

Chronic Prenatal Ethanol Exposure Increases GABA_A Receptor Subunit Protein Expression in the Adult Guinea Pig Cerebral Cortex

Craig D. C. Bailey, James F. Brien, and James N. Reynolds

Department of Pharmacology and Toxicology, Faculty of Health Sciences, Queen's University, Kingston, Ontario, Canada K7L 3N6

Excessive consumption of ethanol during pregnancy can produce teratogenic effects in offspring and is the leading cause of mental deficiency in the Western world. The objective of this study was to examine the effects of chronic prenatal ethanol exposure on the number of GABA_A receptors and relative protein levels for GABA_A receptor α 1 and β 2/3 subunits in the adult guinea pig cerebral cortex. Timed pregnant Dunkin-Hartley strain guinea pigs were given one of the following oral treatments daily throughout gestation: 4 gm of ethanol per kilogram of maternal body weight, isocaloric-sucrose with pair feeding, or isovolumetric water with *ad libitum* access to food. The ethanol treatment resulted in a peak maternal blood ethanol concentration of 328 ± 55 mg/dl (71.3 ± 12.0 mM) on gestational day 57 (term, ~ 68 d). Chronic prenatal exposure to ethanol resulted in increased spontaneous locomotor activity throughout development and decreased cerebral cortical

weight in adult offspring. The number of cerebral cortical [³H]muscimol binding sites was increased in adult offspring from the ethanol treatment group, and there was a corresponding increase in the amount of GABA_A receptor α 1 and β 2/3 subunit proteins in these same animals. For individual offspring, there were correlations between locomotor activity and cerebral cortical weight, as well as between cerebral cortical weight and GABA_A receptor neurochemistry. There was no effect of chronic prenatal ethanol exposure on [³H]MK-801 binding in this tissue. These data demonstrate that chronic prenatal ethanol exposure has long-term consequences on the regulation of GABA_A receptor expression in the cerebral cortex.

Key words: GABA_A receptor; NMDA receptor; prenatal ethanol exposure; cerebral cortex; guinea pig; radioligand binding; Western blot

Excessive consumption of ethanol during pregnancy produces teratogenic effects in the fetus that can manifest postnatally as fetal alcohol syndrome (FAS) (Jones and Smith, 1973). The most debilitating and persistent characteristic of FAS is CNS dysfunction that can present as impairments in emotion management, attention, learning, memory, and problem solving. (Streissguth et al., 1991; Mattson et al., 1996; Steinhausen and Spohr, 1998). FAS is recognized as the leading cause of mental deficiency in the Western world.

The GABA_A receptor is the major mediator of inhibitory neurotransmission in the CNS and is a heteromeric ion channel complex composed of five subunits. CNS GABA_A receptor subunits are organized into six distinct families based on sequence homology (α , β , γ , δ , ϵ , and θ), and some families contain subunit subtypes (e.g., α 1- α 6). Generally, a combination of α , β , and γ subunits forms fully functional receptors. The GABA_A receptor is an important site of action for benzodiazepines, barbiturates, neuroactive steroids, convulsants, and ethanol (for review, see Rabow et al., 1995; Mehta and Ticku, 1999). Different GABA_A receptor isoforms result from the combination of different subunit subtypes, producing GABA_A receptors with distinct pharmaco-

logical characteristics (Barnard et al., 1998). GABA_A receptor subunit subtype expression varies both developmentally and regionally throughout the CNS. The expression of GABA_A receptor subunit subtypes also may be altered significantly by chronic exposure to xenobiotics, including ethanol (Mhatre et al., 1993; Mhatre and Ticku, 1994; Devaud et al., 1995, 1997).

The effect of chronic exposure to ethanol *in utero* on GABA_A receptor number or subunit subtype protein expression has not been elucidated, although certain GABA_A receptor-mediated behavioral (Zimmerberg et al., 1995; Osborn et al., 1998) and neurochemical (Allan et al., 1998; Hsiao et al., 1998, 1999) dysfunctions have been identified. Data from our laboratory demonstrate that chronic prenatal ethanol exposure increases the number of cerebral cortical benzodiazepine-binding GABA_A receptors and decreases [³H]flunitrazepam affinity for these GABA_A receptors in adult offspring (Bailey et al., 1999). Using the guinea pig as an experimental animal model, the objectives of this study were to determine (1) whether chronic prenatal ethanol exposure increases the number of all GABA_A receptors in the adult cerebral cortex using [³H]muscimol as the ligand for the GABA binding site and (2) whether this increase in GABA_A receptor number occurs concurrently with an increase in the relative protein content of the GABA_A receptor α 1 and β 2/3 subunits, each of which is the most abundant subtype or subtypes of their subunit families in the adult cerebral cortex (Laurie et al., 1992). The effect of chronic prenatal ethanol exposure on the number of NMDA receptors in the adult guinea pig cerebral cortex also was determined, because the function of this excitatory neurotransmitter receptor also has been demonstrated to be

Received Nov. 2, 2000; revised March 28, 2001; accepted March 30, 2001.

This work was supported by the Canadian Institutes of Health Research Operating Grant MT-15150. C.D.C.B. is the recipient of a Doctoral Research Award from the Canadian Institutes of Health Research and the NeuroScience Canada Foundation.

Correspondence should be addressed to Dr. James N. Reynolds, Department of Pharmacology and Toxicology, Queen's University, Kingston, Ontario, Canada K7L 3N6. E-mail: jnr@post.queensu.ca.

Copyright © 2001 Society for Neuroscience 0270-6474/01/214381-09\$15.00/0

Table 1. Comparison of chronic prenatal administration of ethanol, isocaloric-sucrose/pair feeding, and water on offspring body, brain, and cerebral cortical weights

| Weight (gm) | Chronic treatment | | |
|---------------------|----------------------------|-----------------------------|---------------------------|
| | Ethanol (<i>n</i> = 8) | Sucrose (<i>n</i> = 12) | Water (<i>n</i> = 13) |
| P1 body | 93.1 ± 4.3* | 90.8 ± 4.6* | 104.6 ± 1.8 |
| P10 body | 182.5 ± 8.5 | 162.1 ± 8.5 | 176.9 ± 3.1 |
| P20 body | 288.8 ± 12.3 | 267.5 ± 11.2 | 273.5 ± 4.9 |
| P60 body | 609.4 ± 29.2 | 589.6 ± 18.5 | 563.8 ± 14.8 |
| P60 brain | 3.22 ± 0.05** | 3.72 ± 0.04 | 3.74 ± 0.04 |
| P60 cerebral cortex | 1.76 ± 0.03** | 2.02 ± 0.03 | 2.03 ± 0.03 |

**p* < 0.05 compared with the water treatment group.

***p* < 0.05 compared with the isocaloric sucrose and water treatment groups.

altered by chronic prenatal exposure to ethanol (Morrisett et al., 1989; Costa et al., 2000).

MATERIALS AND METHODS

Experimental animals. Nulliparous female Dunkin-Hartley strain guinea pigs (550–600 gm body weight; Charles River Canada, St. Constant, Quebec, Canada) were bred with male guinea pigs of the same strain (650–1000 gm body weight) according to an established procedure (Elvidge, 1972). The last day of full vaginal membrane opening was identified as gestational day 0 (G0) (term, ~G68). On G1, the pregnant animals were housed individually at an ambient temperature of 23°C with a 12 hr light/dark cycle with lights on at 8:00 A.M.. Vaginal membrane status and general health of the pregnant animals were monitored, and body weight was measured daily throughout gestation. All guinea pigs were cared for according to the principles and guidelines of the Canadian Council on Animal Care. The experimental protocol was approved by the Queen's University Animal Care Committee.

Animal treatment regimens. On G2, pregnant guinea pigs were randomly assigned to receive oral administration of one of the following treatment regimens up to and including G67: (1) 4 gm of ethanol per kilogram of maternal body weight per day with *ad libitum* access to pellet food (PMI Nutrition International guinea pig diet 5025) and water; (2) isocaloric-sucrose and pair feeding with *ad libitum* access to water; or (3) isovolumetric water with *ad libitum* access to food and water. The pair-feeding regimen consisted of each sucrose-treated guinea pig being paired to an ethanol-treated guinea pig, receiving sucrose that was isocaloric and isovolumetric to the daily ethanol dose and receiving food in an amount that was equal to that consumed by the ethanol-treated guinea pig on each day of gestation. The daily treatments were given via oral intubation into the mouth and were administered as two equally divided doses 2 hr apart commencing between 9:30 and 11:00 A.M.. The ethanol (30% v/v) and sucrose (42% w/v) solutions were prepared with tap water. On G57, 200 μl of blood was taken from an ear blood vessel of the ethanol-treated pregnant animals at 1 hr after the second divided dose of ethanol for the determination of ethanol concentration by an established gas-liquid chromatographic method (Steenart et al., 1985). Blood was collected from animals of the isocaloric-sucrose and water treatment groups at 1 hr after the second daily dose on G57 to control for the stress of blood sampling. Litters were transferred to large plastic bins with wood chip bedding on the day of birth [postnatal day 0 (P0)]. Beginning at P1, offspring were weighed and monitored daily for general health. The offspring were weaned from their mothers on P17 and were separated based on gender on P22. Offspring of each gender and for all treatment groups were housed randomly in groups of up to four animals per bin.

Locomotor activity testing. On each of P10 (preweaning), P20 (postweaning), and P60 (adulthood), horizontal and vertical spontaneous locomotor activity were measured for 1 hr by placing individual offspring in a 42 × 42 cm open field apparatus (Columbus Instruments, Columbus, OH). Infrared photobeams 3 cm above the floor of the instrument measured movement in the horizontal plane, and photobeams placed at 10 cm (P10 and 20) or 12.5 cm (P60) above the floor measured movement in the vertical plane. The test was recorded by videotape for 1 hr in an isolated, quiet environment. Videotapes were reviewed, and the cumu-

lative number of photobeam breaks in each plane was recorded at each 10 min interval.

Tissue collection. On P61, individual offspring were killed by decapitation under halothane anesthesia. The brains were rapidly excised, weighed, and dissected. Cerebral cortices were isolated from each brain, frozen in liquid N₂, weighed, and stored at -70°C until analyzed. For each animal, the right cerebral cortex was allocated to the radioligand binding assay experiments, and the left cerebral cortex was used in the immunoblot analysis.

Preparation of crude cell membrane fraction for radioligand binding assays. Right cerebral cortex from each animal was thawed and homogenized in 20 ml of homogenization buffer (10 mM Tris-HCl, 300 mM sucrose, and 2 mM EDTA, pH 7.4) per gram of tissue. This homogenate was centrifuged at 1000 × *g* for 10 min at 4°C. The resulting supernatant was centrifuged at 20,000 × *g* for 20 min at 4°C. The resulting pellet was then resuspended in 3 ml of homogenization buffer and frozen at -70°C until analyzed.

Radioligand binding assays. [³H]Muscimol binding to GABA_A receptors was determined by first washing the cell membrane preparation as follows: individual aliquots were diluted with five volumes of wash buffer (50 mM Tris-HCl and 2 mM EDTA, pH 7.4), mixed, and centrifuged at 13,000 × *g* for 10 min at 4°C. This washing procedure was repeated twice, and the final pellet was resuspended in binding assay buffer A (10 mM Tris-HCl and 150 mM NaCl, pH 7.4). The protein concentration of each sample was determined by a spectrophotometric protein dye-binding assay based on the method of Bradford (1976), using bovine serum albumin as the standard. [³H]Muscimol saturation binding was determined in reaction vessels containing 100–150 μg of the washed cell membrane fraction, [³H]muscimol in concentrations ranging from 1 to 60 nM (20 Ci/mmol; New England Nuclear, Boston, MA), and binding assay buffer A to give a total volume of 500 μl. Nonspecific binding of [³H]muscimol was determined in the presence of 100 μM GABA. Samples were incubated for 60 min at 4°C, and the reaction was terminated by rapid vacuum filtration through Whatman GF/B glass fiber filters that were prewet with 5 ml of ice-cold binding assay buffer A. The filters were washed twice with 5 ml of ice-cold binding assay buffer A, and the radioactivity remaining on the filters was quantified by liquid scintillation spectrometry using a Beckman LS 6500 scintillation counter.

[³H]MK-801 binding to NMDA receptors was determined by first washing the cell membrane fraction as described above with the following changes: binding assay buffer B (20 mM HEPES and 1 mM EDTA, pH 7.4) was used, and the samples were incubated for 30 min at 37°C before the first centrifugation. [³H]MK-801 binding was determined in reaction vessels containing 200 μg of the washed cell membrane fraction, 5 nM [³H]MK-801 (21.7 Ci/mmol; New England Nuclear), 100 μM glutamate, 100 μM glycine, and binding assay buffer B to give a total volume of 500 μl. Nonspecific binding of [³H]MK-801 was determined in the presence of 100 μM MK-801. Samples were incubated for 3 hr at 37°C, the reaction was terminated by rapid vacuum filtration, and the samples were quantified as described above.

Isolation of protein for immunoblot analysis. Left cerebral cortex from each animal was thawed and homogenized in 3 ml/gm of lysis buffer [10 mM Tris-HCl, 150 mM NaCl, 5 mM EDTA, 1% (w/v) SDS, 1% (w/v) sodium deoxycholate, and 1% (v/v) Triton X-100 plus protease inhibitors (1 μg of aprotinin, 5 μg of leupeptin, and 10 μg of PMSF/ml of lysis buffer)]. The homogenate was centrifuged at 13,000 × *g* for 30 min at 4°C, and the supernatant was collected. The protein concentration of each supernatant was determined by the Lowry et al. (1951) assay, using bovine serum albumin as the standard. Samples were then frozen and stored at -70°C until analyzed.

Immunoblot analysis. Immunoblot analysis of the GABA_A receptor α1 subunit was performed by SDS-PAGE using a Bio-Rad (Mississauga, Ontario, Canada) Mini Protean III apparatus. Individual protein samples were heated at 100°C for 5 min in loading buffer [65.8 mM Tris-HCl, 10% (v/v) glycerol, 2% (w/v) SDS, 4% (v/v) 2-mercaptoethanol, and 0.025% (w/v) bromophenol blue, pH 6.8]. For each sample, 30 μg of protein in 20 μl of loading buffer was loaded onto a 10% SDS-polyacrylamide gel. Each sample was analyzed in triplicate, and each gel contained samples from three animals (one animal from each of the three treatment groups). The protein samples were separated in the presence of a running buffer [15 mM Tris base, 115 mM glycine, and 0.06% (w/v) SDS, pH 8.3] at room temperature for 60 min at 100 V.

The separated protein samples were then transferred onto a polyvinylidene difluoride carrier membrane (Amersham Pharmacia Biotech, Baie d'Urfe, Quebec, Canada). Protein transfer was performed using the

Bio-Rad Mini Trans Blot system in the presence of ice-cold transfer buffer [25 mM Tris base, 192 mM glycine, and 20% (v/v) methanol, pH 8.3] for 60 min at 100 V. Nonspecific sites were blocked by incubating the membrane in blocking solution [5% (w/v) skim milk powder, Tris-buffered saline containing Tween 20 (TBS-T) (50 mM Tris-HCl, 400 mM NaCl, 0.05% (v/v) Tween 20, pH 7.6)] for 1 hr at room temperature. The membrane was washed with TBS-T and then incubated with 2 μ g/ml of a rabbit polyclonal, affinity-purified antibody raised against amino acids 1–15 of the rat GABA_A receptor α 1 subunit (Upstate Biotechnology, Lake Placid, NY) in blocking solution overnight at 4°C. The membrane was washed with TBS-T and then incubated with a 1:4000 dilution in blocking solution of a goat anti-rabbit IgG (H+L) secondary antibody conjugated with horseradish peroxidase (Bio-Rad) for 1 hr at room temperature. The membrane was washed thoroughly with TBS-T and developed using the ECL chemiluminescence detection system and Hyperfilm ECL x-ray film (Amersham Pharmacia Biotech). The amount of total protein loaded on each gel was within the linear range of optical density for all proteins examined in this study.

To control for differences in protein loading, each membrane was stripped and reprobed for β -actin as follows: the membrane was washed with TBS-T and incubated in stripping solution (200 mM glycine, pH 2.6) for 1 hr at room temperature. The blot was washed with TBS-T and probed for β -actin as described above. A 1:4000 dilution of a mouse monoclonal anti- β -actin antibody was used (Sigma-Aldrich Canada, Oakville, Ontario, Canada), followed by a 1:4000 dilution of a goat anti-mouse IgG (H+L) secondary antibody (Bio-Rad).

The immunoblot bands were quantified by measuring their relative optical density (ROD) using Corel (Ottawa, Ontario, Canada) PhotoPaint 9. ROD measurements for the GABA_A receptor subunits were normalized to the ROD of the β -actin signal within each lane, because chronic prenatal ethanol exposure did not affect β -actin content in the cerebral cortex (data not shown).

Immunoblot analysis of the GABA_A receptor β 2/3 subunits was performed as described above for the α 1 subunit with the following alterations. For each sample, 20 μ g of protein was loaded onto the SDS-polyacrylamide gel after incubation in loading buffer (without 2-mercaptoethanol) for 30 min at 37°C. A mouse monoclonal antibody raised against whole GABA_A receptor β 2 and β 3 subunits was used at 2 μ g/ml (Upstate Biotechnology), and the goat anti-mouse IgG (H+L) secondary antibody was used at a 1:4000 dilution. After detection and stripping, the membrane was probed for β -actin using a 1:2000 dilution of the mouse monoclonal anti- β -actin antibody and a 1:4000 dilution of the goat anti-mouse secondary antibody.

Data analysis. The locomotor activity, body, brain, and cerebral cortical weight data are presented as the mean \pm SEM of 8–13 offspring obtained from five different litters for each treatment group. The radioligand binding and immunoblot data are presented as the mean \pm SEM of eight animals from five different litters for each treatment group. The B_{max} and K_D for [³H]muscimol binding were obtained by nonlinear regression analysis of individual saturation binding isotherms (Prism 2.0; GraphPad Software, San Diego, CA). Statistical analysis of the locomotor activity data was performed using two-way ANOVA (Prism 2.0). All other parametric statistical analyses were performed using one-way, randomized-design ANOVA, followed by the Newman–Keuls *post hoc* test for a statistically significant *F* statistic (Prism 2.0). The data were analyzed for homogeneity of variance before conducting the ANOVA tests. Correlational analyses were performed using the Pearson correlation test (Prism 2.0). The level for a statistical difference in the data was set at $p < 0.05$. Unless otherwise noted, statistical difference is cited in the text when the data for the ethanol treatment group were different from the data for both the isocaloric-sucrose/pair-fed and water treatment groups. Because there was no effect of gender on any of the observed parameters in any treatment group, the data of the male and female offspring in each treatment group were combined for the purpose of statistical analysis.

RESULTS

Maternal food intake and weight gain

Maternal average daily caloric intake and weights throughout gestation are displayed in Figure 1. Pregnant guinea pigs in the isocaloric-sucrose/pair-fed treatment group consumed food that was equal to that of the ethanol treatment group because of the pair-feeding regimen. The maternal average daily caloric intake

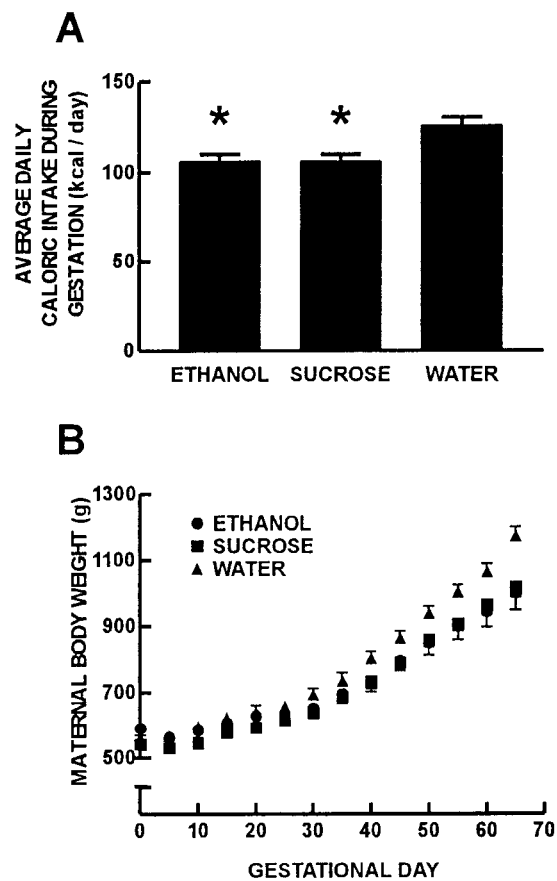


Figure 1. Pregnant guinea pig nutritional data for the five dams in each treatment group. Maternal average daily caloric intake during gestation (*A*). Pregnant guinea pigs in the ethanol treatment group consumed less food than those in the water treatment group ($*p < 0.05$). Pregnant animals in the isocaloric-sucrose/pair-fed treatment group were given and consumed food equal to that of the ethanol treatment group by nature of the pair-feeding regimen. *B*, Maternal body weight for the three treatment groups throughout gestation. Circles represent ethanol, squares represent isocaloric-sucrose/pair-fed, and triangles represent water treatment groups. Pregnant guinea pigs in the ethanol and isocaloric-sucrose/pair-fed treatment groups gained less weight than those in the water treatment group ($p < 0.05$).

for animals in the ethanol and isocaloric-sucrose/pair-fed treatment groups was 106.5 ± 7.8 kcal/d compared with 125.8 ± 9.7 kcal/d for animals in the water treatment group (Fig. 1*A*) ($p < 0.05$). There was no difference in maternal body weight at G0. However, pregnant guinea pigs in the ethanol and isocaloric-sucrose/pair-fed treatment groups gained significantly less weight during gestation than the pregnant guinea pigs in the water treatment group (Fig. 1*B*) ($p < 0.05$).

Maternal blood ethanol concentration

The maternal blood ethanol concentration of the guinea pigs in the ethanol treatment group 1 hr after the second divided dose administered on G57 was 328 ± 55 mg/dl (71.3 ± 12.0 mM). This maternal blood ethanol concentration value was similar to that found in previous experiments from our laboratory using the same chronic ethanol treatment regimen in the guinea pig, in which specific behavioral and neurochemical teratogenic effects were observed (Abdollah et al., 1993; Catlin et al., 1993; Kimura and Brien, 1998; Gibson et al., 2000).

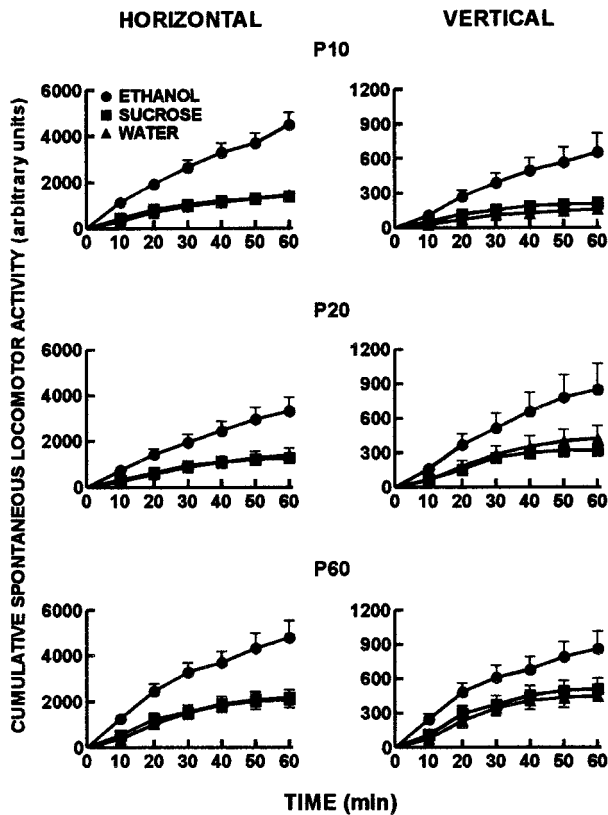


Figure 2. Offspring cumulative horizontal and vertical spontaneous locomotor activity measured in an open-field apparatus for a 60 min period. Activity was measured on each of P10 (preweaning), P20 (postweaning), and P60 (adulthood). *Circles* represent ethanol, *squares* represent isocaloric-sucrose/pair-fed, and *triangles* represent water treatment groups. The cumulative horizontal and vertical spontaneous locomotor activity data of the offspring in the ethanol treatment group were greater compared with the data for the isocaloric-sucrose/pair-fed and water treatment groups at each postnatal age ($p < 0.05$).

Pregnancy outcome

There was no maternal lethality or spontaneous abortion in any treatment group. Chronic maternal ethanol administration had no effect on gestation length, litter size, or the ratio of male to female offspring. There were three incidents of perinatal death (death at parturition) in the ethanol treatment group and one incident in the water treatment group.

Locomotor activity

Offspring horizontal and vertical spontaneous locomotor activity data are presented in Figure 2. At each of P10, 20, and 60, chronic prenatal ethanol administration resulted in increased cumulative spontaneous locomotor activity in both the horizontal and vertical planes compared with the isocaloric-sucrose/pair-fed and water treatment groups ($p < 0.05$).

Body, brain, and cerebral cortical weights

Offspring body, brain, and cerebral cortical weights are presented in Table 1. Both chronic prenatal ethanol treatment and isocaloric-sucrose administration with pair feeding resulted in decreased body weight at P1 compared with the water treatment group ($p < 0.05$). There was no effect of chronic prenatal treatment on body weight at any other postnatal age examined. At P61, chronic prenatal ethanol treatment resulted in decreased brain

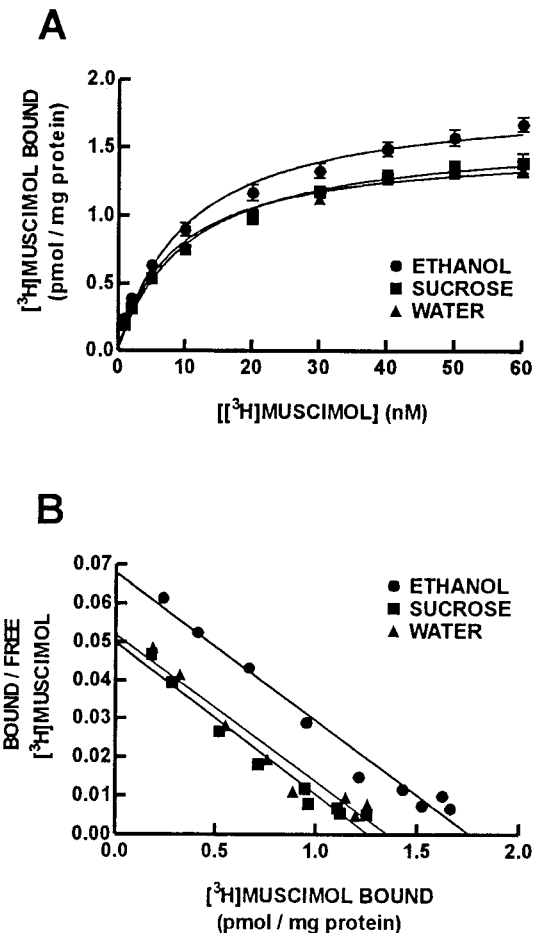


Figure 3. Saturation binding isotherms for [³H]muscimol binding to the high-affinity site on the GABA_A receptor (*A*). Data are presented as the mean specific binding of [³H]muscimol \pm SEM for eight offspring in each treatment group. *Circles* represent ethanol, *squares* represent isocaloric-sucrose/pair-fed, and *triangles* represent water treatment groups. The number of [³H]muscimol binding sites (B_{max}) was increased for offspring in the ethanol treatment group (Fig. 4) ($p < 0.05$), whereas the affinity of [³H]muscimol for the GABA_A receptor was not affected by prenatal ethanol treatment. Representative Scatchard transformations of the [³H]muscimol saturation data also are presented (*B*).

and cerebral cortical weights compared with the isocaloric-sucrose/pair-fed and water treatment groups ($p < 0.05$).

[³H]Muscimol binding to GABA_A receptors

Mean binding isotherm curves for [³H]muscimol binding to cerebral cortical GABA_A receptors at P61 are presented in Figure 3*A*. Representative Scatchard transformations are presented in Figure 3*B*. Chronic prenatal ethanol treatment increased the B_{max} for [³H]muscimol binding to the high-affinity GABA binding site on GABA_A receptors compared with the isocaloric-sucrose/pair-fed and water treatment groups (Fig. 4*A*) ($p < 0.05$). There was no effect of prenatal ethanol treatment on the K_D (affinity) for [³H]muscimol binding to these binding sites. The [³H]muscimol K_D values were 11.9 ± 1.3 nM (ethanol group), 12.1 ± 1.4 nM (isocaloric-sucrose/pair-fed group), and 10.1 ± 1.4 nM (water group).

GABA_A receptor $\alpha 1$ and $\beta 2/3$ subunit proteins

GABA_A receptor $\alpha 1$ subunit protein immunoblot analysis resulted in a strong signal at 51 kDa. Representative x-ray immu-

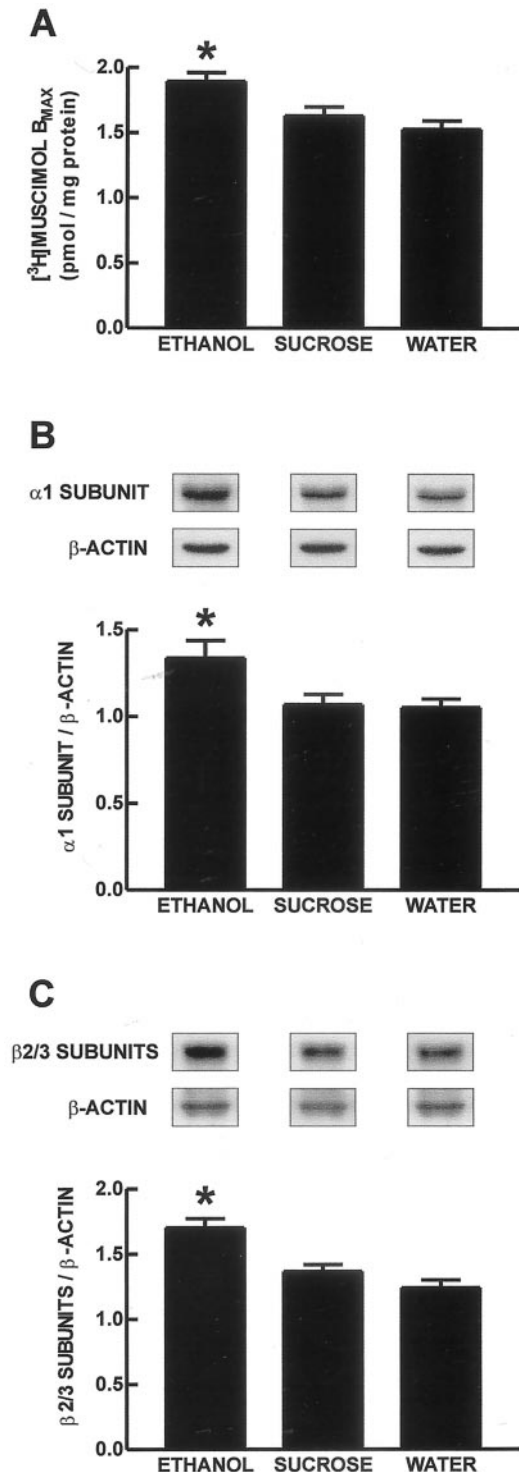


Figure 4. GABA_A receptor [³H]muscimol binding and relative protein content for GABA_A receptor α1 and β2/3 subunits. *A*, B_{max} values for [³H]muscimol binding to the high-affinity binding site of the GABA_A receptor in the adult guinea pig (P61) cerebral cortical cell membrane protein. The data are presented as the mean ± SEM of eight offspring. Representative immunoblot bands and mean ± SEM (*n* = 8 offspring in each treatment group) data for GABA_A receptor α1 (*B*) and β2/3 (*C*) subunit proteins in adult guinea pig (P61) cerebral cortex. The data are expressed as relative amount of GABA_A receptor subunit protein normalized to β-actin protein. **p* < 0.05 for the increase in each of [³H]muscimol B_{max} (*A*), and the relative amounts of protein for the GABA_A receptor α1 (*B*) and β2/3 (*C*) subunits in the ethanol treatment group compared with the isocaloric-sucrose/pair-fed and water treatment groups.

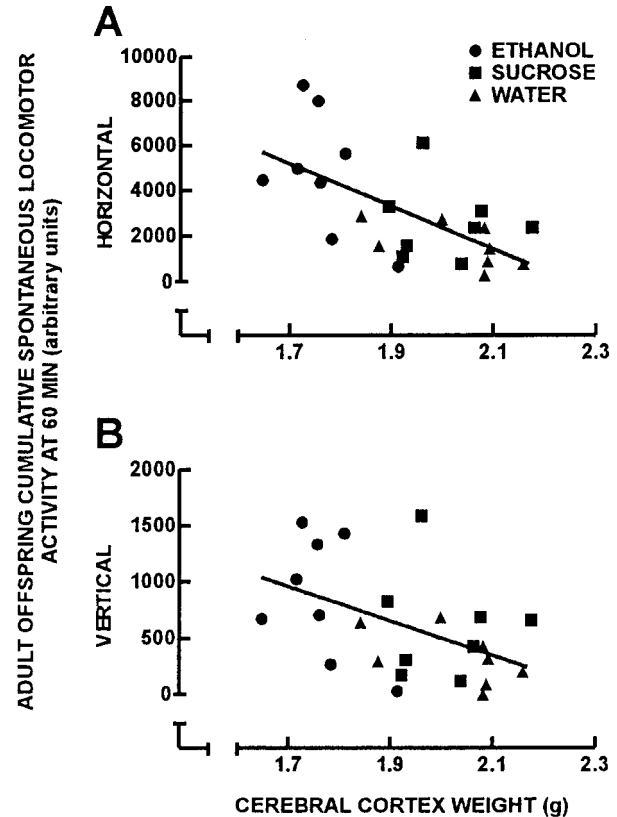


Figure 5. Linear-linear plots for cerebral cortical weight and cumulative horizontal (*A*) and vertical (*B*) spontaneous locomotor activity data at 60 min of the 24 adult offspring of the three treatment groups. Circles represent ethanol, squares represent isocaloric-sucrose/pair-fed, and triangles represent water treatment groups. There was a negative correlation between cerebral cortical weight and spontaneous locomotor activity in both the horizontal (*A*) ($r = 0.629$; $p < 0.05$) and vertical (*B*) ($r = 0.489$; $p < 0.05$) planes.

noblots and group data for the GABA_A receptor α1 subunit protein are presented in Figure 4*B*. Chronic prenatal ethanol administration increased the relative amount of GABA_A receptor α1 subunit protein normalized to β-actin (1.34 ± 0.10 ; $n = 8$) compared with the isocaloric-sucrose/pair-fed (1.07 ± 0.06 ; $n = 8$) and water (1.05 ± 0.05 ; $n = 8$) treatment groups ($p < 0.05$).

Immunoblot analysis of the GABA_A receptor β2/3 subunit proteins resulted in a signal with a molecular weight of 55–57 kDa. Representative immunoblot bands and data are presented in Figure 4*C*. Chronic prenatal ethanol administration increased the relative amount of these proteins (1.70 ± 0.07 ; $n = 8$) compared with the isocaloric-sucrose/pair-fed (1.36 ± 0.05 ; $n = 8$) and water (1.24 ± 0.06 ; $n = 8$) treatment groups ($p < 0.05$).

[³H]MK-801 binding to NMDA receptors

The amount of [³H]MK-801 bound to cerebral cortical NMDA receptors at P61 was not different among the three prenatal treatment groups. The amounts of [³H]MK-801 bound at 5 nM concentration were 751.4 ± 38.6 fmol/mg of protein (ethanol group), 784.9 ± 26.7 fmol/mg of protein (isocaloric-sucrose/pair-fed group), and 810.1 ± 38.1 fmol/mg of protein (water group).

Correlational analyses

Correlational analyses were performed to compare cerebral cortical weight with cumulative horizontal and vertical spontaneous locomotor activity at P60 for individual offspring in which a

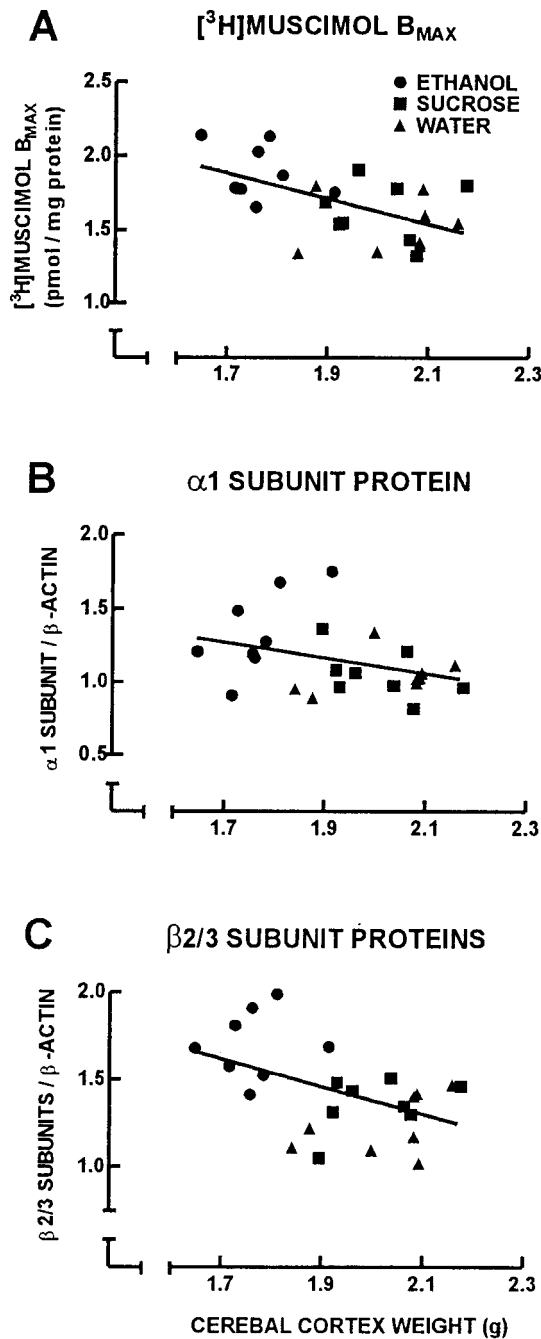


Figure 6. Linear-linear plots for cerebral cortical weight and [³H]muscimol B_{max} (A), and relative amounts of protein for the GABA_A receptor α1 (B) and β2/3 subunits (C) of the 24 adult offspring of the ethanol (circles), isocaloric-sucrose/pair-fed (squares), and water (triangles) treatment groups. There was a negative correlation between cerebral cortical weight and [³H]muscimol B_{max} (A) ($r = 0.550$; $p < 0.05$), and between cerebral cortical weight and the relative amount of protein for the GABA_A receptor β2/3 subunits (C) ($r = 0.472$; $p < 0.05$).

complete set of behavioral, cerebral cortical weight, and neurochemical data were obtained (Fig. 5). There was a negative correlation between adult offspring cerebral cortical weight and the cumulative horizontal (Fig. 5A) ($r = 0.629$; $p < 0.05$) and vertical (Fig. 5B) ($r = 0.489$; $p < 0.05$) spontaneous locomotor activity. Correlational analyses were performed to compare adult guinea pig cerebral cortical weight with GABA_A receptor binding

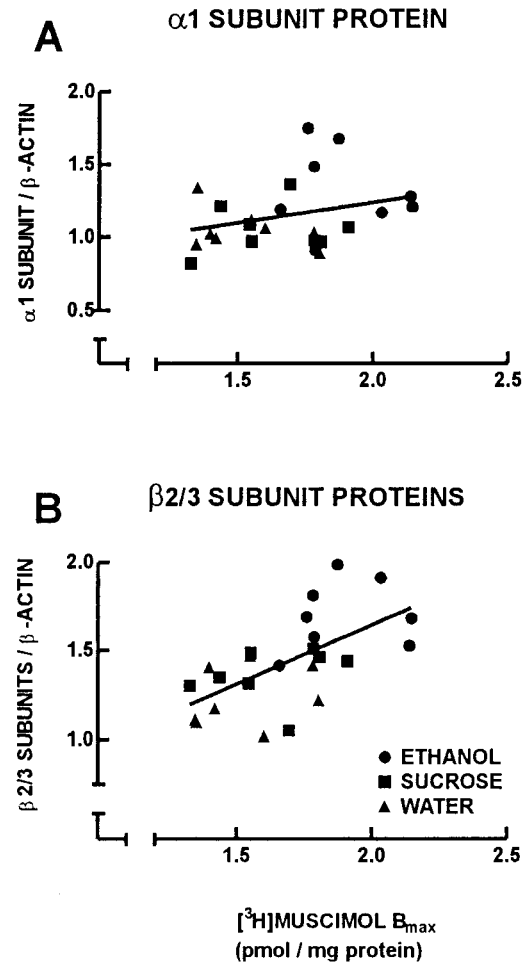


Figure 7. Linear-linear plots for [³H]muscimol B_{max} and relative amounts of protein for the GABA_A receptor α1 (A) and β2/3 (B) subunits of the 24 adult offspring of ethanol (circles), isocaloric-sucrose/pair-fed (squares), and water (triangles) treatment groups. There was a positive correlation between [³H]muscimol B_{max} and the relative amount of protein for the GABA_A receptor β2/3 subunits (B) ($r = 0.622$; $p < 0.05$).

and GABA_A receptor subunit protein content (Fig. 6). There was a negative correlation between cerebral cortical weight and both GABA_A receptor number ([³H]muscimol B_{max}) (Fig. 6A) ($r = 0.550$; $p < 0.05$) and GABA_A receptor β2/3 subunit protein content (Fig. 6C) ($r = 0.472$; $p < 0.05$). There was no correlation between cerebral cortical weight and GABA_A receptor α1 subunit protein content (Fig. 6B) ($r = 0.343$). Correlational analysis also was performed to identify relationships between the B_{max} for [³H]muscimol and the relative amount of protein for the GABA_A receptor α1 and β2/3 subunits (Fig. 7). There was no correlation between [³H]muscimol B_{max} and the relative amount of GABA_A receptor α1 subunit protein (Fig. 7A) ($r = 0.284$). However, there was a positive correlation between the B_{max} for [³H]muscimol and the relative amount of GABA_A receptor β2/3 subunit proteins (Fig. 7B) ($r = 0.622$; $p < 0.05$).

DISCUSSION

Chronic prenatal exposure to ethanol throughout gestation, via maternal ethanol administration, resulted in an increase in GABA_A receptor number in the adult guinea pig cerebral cortex. Concurrent with this chronic prenatal ethanol-induced increase in receptor number was an increase in the relative expression of

protein for cerebral cortical GABA_A receptor $\alpha 1$ and $\beta 2/3$ subunits in these same animals. This effect was found in the adult offspring 61 d after they were born, which demonstrates that prenatal exposure to ethanol can produce long-term neurochemical alterations. Indeed, young adults diagnosed with FAS during childhood have persistent behavioral deficits, including impairments in attention and memory (Kerns et al., 1997).

The decrease in maternal average daily food intake and corresponding decrease in maternal body weight gain in the ethanol and isocaloric-sucrose/pair-fed treatment groups compared with the water treatment group indicates that the isocaloric-sucrose/pair-fed treatment group was an appropriate nutritional control in this study. Moreover, offspring in the isocaloric-sucrose/pair-fed treatment group had birth weights that were similar to those in the ethanol treatment group and less than those in the water treatment group (Table 1). Although offspring body weights normalized by P10, chronic prenatal ethanol exposure resulted in persistent effects on decreasing brain and cerebral cortical weights, as seen at P61 (Table 1).

In this study, the persistent long-term effects of chronic prenatal ethanol exposure to increase locomotor activity and to decrease brain and cerebral cortical weights were of similar magnitude to results found in previous experiments from our laboratory using the same dosing regimen (Abdollah et al., 1993; Catlin et al., 1993; Butters et al., 2000). These changes are considered to be key manifestations of ethanol neurobehavioral teratogenicity. In this current study we found a strong negative correlation between cerebral cortical weight and locomotor activity and negative correlations between cerebral cortical weight and both the GABA_A receptor number and GABA_A receptor $\beta 2/3$ subunit protein expression. These data demonstrate a relationship between brain injury and altered behavior in this animal model and suggest that the magnitude of changes in GABA_A receptor expression are related to the extent of brain injury after chronic prenatal ethanol exposure.

Previous studies from our laboratory have shown that chronic prenatal ethanol exposure increases the number of benzodiazepine-binding GABA_A receptors in the adult (P61) guinea pig cerebral cortex but not at P11 or P21 (Bailey et al., 1999). GABA_A receptors normally contain α and β subunits but also require γ (with or without θ) subunits to confer benzodiazepine binding (Barnard et al., 1998; Bonnert et al., 1999). Using [³H]muscimol, a ligand that is selective for the high-affinity GABA binding site on the GABA_A receptor, it was possible in the present study to determine the relative number of all GABA_A receptors in the adult guinea pig cerebral cortex (i.e., those with and without benzodiazepine binding sites). The $\alpha 1$ and $\beta 2/3$ subunits were chosen for examination in this study because they are the most predominant GABA_A receptor subunit subtypes in their respective subunit families in the adult cerebral cortex (Laurie et al., 1992) and would therefore be the most likely receptor subunits responsible for an increase in GABA_A receptor number. This hypothesis was supported for the $\beta 2/3$ subunits, for which there was a positive correlation between the number of [³H]muscimol binding sites and the relative amount of protein for the GABA_A receptor $\beta 2/3$ subunits of the individual animals of the ethanol, isocaloric-sucrose/pair-fed, and water treatment groups. The $\alpha 1$ subunit protein expression did not correlate with [³H]muscimol binding, which suggests that other α subunits also contribute to the observed increased number of [³H]muscimol binding sites. This idea is also supported by our previous study (Bailey et al., 1999), in which we reported that chronic prenatal

ethanol exposure produced a decrease in the relative proportion of zolpidem-sensitive GABA_A receptors (putative $\alpha 1$ subunit-containing GABA_A receptors) in the adult guinea pig cerebral cortex.

GABA_A receptor expression has been demonstrated to increase in response to chronic GABA_A receptor hypofunction *in vivo* (Miller et al., 1989). The observed increase in GABA_A receptor number in the adult guinea pig cerebral cortex produced by chronic prenatal ethanol exposure may therefore be a compensatory mechanism for a potential decrease in GABAergic neurotransmission and/or GABA_A receptor function at earlier time points in development. Chronic prenatal exposure to ethanol in a rat model has been demonstrated to decrease the whole-cell patch clamp response to GABA in medial septum/diagonal band neurons in juvenile offspring but not in adult offspring (Hsiao et al., 1998). The anxiolytic response to a naturally occurring neuroactive steroid, 3 α -hydroxy-5 α -pregnan-20-one (allo-pregnanolone), which potentiates GABA_A receptor function, was also decreased in neonatal rats that were exposed to ethanol during gestation (Zimmerberg et al., 1995). Interestingly, GABA_A receptor function has been demonstrated to be increased in adult offspring that were prenatally exposed to ethanol. The behavioral effect of benzodiazepines to reduce anxiety was found to be increased in adult rats that were exposed to ethanol *in utero* (Osborn et al., 1998). Adult rats that were exposed to ethanol during the last third of gestation exhibited an increased electrophysiological response to GABA in frontal and somatosensory cortical neurons (Janiri et al., 1994). One report demonstrated that prenatal exposure to ethanol did not affect GABA-elicited Cl⁻ flux through GABA_A receptors in adult rat offspring but decreased modulation of Cl⁻ flux by benzodiazepines and neuroactive steroids in the cerebral cortex (Allan et al., 1998). The disagreement among these studies in adult offspring may reflect differences in prenatal ethanol treatment regimens and differences in blood ethanol concentrations achieved by these different regimens, but also may suggest that there are brain regional differences in the long-term effects of prenatal exposure to ethanol.

One possible morphological explanation for a decrease in GABAergic input and subsequent upregulation of GABA_A receptors is a decrease in the number of cells that express GABA_A receptors and/or use GABA as a neurotransmitter. Affected cell types in the cerebral cortex may include GABAergic interneurons, glutamatergic pyramidal cells, or glial cells (Bureau et al., 1995; Hornung and Fritschy, 1996). Chronic prenatal exposure to ethanol in the rat decreases neuronal and glial cell number in the somatosensory cortex of adult offspring (Miller and Potempa, 1990) and, more specifically, in the number of parvalbumin-positive neurons in the cingulate cortex, an index of normally functioning GABAergic neurons (Moore et al., 1998). Immunohistochemical experiments are being conducted in our laboratory to identify the cell types that may be affected by chronic prenatal ethanol exposure in the adult guinea pig somatosensory cortex. A decrease in GABAergic input also could result from a decrease in GABA synthesis and/or release from existing GABAergic interneurons. However, preliminary experiments in our laboratory in the adult guinea pig cerebral cortex indicate that protein content for glutamic acid decarboxylase, the enzyme responsible for GABA synthesis from glutamate, is not affected by chronic prenatal ethanol exposure (data not shown). The effect of chronic prenatal ethanol exposure on synaptic GABA release in the cerebral cortex has not been examined.

In this study, adult guinea pig cerebral cortical NMDA receptor number was not statistically different after chronic prenatal ethanol exposure. In the adult rat, chronic prenatal ethanol exposure decreases NMDA receptor number and produces electrophysiologic deficits in the hippocampus (Morrisett et al., 1989; Savage et al., 1991). Previous studies in our laboratory suggest that chronic prenatal ethanol exposure decreases glutamate and NMDA binding in the near-term fetal hippocampus (Abdollah and Brien, 1995) but increases MK-801 binding in the near-term fetal cerebral cortex (Chiu et al., 1999). Collectively, these data indicate that there may be brain regional and developmental age selectivity for the effects of prenatal ethanol exposure on NMDA receptors. Recently, it was proposed that ethanol acts by a dual mechanism of blockade of NMDA receptors and excessive activation of GABA_A receptors to produce apoptotic neurodegeneration in the developing rat forebrain (Ikonomidou et al., 2000). The results of the present study demonstrate that there are differential long-term effects of chronic prenatal ethanol exposure on the NMDA and GABA_A receptors in the adult cerebral cortex.

In conclusion, these findings in the guinea pig are novel and could have an important impact on elucidating the long-term neurobehavioral effects of chronic prenatal ethanol exposure. The GABA_A receptor system is the major inhibitory neurotransmitter system in the cerebral cortex and is present in 20–50% of cerebral cortical synapses (Bloom and Iversen, 1971; Halasy and Somogyi, 1993). The results of this study suggest that there may therefore be altered inhibitory neurotransmission and GABA_A receptor pharmacology within this brain region both during normal physiological activity and in response to neuroactive xenobiotics. The observed increases in GABA_A receptor number and expression of GABA_A receptor $\alpha 1$ and $\beta 2/3$ subunit proteins may be causally related, at least in part, to the known consequences of chronic prenatal ethanol exposure in the human, including deficits in cognition, attention, learning, and problem solving. This neurotransmitter receptor system may therefore be a potential target for therapeutic intervention to lessen the long-term neurobehavioral effects of prenatal ethanol exposure.

REFERENCES

- Abdollah S, Brien JF (1995) Effect of chronic maternal ethanol administration on glutamate and *N*-methyl-D-aspartate binding sites in the hippocampus of the near-term fetal guinea pig. *Alcohol* 12:377–382.
- Abdollah S, Catlin MC, Brien JF (1993) Ethanol neuro-behavioural teratogenesis in the guinea pig: behavioural dysfunction and hippocampal morphological change. *Can J Physiol Pharmacol* 71:776–782.
- Allan AM, Wu H, Paxton LL, Savage DD (1998) Prenatal ethanol exposure alters the modulation of the γ -aminobutyric acid_A receptor-gated chloride ion channel in adult rat offspring. *J Pharmacol Exp Ther* 284:250–257.
- Bailey CDC, Brien JF, Reynolds JN (1999) Altered GABA_A-benzodiazepine receptor number and pharmacology in the adult guinea pig cerebral cortex after chronic prenatal ethanol exposure. *Alcohol Clin Exp Res* 23:1816–1824.
- Barnard EA, Skolnick P, Olsen RW, Mohler H, Sieghart W, Biggio G, Braestrup C, Bateson AN, Langer SZ (1998) International Union of Pharmacology. XV. Subtypes of γ -aminobutyric acid_A receptors: classification on the basis of subunit structure and receptor function. *Pharmacol Rev* 50:291–313.
- Bloom FE, Iversen LL (1971) Localizing ³H-GABA in nerve terminals of rat cerebral cortex by electron microscopic autoradiography. *Nature* 229:628–630.
- Bonnert TP, McKernan RM, Farrar S, le Bourdelles B, Heavens RP, Smith DW, Hewson L, Rigby MR, Sirinathsinghji DJS, Brown N, Wafford KA, Whiting PJ (1999) θ , a novel γ -aminobutyric acid type A receptor subunit. *Proc Natl Acad Sci USA* 96:9891–9896.
- Bradford M (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72:248–254.
- Bureau M, Laschet J, Bureau-Heeren M, Hennuy B, Minet A, Wins P, Grisar T (1995) Astroglial cells express large amounts of GABA_A receptor proteins in mature brain. *J Neurochem* 65:2006–2015.
- Butters NS, Gibson MAS, Reynolds JN, Brien JF (2000) Effects of chronic prenatal ethanol exposure on hippocampal glutamate release in the postnatal guinea pig. *Alcohol* 21:1–9.
- Catlin MC, Abdollah S, Brien JF (1993) Dose-dependent effects of prenatal ethanol exposure in the guinea pig. *Alcohol* 10:109–115.
- Chiu J, Brien JF, Wu P, Eubanks JH, Zhang L, Reynolds JN (1999) Chronic ethanol exposure alters MK-801 binding sites in the cerebral cortex of the near-term fetal guinea pig. *Alcohol* 17:215–221.
- Costa ET, Olivera DS, Meyer DA, Ferreira VM, Soto EE, Frausto S, Savage DD, Browning MD, Valenzuela CF (2000) Fetal alcohol exposure alters neurosteroid modulation of hippocampal *N*-methyl-D-aspartate receptors. *J Biol Chem* 275:38268–38274.
- Devaud LL, Smith FD, Grayson DR, Morrow AL (1995) Chronic ethanol consumption differentially alters the expression of γ -aminobutyric acid_A receptor subunit mRNAs in rat cerebral cortex: competitive, quantitative reverse transcriptase-polymerase chain reaction analysis. *Mol Pharmacol* 48:861–868.
- Devaud LL, Fritschy JM, Sieghart W, Morrow AL (1997) Bidirectional alterations of GABA_A receptor subunit peptide levels in rat cortex during chronic ethanol consumption and withdrawal. *J Neurochem* 69:126–130.
- Elvidge H (1972) Production of dated pregnant guinea pigs without postpartum matings. *J Inst Animal Tech* 23:111–117.
- Gibson MAS, Butters NS, Reynolds JN, Brien JF (2000) Effects of chronic prenatal ethanol exposure on locomotor activity, and hippocampal weight, neurons, and nitric oxide synthase activity of the young postnatal guinea pig. *Neurotoxicol Teratol* 22:183–192.
- Halasy K, Somogyi P (1993) Distribution of GABAergic synapses and their targets in the dentate gyrus of rat: a quantitative immunoelectron microscopic analysis. *J Hirnforsch* 34:299–308.
- Hornung J-P, Fritschy J-M (1996) Developmental profile of GABA_A-receptors in the Marmoset monkey: expression of distinct subtypes in pre- and postnatal brain. *J Comp Neurol* 367:413–430.
- Hsiao SH, Mahoney JC, West JR, Frye GD (1998) Development of GABA_A receptors on medial septum/diagonal band (MD/DB) neurons after postnatal ethanol exposure. *Brain Res* 810:100–113.
- Hsiao SH, West JR, Mahoney JC, Frye GD (1999) Postnatal ethanol exposure blunts upregulation of GABA_A receptor currents in Purkinje neurons. *Brain Res* 832:124–135.
- Ikonomidou C, Bittigau P, Ishimaru MJ, Wozniak DF, Koch C, Genz K, Price MT, Stefovskaya V, Horster F, Tenkova T, Dikranian K, Olney JW (2000) Ethanol-induced apoptotic neurodegeneration and fetal alcohol syndrome. *Science* 287:1056–1060.
- Janiri L, Gobbi G, Persico AM, Santarelli M, Minciacchi D, Tempesta E (1994) Alterations of neocortical neuronal responses to acetylcholine and GABA in rats born to alcohol-dependent mothers. *Alcohol Alcohol* 29:611–619.
- Jones KL, Smith DW (1973) Recognition of the fetal alcohol syndrome in early infancy. *Lancet* 2:999–1001.
- Kerns KA, Don A, Mateer CA, Streissguth AP (1997) Cognitive deficits in nonretarded adults with fetal alcohol syndrome. *J Learn Disabil* 30:685–693.
- Kimura KA, Brien JF (1998) Hippocampal nitric oxide synthase in the fetal guinea pig: effects of chronic prenatal ethanol exposure. *Dev Brain Res* 106:39–46.
- Laurie DJ, Wisden W, Seeburg PH (1992) The distribution of thirteen GABA_A receptor subunit mRNAs in the rat brain. III. Embryonic and postnatal developments. *J Neurosci* 12:4151–4172.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with the folin phenol reagent. *J Biol Chem* 193:265–275.
- Mattson SN, Riley EP, Delis DC, Stern C, Jones KL (1996) Verbal learning and memory in children with fetal alcohol syndrome. *Alcohol Clin Exp Res* 20:810–816.
- Mehta AK, Ticku MK (1999) An update on GABA_A receptors. *Brain Res* 832:164–167.
- Mhatre MC, Ticku MK (1994) Chronic ethanol treatment up-regulates the GABA receptor β subunit expression. *Mol Brain Res* 23:246–252.
- Mhatre MC, Pena G, Sieghart W, Ticku MK (1993) Antibodies specific for GABA_A receptor alpha subunits reveal that chronic alcohol treatment down-regulates alpha-subunit expression in rat brain regions. *J Neurochem* 61:1620–1625.
- Miller LG, Greenblatt DJ, Roy RB, Gaver A, Lopez F, Shader RI (1989) Chronic benzodiazepine administration. III. Upregulation of gamma-aminobutyric acid_A receptor binding and function associated with chronic benzodiazepine antagonist administration. *J Pharmacol Exp Ther* 248:1096–1101.
- Miller MW, Potempa G (1990) Numbers of neurons and glia in mature rat somatosensory cortex: effects of prenatal exposure to ethanol. *J Comp Neurol* 293:92–102.
- Moore DB, Quintero MA, Ruygrok AC, Walker DW, Heaton MB (1998) Prenatal ethanol exposure reduces parvalbumin-immunoreactive GABAergic neuronal number in the adult rat cingulate cortex. *Neurosci Lett* 249:25–28.

- Morrisett RA, Martin D, Wilson WA, Savage DD, Swartzwelder HS (1989) Prenatal exposure to ethanol decreases the sensitivity of the adult rat hippocampus to *N*-methyl-D-aspartate. *Alcohol* 6:415–420.
- Osborn JA, Yu C, Gabriel K, Weinberg J (1998) Fetal ethanol effects on benzodiazepine sensitivity measured by behavior on the elevated plus-maze. *Pharmacol Biochem Behav* 60:625–633.
- Rabow LE, Russek SJ, Farb DH (1995) From ion currents to genomic analysis: recent advances in GABA_A receptor research. *Synapse* 21:189–274.
- Savage DD, Montano CY, Otero MA, Paxton LL (1991) Prenatal ethanol exposure decreases hippocampal NMDA-sensitive [³H]-glutamate binding site density in 45-day-old rats. *Alcohol* 8:193–201.
- Steenart NAE, Clarke DW, Brien JF (1985) Gas-liquid chromatographic analysis of ethanol and acetaldehyde in blood with minimal artifactual acetaldehyde formation. *J Pharmacol Meth* 14:199–212.
- Steinhausen HC, Spohr HL (1998) Long-term outcome of children with fetal alcohol syndrome: psychopathology, behavior, and intelligence. *Alcohol Clin Exp Res* 22:334–338.
- Streissguth AP, Aase JM, Clarren SK, Randels SP, LaDue RA, Smith DF (1991) Fetal alcohol syndrome in adolescents and adults. *JAMA* 265:1961–1967.
- Zimmerberg B, Drucker PC, Weider JM (1995) Differential behavioral effects of the neuroactive steroid allopregnanolone on neonatal rats prenatally exposed to alcohol. *Pharmacol Biochem Behav* 51:463–468.