilities of discovering novel metabolites in the fungal community. From a large number of medicinal plants indigenous to South Africa, *Pelargonium sidoides* DC. have been reported to be the most traditional use plant for primary health care [6]. Due to the aptitude of this plant to produce secondary metabolites, it can be expected that endophytic fungi possessing some antimicrobial properties can be isolated [7].

For decades, bacteria have emerged as important healthcare-associated pathogens. The rapid spread of enterococci with resistance to vancomycin (VRE) has been of particular concern worldwide [7] when a substantial percentage of the population is immune compromised patients [8]. Although, *Enterococcus faecium* is the leading bacteria

responsible for medical intensive care units' device-associated infections, other enterococcal species such as *E. avium*, *E. gallinarum*, *E. casseliflavus*, are of clinical concerns. Opportunistic diseases in developing countries are a major cause of human mortality due to inadequate sanitation, a lack of safe drinking water, malnourishment, war, and famine claiming approximately 2 million lives a year [9]. While most coliforms are harmless to human health, the presence of *Escherichia coli*, can be accountable for outbreaks of infectious diarrhoea and held responsible for a number of death in developing countries.

Manganyi et al. [10] reported in depth the biodiversity and phylogenetic relationship of the endophytic fungi isolated from *Pelargonium sidoides* DC. The primary objective of the current study was to screen for the antibacterial properties of fungal extracts against seven selected bacteria of clinical interest. And finally to determine the chemical profile of the most abundant bioactive compounds using gas chromatography mass spectrophotometry (GC-MS).

2. Materials and methods

2.1. Endophytic fungi isolated from *Pelargonium sidoides* DC

One hundred and thirty three ($n=133$) endophytic fungi were successfully isolated from healthy leave and roots of *Pelargonium sidoides*. Morphological and molecular identification were performed using internal transcribe spacer (ITS) region as describe by [10]. The pure cultures were preserved in the Agricultural Research Council (ARC, Mycology) on water, slant, and freeze dry for future use.

2.2. Production of secondary metabolites

The fungal isolates were revived by culturing them on Potato Dextrose agar (PDA, Merck, Darmstadt, Germany) and incubated at 25 °C for 10 days. A plug of active mycelia was inoculated into a 250 mL Erlenmeyer flask containing 50 mL of malt extract broth (MEB; Merck, Darmstadt, Germany). The numbers of spores were counted with a hemocytometer (Merck, Johannesburg, South Africa) and adjusted to 1×10^6 conidia/mL. The secondary metabolites were produced by fermentation as described by Premjanu and Jaynthy [11] and each fermentation performed in triplicate. Briefly, fungal cell mass was removed by filtration through a 0.45 μm syringe filter and the resulting filtrate stored in sterile conical flasks at 4 °C, until further use.

2.3. Bacteria strains

The target bacterial strains used were both environmental strains and control strain (American Type Culture Collection, ATCC; Table 1) with potential clinical implications and are well-known to be resistance against modern antibiotics. Bacteria were cultured in nutrient broth (NB; Merck, Darmstadt, Germany) for 24 h at 37 °C to reach a final suspension of 1×10^7 cells/mL.

2.4. Antibacterial properties

One hundred and thirty-three (133) extracts were screened for their antibacterial activities against six targeted bacteria strains. The disk diffusion assay was used as described by Ahmad et al. [12] and the experiment done in triplicate. The zone inhibition as the degree of activity was expressed as diameter (mm).

2.5. Characterization by gas-chromatography mass spectrometry

The most active fungal extracts were selected to undergo secondary metabolites identification using gas-chromatography mass spectrometry GCMS, (GC-MS TQ8050; Shimadzu, Johannesburg, South Africa) equipped with a Multifunctional Autosampler (AOC-6000), a capillary column (RTX-5, 60 m \times 0.25 mm \times 0.25 μm , New Delhi, India) as described by Sharma et al. [7]. The identities of the compounds were determined by searching known molecules in databases of NIST05; WILEY 8, and FFNSC1.3 libraries.

3. Results and discussion

3.1. Diversity of fungal extracts with antibacterial activity

From all the fungal isolates ($n=133$) tested; only 16 displayed inhibition activity against the selected bacteria (Table 2). The results (Figure 1) revealed that approximately 25% were *Penicillium* sp. which was the most dominant genera followed by *Fusarium* sp (19%), *Alternaria* sp. (13%), and *Aspergillus* sp. (7%). These genera belong to the ascomycete's class, which is reported to be one of the two larger class of endophytes [13]. The results of *Fusarium* sp. being the second most prevalent genera is not surprising as the *Fusaria* genus is the largest group of filamentous fungi [14]. *Geotrichum* sp. which falls under the same division (Ascomycota) as fungi, has been isolated from clones cocoa resistant VSD M.05 [15] and has been associated with the growth promoting protection capabilities of the plant hosts from pests and diseases.

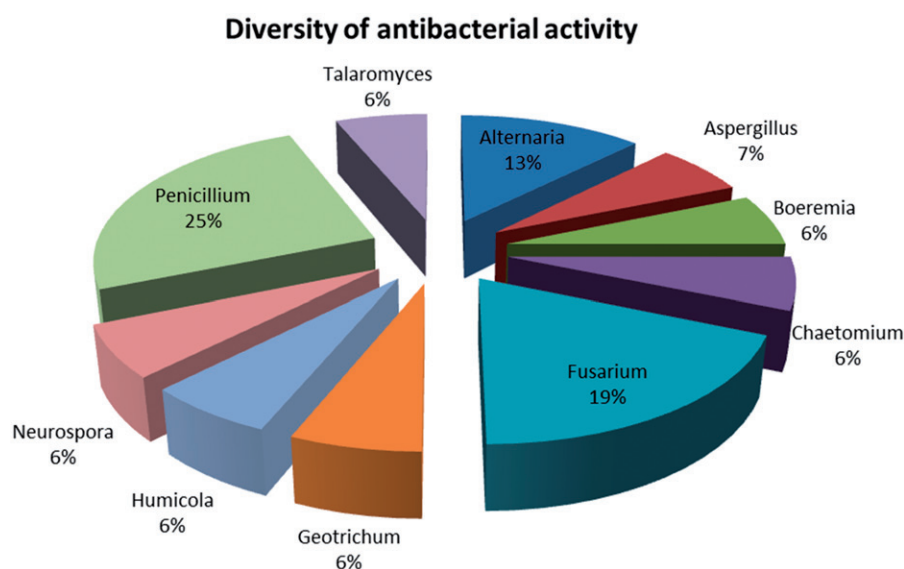
Table 1. Target bacteria with their origin and accession number.

Target bacteria	Accession no.	Origin
<i>Escherichia coli</i>	ATCC 25922	ATCC collection
<i>Escherichia coli</i>	ID = O177	Environmental isolate from cattle faeces
<i>Bacillus cereus</i>	ATCC 10876	ATCC collection
<i>Enterococcus faecalis</i>	ATCC S1299	Environmental isolate from ground water
<i>Enterococcus faecium</i>	ATCC 700221	Environmental isolate from ground water
<i>Enterococcus gallinarum</i>	ATCC 700425	ATCC collection

ID: Identified as

Table 2. Antimicrobial activity of extracts produced by endophytic fungal isolated from *Pelargonium sidoides*.

Sample. No	Sample ID	Probable ID	Zone of inhibition (mm)					
			<i>E. coli</i> ATCC 25922	<i>E. coli</i> ATCC 0177	<i>B. cereus</i> ATCC 10876	<i>E. faecalis</i> ATCC S1299	<i>E. faecium</i> ATCC 700221	<i>E. gallinarum</i> ATCC 700425
1	RNK 001	<i>Talaromyces</i> sp.	+ (9)	-	-	-	+ (6)	-
2	RNK 004	<i>Penicillium glabrum</i>	++ (11)	-	-	-	-	-
3	RNK 016	<i>Alternaria tenuissima</i>	-	-	+ (9)	-	-	-
4	PG 9	<i>Chaetomium subaffine</i>	-	+ (9)	+ (6)	-	-	-
5	PG 10	<i>Humicola</i> sp.	-	-	+ (6)	-	-	-
6	END 015	<i>Boeremia exigua var. pseudolilacis</i>	-	-	-	++ (11)	-	-
7	END 017,1	<i>Penicillium</i> sp.	+ (9)	-	-	-	-	-
8	END 021	<i>Penicillium commune</i>	+ (10)	-	-	-	-	-
9	MHE 001	<i>Fusarium solani</i>	-	++ (11)	-	-	-	-
10	MHE 010	<i>Neurospora crassa</i>	+ (9)	+ (8)	-	-	-	-
11	MHE 011	<i>Penicillium</i> sp.	+ (9)	-	-	-	-	-
12	MHE 033	<i>Aspergillus</i> sp.	+ (2)	+ (9)	-	-	-	-
13	MHE 055	<i>Fusarium solani</i>	-	++ (12)	-	-	-	-
14	MHE 056	<i>Fusarium</i> sp.	-	-	+ (8)	-	++ (11)	-
15	MHE 059	<i>Geotrichum candidum</i>	+ (9)	-	-	-	-	-
16	MHE 068	<i>Alternaria</i> sp.	-	-	+ (8)	-	++ (11)	++ (12)

**Figure 1.** Diversity of fungal extracts displaying antibacterial activity.

3.2. Antibacterial activity against selected bacteria

Furthermore, the results showed that most endophytic fungal extracts from medicinal plants have limited antibacterial activity. None of the isolates tested were able to control all six pathogenic bacteria. Only MHE 068 isolate, identified as *Alternaria* sp. (Figure 2) displayed significant antibacterial activity against three bacterial strains (*B. cereus*, *E. faecium* (ATCC 700221) and *E. gallinarum*). These

findings are supported by [2], who reported similar results about *Alternaria* sp. exhibiting antibacterial activities against *Bacillus* sp., *Staphylococcus aureus*, *E. faecalis*, and *E. coli*. Despite of the overall 25% activity of *Penicillium* genera, the results show that *Penicillium* sp. could only inhibit *E. coli* (ATCC 25922) and nothing else. The discovery of antibiotics started with *Penicillium* strain producing bioactive compounds with significant biological properties which revolutionized medicine and

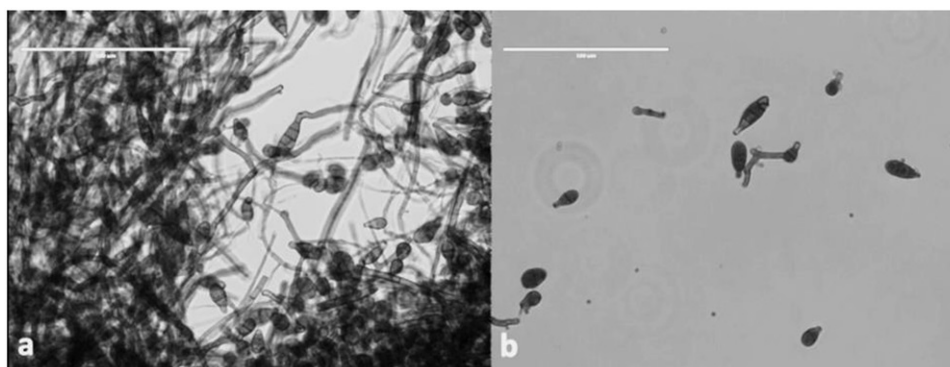


Figure 2. Endophytic fungi *Alternaria* (a) Conidia structure wrapped in mycelia (b) individual conidia structure (scale bars: 100 µm).

Table 3. Main compounds identified in fungal extracts (Sample MHE 68).

	Name	Retention Time (min)	Height	Area
1	Tetradecamethyl hexasiloxane	21.9	49846	107935
2	Tetradecamethyl hexasiloxane	24.1	70746	134156
3	Group of octadecadienoic acid	25.9	84122	146379
4	Group of octadecadienoic acid	26.6	178016	271510
5	Group of octadecadienoic acid	26.7	304562	593966
6	Group of octadecadienoic acid	26.7	172773	340337
7	Group of octadecadienoic acid	26.9	96415	287016
8	Group of octadecadienoic acid	27.1	779997	1597988
9	Eicosamethyl cyclodecasiloxane	27.2	114397	250984
10	1H-Purin-6-amine, N-((3-fluorophenyl)methyl)-6-(3-fluorobenzylamino)purine	27.6	22586	68648
11	Eicosamethyl cyclodecasiloxane	28.4	108477	292869
12	Tetradecamethyl hexasiloxane	29.6	99113	272854
13	1H-Purin-6-amine, N-((3-fluorophenyl)methyl)-6-(3-fluorobenzylamino)purine	29.9	13195	78786
14	Eicosamethyl cyclodecasiloxane	30.6	102788	244851
15	1H-Purin-6-amine, N-((3-fluorophenyl)methyl)-6-(3-fluorobenzylamino)purine	30.8	23772	61412
16	Eicosamethyl cyclodecasiloxane	31.6	50978	107960
17	Propanoic acid	32.6	26327	64355
18	Methyl 2,3,4-tri-O-acetyl-6-deoxy-6-iodo- α -D-glucopyranoside	33.2	19795	84775
19	1,2-Benzenediol	33.7	25879	94106
20	6-Decylsulfonylhexane-1,2,3,4,5-pentol	34.6	25345	107673

pharmaceutical products. The *Penicillium* extracts in this study exhibited narrow spectrum of activity.

In addition, it can be noted that the three *Fusarium* isolates displaying some antibacterial activities against *B. cereus*, *E. coli* (ATCC 0177), and *E. faecium* (ATCC 700221). As reported by [16], the endophytic *Fusarium* sp. is primarily known to exhibit good antibacterial activities against *E. coli*.

3.3. Characterization of bioactive compounds by GC-MS analysis

As previously stated, only the most effective extract (MHE 68, *Alternaria* sp.) was further analysed by GC-MS. Out of twenty compounds, separated and preliminary identified, the fatty acid, 9,12-octadecadienoic acid (Z,Z) (34%) commonly known as linoleic acid, was detected as dominant compound followed by several peaks initially identified as a cyclic volatile, eicosamethyl-cyclodecasiloxane oligomers (Table 3). Like several endophytic fungi, *Alternaria* sp. has been reported to exhibit significant level antibacterial activity against Gram positive

and negative bacteria [17]. The antibacterial activity can be attributed to the high level of linoleic acid which has been reported to inhibit the binding of *E. coli* heat-labile enterotoxin (LT) to the receptor ganglioside GM1 in rabbit [18].

Furthermore, the antibacterial activity of plant volatile oils including cyclic volatiles have been demonstrated its activity against 25 different genera of bacteria such as *E. coli* and *E. faecalis* (NCTC 775) [19]. In conclusion, the study confirmed the potential use of endophytes from untapped indigenous medicinal plant for the control of opportunistic pathogens responsible for the mortality rate in developing countries. The extract from the *Alternaria* strain clearly confirmed the presence of linoleic acid. However, further studies on the optimization of the fermentation process and purification of the compounds are needed. As well as further bio-guided fractionation by nuclear magnetic resonance (NMR) and MS spectroscopy is necessary in future studies. This does not omit that the data obtained is critical in the investigation of novel bioactive compounds against bacterial strains of clinical

importance. This is the first report on endophytic fungi isolated from *Pelargonium sidoides* DC. which were screened for their antibacterial activities.


Disclosure statement


No potential conflict of interest was reported by the authors.

Funding

We would like to express our sincere gratitude to North West University, the National Research Foundation (NRF) and the Food Security and Safety for the financial support.

ORCID

Christ-Donald K. Tchatchouang  <http://orcid.org/0000-0002-6792-9805>

Cornelius Carlos Bezuidenhout  <http://orcid.org/0000-0002-6047-4991>

Collins Njie Ateba  <http://orcid.org/0000-0003-1230-5138>

References

- [1] Thatoi H, Behera BC, Mishra RR. Ecological role and biotechnological potential of mangrove fungi. *Mycology*. 2013;4:54–71.
- [2] Smith RA, M'ikanatha NM, Read AF. Antibiotic resistance: a primer and call to action. *Health Commun*. 2015;30:309–314.
- [3] Khorasani M. *Cylindrocarpon* species in Pacific Northwest Douglas-fir Nurseries, diversity and effects of temperature and fungicides on mycelial growth [MSc dissertation]. Seattle, WA: University of Washington; 2013.
- [4] Malhadas C, Malheiro R, Pereira JA. Antimicrobial activity of endophytic fungi from olive tree leaves. *World J Microbiol Biotechnol*. 2017;33:46.
- [5] Chatterjee A, Chowdhury R. Bile and unsaturated fatty acids inhibit the binding of cholera toxin and *Escherichia coli* heat-labile enterotoxin to GM1 receptor. *Antimicrob Agents Chemother*. 2008;52:220–224.
- [6] Dorman HJD, Deans SG. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J Appl Microbiol*. 2000;88:308–316.
- [7] Sharma A, Kumar V, Kanwar MK, et al. Phytochemical profiling of the leaves of *Brassica juncea* L. using GC-MS. *Int Food Res J*. 2017;24:547–551.
- [8] Nelson PE, Dignani MC, Anaissie EJ. Taxonomy, biology, and clinical aspects of *Fusarium* species. *Clin Microbiol Rev*. 1994;7:479–504.
- [9] Kumar CG, Mongolla P, Joseph J, et al. Chemical. Antimicrobial activity from the extracts of fungal isolates of soil and dung samples from Kaziranga National Park, Assam. *J Mycol Med*. 2010;20:283–289.
- [10] Manganyi MC, Regnier T, Kumar A. Biodiversity and antibacterial screening of endophytic fungi isolated from *Pelargonium sidoides*. *S Afr J Bot*. 2018;116:192–199.
- [11] Sabol K, Patterson JE, Lewis JII, et al. Emergence of daptomycin resistance in *Enterococcus faecium* during daptomycin therapy. *Antimicrob Agents*. 2005;49:1664–1665.
- [12] Ahmad S, Khan MA, Ayaz S, et al. Antibacterial and antifungal studies of the crude extract and solvent fractions of *Onosma khyberianum*. *Pharmacologia*. 2013;4:525–528.
- [13] Hénock BNY, Dovie DB. Diarrheal diseases in the history of public health. *Arch Med Res*. 2007;38:159–163.
- [14] Ratnaweera PB, De Silva ED, Williams DE, et al. Antimicrobial activities of endophytic fungi obtained from the arid zone invasive plant *Opuntia dillenii* and the isolation of equisetin, from endophytic *Fusarium* sp. *BMC Complement Altern Med*. 2015;15:1–7.
- [15] Amin N, Salam M, Junaid M, et al. Isolation and identification of endophytic fungi from cocoa plant resistant VSD M.05 and cocoa plant Susceptible VSD M.01 in South Sulawesi. Indonesia *Int J Curr Microbiol App Sci*. 2014;3:459–467.
- [16] Sadrati N, Daoud H, Zerroug A. Screening of antimicrobial and antioxidant secondary metabolites from endophytic fungi isolated from wheat (*Triticum Durum*). *J Plant Prot Res*. 2013;53:1–9.
- [17] Premjanu N, Jaynthy C. Identification and characterization of antimicrobial metabolite from an endophytic fungus, *Colletotrichum gloeosporioides* isolated from *Lannea corammendalica*. *Int J Chem Tech Res*. 2015;07:369–374.
- [18] Department of Agriculture, Forestry and Fisheries. Medicinal Plants of South Africa. 2013; [cited 2017 May 29]. Available from: <http://www.daff.gov.za/Daffweb3/Portals/0/Brochures%20and%20Production%20guidelines/Brochure%20Medical%20Plants%20Of%20South%20Africa.pdf>
- [19] Gouda S, Das G, Sen SK, et al. Treasure house of bioactive compounds of medicinal importance. *Front Microbiol*. 2016;7:1–8.