

# Roles of Glutamate Receptor $\delta 2$ Subunit (GluR $\delta 2$ ) and Metabotropic Glutamate Receptor Subtype 1 (mGluR1) in Climbing Fiber Synapse Elimination during Postnatal Cerebellar Development

Kouichi Hashimoto,<sup>1</sup> Ryoichi Ichikawa<sup>2</sup>, Hajime Takechi,<sup>3</sup> Yoshiro Inoue,<sup>4</sup> Atsu Aiba,<sup>5</sup> Kenji Sakimura,<sup>6</sup> Masayoshi Mishina,<sup>7</sup> Tsutomu Hashikawa,<sup>8</sup> Arthur Konnerth,<sup>9</sup> Masahiko Watanabe,<sup>4</sup> and Masanobu Kano<sup>1</sup>

<sup>1</sup>Department of Physiology, Kanazawa University School of Medicine, Takara-machi, Kanazawa 920-8640, Japan, <sup>2</sup>Department of Anatomy, Sapporo Medical University, Sapporo 060-8556, Japan, <sup>3</sup>Department of Gerontology, Kyoto University Faculty of Medicine, Kyoto 606-8507, Japan, <sup>4</sup>Department of Anatomy, Hokkaido University School of Medicine, Sapporo 060-8638, Japan, <sup>5</sup>Division of Cell Biology, Department of Molecular and Cellular Biology, Kobe University Graduate School of Medicine, Kobe 650-0017, Japan, <sup>6</sup>Department of Cellular Neurobiology, Brain Research Institute, Niigata University, Niigata 951-8585, Japan, <sup>7</sup>Department of Molecular Neurobiology and Pharmacology, Graduate School of Medicine, University of Tokyo, and Cure Research for Evolutional Science and Technology (CREST), Japan Science and Technology Corporation, Tokyo 113-0033, Japan, <sup>8</sup>Laboratory for Neural Architecture, Brain Science Institute, RIKEN, Wako-shi, Saitama 351-0198, Japan, and <sup>9</sup>Institut für Physiologie, Ludwig-Maximilians-Universität München, 80802 München, Germany

Climbing fiber (CF) synapse formation onto cerebellar Purkinje cells (PCs) is critically dependent on the synaptogenesis from parallel fibers (PFs), the other input to PCs. Previous studies revealed that deletion of the glutamate receptor  $\delta 2$  subunit (GluR $\delta 2$ ) gene results in persistent multiple CF innervation of PCs with impaired PF synaptogenesis, whereas mutation of the metabotropic glutamate receptor subtype 1 (mGluR1) gene causes multiple CF innervation with normal PF synaptogenesis. We demonstrate that atypical CF-mediated EPSCs (CF-EPSCs) with slow rise times and small amplitudes coexisted with typical CF-EPSCs with fast rise times and large amplitudes in PCs from GluR $\delta 2$  mutant cerebellar slices. CF-EPSCs in mGluR1 mutant and wild-type PCs had fast rise times. Atypical slow CF responses of GluR $\delta 2$  mutant PCs were associated with voltage-dependent  $Ca^{2+}$  signals that were confined to PC distal dendrites. In the wild-type and mGluR1 mutant PCs, CF-induced

$Ca^{2+}$  signals involved both proximal and distal dendrites. Morphologically, CFs of GluR $\delta 2$  mutant mice extended to the superficial regions of the molecular layer, whereas those of wild-type and mGluR1 mutant mice did not innervate the superficial one-fifth of the molecular layer. It is therefore likely that surplus CFs of GluR $\delta 2$  mutant mice form ectopic synapses onto distal dendrites, whereas those of wild-type and mGluR1 mutant mice innervate proximal dendrites. These findings suggest that GluR $\delta 2$  is required for consolidating PF synapses and restricting CF synapses to the proximal dendrites, whereas the mGluR1-signaling pathway does not affect PF synaptogenesis but is involved in eliminating surplus CF synapses at the proximal dendrites.

**Key words:** climbing fiber; parallel fiber; cerebellum; Purkinje cell; synapse; glutamate receptor; postnatal development; mutant mouse

In the adult cerebellum, each Purkinje cell (PC) is innervated by a single climbing fiber (CF) that originates in the inferior olive and forms strong excitatory synapses onto proximal PC dendrites (Palay and Chan-Palay, 1974; Ito, 1984; Strata and Rossi, 1998). This one-to-one relationship is preceded by a developmental stage of redundant innervation of PCs by multiple CFs (Crépel, 1982). Elimination of supernumerary CFs proceeds in parallel with synaptogenesis onto PCs from parallel fibers (PFs), bifur-

cated axons of granule cells and the other excitatory input to PCs (Ito, 1984). The importance of PF–PC synapses in CF synapse elimination has been substantiated by the fact that multiple CF innervation persists in animals with impaired PF–PC synaptogenesis, such as x-irradiated rats (Woodward et al., 1974; Crépel and Delhaye-Bouchaud, 1979; Bravin et al., 1995; Sugihara et al., 2000) and agranular cerebellar mutants (*weaver*, *reeler*, and *staggerer* mice) (Crépel and Mariani, 1976; Mariani et al., 1977; Crépel et al., 1980; Mariani and Changeux, 1980).

Glutamate or its related amino acids is considered to be released from CF and PF terminals and to activate glutamate receptors (GluRs) on PCs. The GluR  $\delta 2$  subunit (GluR $\delta 2$ ) is highly enriched in PCs (Araki et al., 1993; Lomeli et al., 1993) that are localized at the junctional site of PF–PC synapses but not at the site of CF–PC synapses (Landsend et al., 1997). In PCs of GluR $\delta 2$ -deficient mice, the number of PF–PC synapses is reduced to nearly one-half that of wild-type mice, leaving approximately one-third of PC dendritic spines free of innervation (Kurihara et al., 1997). GluR $\delta 2$ -deficient mice display persistent multiple CF innervation, impaired long-term depression (LTD), and motor

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Correspondence should be addressed to Masanobu Kano, Department of Physiology, Kanazawa University School of Medicine, Takara-machi, Kanazawa 920-8640, Japan. E-mail: mkano@med.kanazawa-u.ac.jp.

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discoordination (Hirano et al., 1995; Kashiwabuchi et al., 1995). The metabotropic GluR subtype 1 (mGluR1) is also abundant in PCs that are localized at the perijunctional site of both the PF–PC and CF–PC (Baude et al., 1993; Nusser et al., 1994). Inactivation of the mGluR1 gene results in persistent multiple CF innervation, impaired LTD, and motor discoordination, without apparent defects in PF–PC synaptogenesis (Aiba et al., 1994; Conquet et al., 1994; Kano et al., 1997; L  v  n  s et al., 1997; Ichise et al., 2000).

To differentiate the roles of these two GluR molecules in CF synapse formation and elimination, we comparatively examined the GluR $\delta$ 2 and mGluR1 mutant mice by recording CF-mediated EPSCs (CF-EPSCs), imaging CF-induced Ca<sup>2+</sup> signals in PCs, and labeling CFs anterogradely. We present evidence that surplus CFs form ectopic synapses onto distal dendrites of GluR $\delta$ 2 mutant PCs, whereas CFs innervate proximal dendrites of mGluR1 mutant PCs. These findings suggest that GluR $\delta$ 2 is required for consolidating PF synapses and restricting CF synapses to the proximal dendritic domain, whereas the mGluR1-signaling pathway is involved in eliminating excess CF synapses at the proximal dendrites.

## MATERIALS AND METHODS

**Animals.** The GluR $\delta$ 2 mutant mice (Kashiwabuchi et al., 1995) and the mGluR1 mutant mice (Aiba et al., 1994) were produced as described. The GluR $\delta$ 2 mutant mice and their littermates were of C57BL/6  $\times$  TT2 genetic background (Yagi et al., 1993). The mGluR1 mutant mice and their littermates were of the 129/Sv  $\times$  C57BL/6 genetic background. Mice were kept in the same room at the animal facility with a 12 hr light/dark cycle. Because we have not found any electrophysiological or morphological difference so far between the wild-type (+/+) mice of C57BL/6  $\times$  TT2 genetic background and those of 129/Sv  $\times$  C57BL/6 genetic background, nonmutant (+/+) littermates including both genotypes were used in electrophysiological analyses as wild-type controls.

**Electrophysiology.** Sagittal cerebellar slices of 200–250  $\mu$ m thickness were prepared from wild-type, GluR $\delta$ 2 mutant, or mGluR1 mutant mice as described previously (Edwards et al., 1989; Llano et al., 1991; Kano and Konnerth, 1992; Aiba et al., 1994). Whole-cell recordings were made from visually identified PCs using a 40 $\times$  water-immersion objective attached to either an Olympus (BH-2 or BX50WI, Olympus Optical, Tokyo, Japan) or a Zeiss (Axioskop) upright microscope (Edwards et al., 1989; Llano et al., 1991) at 32°C. The resistance of patch pipettes was 3–6 M $\Omega$  when filled with an intracellular solution composed of (in mM): 60 CsCl, 30 Cs D-gluconate, 20 TEA-Cl, 20 BAPTA, 4 MgCl<sub>2</sub>, 4 ATP, and 30 HEPES; pH 7.3, adjusted with CsOH. The composition of standard bathing solution was (in mM): 125 NaCl, 2.5 KCl, 2 CaCl<sub>2</sub>, 1 MgSO<sub>4</sub>, 1.25 NaH<sub>2</sub>PO<sub>4</sub>, 26 NaHCO<sub>3</sub>, and 20 glucose, bubbled continuously with a mixture of 95% O<sub>2</sub> and 5% CO<sub>2</sub>. Bicuculline (10  $\mu$ M) was always present in the saline to block spontaneous IPSCs (Konnerth et al., 1990; Kano et al., 1992). Ionic currents were recorded with either an Axopatch-1D (Axon Instruments, Foster City, CA) or an EPC-9 (HEKA, Lambrecht, Germany) patch-clamp amplifier. The pipette access resistance was compensated as described by Llano et al. (1991). Stimulation and on-line data acquisition were performed using the PULSE software on a Macintosh computer (HEKA). Signals were filtered at 3 kHz and digitized at 20 kHz. The decay phase of EPSCs was fitted with the PULSE-FIT software (HEKA). A glass pipette with a 5- to 10- $\mu$ m-diameter tip that was filled with standard saline was used for stimulating CFs. To search for multiple CFs innervating the recorded PC, the stimulation pipette was moved systematically in the granule cell layer, and the stimulus intensity was gradually increased at each stimulation site (pulse width 0.1 msec; strength 0–100 V).

**Ca<sup>2+</sup> imaging.** PCs were loaded for at least 20 min with a Ca<sup>2+</sup> indicator (Oregon Green 488 BAPTA-1, Molecular Probes, Eugene, OR; 100  $\mu$ M) through the patch pipette at room temperature. A high-speed confocal laser-scanning microscope (Noran Odyssey or Oz) attached to an upright microscope (Zeiss Axioskop or Olympus BX50WI) was used to acquire fluorescence images at 30 or 60 Hz in parallel to the whole-cell recordings (Eilers et al., 1995a; Takechi et al., 1998). Full-frame images were recorded on an optical disk (TQ2026F, Panasonic) and analyzed

off-line with the Image-1 software (Universal Image). In some experiments, fluorescence images were acquired at 30 Hz by using a cooled CCD camera system (IMAGO, T.I.L.L. Photonics, Gr  felfing, Germany) attached to an upright microscope (Zeiss Axioskop). The Ca<sup>2+</sup>-dependent fluorescence signals from selected regions of interest were background corrected and expressed as increases in fluorescence divided by the prestimulus fluorescence values ( $\Delta F/F_0$ ) using Igor Pro software (Wavemetrics). In PCs with multiple CF innervation, it was often difficult to detect CF-induced Ca<sup>2+</sup> signals under normal resting potentials with an extracellular Ca<sup>2+</sup> concentration of 2 mM. This was presumably because the CF-induced EPSPs were too small to evoke spikes or, even if they induced spikes, the associated Ca<sup>2+</sup> entry was too small to be detected. In these cases, we depolarized the PCs by constant current injection so that CF-EPSPs evoked spikes and increased the extracellular Ca<sup>2+</sup> concentration to 3 mM.

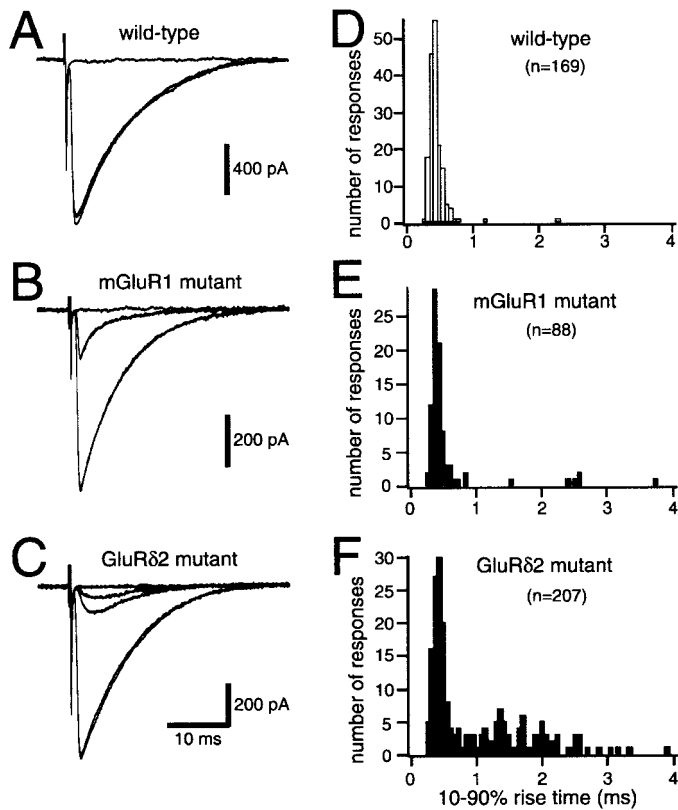
**Anterograde labeling of CFs.** Under deep anesthesia with chloral hydrate (350 mg/kg of body weight), a glass pipette (inner tip diameter 10–20  $\mu$ m) filled with 2–3  $\mu$ l of a 10% solution of biotinylated dextran amine (10,000 molecular weight; Molecular Probes) in 0.01 M PBS, pH 7.4, was inserted into the right inferior olive by dorsal approach (Rossi et al., 1995). Biotinylated dextran amine was injected iontophoretically by positive current (7  $\mu$ A for 30 min with a protocol of 700 msec on and 1300 msec off). After 6–8 d of survival, mice were anesthetized with chloral hydrate (350 mg/kg) and perfused transcardially with 4% paraformaldehyde in 0.1 M phosphate buffer, pH 7.4. Coronal brainstem sections were examined to check the injection site of biotinylated dextran amine. Mice with the injection site restricted to the caudal part of the medial accessory olive were chosen for the preparation of microslicer cerebellar sections (50  $\mu$ m in thickness). Parasagittal cerebellar sections were incubated overnight in avidin–peroxidase (Amersham Biosciences, Buckinghamshire, UK) diluted with PBS containing 1% Tween 20 and visualized with diaminobenzidine and cobalt. Low-power photographs were taken with a Normarski interference contrast microscope (Axio-photo, Zeiss), whereas high-power ones were taken with a bright-field microscope (AX-80, Olympus). To quantify the extent of arbors of labeled CFs, the relative vertical height of the most distal tip was measured for at least 30 CFs in each mouse and expressed as the percentage of the molecular layer thickness. In each genotype, the mean value and SEM were obtained from three mice, and the difference between the genotypes was tested by Student's *t* test.

## RESULTS

### A novel slow time course CF-EPSC in GluR $\delta$ 2 mutant PCs

In parasagittal cerebellar slices, PCs were recorded in the whole-cell configuration, and CFs were stimulated with a glass pipette placed in the granule cell layer near the recorded PCs (Aiba et al., 1994; Kano et al., 1995, 1997). To search PCs with multiple CF innervation, the stimulation pipette was systematically moved by  $\sim$ 20  $\mu$ m steps, and the stimulus intensity was gradually increased at each stimulation site (pulse width 0.1 msec; strength 0–100 V) (Kano et al., 1995, 1997, 1998; Offermanns et al., 1997; Watase et al., 1998; Ichise et al., 2000). EPSCs were elicited in an all-or-none fashion in most of wild-type PCs (Fig. 1A), whereas EPSCs with more than one discrete step were recorded in mGluR1 mutant PCs (Fig. 1B) with much higher incidence than in wild-type PCs (Kano et al., 1997; L  v  n  s et al., 1997; Ichise et al., 2000). These EPSCs displayed clear paired-pulse depression (PPD) to a stimulus pair with varying interpulse intervals (Fig. 2A,B, *wild-type*) (Konnerth et al., 1990; Aiba et al., 1994; Kano et al., 1995, 1997; Hashimoto and Kano, 1998).

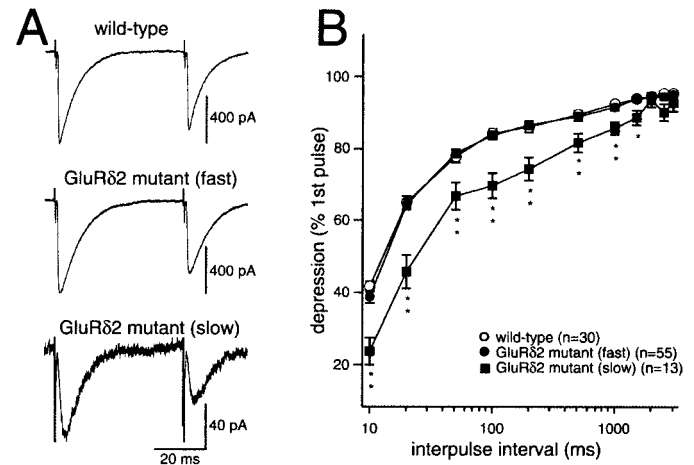
In the GluR $\delta$ 2 mutant mice, stimulation in the granule cell layer elicited multiple EPSCs in 88 of 122 PCs (Fig. 1C). Each of these multiple EPSCs displayed clear PPD to a similar or greater extent than that of wild-type CF-EPSCs (Fig. 2A,B, *GluR $\delta$ 2 mutant*). These multiple EPSCs in GluR $\delta$ 2 mutant PCs were judged to be elicited by stimulating CFs and not by activating mossy fiber–granule cell-PF circuits, because EPSCs elicited by PF stimulation display clear paired-pulse facilitation in GluR $\delta$ 2



**Figure 1.** Time course of CF-EPSCs in wild-type, mGluR1 mutant, and GluR $\delta$ 2 mutant PCs. *A–C*, EPSCs elicited by stimulation of CFs in the granule cell layer in a wild-type (*A*, P28), a mGluR1 mutant (*B*, P49), and a GluR $\delta$ 2 mutant (*C*, P64) PC. With gradually increasing stimulus intensities, EPSCs of the wild-type were elicited in an all-or-none fashion (*A*), whereas those of the mGluR1 mutant (*B*) and the GluR $\delta$ 2 mutant (*C*) occurred at multiple discrete steps, indicating that they were innervated by multiple CFs. Note that the rise time of the largest CF-EPSC of the GluR $\delta$ 2 mutant is similar to that of the wild-type or the mGluR1 mutant CF-EPSC, whereas the rise times of the smaller two CF-EPSCs of the GluR $\delta$ 2 mutant are significantly slower. One to three traces are superimposed at each threshold intensity. Stimuli were applied at 0.2 Hz. Holding potentials were  $-10$  mV for both the wild-type and GluR $\delta$ 2 mutant PCs and  $-20$  mV for the mGluR1 mutant PC to inactivate voltage-dependent conductances. *D–F*, Summary histograms showing the 10–90% rise time of CF-EPSCs in wild-type (*D*), mGluR1 mutant (*E*), and GluR $\delta$ 2 mutant (*F*) PCs. Data were obtained from mice at P23–P70.

mutant PCs as in wild-type PCs (Kashiwabuchi et al., 1995; Kurihara et al., 1997). These results indicate that most of the GluR $\delta$ 2 mutant PCs were innervated by multiple CFs.

In the multiple CF-EPSCs of the GluR $\delta$ 2 PC shown in Figure 1*C*, the time course was significantly slower for the smaller two EPSCs than for the largest EPSC. The 10–90% rise time of the wild-type CF-EPSCs displayed a normal distribution with a peak around 0.5 msec ( $0.46 \pm 0.11$  msec; mean  $\pm$  SD;  $n = 105$ ) (Fig. 1*D*). That of the mGluR1 mutant had a similar distribution ( $0.57 \pm 0.58$  msec; mean  $\pm$  SD;  $n = 88$ ) (Fig. 1*E*). In contrast, the rise time of GluR $\delta$ 2 mutant CF-EPSCs segregated into two distinct populations (Fig. 1*F*). The mean value for the fast ( $<1.0$  msec) population was 0.48 msec ( $\pm 0.15$  msec SD;  $n = 127$ ), which was similar to wild-type, whereas that for the slow ( $>1.0$  msec) population was 1.81 msec ( $\pm 0.59$  msec SD;  $n = 80$ ). These results suggest that GluR $\delta$ 2 mutants have two distinct populations of CF inputs in terms of the EPSC rise time. The CF-EPSCs with fast

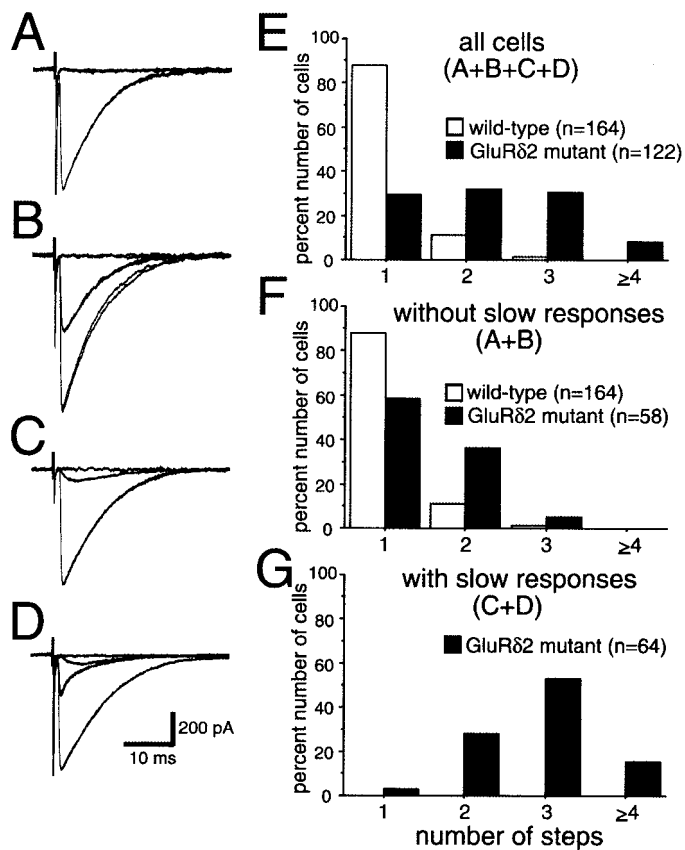


**Figure 2.** Paired-pulse depression of CF-EPSCs in the wild-type and GluR $\delta$ 2 mutant PCs. *A*, Single-sweep examples of paired-pulse depression (interpulse interval of 50 msec) of CF-EPSCs from a wild-type (*top panel*, P28) and a GluR $\delta$ 2 mutant (*middle and bottom panels*, P62) PC. The fast (*middle panel*) and slow (*bottom panel*) CF-EPSCs were elicited separately from the same GluR $\delta$ 2 mutant PC. Stimulus pairs were applied at 0.2 Hz. Holding potential was  $-10$  mV for all cells. *B*, Summary graphs showing paired-pulse depression for the wild-type ( $\circ$ ), the fast GluR $\delta$ 2 mutant ( $\bullet$ ), and the slow GluR $\delta$ 2 mutant ( $\blacksquare$ ) CF-EPSCs. The second response (expressed as a percentage of response to the first pulse; mean  $\pm$  SEM) is plotted as a function of interpulse interval. Stimulus pairs were applied at 0.2 Hz.  $**p < 0.01$ ;  $*p < 0.05$ , compared with the values of the wild-type CF-EPSCs (Student's *t* test).

rise time were similar to those of wild-type and mGluR1 mutant mice, whereas the slower CF-EPSCs appear to be unique to GluR $\delta$ 2 mutant PCs. We also measured the decay time constant of CF-EPSCs in the three strains of mice by fitting the CF-EPSC decay with single exponentials (Llano et al., 1991; Aiba et al., 1994; Kano et al., 1995, 1997). The decay time constants of CF-EPSCs in wild-type and GluR $\delta$ 2 mutant mice were not significantly different, being 6.73 msec ( $\pm 2.04$  msec;  $n = 84$ ) for wild-type and 6.48 msec ( $\pm 2.92$  msec;  $n = 174$ ) for the GluR $\delta$ 2 mutant. The decay time constants were 4.75 msec ( $\pm 2.36$  msec;  $n = 88$ ) for the mGluR1 mutant, which is significantly shorter than wild-type mice as reported previously (Aiba et al., 1994; Kano et al., 1997).

The GluR $\delta$ 2 mutant PCs could be classified into four categories in terms of the pattern of CF innervation. These are as follows: (1) PC innervated by a single CFs that generates typical fast CF-EPSCs (Fig. 3*A*); (2) PC multiple-innervated by more than one CF that generates fast CF-EPSCs (Fig. 3*B*); (3) PC multiple-innervated by one CF that generates fast CF-EPSCs and one or more CFs that generate slow CF-EPSCs (Fig. 3*C*); and (4) PC multiple-innervated by more than one CF that generates fast CF-EPSCs and one or more CFs that generate slow CF-EPSCs (Fig. 3*D*). Frequency distribution of all PCs in terms of number of CF-EPSC steps indicates that 72.1% of PCs in GluR $\delta$ 2 mutant mice were innervated by multiple CFs (Fig. 3*E*). The histogram constructed only from PCs without slow CF-EPSCs indicates that 36.2% of such PCs were innervated by two CFs and 5.2% by three CFs (Fig. 3*F*). On the other hand, the histogram constructed only from PCs with slow CF-EPSCs shows that most of the PCs were multiple-innervated (Fig. 3*G*). In particular, 68.7% are innervated by three or more CFs (Fig. 3*G*). These results indicate that the existence of the slow CF-EPSCs significantly contributes to





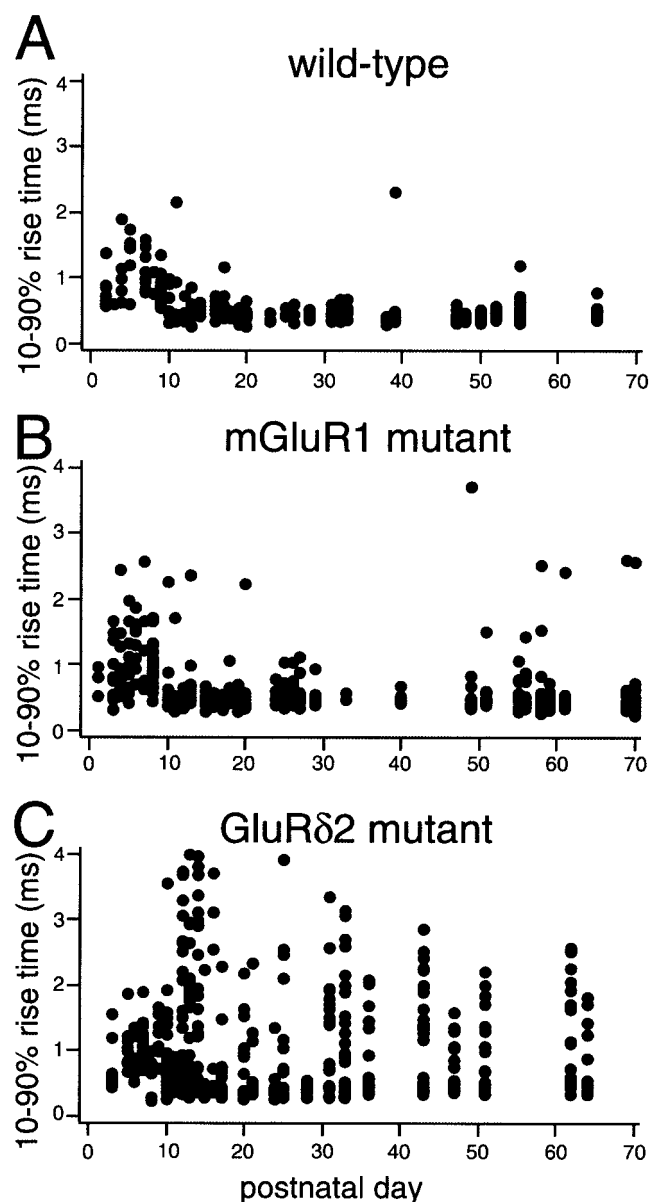
**Figure 3.** Four patterns of CF innervation in GluR $\delta$ 2 mutant PCs. *A–D*, Sample records of GluR $\delta$ 2 mutant CF-EPSCs representing four distinct CF innervation patterns. *A*, Single fast CF-EPSC; *B*, multiple fast CF-EPSCs; *C*, single fast plus one or more slow CF-EPSCs; *D*, multiple fast plus one or more slow CF-EPSCs. One to three traces are superimposed at each threshold intensity. Stimuli were applied at 0.2 Hz. Holding potentials were  $-10$  mV for *A*, *C*, and *D* and  $-20$  mV for *B*. *E–G*, Summary histograms showing the number of discrete steps of CF-EPSCs for all PCs (*E*), for PCs without slow EPSCs (*F*), and for PCs with slow EPSCs (*G*). *Open* and *filled columns* represent the wild-type and GluR $\delta$ 2 mutant PCs, respectively. Data were obtained from mice at P24–P64.

the high percentage of multiple-innervated PCs in the GluR $\delta$ 2 mutant mice.

#### Slow CF-EPSCs of GluR $\delta$ 2 mutant PCs appear during the second postnatal week

In rodents, PF synaptogenesis onto PC dendrites is known to continue postnatally up to approximately postnatal day (P) 20 (Altman and Bayer, 1997). The structure of the PF–PC synapse is immature around P7, when no significant differences in the morphology and electrophysiology are found between the wild-type and GluR $\delta$ 2 mutant mice (Kurihara et al., 1997). Impaired PF synapse formation is obvious in the GluR $\delta$ 2 mutant mouse at P14 (Kurihara et al., 1997). In contrast, PF–PC synapse formation appears normal in mGluR1 mutant mice (Kano et al., 1997). We followed the developmental course of CF-EPSCs in the three strains of mice to examine whether the impaired PF synapse formation correlated with the appearance of atypical slow CF-EPSCs in GluR $\delta$ 2 mutant mice.

The 10–90% rise time of the wild-type CF-EPSC was longer during P3–P9 than at later developmental stages (Fig. 4*A*) ( $1.0 \pm 0.4$  msec; mean  $\pm$  SD;  $n = 43$ ). The rise times of the mGluR1 mutant and GluR $\delta$ 2 mutant CF-EPSCs from P3 to P9 had a

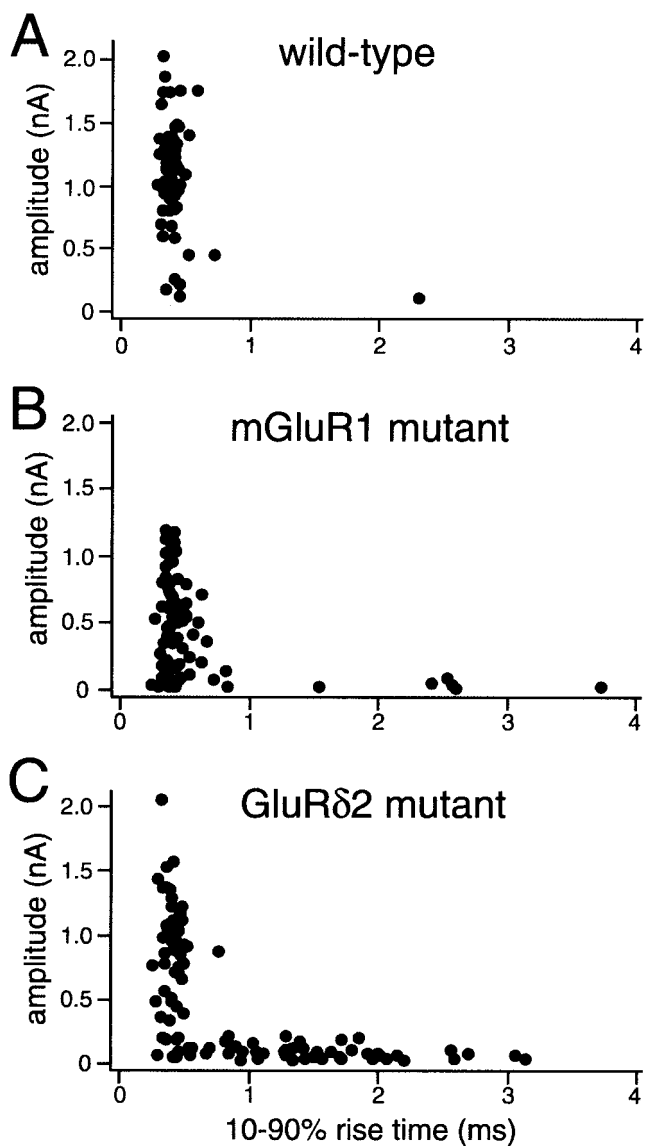


**Figure 4.** Postnatal change of CF-EPSC time course in the three strains of mice. The 10–90% rise times of CF-EPSCs are plotted against postnatal day for wild-type (*A*), mGluR1 mutant (*B*), and GluR $\delta$ 2 mutant (*C*) PCs. Note that CF-EPSCs have similar rise times in the three strains of mice during P3–P9, whereas those with significantly slower rise times are obvious after P10 in the GluR $\delta$ 2 mutant.

similar distribution ( $1.0 \pm 0.5$  msec,  $n = 71$  for the mGluR1 mutant;  $0.9 \pm 0.4$  msec,  $n = 64$  for the GluR $\delta$ 2 mutant) (Fig. 4*B,C*) to that of wild-type. During P10–P14, the rise times of the wild-type and mGluR1 mutant CF-EPSCs decreased to adult levels (Fig. 4*A,B*). In contrast, CF-EPSCs with significantly slower rise times appeared during P10–P14 in the GluR $\delta$ 2 mutant (Fig. 4*C*). These data indicate that atypical slow CF-EPSCs become evident during the second postnatal week when a defect in the PF to PC synapse formation occurs in the GluR $\delta$ 2 mutant mouse.

#### Electrophysiological properties of slow CF-EPSC in GluR $\delta$ 2 mutant PCs

To examine whether the CF-EPSC amplitudes of mGluR1 mutant and GluR $\delta$ 2 mutant mice differ from those of wild-type mice, the



**Figure 5.** Two distinct populations of CF-EPSCs in the GluR $\delta$ 2 mutant mice. The amplitudes of CF-EPSCs (measured at a holding potential of  $-10$  mV) are plotted against the 10–90% rise time for wild-type (*A*), mGluR1 mutant (*B*), and GluR $\delta$ 2 mutant (*C*) mice. Note that two distinct populations of CF-EPSCs, those with a fast rise time and relatively large amplitude and those with a slow rise time and small amplitude, were evident in *C*.

peak amplitudes of CF-EPSCs were measured at a fixed holding potential ( $-10$  mV). In cases of multiple-innervated PCs (Fig. 1*B,C*), the amplitude of a given CF-EPSC was measured by digitally subtracting other CF-EPSCs that were induced by lower stimulus strength. The peak CF-EPSC amplitude was plotted against the 10–90% rise time. CF-EPSCs in wild-type (Fig. 5*A*) and mGluR1 mutant (Fig. 5*B*) mice were in a single population with only a few CF-EPSCs with slow rise time and small amplitude. In contrast, two distinct populations of CF-EPSCs, those with fast rise time and relatively large amplitude and those with slow rise time and small amplitude, were present in GluR $\delta$ 2 mutant mice (Fig. 5*C*).

We examined other electrophysiological and pharmacological properties of atypical slow CF-EPSCs of GluR $\delta$ 2 mutant PCs. They displayed a linear current–voltage ( $I$ – $V$ ) relationship (data

**Table 1. Passive properties of PCs**

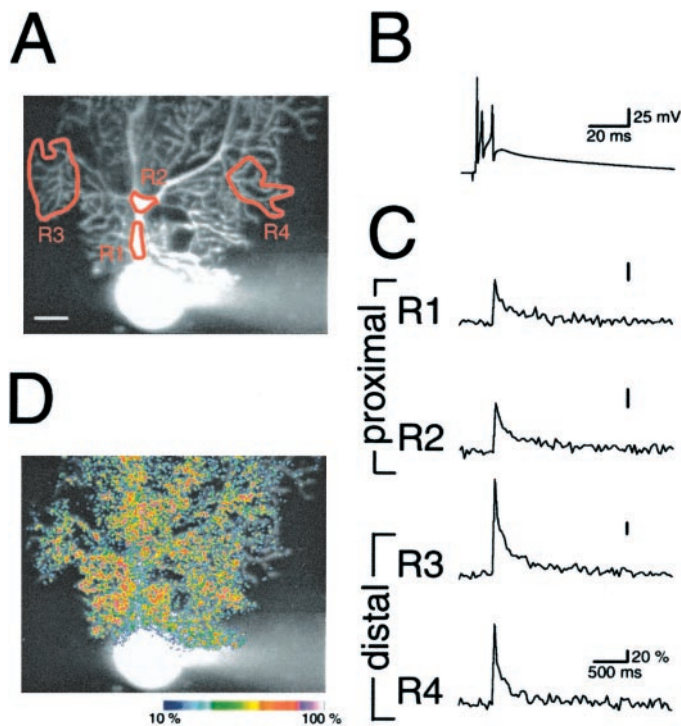
	Wild-type ( $n = 9$ )	GluR $\delta$ 2 mutant ( $n = 11$ )
C1 (pF)	$73.8 \pm 12.9$	$67.2 \pm 20.3$
C2 (pF)	$684 \pm 101^{**}$	$513 \pm 75.2^{**}$
R1 (M $\Omega$ )	$9.3 \pm 1.3$	$7.9 \pm 2.4$
R2 (M $\Omega$ )	$10.6 \pm 2.7^*$	$14.9 \pm 4.6^*$
R3 (M $\Omega$ )	$145.7 \pm 41.5^{**}$	$212.8 \pm 47.1^{**}$

Parameters were calculated on the basis of the model described by Llano et al. (1991), which distinguishes two regions of PCs: region 1 representing the soma and the main proximal dendrites, and region 2 representing the dendritic tree. C1 represents the lumped membrane capacitance of region 1, and R1 represents the pipette access resistance. Region 2 is linked to region 1 by resistor R2, which represents the lumped resistance between the main proximal dendrite and each membrane region of the distal dendrites. C2 and R3 represent the lumped capacitance and resistance, respectively, of the dendritic tree of PCs. Data are expressed as mean  $\pm$  SD.  $^{**}p < 0.01$ ;  $^*p < 0.05$  (two-tailed  $t$  test).

not shown). In  $Mg^{2+}$ -free saline containing glycine ( $10 \mu M$ ), atypical slow CF-EPSCs were not affected by an NMDA receptor antagonist, DL-2-amino-5-phosphonopentanoate ( $50 \mu M$ ) but were totally suppressed by a non-NMDA antagonist, 6-cyano-7-nitroquinoxaline-2,3-dione ( $10 \mu M$ ) (data not shown). These results indicate that atypical CF-EPSCs of GluR $\delta$ 2 mutant PCs are mediated exclusively by non-NMDA receptors, which is similar to typical CF-EPSCs of wild-type and those of GluR $\delta$ 2 PCs (Kashiwabuchi et al., 1995).

#### Passive membrane properties of GluR $\delta$ 2 mutant PCs

It has been shown morphologically that CF synapses are located on the proximal dendrites of PCs, whereas PF synapses oppose spines at the distal dendrites (Palay and Chan-Palay, 1974; Ito, 1984; Strata and Rossi, 1998). The 10–90% rise times of PF-EPSCs are significantly slower than those of CF-EPSCs in both the rat (Llano et al., 1991) and mouse (Aiba et al., 1994; Kano et al., 1995, 1997), suggesting that the EPSC rise time in PCs reflects the electrotonic length from the soma to the site of the synapse. It is likely, therefore, that the atypical slow CF-EPSCs in GluR $\delta$ 2 mutant PCs arise from synapses located electrotonically distant from the soma, presumably on the distal dendrites. Because the EPSC rise time depends on the passive membrane properties, we compared several parameters of wild-type and GluR $\delta$ 2 mutant PCs in response to hyperpolarizing voltage steps from  $-70$  to  $-80$  mV. As reported previously in rat PCs (Llano et al., 1991), the decay of the current was biphasic and could be described by the sum of two exponentials in both wild-type and GluR $\delta$ 2 mutant PCs (data not shown). The time constants for the fast ( $\tau_1$ ) and slow ( $\tau_2$ ) components were  $0.35 \pm 0.04$  and  $13.4 \pm 1.9$  msec (mean  $\pm$  SD;  $n = 9$ ) for the wild-type and  $0.33 \pm 0.11$  and  $11.6 \pm 2.0$  msec ( $n = 11$ ) for the GluR $\delta$ 2 mutant. Both  $\tau_1$  and  $\tau_2$  values were not significantly different between the wild-type and the GluR $\delta$ 2 mutant ( $p > 0.05$ ; Student's  $t$  test). We then calculated several parameters representing passive properties of PCs based on the model equivalent circuit of PCs described by Llano et al. (1991) (Table 1). As shown in Table 1, the lumped dendritic capacitance (C2) was smaller ( $p < 0.01$ ; Student's  $t$  test) and lumped resistances were larger ( $p < 0.05$  for R2;  $p < 0.01$  for R3; Student's  $t$  test) in the GluR $\delta$ 2 mutant than in the wild-type mice. These results suggest that the average total membrane area of the dendritic tree of the GluR $\delta$ 2 mutant PCs is smaller than that of wild-type PCs. This is consistent with morphological data that the molecular layer of the GluR $\delta$ 2 mutant is thinner than that of



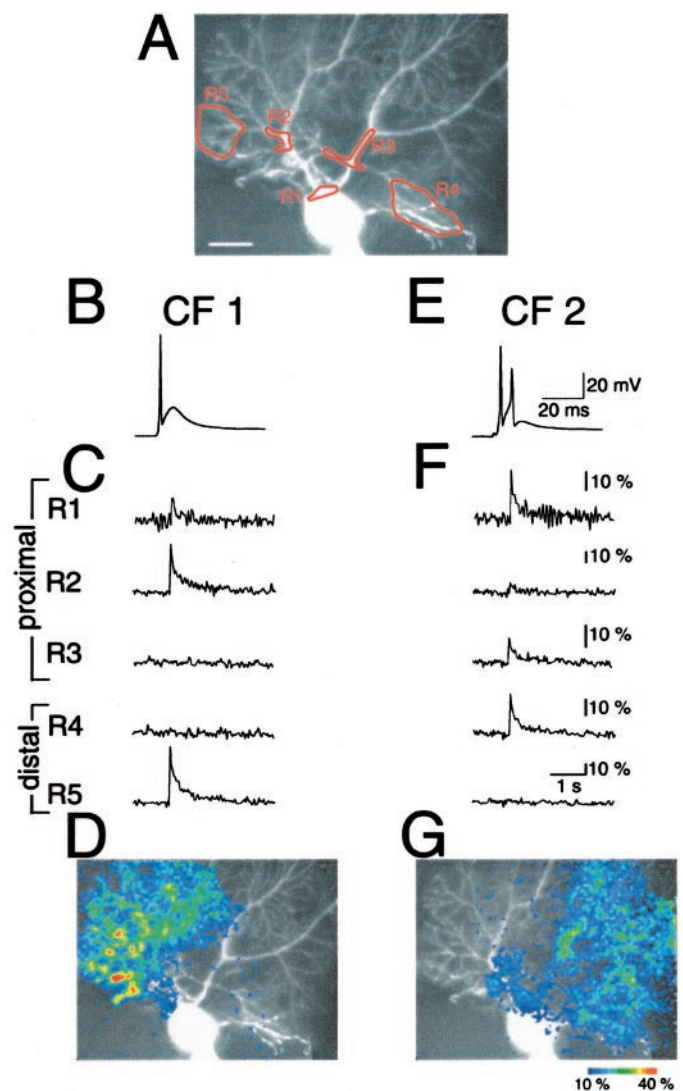
**Figure 6.** CF-induced  $\text{Ca}^{2+}$  transients in a mono-innervated wild-type PC. *A*, Reconstructed image of a wild-type PC. R1–R4 indicate the dendritic regions from which the  $\text{Ca}^{2+}$  signals shown in *C* are measured. Scale bar, 20  $\mu\text{m}$ . *B*, Complex spike induced by CF stimulation under the current-clamp recording mode. *C*, Time course of CF-induced  $\text{Ca}^{2+}$  signals measured in the regions indicated in *A*. *D*, Pseudocolor image showing the relative increase in  $\text{Ca}^{2+}$ -dependent fluorescence ( $\Delta F/F_0$ ) evoked by CF stimulation. For this and the following figures (Figs. 7–9), the  $\Delta F/F_0$  values for pseudocolor images were calculated from the pre-stimulus and poststimulus fluorescent values that were obtained by averaging five consecutive images (132 msec periods) just before and after CF stimulation.

the wild-type cerebellum (Kurihara et al., 1997). These results indicate that the slow rise times of a subpopulation of GluR $\delta$ 2 mutant CF-EPSCs cannot be explained by the altered passive properties of the PC dendrites.

#### Imaging CF-induced $\text{Ca}^{2+}$ transients suggests that slow CF-EPSCs of GluR $\delta$ 2 mutant PCs arise from synapses located at distal dendrites

We measured  $\text{Ca}^{2+}$  signals associated with CF responses by using a high-speed confocal laser-scanning microscope or a cooled CCD camera system. Whole-cell recording from PCs was conducted simultaneously with the  $\text{Ca}^{2+}$  imaging. The  $\text{Ca}^{2+}$  signals were measured in small regions at three different levels of PC dendritic arborization: the primary smooth branch, the tertiary smooth branch, and terminal branchlets. In the following figures, the primary and tertiary smooth branches are collectively termed “proximal dendrites,” and the terminal branchlets are termed “distal dendrites.” In wild-type animals, it is shown morphologically that CFs form extensive synaptic contacts onto proximal dendrites, whereas PFs form *en passant* synapses onto spines of distal dendrites (Palay and Chan-Palay, 1974; Ito, 1984; Strata and Rossi, 1998).

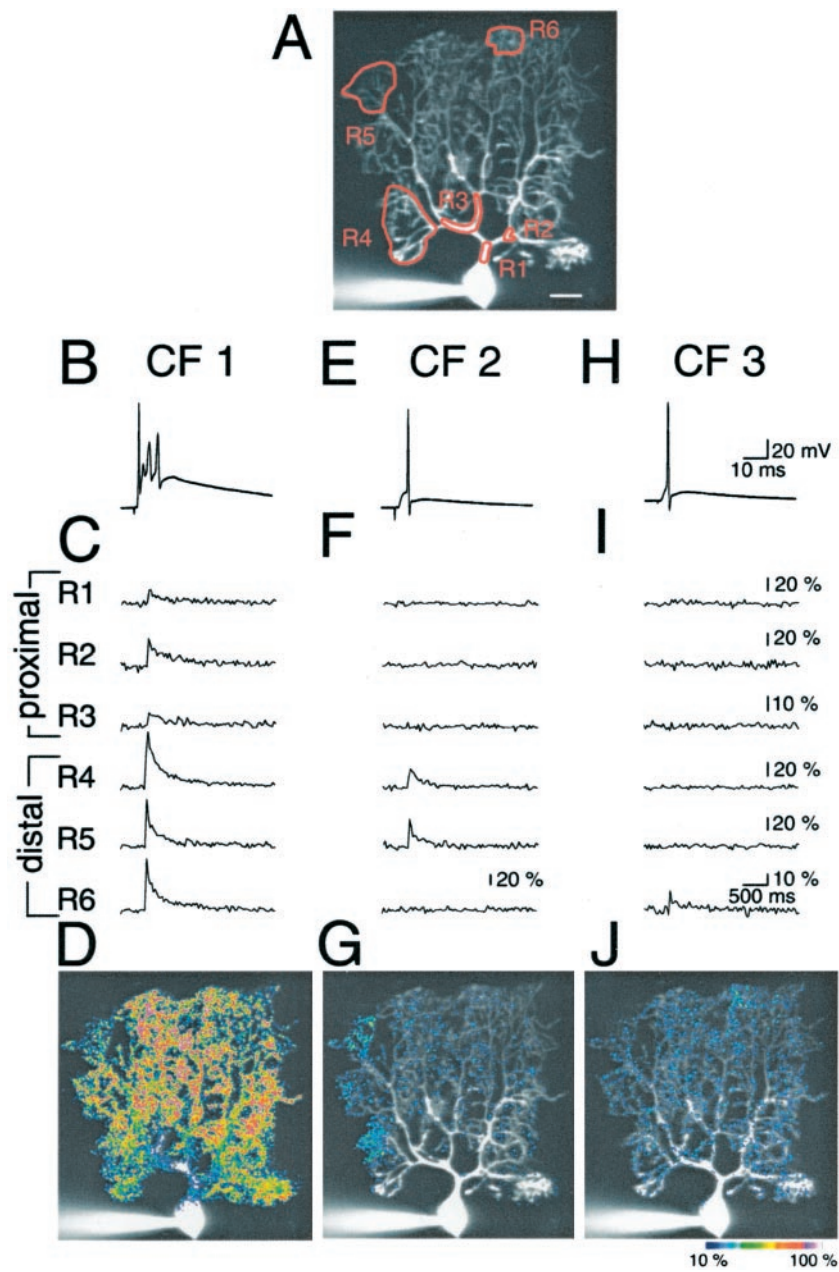
In the mono-innervated wild-type PC (Fig. 6*A*), stimulation of the CF induced a typical complex spike (Fig. 6*B*) that accompanied a clear  $\text{Ca}^{2+}$  signal over the entire dendritic tree (Fig. 6*C,D*).



**Figure 7.** CF-induced  $\text{Ca}^{2+}$  transients in a double-innervated mGluR1 mutant PC. *A*, Reconstructed image of a mGluR1 mutant PC. R1–R5 indicate the dendritic regions from which the  $\text{Ca}^{2+}$  signals shown in *C* and *F* are measured. Scale bar, 20  $\mu\text{m}$ . *B*, *E*, Responses induced by stimulating two different CFs, CF1 (*B*) and CF2 (*E*), under the current-clamp recording mode. *C*, *F*, Time course of  $\text{Ca}^{2+}$  signals measured in the regions indicated in *A* in response to stimulation of CF1 (*C*) and CF2 (*F*). *D*, *G*, Pseudocolor images showing the relative increase in  $\text{Ca}^{2+}$ -dependent fluorescence ( $\Delta F/F_0$ ) evoked by CF1 (*D*) and CF2 (*G*) stimulation.

The  $\text{Ca}^{2+}$  rose in both proximal (*R1*, *R2*) and distal (*R3*, *R4*) dendrites (Fig. 6*C,D*). In the double-innervated mGluR1 mutant PC (Fig. 7*A*), stimulation of each CF induced a large EPSP with spike(s) (Fig. 7*B,E*) and induced a clear  $\text{Ca}^{2+}$  transient (Fig. 7*C,F*). Stimulation of one CF (*CF1*) caused elevation of  $\text{Ca}^{2+}$  that was confined to the left dendritic tree (Fig. 7*C,D*), whereas stimulation of the other CF (*CF2*) caused  $\text{Ca}^{2+}$  transients in the right dendritic tree (Fig. 7*F,G*). It should be noted that  $\text{Ca}^{2+}$  was elevated in both proximal and distal portions of each dendritic tree. In 14 multiple-innervated mGluR1 mutant PCs, CF-induced  $\text{Ca}^{2+}$  signals always involved proximal dendrites. In 16 mono-innervated PCs from mGluR1 mutant mice, the pattern of CF-induced  $\text{Ca}^{2+}$  signals was identical to that of the mono-innervated wild-type PCs (data not shown). These results suggest



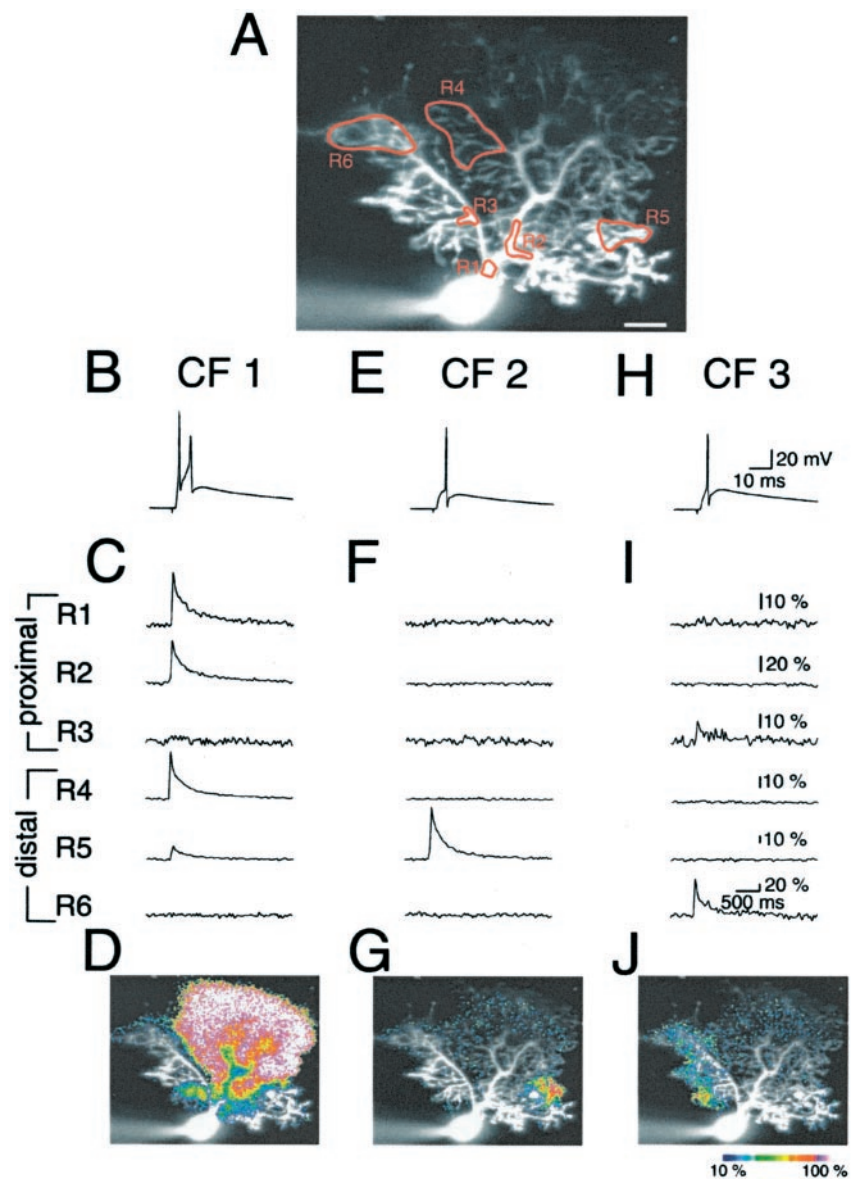


**Figure 8.** CF-induced  $\text{Ca}^{2+}$  transients in a multiple-innervated GluR $\delta$ 2 mutant PC. *A*, Reconstructed image of a GluR $\delta$ 2 mutant PC. R1–R6 indicate the dendritic regions from which the  $\text{Ca}^{2+}$  signals shown in *C*, *F*, and *I* are measured. Scale bar, 20  $\mu\text{m}$ . *B*, *E*, *H*, Responses induced by stimulating three different CFs, CF1 (*B*), CF2 (*E*), and CF3 (*H*), under the current-clamp recording mode. *C*, *F*, *I*, Time course of  $\text{Ca}^{2+}$  signals measured in the regions indicated in *A* in response to stimulation of CF1 (*C*), CF2 (*F*), or CF3 (*I*). *D*, *G*, *J*, Pseudocolor images showing the relative increase in  $\text{Ca}^{2+}$ -dependent fluorescence ( $\Delta F/F_0$ ) evoked by CF1 (*D*), CF2 (*G*), or CF3 (*J*) stimulation.

that mGluR1 mutant PCs are innervated by CFs at proximal dendrites as wild-type PCs whether they are mono- or multiple-innervated. The  $\text{Ca}^{2+}$  signals in the distal dendrite presumably reflected the spread of depolarization and activation of voltage-gated  $\text{Ca}^{2+}$  channels.

In GluR $\delta$ 2 mutant PCs, we detected different pattern of CF-induced  $\text{Ca}^{2+}$  signals from those seen in wild-type or mGluR1 mutant PCs. The GluR $\delta$ 2 mutant PC in Figure 8 was innervated by at least three distinct CFs. Stimulation of one CF (CF1) generated typical complex spike (Fig. 8*B*), whereas stimulation of other two CFs (CF2 and CF3) elicited slow responses (Fig. 8*E*,*H*). Stimulation of CF1 induced  $\text{Ca}^{2+}$  transients that spread over the entire dendritic tree involving both proximal and distal dendrites (Fig. 8*C*,*D*, R1–R6). In contrast, stimulation of CF2 induced  $\text{Ca}^{2+}$  transients that were confined to the distal portions (R4, R5) of the left dendritic branch (Fig. 8*F*,*G*). Stimulation of CF3 induced small  $\text{Ca}^{2+}$  transients that were confined to the

most distal dendrites (R6) near the pial surface (Fig. 8*I*,*J*). Another GluR $\delta$ 2 mutant PC shown in Figure 9 was innervated by at least three distinct CFs that generated a fast response (Fig. 9*B*, CF1) and slow responses (Fig. 9*E*,*H*, CF2 and CF3). Stimulation of CF1 induced  $\text{Ca}^{2+}$  transients that spread over the middle dendritic tree and spared the small trees in the left and the right branches (Fig. 9*C*,*D*). The  $\text{Ca}^{2+}$  rose in both proximal (R1, R2) and distal (R4, R5) portions of the middle dendritic tree (Fig. 9*C*,*D*). Stimulation of CF2 induced  $\text{Ca}^{2+}$  transients that were confined to the distal portions (R5) of the right dendritic tree and spared its proximal (R1, R2) portions (Fig. 9*F*,*G*). Stimulation of CF3 induced  $\text{Ca}^{2+}$  transients that were confined to the left dendritic branch (Fig. 9*I*,*J*). The  $\text{Ca}^{2+}$  rose slightly in the tertiary dendrite (R3) and clearly in distal dendrites (R6) but showed no change in the primary dendrite (R1). In 24 GluR $\delta$ 2 mutant PCs, we found that slow CF responses were always associated with  $\text{Ca}^{2+}$  transients that were confined to the distal dendrites. In a



**Figure 9.** CF-induced  $\text{Ca}^{2+}$  transients in another multiple-innervated GluR $\delta$ 2 mutant PC. Reconstructed image of the PC (*A*), responses to CF stimulation (*B*, *E*, *H*), time course of  $\text{Ca}^{2+}$  signals (*C*, *F*, *I*), and pseudocolor images (*D*, *G*, *J*) are illustrated in a manner similar to Figure 8. Scale bar, 20  $\mu\text{m}$ .

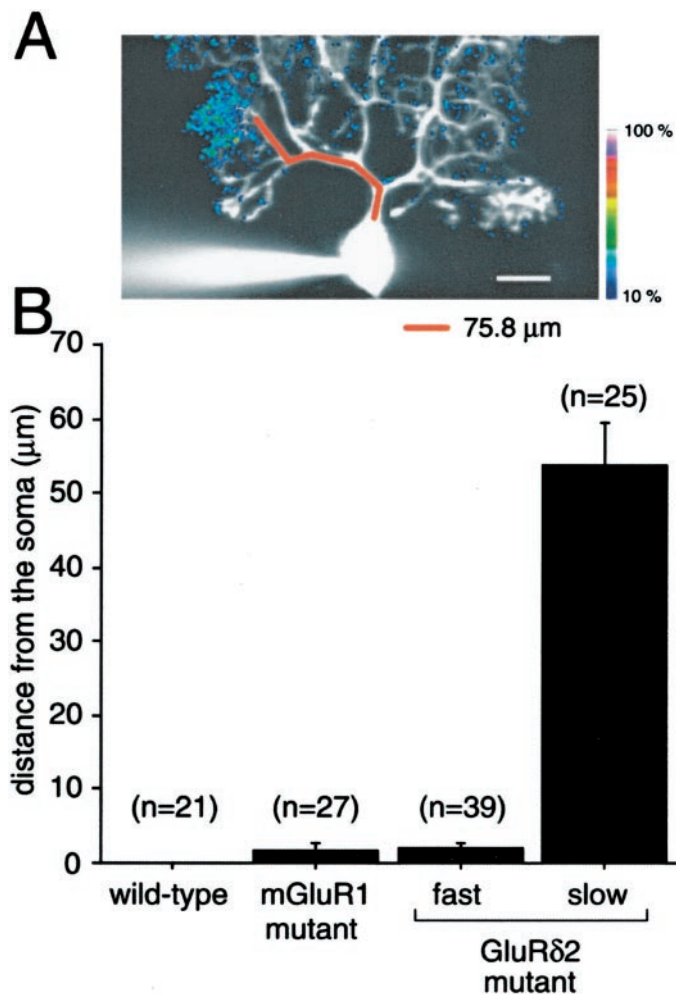
few slow CF responses, small  $\text{Ca}^{2+}$  signals could be detected also in the tertiary dendrites (Fig. 9, *CF3*); however, the  $\text{Ca}^{2+}$  transients never involved the primary dendrites. This pattern of CF-induced  $\text{Ca}^{2+}$  transients was observed in no PCs from the wild-type and in only one PC from the mGluR1 mutant.

To quantify the differences in CF innervation patterns, the distance from the soma to the dendrites with detectable  $\text{Ca}^{2+}$  signals was measured along the main dendrites (Fig. 10). As exemplified in Figure 10*A*, the actual dendritic branches were traced, and the shortest distance was measured from the origin of the primary dendrite to the most proximal point of the dendritic tree where the  $\text{Ca}^{2+}$  signal was detected. In CF responses of wild-type and mGluR1 mutant mice, the distance was almost 0  $\mu\text{m}$  (Fig. 10*B*), indicating that the  $\text{Ca}^{2+}$  rose in the primary dendrite. In GluR $\delta$ 2 mutant mice, the length was almost 0  $\mu\text{m}$  for the fast response, whereas it was  $\sim 50$   $\mu\text{m}$  on an average for the slow response (Fig. 10*B*). These results suggest that atypical slow CF responses of GluR $\delta$ 2 PCs arise from synapses located on the distal dendrites.

### Morphology of CFs from GluR $\delta$ 2- and mGluR1-deficient mice

To investigate the CF innervation patterns morphologically, anterogradely labeled CFs were visualized in parasagittal cerebellar sections (Fig. 11). Genotypic differences were noted in the extent of CF arbors within the molecular layer. Labeled CFs of GluR $\delta$ 2 mutant mice extended up to more superficial regions of the molecular layer than those of wild-type and mGluR1 mutant mice (Fig. 11*A–C*). Because the thickness of the molecular layer is slightly reduced in GluR $\delta$ 2 mutant mice (Kurihara et al., 1997), we evaluated the extent of CF arbors by measuring the height of the most distal tip of each labeled CF relative to the thickness of the molecular layer. We selected cerebellar sections from each mouse that contained staining of entire CF arbors from the Purkinje cell layer to the top of the molecular layer. Then, we examined at least 30 labeled CFs in the straight portion of the lobule VI. The most distal tip of CFs in GluR $\delta$ 2 mutant mice reached  $95.0 \pm 0.1\%$  (mean  $\pm$  SEM;  $n = 3$ ) of the molecular layer thickness. This score was significantly higher than that of the





**Figure 10.** Distance from the soma to the dendrites with detectable  $\text{Ca}^{2+}$  signals. *A*, Pseudocolor image from the GluR $\delta$ 2 mutant PC shown in Figure 9 exemplifying how the distance was measured. The actual dendritic branches were traced (thick red line), and the shortest distance was measured from the origin of the primary dendrite to the most proximal point of the dendritic tree where the  $\text{Ca}^{2+}$  signal was detected. The distance was 75.8  $\mu\text{m}$  in this case. Scale bar, 20  $\mu\text{m}$ . *B*, Summary histogram of the distance from the soma for the three strains of mice.

wild-type ( $83.1 \pm 0.5\%$ ;  $n = 3$ ;  $p < 0.0001$ ) or the mGluR1 mutant ( $77.8 \pm 0.7\%$ ;  $n = 3$ ;  $p < 0.0001$ ) mouse. Our previous study indicates that PC dendrites extend to the surface of the molecular layer in GluR $\delta$ 2 mutant mice as well as wild-type mice (Kurihara et al., 1997). Taken together, these results suggest that CFs of GluR $\delta$ 2 mutant mice innervate more distal portions of PC dendrites than those of wild-type and mGluR1 mutant mice.

## DISCUSSION

### Distinct CF innervation patterns in GluR $\delta$ 2 mutant and mGluR1 mutant PCs

GluR $\delta$ 2 mutant PCs had atypical CF-EPSCs with slow 10–90% rise times that coexisted with typical fast rise time CF-EPSCs, suggesting that slow CF-EPSCs arise from synapses located electrotonically distant from the soma. Imaging of CF-induced  $\text{Ca}^{2+}$  signals through voltage-gated  $\text{Ca}^{2+}$  channels revealed that GluR $\delta$ 2 mutant PCs had atypical small CF responses with  $\text{Ca}^{2+}$  signals that were confined to PC distal dendrites. In contrast, CF-induced  $\text{Ca}^{2+}$  signals involved both proximal and distal den-

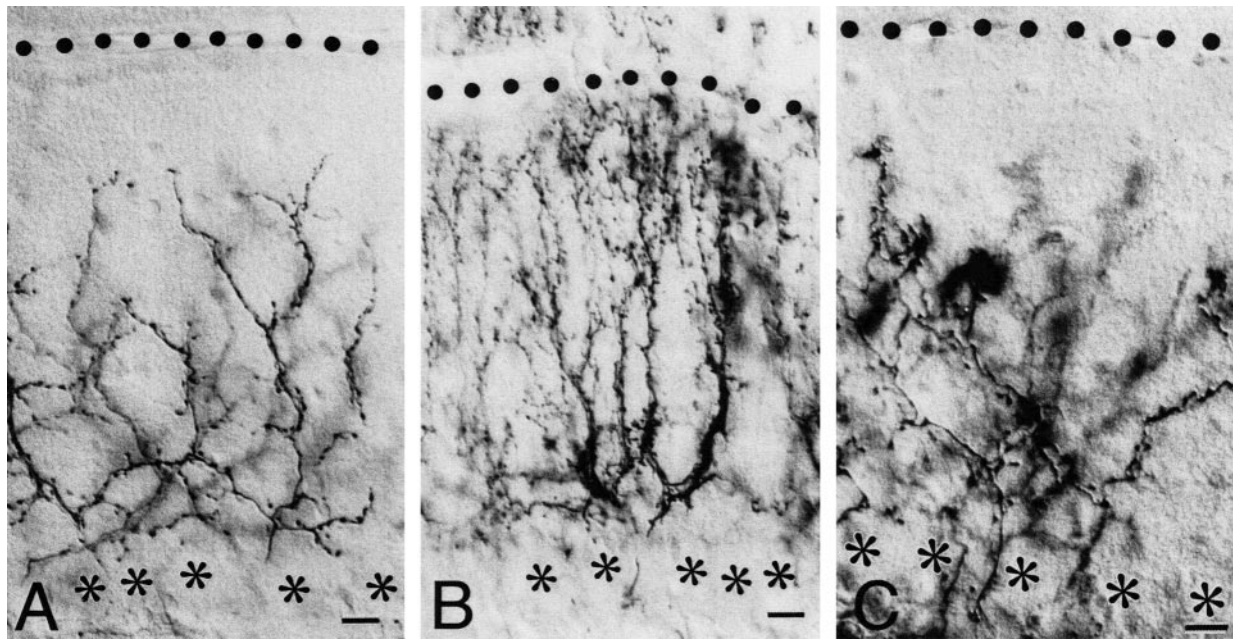
drites in wild-type PCs. Morphologically, CFs of GluR $\delta$ 2 mutant mice extended to the superficial regions of the molecular layer, whereas CFs of wild-type mice extended to the four-fifths of the molecular layer and spared the external one-fifth. On the other hand, the CF innervation pattern of mGluR1 mutant mice resembled that of the wild-type mice in the following respects: (1) CF-EPSCs had fast rise times ( $\leq 1$  msec), (2) CF-induced  $\text{Ca}^{2+}$  signals involved proximal dendrites, and (3) anterogradely labeled CFs extended to the four-fifths of the molecular layer and spared the external one-fifth.

Previous electron microscopic studies in mouse and rat indicate that CFs form extensive synaptic contacts on protuberance of the proximal dendrites of PCs, whereas PFs form *en passant* synapses on spines of the distal dendrites of PCs (Larramendi and Victor, 1967; Palay and Chan-Palay, 1974; Ito, 1984). Staining of CFs in rats also revealed that CFs innervate PC proximal dendrites (Strata and Rossi, 1998; Sugihara et al., 1999; Kreitzer et al., 2000). The range of CF-induced  $\text{Ca}^{2+}$  signals, however, does not precisely reflect the location of CF synapses on PC dendritic trees. Previous studies consistently indicate that CF-induced  $\text{Ca}^{2+}$  transients spread over the entire dendritic tree of PCs involving both proximal and distal dendrites (Ross and Werman, 1987; Kano et al., 1992; Konnerth et al., 1992; Miyakawa et al., 1992; Eilers et al., 1995b). The  $\text{Ca}^{2+}$  transients in distal dendrites are thought to reflect the spread of depolarization from proximal dendrites. CF-EPSPs are generated at proximal dendrites of PCs. However, because they are quite large in size, they can readily activate voltage-gated  $\text{Ca}^{2+}$  channels not only at proximal dendrites but also at distal dendrites. In contrast, the  $\text{Ca}^{2+}$  transients induced by PF stimulation are confined to the distal dendrites, and the range of the  $\text{Ca}^{2+}$  signals reflects the site of PF synapses (Eilers et al., 1995a; Finch and Augustine, 1998; Takechi et al., 1998; Wang et al., 2000). This is presumably because PF-EPSPs are relatively small in size and can only activate voltage-gated  $\text{Ca}^{2+}$  channels locally at the site of PF-EPSP generation. The atypical CF responses in GluR $\delta$ 2 mutant mice were also relatively small in size and presumed to induce  $\text{Ca}^{2+}$  signals that reflect the site of their generation. In these respects, the atypical CF responses resemble PF-mediated responses and presumably reflect synaptic activity at PC distal dendrites.

Taken together, the results of the present study suggest that surplus CFs persist in GluR $\delta$ 2 mutant mice by forming ectopic synapses onto distal dendrites of PCs, whereas CFs of mGluR1 mutant mice form synapses onto PC proximal dendrites similar to wild-type mice.

### Roles of GluR $\delta$ 2 in PF and CF synapse formation

Our previous study in the GluR $\delta$ 2 mutant indicates that the abnormality of PF synaptogenesis is not found at P7 but is evident at P14 and thereafter (Kurihara et al., 1997). Thus, the appearance of atypical slow CF-EPSCs coincides temporally with the manifestation of impaired PF synaptogenesis. Previous studies indicate that the developmental change in CF from multiple- to mono-innervation of cerebellar PCs depends on the presence of intact granule cells and the formation of PF to PC synapses (Woodward et al., 1974; Crépel and Mariani, 1976; Mariani et al., 1977; Crépel and Delhaye-Bouchaud, 1979; Crépel et al., 1980; Mariani and Changeux, 1980; Mariani et al., 1990; Bravin et al., 1995; Sugihara et al., 2000). Thus, impairment of PF to PC synapse formation in GluR $\delta$ 2 mutant mice (Kashiwabuchi et al., 1995; Kurihara et al., 1997) can cause persistent multiple CF innervation. In wild-type mice, massive formation of PF to PC



**Figure 11.** Anterogradely labeled CFs in the cerebellar lobule VI of the wild-type (*A*), GluR $\delta$ 2 mutant (*B*), and mGluR1 mutant (*C*) mice. Dotted lines in *A–C* indicate the pial surface of the molecular layer. Asterisks indicate PC somata. Scale bars, 10  $\mu$ m.

synapses occurs onto dendritic spines of PC distal dendrites during the second and third postnatal weeks. CF synapses may not be maintained at distal dendrites and become confined to the thick proximal dendrites. Then, a competition may follow between the remaining CFs for postsynaptic sites on the proximal dendrites, and finally, a single CF may survive to innervate the PC. In GluR $\delta$ 2 mutant mice, CF synapses may remain at distal dendrites because of the impairment of PF to PC synapse formation during the second and third postnatal weeks. However, the competition may occur between CFs for postsynaptic sites on the proximal dendrites, and in the majority of PCs, one CF may win to form extensive synapses on the proximal dendrites. As a consequence, many PCs lacking GluR $\delta$ 2 have one typical CF that generated fast CF-EPSCs and additional atypical CFs that generated slow CF-EPSCs.

A previous study indicates that the  $\delta$  receptor immunoreactivity is prominent at CF synapses at P10 and P14 and then becomes weaker at later developmental stages (Zhao et al., 1998). On the other hand,  $\delta$  receptors are abundant on postsynaptic membranes at PF synapses from P10 to adult (Takayama et al., 1995, 1996; Landsend et al., 1997; Zhao et al., 1998). Thus, the developmental stage at which the impaired PF synapse formation becomes obvious in GluR $\delta$ 2 mutant mice corresponds well with the period during which the  $\delta$  receptors are transiently expressed at CF synapses. These results suggest that GluR $\delta$ 2 may also play active roles in CF synapse formation during P10–P14 in addition to stabilizing PF synapses.

It is reported that the spontaneously occurring ataxic mouse mutation *lurcher* is a gain-of-function mutation in the GluR $\delta$ 2 gene (Zuo et al., 1997). Mutated GluR $\delta$ 2 channels behave as depolarizing leak channels independent of glutamate. Consequently, PCs display a very high membrane conductance and are constitutively depolarized and, perhaps, permit continuous  $Ca^{2+}$  inflow. This appears to disturb various signal transduction cascades within PCs, including the pathway from mGluR1 to the  $\gamma$  isoform of protein kinase C (PKC $\gamma$ ) that is shown to be required

for CF synapse elimination (Kano et al., 1995, 1997, 1998; Offermanns et al., 1997; Hashimoto et al., 2000). This may be a cause of persistent multiple CF innervation in *lurcher* mutant mice (Rabacchi et al., 1992). We studied the kinetics of CF-EPSCs in *lurcher* mice, but atypical slow CF-EPSCs abundant in GluR $\delta$ 2 mutant mice were not found (K. Hashimoto and M. Kano, unpublished observation).

### Roles of mGluR1 in CF synapse elimination

We have reported that multiple CF innervation persists into adulthood in four strains of mutant mice lacking mGluR1 (Kano et al., 1997), G-protein  $\alpha_q$  subunit ( $G\alpha_q$ ) (Offermanns et al., 1997; Hashimoto et al., 2000), phospholipase C  $\beta_4$  (PLC  $\beta_4$ ) (Kano et al., 1998; Hashimoto et al., 2000), and PKC $\gamma$  (Kano et al., 1995) caused by the defects in CF synapse elimination during the third postnatal week. In contrast to GluR $\delta$ 2 mutant mice, these mutant mice have normal PF–PC synaptogenesis and normal rise time of the CF-EPSCs. We also found that blockade of NMDA receptors during P15 and P16, but not before or after this period, resulted in a higher incidence of multiple CF innervation and caused a mild but persistent loss of motor coordination (Kakizawa et al., 2000). Neither basic synaptic functions nor cerebellar morphology was affected by this manipulation. During P15 and P16, NMDA receptors were not functional at either PF to PC or CF to PC synapses but were rich in mossy fiber to granule cell synapses (Kakizawa et al., 2000). We also demonstrated that the mGluR1 to PKC $\gamma$  cascade must work within PCs to complete CF synapse elimination (Ichise et al., 2000). We introduced the mGluR1 transgene into mGluR1-null mutant mice using a PC-specific promoter, L7. The mGluR1-rescue mice showed normal regression of multiple CF innervation and apparently normal motor coordination (Ichise et al., 2000). These results suggest that the signal transduction cascade from mGluR1 to PKC $\gamma$  is activated at PF–PC synapses (Kano et al., 1995, 1997) in a manner dependent on NMDA receptor-mediated activity at mossy fiber to granule cell synapses (Kakizawa et al., 2000).



In GluR $\delta$ 2 mutant mice a substantial number of PCs had multiple CF-EPSCs with fast rise-times in addition to atypical slow CF-EPSCs (Fig. 3). It is possible that the impaired PF-PC synapse formation in GluR $\delta$ 2 mutant leads to a reduction of PF-induced drive of mGluR1 and causes persistent multiple CF innervations that are similar to those of the mGluR1, G $\alpha$ q, PLC $\beta$ 4, and PKC $\gamma$  mutant PCs.

In conclusion, we propose that GluR $\delta$ 2 is required for consolidating PF synapses onto PC distal dendrites and restricting CF synapses to the proximal dendritic domain, whereas the mGluR1 to PKC $\gamma$  signaling pathway is involved in eliminating excess CF synapses at the proximal dendrites. By the actions of these two glutamate receptors, synaptic connections of PCs mature normally during postnatal cerebellar development.

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