



Complete Genome Sequence of *Xanthomonas* Phage Pagan

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ABSTRACT The T7-like podophage Pagan infects *Xanthomonas* sp. strain ATCC PTA-13101, which was isolated from rice. The 44-kbp Pagan genome contains direct terminal repeats and contains 59 genes, 27 of which have a predicted function. Pagan is most closely related to *Xanthomonas* phage phi Xc10 and *Xylella* phage Prado.

The genus *Xanthomonas* consists of many diverse species of Gram-negative, rod-shaped phytopathogenic gammaproteobacteria (1). Members of the genus *Xanthomonas* are ubiquitous and cause a variety of diseases in many plants, including bacterial spot disease of tomatoes and peppers and bacterial leaf blight of rice (2, 3). The resulting diseased crop is unusable, which leads to economically important crop yield loss, which is as high as 50% in rice cultivation (2). *Xanthomonas* and *Xylella* are closely related genera, and *Xylella fastidiosa* causes Pierce's disease of grapevines (4). Phage-based therapeutics utilizing isolated virulent *Xylella* phages have exhibited significant potential for treating Pierce's disease (5).

Phage Pagan was isolated from filtered (pore size, 0.2 μ m), mixed freshwater collected in Vidor, TX, and infects the rice-associated *Xanthomonas* sp. strain ATCC PTA-13101. The host was grown aerobically at 28°C in tryptone nutrient broth/agar (BD), and phages were propagated via the soft agar overlay method described previously (6, 7). Full genomic DNA was purified following the shotgun library preparation protocol described by Summer (8), prepared as Illumina TruSeq libraries with the Nano low-throughput kit, and sequenced on an Illumina MiSeq instrument with paired-end 250-bp reads using v2 500-cycle chemistry. The quality of the 28,419 total sequence reads from the index containing the phage genome was evaluated using FastQC (www.bioinformatics.babraham.ac.uk/projects/fastqc). Sequence reads were then trimmed using the FASTX-Toolkit v0.0.14 (http://hannonlab.cshl.edu/fastx_toolkit/). The genome was assembled through SPAdes v3.5.0 (using default parameters) with a sequencing coverage of 6.3-fold (9). Next, the genome was closed by PCR amplification off the contig ends (forward primer, 5'-GGTAGGTGATGTGGCAGTC-3'; reverse primer, 5'-GTGTTCAACGTAGGTGAGAAGG-3') and subsequent Sanger sequencing. Genome annotation and bioinformatic analysis were conducted through software tools available in the Center for Phage Technology Galaxy and Web Apollo instances (<https://cpt.tamu.edu/galaxy-pub/>) (10, 11). Structural annotation of the genome was accomplished using Glimmer v3.0 and MetaGeneAnnotator v1.0 and with ARAGORN v2.36 for tRNAs (12–14). Gene function was then predicted using default settings for conserved domain searches with InterProScan v5.33-72 and sequence similarity searches of the NCBI nonredundant or UniProtKB Swiss-Prot/TrEMBL databases with BLAST v2.2.31 at a 0.001 maximum expectation value (15–17). Likely transmembrane domains were identified by TMHMM v2.0, and lipoboxes were identified by LipoP v1.0 (18, 19). Rho-independent termination sites were annotated with TransTermHP v2.09 (20). progressiveMauve v2.4.0 was used to calculate genome-wide DNA sequence similarity (21). Phage morphology was assessed by negatively staining samples with 2% (wt/vol) uranyl acetate and viewing through transmission electron microscopy at the Texas A&M Microscopy and Imaging Center (22).

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The 44,448-bp podophage Pagan genome has a coding density of 96.95% and contains 59 predicted genes but no tRNAs. The genome of Pagan displayed a G+C content of 62.3%, near the host *Xanthomonas* genome average of 64% G+C content (4). The contig was reopened at the direct terminal repeat boundary predicted by PhageTerm and that is syntenic with T7-like phages (23). Pagan is most closely related to *Xanthomonas* phage phi Xc10 (GenBank accession number [MF375456](#)) and *Xylella* phage Prado (accession number [KF626667](#)), with 93.46% and 69.55% nucleotide sequence identity and 53 and 48 shared proteins, respectively (24). These similarities contributed to lysis gene identification, including that of a class II pinholin, signal-arrest-release (SAR) endolysin, and overlapping spanin genes.

Data availability. The genome sequence and associated data for phage Pagan were deposited under GenBank accession number [MK903278](#), BioProject accession number [PRJNA222858](#), SRA accession number [SRR8892144](#), and BioSample accession number [SAMN11408680](#).

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REFERENCES

- Vauterin L, Rademaker J, Swings J. 2000. Synopsis on the taxonomy of the genus *Xanthomonas*. *Phytopathology* 90:677–682. <https://doi.org/10.1094/PHYTO.2000.90.7.677>.
- Lee B-M, Park Y-J, Park D-S, Kang H-W, Kim J-G, Song E-S, Park I-C, Yoon U-H, Hahn J-H, Koo B-S, Lee G-B, Kim H, Park H-S, Yoon K-O, Kim J-H, Jung C-H, Koh N-H, Seo J-S, Go S-J. 2005. The genome sequence of *Xanthomonas oryzae* pathovar *oryzae* KACC10331, the bacterial blight pathogen of rice. *Nucleic Acids Res* 33:577–586. <https://doi.org/10.1093/nar/gki206>.
- Thieme F, Koebnik R, Bekel T, Berger C, Boch J, Büttner D, Caldana C, Gaigalat L, Goesmann A, Kay S, Kirchner O, Lanz C, Linke B, McHardy AC, Meyer F, Mittenhuber G, Nies DH, Niesbach-Klösgen U, Patschkowski T, Rückert C, Rupp O, Schneiker S, Schuster SC, Vorhölter F-J, Weber E, Pühler A, Bonas U, Bartels D, Kaiser O. 2005. Insights into genome plasticity and pathogenicity of the plant pathogenic bacterium *Xanthomonas campestris* pv. *vesicatoria* revealed by the complete genome sequence. *J Bacteriol* 187:7254–7266. <https://doi.org/10.1128/JB.187.21.7254-7266.2005>.
- Monteiro-Vitorello CB, de Oliveira MC, Zerillo MM, Varani AM, Civerolo E, Van Sluys M-A. 2005. *Xylella* and *Xanthomonas* Mobil'omics. *OMICS* 9:146–159. <https://doi.org/10.1089/omi.2005.9.146>.
- Das M, Bhowmick TS, Ahern SJ, Young R, Gonzalez CF. 2015. Control of Pierce's disease by phage. *PLoS One* 10:e0128902. <https://doi.org/10.1371/journal.pone.0128902>.
- Gill JJ, Summer EJ, Russell WK, Cologna SM, Carlile TM, Fuller AC, Kitsopoulos K, Mebane LM, Parkinson BN, Sullivan D, Carmody LA, Gonzalez CF, LiPuma JJ, Young R. 2011. Genomes and characterization of phages Bcep22 and BcepL02, founders of a novel phage type in *Burkholderia cenocepacia*. *J Bacteriol* 193:5300–5313. <https://doi.org/10.1128/JB.05287-11>.
- Adams MH. 1956. *Bacteriophages*. Interscience Publishers, Inc., New York, NY.
- Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. *Methods Mol Biol* 502:27–46. https://doi.org/10.1007/978-1-60327-565-1_4.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Grüning BA, Guerler A, Hillman-Jackson J, Hiltmann S, Jalili V, Rasche H, Soranzo N, Goecks J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. *Nucleic Acids Res* 46:W537–W544. <https://doi.org/10.1093/nar/gky379>.
- Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elisk CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. *Genome Biol* 14:R93. <https://doi.org/10.1186/gb-2013-14-8-r93>.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Jones P, Binns D, Chang H-Y, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjov M, Yong S-Y, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledge-base. *Nucleic Acids Res* 46:2699. <https://doi.org/10.1093/nar/gky092>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- Krogh A, Larsson B, Heijne von G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- Juncker AS, Willenbrock H, Heijne Von G, Brunak S, Nielsen H, Krogh A. 2003.

- Prediction of lipoprotein signal peptides in Gram-negative bacteria. *Protein Sci* 12:1652–1662. <https://doi.org/10.1110/ps.0303703>.
20. Kingsford CL, Ayanbule K, Salzberg SL. 2007. Rapid, accurate, computational discovery of rho-independent transcription terminators illuminates their relationship to DNA uptake. *Genome Biol* 8:R22. <https://doi.org/10.1186/gb-2007-8-2-r22>.
 21. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
 22. Valentine RC, Shapiro BM, Stadtman ER. 1968. Regulation of glutamine synthetase. XII. Electron microscopy of the enzyme from *Escherichia coli*. *Biochemistry* 7:2143–2152. <https://doi.org/10.1021/bi00846a017>.
 23. Garneau JR, Depardieu F, Fortier L-C, Bikard D, Monot M. 2017. PhageTerm: a tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. *Sci Rep* 7:8292. <https://doi.org/10.1038/s41598-017-07910-5>.
 24. Ahern SJ, Das M, Bhowmick TS, Young R, Gonzalez CF. 2014. Characterization of novel virulent broad-host-range phages of *Xylella fastidiosa* and *Xanthomonas*. *J Bacteriol* 196:459–471. <https://doi.org/10.1128/JB.01080-13>.