

# Reduction in the Density and Expression, But Not G-Protein Coupling, of Serotonin Receptors (5-HT<sub>1A</sub>) in 5-HT Transporter Knock-Out Mice: Gender and Brain Region Differences

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The aim of the present study was to investigate the mechanisms underlying the desensitization of 5-HT<sub>1A</sub> receptors in the dorsal raphe and hypothalamus of serotonin (5-HT) transporter knock-out mice (5-HTT<sup>-/-</sup>). The density of 5-HT<sub>1A</sub> receptors in the dorsal raphe was reduced in both male and female 5-HTT<sup>-/-</sup> mice. This reduction was more extensive in female than in male 5-HTT<sup>-/-</sup> mice. 8-OH-DPAT-induced hypothermia was absent in female 5-HTT<sup>-/-</sup> and markedly attenuated in 5-HTT<sup>+/-</sup> mice. The density of 5-HT<sub>1A</sub> receptors also was decreased significantly in several nuclei of the hypothalamus, amygdala, and septum of female 5-HTT<sup>-/-</sup> mice. 5-HT<sub>1A</sub> receptor mRNA was reduced significantly in the dorsal raphe region, but not in the hypothalamus or hippocampus, of female 5-HTT<sup>+/-</sup> and 5-HTT<sup>-/-</sup> mice. G-protein coupling to 5-HT<sub>1A</sub> receptors and G-protein levels in most brain regions were not reduced significantly, ex-

cept that G<sub>o</sub> and G<sub>i1</sub> proteins were reduced modestly in the midbrain of 5-HTT<sup>-/-</sup> mice. These data suggest that the desensitization of 5-HT<sub>1A</sub> receptors in 5-HTT<sup>-/-</sup> mice may be attributable to a reduction in the density of 5-HT<sub>1A</sub> receptors. This reduction is brain region-specific and more extensive in the female mice. The reduction in the density of 5-HT<sub>1A</sub> receptors may be mediated partly by reduction in the gene expression of 5-HT<sub>1A</sub> receptors in the dorsal raphe, but also by other mechanisms in the hypothalamus of 5-HTT<sup>-/-</sup> female mice. Finally, alterations in G-protein coupling to 5-HT<sub>1A</sub> receptors are unlikely to be involved in the desensitization of 5-HT<sub>1A</sub> receptors in 5-HTT<sup>-/-</sup> mice.

**Key words:** 5-HT<sub>1A</sub> receptors; 5-HT<sub>1A</sub> mRNA; 5-HT transporter knock-out mice; G-protein coupling; hypothermia; gender difference; autoradiography; competitive RT-PCR; in situ hybridization

Increasing evidence suggests that the function of serotonin (5-HT) transporter is important in the regulation of emotional states. For example, polymorphisms in the 5' regulatory region and intron 2 of the 5-HT transporter (5-HTT) gene, which affects 5-HTT expression, may be related to neuroticism and some affective disorders (Lesch et al., 1996; Mazzanti et al., 1998; MacKenzie and Quinn, 1999; Greenberg et al., 2000; Stoltenberg and Burmeister, 2000). Also, the 5-HTT is the target of a widely used class of antidepressant drugs: the selective serotonin reuptake inhibitors (SSRIs) such as fluoxetine (Prozac). The 5-HTT removes released 5-HT from serotonergic nerve terminals and along axons (Zhou et al., 1998). In doing so, the 5-HTT terminates the activation of postsynaptic 5-HT receptors by extracellular 5-HT. It is believed that the effects of 5-HTT on the regulation of emotion are mediated by adaptive changes in the serotonergic system induced by alterations in extracellular 5-HT concentration. Therefore, studying the mechanisms underlying the effects of 5-HTT on emotion will have a significant impact on our understanding of the etiology of psychiatric disorders and should help to develop better therapeutic approaches for psychiatric disorders.

5-HT<sub>1A</sub> receptors play a role in anxiety and probably also in depression. Previous studies showed that disruption of 5-HTT function either by chronic SSRIs or by knock-out of the 5-HTT gene produces a desensitization (decrease in physiological responses to the stimulation of 5-HT<sub>1A</sub> receptors) of 5-HT<sub>1A</sub> receptors in the hypothalamus and dorsal raphe nucleus (Le Poul et al., 1995; Li et al., 1996, 1997a, 1999; Blier et al., 1998). Behaviorally, 5-HTT knock-out mice (5-HTT<sup>-/-</sup> mice) are more anxious

relative to 5-HTT<sup>+/+</sup> mice (as examined by the elevated zero maze and light/dark box) (Murphy et al., 1999; C. Wichems, unpublished data). Interestingly, these behavioral alterations also are observed in 5-HT<sub>1A</sub> receptor knock-out mice (Heisler et al., 1998; Parks et al., 1998; Ramboz et al., 1998; Zhuang et al., 1999). These results suggest that desensitization of 5-HT<sub>1A</sub> receptors may play an important role in the effects of 5-HTT on emotion. In fact, several clinical studies have reported that the combined administration of 5-HT<sub>1A</sub> antagonists with SSRIs produces an earlier therapeutic effect than SSRIs alone, suggesting that desensitization of 5-HT<sub>1A</sub> receptors may contribute to the therapeutic effects of SSRIs. Therefore, studying the mechanisms underlying the desensitization of 5-HT<sub>1A</sub> receptors induced by disruption of the function of 5-HTT should help us to understand the effects of 5-HTT on the regulation of emotion.

The purpose of the present study was to investigate the mechanisms underlying the desensitization of 5-HT<sub>1A</sub> receptors in 5-HTT mutant mice. Because the desensitization of 5-HT<sub>1A</sub> receptors can be attributable to reduction in the density or the G-protein coupling to 5-HT<sub>1A</sub> receptors, we examined the density of 5-HT<sub>1A</sub> receptors, their mRNA, and G-protein coupling. To determine whether the desensitization of 5-HT<sub>1A</sub> receptors is mediated by a decrease in the concentration of G-proteins that are coupled with 5-HT<sub>1A</sub> receptors, we also examined several G-proteins in various brain regions.

## MATERIALS AND METHODS

### Animals

5-HTT mutant mice from a CD-1x129sv/ev background were created by homologous recombination as previously reported (Bengel et al., 1998). The 5-HTT mutant mice were from the F5 generation of backcross mating with CD1 mice and were 3–5 months of age, with body weights of 30–40 gm. The mice were housed in groups of four to five per cage in a light- (12 hr light/dark, lights on at 6 A.M.), humidity-, and temperature-controlled room. Food and water were available *ad libitum*. In all of the experiments, five to eight male or female mice, as noted, were included in each group. All animal procedures were approved by the National Institute of Mental Health Animal Care and Use Committee.

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## Materials

(+)-8-OH-DPAT and (±)-8-OH-DPAT [8-hydroxy-2-(di-*n*-propylamino)-tetraline] were purchased from Research Biochemicals International (RBI, Natick, MA). <sup>125</sup>I-MPPI [4-(2'-methoxyphenyl)-1-[2'-[N-(2'-pyridinyl)-iodo-benzamido]ethyl]piperazine] and <sup>35</sup>S-GTP-γ-S were purchased from NEN Life Science Products (Boston, MA). Antibodies for G<sub>o</sub> and G<sub>z</sub> proteins were purchased from Santa Cruz Biotechnology (Santa Cruz, CA). Anti-G<sub>11/2</sub> serum, AS7, was purchased from NEN Life Science Products. Anti-G<sub>13</sub> serum was purchased from Upstate Biotechnology (Lake Placid, NY). <sup>35</sup>S-UTP and RNA labeling kits were purchased from Amersham (Arlington Heights, IL). <sup>32</sup>P-γ-ATP was purchased from ICN Biochemicals (Costa Mesa, CA).

## Autoradiographic studies

### Preparation of brain sections

Male and female mice with intact (+/+), heterozygous (+/-), and homozygous knock-out (-/-) 5-HTT genes were decapitated. The whole brains were removed and frozen immediately in dry, ice-cooled isopentyl alcohol for 10 sec. Then the brains were placed on dry ice for 10 min until they were frozen completely. Brains were stored at -80°C until they were studied.

The mouse brain was cut into 15-μm-thick coronal sections in a cryostat. The sections were thaw-mounted onto chromalum/gelatin-coated glass slides and stored at -80°C for studies within 1 month. Each slide contained brain sections from three mice (one genotype each) so that we could limit variation between slides. Five levels of sections were collected: frontal cortex (bregma 2.46–2.8 mm), striatum (bregma 0.98–0.50 mm), medial hypothalamus (bregma 0.7 to -1.06 mm), caudal hypothalamus (bregma -1.34 to -1.94 mm), and midbrain (bregma -4.36 to -4.84 mm) according to a mouse brain atlas (Franklin and Paxinos, 1997). Adjacent sections were used in <sup>125</sup>I-MPPI binding, *in situ* hybridization, and <sup>35</sup>S-GTP-γ-S binding studies.

### Autoradiography of <sup>125</sup>I-MPPI binding

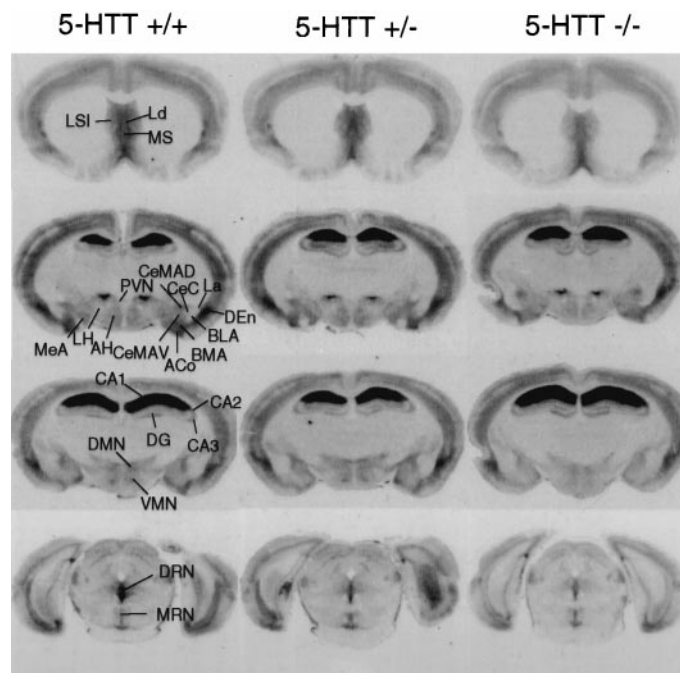
<sup>125</sup>I-MPPI binding sites in the brain sections were determined by autoradiographic assay as described (Kung et al., 1995) with slight modification. Briefly, the slides were thawed and dried in a desiccator at room temperature before assay. The brain sections were preincubated for 30 min at room temperature in assay buffer (50 mM Tris-HCl, pH 7.4, containing 2 mM MgCl<sub>2</sub>). Then the slides were incubated with <sup>125</sup>I-MPPI (0.14 nM in assay buffer) for 2 hr at room temperature. Nonspecific binding was defined in the presence of 10<sup>-5</sup> M 5-HT. Then the slides were washed twice with assay buffer at 4°C for 15 min and rinsed with cold ddH<sub>2</sub>O. After being air-blow-dried, the slides were exposed to <sup>3</sup>H-Hyperfilm (Amersham) for 1 or 3 d. The <sup>125</sup>I-MPPI binding sites in the hippocampus and dorsal raphe were measured by using films that were exposed for 1 d. The remainders of the brain regions were analyzed by using films exposed for 3 d. A set of <sup>125</sup>I microscale (Amersham) was exposed with the slides to calibrate the optic density into fmol/mg of tissue equivalent.

### In situ hybridization for 5-HT<sub>1A</sub> mRNA

**Preparation of riboprobes.** A DNA fragment encoding the third intracellular loop of mouse 5-HT<sub>1A</sub> receptor (695–1110 bp) was inserted into PCRscript vector (Stratagene, La Jolla, CA). After linearizing the plasmid with *Sac*I or *Kpn*I for sense and antisense, respectively, we performed an *in vitro* transcription with an RNA labeling kit (Amersham) and <sup>35</sup>S-UTP (1000 Ci/mmol, 20 mCi/ml; Amersham). The template DNA was removed by incubating the RNase-free DNase (10 U) at 37°C for 30 min. After the reaction was stopped by adding 25 μl of STE (TE contains 150 mM NaCl, pH 8.0), the <sup>35</sup>S-labeled riboprobes were purified by a ProbQuant G-50 micro column (Pharmacia, Piscataway, NJ). Then the collected riboprobe (~50 μl) was precipitated by ethanol (0.1 vol of 5 M ammonium acetate and 3 vol of 100% ethanol) and resuspended in 50 μl of DEPC-treated H<sub>2</sub>O after the pellets were washed with 75% ethanol. Another riboprobe encoding the 3' noncoding region of 5-HT<sub>1A</sub> mRNA (1481–1860 bp) was prepared to evaluate the selectivity of the probes.

**Hybridization.** Brain slides were thawed and dried as described above. Then the brain sections were fixed by 4% paraformaldehyde for 10 min. After being washed twice with PBS, the sections were treated with 0.25% acetic anhydride in 1 M triethanolamine for 10 min and dehydrated. After the sections were air-dried, 150 μl of hybridization solution (20 mM Tris, pH 7.4, 50% formamide, 0.3 M NaCl, 1 mM EDTA, 1× Denhardt's solution, 10% dextran sulfate, 150 mM DTT, 0.2% SDS, 50 μg/ml salmon sperm, 0.25 mg/ml tRNA, and 20,000–40,000 cpm/μl <sup>35</sup>S-labeled probe) was added on each slide, and the sections were covered with a coverslip. The slides were incubated at 54°C overnight in humidified chambers (a plastic box with two layers of filter paper wetted by 50% formamide in 2× SSC).

Then the slides were washed four times with 4× SSC for 5 min at room temperature, followed by incubation of the slides with 40 μg/ml of RNase A solution (0.01 M Tris, pH 8.0, 0.5 M NaCl, and 1 mM EDTA) at 37°C for 30 min. After being washed with 1, 0.5, and 0.1× SSC at room temperature for 5 min with shaking, the slides were incubated four times in 0.1× SSC containing 2 mM DTT at 65°C for 15 min (DTT was added immediately before incubation). Then the slides were incubated in 0.1× SSC and 2 mM DTT at room temperature for 1 min, followed by dehydration with 50, 70,



**Figure 1.** Autoradiography of <sup>125</sup>I-MPPI binding in 5-HTT mutant mice. *Left*, Wild-type 5-HTT mice (5-HTT +/+). *Middle*, Heterozygous 5-HTT knock-out mice (5-HTT +/-). *Right*, Homozygous 5-HTT knock-out mice (5-HTT -/-). The brain sections from *top* to *bottom* are striatum (bregma 0.98–0.50 mm), medium hypothalamus (bregma 0.7 to -1.06 mm), caudal hypothalamus (bregma -1.34 to -1.94 mm), and midbrain (bregma -4.36 to -4.84 mm). *ACo*, Anterior cortical amygdaloid nucleus; *AH*, anterior hypothalamic nucleus; *BLA*, basolateral amygdaloid nucleus, anterior; *BMA*, basomedial amygdaloid, anterior; *CA1–CA3*, CA1–CA3 field of hippocampus; *CeC*, central amygdaloid nucleus, capsular division; *CeMAD*, central amygdaloid nucleus, medial anterodorsal; *CeMAV*, central amygdaloid nucleus, medial anteroventral; *DEN*, dorsal endopiriform nucleus; *DG*, dentate gyrus of hippocampus; *DMN*, dorsomedial hypothalamic nucleus; *DRN*, dorsal raphe nucleus; *La*, lateral amygdaloid nucleus; *Ld*, lambdaoid septal zone; *LH*, lateral hypothalamic nucleus; *LSI*, lateral septal nucleus, intermediate; *MeA*, medial amygdaloid nucleus, anterodorsal; *MRN*, medial raphe nucleus; *MS*, medial septal nucleus; *PVN*, paraventricular hypothalamic nucleus; *VMN*, ventromedial hypothalamic nucleus.

90, and 95% ethanol containing 300 mM NH<sub>4</sub>Ac. After being rinsed with 100% ethanol, the slides were air-dried and exposed, along with a <sup>14</sup>C-microscale (Amersham), to <sup>3</sup>H-Hyper film for 7–14 d.

### Autoradiography of 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding

8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding was performed as previously described (Sim et al., 1997; Waerber and Moskowitz, 1997) with slight modification. Briefly, the brain slides were thawed and dried in a desiccator for 1–2 hr (no more than 2 hr) at room temperature. After preincubation in the assay buffer [containing (in mM) 50 Tris-HCl, pH 7.4, 3 MgCl<sub>2</sub>, 0.2 EGTA, 100 NaCl, and 0.2 DTT] for 15 min at room temperature, the slides were incubated in 2 mM GDP (in assay buffer) for 15 min at room temperature. Then the slides were incubated with 50 pM <sup>35</sup>S-GTP-γ-S (in assay buffer containing 2 mM GDP) in the absence or presence of 10<sup>-5</sup> M (+)-8-OH-DPAT for 60 min at 30°C, defined as basal or stimulated <sup>35</sup>S-GTP-γ-S binding, respectively. Nonspecific binding was defined in the presence of 10<sup>-5</sup> M GTP-γ-S. The slides were washed twice in 50 mM Tris buffer, pH 7.4, for 3 min at 4°C. After being rinsed with cold ddH<sub>2</sub>O, the slides were air-dried and exposed to <sup>3</sup>H-Hyperfilm film for 3–7 d. A <sup>14</sup>C microscale set was exposed along with the slides to calibrate the density into fmol/mg of tissue equivalent.

### Data analysis

Brain images were captured and analyzed with the National Institutes of Health Image program. The gray scale density readings were calibrated to fmol/mg of tissue equivalent by using the microscale (<sup>125</sup>I microscale for <sup>125</sup>I-MPPI binding and <sup>14</sup>C microscale for *in situ* hybridization and <sup>35</sup>S-GTP-γ-S binding). Brain regions were identified according to a mouse atlas (Franklin and Paxinos, 1997). The adjacent brain sections were used for all three autoradiographic studies. The data for a brain region of each mouse represent the mean of four adjacent brain sections.

Specific <sup>125</sup>I-MPPI binding in each brain region was determined by subtracting the nonspecific binding sites from the total binding sites in each

**Table 1.** Autoradiography of <sup>125</sup>I-MPPI binding in 5-HTT mutant mice

	Density of <sup>125</sup> I-MPPI-labeled 5-HT <sub>1A</sub> receptors (fmol/mg tissue equivalent)					
	Female			Male		
	5-HTT +/+	5-HTT +/-	5-HTT -/-	5-HTT +/+	5-HTT +/-	5-HTT -/-
<b>Frontal cortex</b>						
M2 I-IV	43 ± 3	45 ± 3	39 ± 3	40 ± 4	45 ± 2	39 ± 2
M2 V	92 ± 6	100 ± 4	74 ± 10	101 ± 7	110 ± 3	105 ± 5
M2 VI	88 ± 6	94 ± 4	78 ± 14	89 ± 6	102 ± 4	100 ± 2
LO I-IV	66 ± 7	58 ± 5	67 ± 14	91 ± 11	93 ± 6	110 ± 7
LO V	142 ± 11	140 ± 6	128 ± 21	163 ± 11	159 ± 7	187 ± 4*
<b>Septum</b>						
LSI	110 ± 6	108 ± 6	77 ± 6*	123 ± 13	126 ± 5	130 ± 16
Ld	230 ± 16	229 ± 13	165 ± 8*	256 ± 16	260 ± 15	238 ± 19
MS	230 ± 19	241 ± 12	196 ± 24	278 ± 23	282 ± 25	302 ± 20
<b>Hypothalamus</b>						
PVN	28 ± 3	29 ± 2	20 ± 1*	40 ± 4	41 ± 4	38 ± 3
AHN	64 ± 2	70 ± 3	51 ± 2*	73 ± 6	76 ± 5	65 ± 9
LHN	29 ± 2	32 ± 1	22 ± 3*	31 ± 2	30 ± 2	26 ± 3
VMN	42 ± 3	39 ± 2	23 ± 5*	69 ± 6	65 ± 3	60 ± 4
DM	45 ± 2	41 ± 1	27 ± 3*	81 ± 4	75 ± 3	69 ± 6
<b>Amygdala</b>						
La	29 ± 2	32 ± 3	24 ± 2	32 ± 2	30 ± 3	32 ± 2
BLa	30 ± 2	27 ± 2	23 ± 3	27 ± 1	27 ± 1	26 ± 2
CeC	41 ± 2	37 ± 2	28 ± 4*	39 ± 2	41 ± 2	42 ± 5
CeMAD	130 ± 13	131 ± 10	98 ± 9	153 ± 21	144 ± 7	117 ± 3
CeMAV	90 ± 5	80 ± 5	58 ± 6*	109 ± 14	105 ± 2	89 ± 7
BMA	150 ± 4	118 ± 11*	89 ± 8*	136 ± 14	141 ± 15	119 ± 14
MeAD	79 ± 3	79 ± 5	54 ± 2*	87 ± 3	94 ± 5	73 ± 6
Aco	134 ± 5	128 ± 10	92 ± 2*	136 ± 10	156 ± 16	120 ± 11
<b>Dorsal endopiriform (DEn)</b>						
(DEn)	327 ± 14	330 ± 19	261 ± 22*	316 ± 18	317 ± 13	320 ± 16
<b>Hippocampus</b>						
CA1	300 ± 16	291 ± 21	322 ± 40	376 ± 21	362 ± 12	367 ± 19
CA2	34 ± 2	30 ± 2	32 ± 3	81 ± 4	71 ± 4	70 ± 4
CA3	41 ± 4	35 ± 2	43 ± 6	99 ± 6	83 ± 5	82 ± 4
DG	66 ± 15	46 ± 5	67 ± 19	170 ± 20	126 ± 10	107 ± 20
<b>Midbrain</b>						
DR	677 ± 78	484 ± 63	257 ± 11*	664 ± 57	573 ± 42	387 ± 43*
MR	352 ± 30	353 ± 19	241 ± 26*	374 ± 12	511 ± 34*	315 ± 34

The data are represented as mean ± SEM of five to seven mice.

\*Significant difference from 5-HTT +/+ mice;  $p < 0.05$ .

The abbreviations are the same as listed in Figure 1. In addition, LO I-V, lateral orbital cortex layers I-V; M2 I-VI, secondary motor cortex layers I-VI.

region. 5-HT<sub>1A</sub> mRNA levels in *in situ* hybridization were determined by subtracting the hybridization of sense probe from that of antisense probe. No difference was observed in the distribution and levels of 5-HT<sub>1A</sub> mRNA when *in situ* hybridization that used the probes encoding the third intracellular loop was compared with that using the probes encoding the 3' noncoding region (data not shown).

8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding sites were determined by subtracting the basal <sup>35</sup>S-GTP-γ-S binding (in the absence of 8-OH-DPAT) from the stimulated <sup>35</sup>S-GTP-γ-S binding (in the presence of 8-OH-DPAT). Because a decrease either in the density of 5-HT<sub>1A</sub> receptors or in the G-protein coupling of 5-HT<sub>1A</sub> receptors would reduce the 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding sites, the ratio of <sup>125</sup>I-MPPI binding sites and 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding sites was used to express the G-protein coupling of 5-HT<sub>1A</sub> receptors.

#### Competitive RT-PCR for 5-HT<sub>1A</sub> receptors

**Tissue preparation and total RNA isolation.** Female 5-HTT +/+, +/-, and -/- mice ( $n = 8$ ) were decapitated, and the whole brain was removed from each. The hypothalamus, hippocampus, and dorsal raphe regions were dissected. The tissues were placed immediately in 1 ml of TriReagent (Sigma, St. Louis, MO) at 4°C and homogenized with a Tissue Tearer homogenizer at high speed for 10 sec in ice. The homogenates were stored at -80°C until RNA isolation. Total RNA was isolated according to the protocol of TriReagent and dissolved in 100 μl of DEPC-treated H<sub>2</sub>O. The

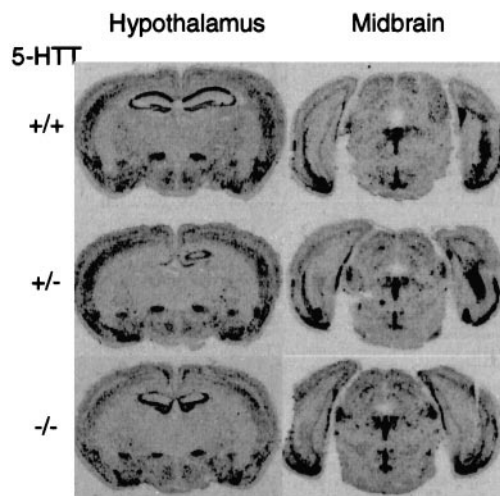
concentration of the RNA solution was determined by absorbance at 260 nm. After treatment with DNase (Life Technologies, Gaithersburg, MD), the RNA solutions then were diluted to 10 ng/μl for dorsal raphe and 20 ng/μl for hypothalamus and hippocampus before use. The mRNA concentrations of the samples were evaluated by RT-PCR of two housekeeping genes, cyclophilin and glyceraldehyde-3-phosphate dehydrogenase (GPHAD). These genes were not significantly different among the three genotypes of 5-HTT mutant mice (data not shown).

**Preparation of 5-HT<sub>1A</sub> standard.** A plasmid containing a DNA fragment encoding the third intracellular loop of 5-HT<sub>1A</sub> receptor was digested by *Pfu*MI. The resulting cohesive ends were excised by the Klenow fragment of DNA polymerase. After religation the plasmid was transfected into DH 10B *Escherichia coli* cells. A colony with site mutation (four base pairs shorter than the original plasmid) was selected and stored at -80°C.

To prepare 5-HT<sub>1A</sub> standard RNA, we linearized the plasmid with *Sac*I and transcribed it with T7 RNA polymerase. After treatment with DNase the RNA was purified by phenol/chloroform extraction, followed by ethanol precipitation. The pellet was dissolved in DEPC-treated H<sub>2</sub>O. RNA concentrations were determined by absorbance at 260 nm. A series of the 5-HT<sub>1A</sub> standard solutions was prepared by sequential dilutions of the 5-HT<sub>1A</sub> standard RNA stock solution.

**Competitive RT-PCR.** The competitive RT-PCR was modified from the method described by Chun et al. (1996). First-strand cDNA was synthesized with the ThermoScript RT-PCR system (Life Technologies). Briefly,

5-HT<sub>1A</sub> standard RNA (1  $\mu$ l) and total RNA (1  $\mu$ l) were incubated with a final concentration of 10 nM 5-HT<sub>1A</sub> primer (5'-CAGTGTCTTCA CTGTCTTCCT-3', which encoded mouse 5-HT<sub>1A</sub> antisense DNA 1110–1090 bp) at 65°C for 5 min in a total volume of 5  $\mu$ l. Then the denatured RNA was incubated with 5  $\mu$ l of RT transcription master solution at 65°C for 45 min. The reaction was terminated by incubation of the samples at 85°C for 5 min. Two microliters of the cDNA solution were used for subsequent PCR. The PCR was performed in 10  $\mu$ l of buffer solution containing 1 $\times$  PCR buffer, 1.5 mM MgCl<sub>2</sub>, 0.2 mM dNTP, 0.8 U platinum Taq DNA polymerase, 0.5  $\mu$ M sense and antisense primers (sense, encoding 695–715 bp: TGCTCATGCGGTCCTCTAT; antisense, encoding 874–857 bp: 5'-TCTCAGCACTGCGCCTGC-3'), and the 5-HT<sub>1A</sub> cDNA. After incubation at 94°C for 5 min, 28 cycles were performed at denaturation, annealing, and extension of 94°C for 30 sec, 57°C for 30 sec, and 72°C for 30 sec, respectively. The reaction was terminated by incubation at 72°C for 10 min and by the addition of 2  $\mu$ l of loading dye. To visualize PCR products, we end-labeled the antisense primer (2.5  $\mu$ M) by incubating it with <sup>32</sup>P-[ $\gamma$ ]-ATP (ICN Biochemicals) and T<sub>4</sub> polynucleotide kinase (Epicenter Technologies, Madison, WI) for 30 min at 37°C, followed by termination of the reaction at 80°C for 5 min. The PCR products (2  $\mu$ l) were resolved on 8% polyacrylamide DNA sequencing gel (Sequagel-8, National Diagnostics, Atlanta, GA). The gel was dried and exposed to Kodak BioMax MR film. The films were analyzed densitometrically with the National Institutes of Health Image analysis program. The gray scale density was calibrated by a Control Scale T-14 (Kodak). The ratio between the density of 5-HT<sub>1A</sub> mRNA and of 5-HT<sub>1A</sub> standard RNA was calculated. The logarithm of the ratio was plotted with the logarithm of concentration of 5-HT<sub>1A</sub> standard RNA by using linear regression. The x-axis intercept of the plot was calculated by a computer program, Prism (Graphpad Software, San Diego, CA), to determine the concentration of the 5-HT<sub>1A</sub> mRNA.



**Figure 2.** Autoradiography of *in situ* hybridization for 5-HT<sub>1A</sub> receptor mRNA in 5-HTT mutant mice. The brain sections were hybridized with <sup>32</sup>P-riboprobe encoding the third intracellular loop, as described in Materials and Methods. The brain sections were adjacent to the medial hypothalamus and midbrain sections in the <sup>125</sup>I-MPPI binding.

## Immunoblots

The levels of G<sub>i1</sub>, G<sub>i2</sub>, G<sub>i3</sub>, and G<sub>o</sub> proteins in the hypothalamus, midbrain, and frontal cortex were measured by using immunoblots as described in our previous paper with minor modification (Li et al., 1996). Briefly, the solubilized proteins (10–35  $\mu$ g of protein) that were extracted from the membranes of the hypothalamus, hippocampus, midbrain, and frontal cortex (Sternweis and Robinschaw, 1984; Okuhara et al., 1996) were resolved by SDS-polyacrylamide gel electrophoresis [containing 0.1% sodium dodecyl sulfate (SDS), 12% acrylamide/bisacrylamide (30:0.2), 4 M urea, and 375 mM Tris, pH 8.4 (Mullaney and Miligan, 1990)]. Then the proteins were transferred electrophoretically to polyvinylidene difluoride (PVDF) membranes. The membranes were incubated overnight with polyclonal antisera for G<sub>z</sub> (anti-G<sub>z</sub>, 1:2500), G<sub>i1/2</sub> (AS/7, 1:2500 dilution), G<sub>i3</sub> (anti-G<sub>i3</sub>, 1:2000 dilution), or G<sub>o</sub> (anti-G<sub>o</sub>, 1:500 dilution), followed by a secondary antibody for 1 hr (1:25,000 goat anti-rabbit IgG-alkaline phosphatase). After several washes the membranes were placed on a flat surface and incubated with a chemiluminescence substrate solution, CDP-Star substrate (Tropix, Bedford, MA), by pipetting a thin layer of the solution onto the membranes (~3 ml/100 cm<sup>2</sup>) for 5 min. The membranes were wrapped with Saran wrap and then exposed to Kodak x-ray film for 5–30 sec. Films were analyzed densitometrically by the National Institutes of Health Image analysis program as detailed in our previous paper (Li et al., 1996).

## Hypothermic response to a 5-HT<sub>1A</sub> agonist, 8-OH-DPAT in female 5-HTT mutant mice

5-HTT +/+, +/-, and -/- mice were injected with 8-OH-DPAT (0.1 mg/kg, s.c.). Body temperatures of the mice were measured every 10 min, from 20 min before to 60 min after the 8-OH-DPAT injections. The means of the temperatures from 20 and 10 min before 8-OH-DPAT injections were used for the zero time point temperature measures. The body temperatures of the mice were measured by a digital thermometer with a temperature probe (Physitemp BAT-12, Physitemp Instruments, Clifton, NJ) inserted 2–2.5 cm into the rectum, with the mice slightly restrained by the tail. The temperatures were measured in a room with an ambient temperature at 25°C.

## Statistics

The data are presented as group means and the SEM of five to eight mice. The data were analyzed by one-way or two-way ANOVA, followed by Student–Newman–Keuls *post hoc* tests when ANOVA showed a significant difference. Statistics were performed with a computer program (SuperANOVA, Abacus Concepts, Berkeley, CA).

## RESULTS

### Autoradiography of <sup>125</sup>I-MPPI binding: The density of 5-HT<sub>1A</sub> receptors in 5-HTT mutant mice

<sup>125</sup>I-MPPI binding sites were reduced significantly in the dorsal raphe of 5-HTT -/- mice (Fig. 1; Table 1). The reduction in the 5-HT<sub>1A</sub> receptors was ~50% in the female and ~30% in the male mice (Table 1). Although 5-HT<sub>1A</sub> receptors in the medial raphe were decreased significantly in the female 5-HTT mutant mice, a significant reduction was not observed in the male mice (Table 1).

In the female 5-HTT knock-out mice the <sup>125</sup>I-MPPI binding sites were reduced significantly in all of the nuclei of the hypothalamus and several nuclei in the amygdala and septum (Table 1). However, a similar significant reduction was not observed in the

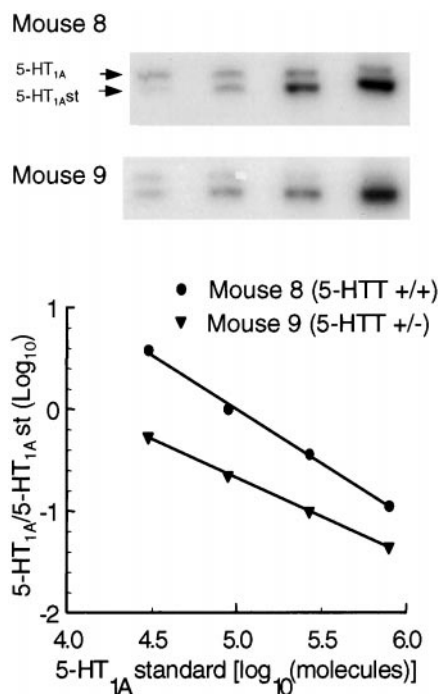
**Table 2.** *In situ* hybridization of 5-HT<sub>1A</sub> receptor mRNA in 5-HTT mutant mice

	5-HT <sub>1A</sub> receptor mRNA (fmol/mg tissue)					
	Female			Male		
	5-HTT +/+	5-HTT +/-	5-HTT -/-	5-HTT +/+	5-HTT +/-	5-HTT -/-
<b>Hypothalamus</b>						
AHN	48 ± 4	39 ± 2	42 ± 4	36 ± 3	44 ± 7	44 ± 3
LHN	13 ± 2	10 ± 1	9 ± 1	6 ± 2	7 ± 2	5 ± 2
VMN	48 ± 6	54 ± 8	41 ± 9	23 ± 4	28 ± 3	34 ± 4
DM	31 ± 4	33 ± 2	29 ± 3	12 ± 2	16 ± 3	16 ± 3
<b>Midbrain</b>						
DR	270 ± 25	231 ± 19	177 ± 9*	371 ± 52	243 ± 26*	221 ± 22*
MR	252 ± 16	254 ± 15	257 ± 34	261 ± 32	278 ± 21	268 ± 31

The data are represented as mean ± SEM of five to seven mice.

\*Significant difference from 5-HTT +/+ mice; *p* < 0.05.

The abbreviations are as same as listed in Figure 1.



**Figure 3.** An example of competitive RT-PCR for 5-HT<sub>1A</sub> receptor mRNA in the dorsal raphe of 5-HTT mutant mice. *A*, Autoradiography of DNA sequence gel that resolves RT-PCR products from 5-HT<sub>1A</sub> mRNA (5-HT<sub>1A</sub>) and 5-HT<sub>1A</sub> RNA standard (5-HT<sub>1Ast</sub>). Four concentrations of 5-HT<sub>1A</sub> RNA standard were used to compete 5-HT<sub>1A</sub> mRNA (10 ng of total RNA). *B*, Linear regression curve for calculation of concentration of 5-HT<sub>1A</sub> mRNA. The intercept of the x-axis represents the concentration of 5-HT<sub>1A</sub> mRNA.

male mice, although there was a tendency toward reduction of the <sup>125</sup>I-MPPI binding sites in these brain regions, e.g., hypothalamus and amygdala (Table 1). In contrast, no significant reduction in <sup>125</sup>I-MPPI binding sites was observed in the hippocampus and frontal cortex of either female or male 5-HTT <sup>-/-</sup> mice (Table 1).

#### **In situ hybridization to determine 5-HT<sub>1A</sub> mRNA in 5-HTT mutant mice**

5-HT<sub>1A</sub> mRNA levels in the nuclei of raphe and hypothalamus of 5-HTT mutant mice were examined by *in situ* hybridization (Fig. 2; Table 2). The distribution of 5-HT<sub>1A</sub> mRNA was consistent with both riboprobes (encoding the third intracellular loop or the 3' noncodon region), suggesting that the riboprobe used in the assay was selective for 5-HT<sub>1A</sub> mRNA. 5-HT<sub>1A</sub> mRNA levels were reduced significantly in the dorsal raphe of 5-HTT +/- and -/- male mice and 5-HTT -/- female mice (Table 2). No significant change was observed in the other brain regions.

#### **Competitive RT-PCR for examination of 5-HT<sub>1A</sub> mRNA in 5-HTT mutant mice**

To confirm the results observed by *in situ* hybridization, we conducted competitive RT-PCR to examine 5-HT<sub>1A</sub> mRNA in the hypothalamus, hippocampus, and dorsal raphe regions. The plots of the competitive RT-PCR showed a high linearity (Fig. 3). The mean of regression coefficient (*r*<sup>2</sup>) was 0.96. The internal variation was 12.4%.

In competitive RT-PCR a significant reduction of 5-HT<sub>1A</sub> mRNA was observed in the dorsal raphe region of female 5-HTT +/- and -/- mice (one-way ANOVA, *F*<sub>(2, 18)</sub> = 5.1, *p* < 0.05; Table 3). This reduction was not found in the hypothalamus or hippocampus (Table 3).

#### **Autoradiography of 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding in 5-HTT mutant mice**

8-OH-DPAT increased <sup>35</sup>S-GTP-γ-S binding ~100% in the CA1 region of hippocampus and 50% in the dorsal raphe (Figs. 4, 5). However, a high basal level of <sup>35</sup>S-GTP-γ-S binding was observed

**Table 3. Competitive RT-PCR for 5-HT<sub>1A</sub> mRNA levels in female 5-HTT mutant mice**

	5-HT <sub>1A</sub> mRNA (10 <sup>4</sup> molecules/ng total RNA)		
	5-HTT +/+	5-HTT +/-	5-HTT -/-
Dorsal raphe	0.64 ± 0.1	0.28 ± 0.02*	0.35 ± 0.1*
Hypothalamus	12.3 ± 1.6	11.3 ± 1.6	12.6 ± 2.4
Hippocampus	5.4 ± 0.5	6.0 ± 0.5	5.4 ± 0.7

Data are represented as mean ± SEM of seven to eight mice.

\*Significant difference from 5-HTT +/+ mice; *p* < 0.05.

in the hypothalamus and amygdala (Fig. 4), which caused a limited increase of <sup>35</sup>S-GTP-γ-S binding in the presence of 8-OH-DPAT. Therefore, 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding sites were examined only in the dorsal raphe and CA1 region of hippocampus.

8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding was reduced significantly in the dorsal raphe of 5-HTT -/- mice (Table 4; Fig. 5). However, the ratio of <sup>125</sup>I-MPPI binding sites and 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding sites was not altered significantly in 5-HTT mutant mice (Table 4). This suggested that the reduction of 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding in the dorsal raphe is primarily attributable to the observed decreased density of 5-HT<sub>1A</sub> receptors and not to a reduction in the G-protein coupling of 5-HT<sub>1A</sub> receptors. On the other hand, 8-OH-DPAT-stimulated <sup>35</sup>S-GTP-γ-S binding was not changed in the CA1 region of hippocampus of 5-HTT mutant mice (Table 4). No difference in the basal level of <sup>35</sup>S-GTP-γ-S was observed among three genotypes of 5-HTT mutant mice.

#### **G-protein levels in brain of 5-HTT mutant mice**

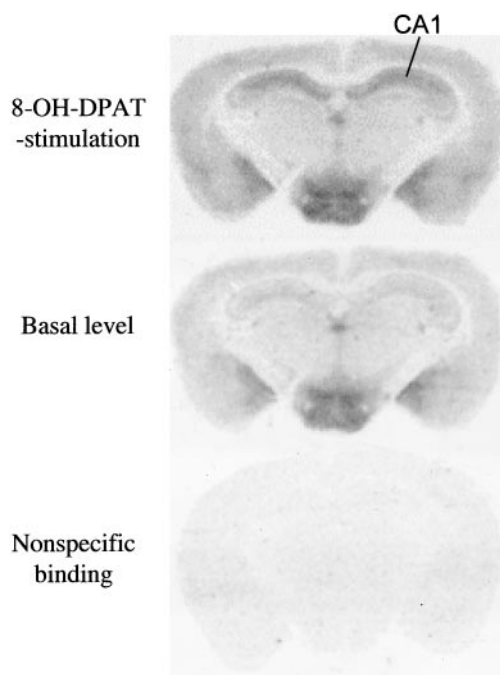
Several G-proteins that are coupled to 5-HT<sub>1A</sub> receptors were examined in the frontal cortex, hypothalamus, hippocampus, and midbrain of female mice with 5-HTT mutations by using immunoblots (Fig. 6). G<sub>11</sub> and G<sub>o</sub> proteins in the midbrain of 5-HTT mice were reduced significantly as compared with their normal littermates. On the other hand, G<sub>z</sub> proteins in the hypothalamus were increased significantly in 5-HTT -/- mice. No significant changes of G<sub>12</sub> and G<sub>13</sub> proteins were observed in any of the brain regions that were examined. None of the G-proteins was altered in the hippocampus and frontal cortex (not shown by data).

#### **Hypothermic response to 8-OH-DPAT in female 5-HTT mutant mice**

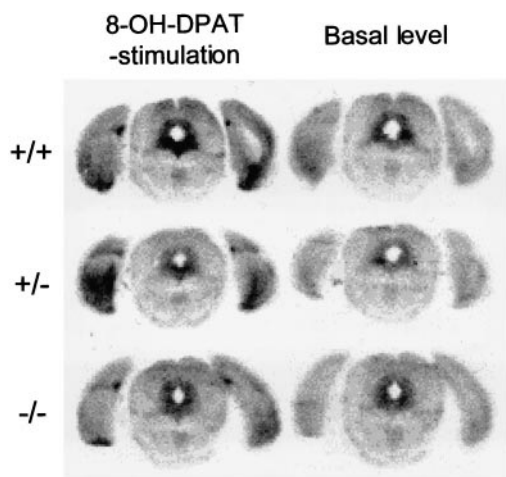
The body temperature of 5-HTT +/+ mice was attenuated significantly 10 min after injection with 8-OH-DPAT (Fig. 7). The peak time of the hypothermia that was induced by 8-OH-DPAT was 20 min. The hypothermic response to 8-OH-DPAT was absent in 5-HTT -/- mice. Although there was a slight attenuation of the body temperature after the injection with 8-OH-DPAT in 5-HTT +/- mice, the difference did not reach statistical significance when compared with 5-HTT +/+ mice (Fig. 7). Furthermore, there was no significant difference between the hypothermic response to 8-OH-DPAT in 5-HTT +/- and that in 5-HTT -/- mice.

#### **DISCUSSION**

The present studies demonstrated that the density of 5-HT<sub>1A</sub> receptors in 5-HTT -/- mice is reduced in a brain region-specific manner. These results confirm our previous observations that the desensitization of 5-HT<sub>1A</sub> receptors in the hypothalamus and in the dorsal raphe nucleus of 5-HTT mutant mice is mediated by a reduction of the density of 5-HT<sub>1A</sub> receptors (Li et al., 1999). Furthermore, we have now shown that the reduction of 5-HT<sub>1A</sub> receptors is more extensive in female than in male 5-HTT mutant mice. This is consistent with the functional examination that the hypothermic response to 8-OH-DPAT is attenuated much more extensively in female than in male 5-HTT +/- mice. In our previous report male 5-HTT +/- mice demonstrated essentially no



**Figure 4.** 8-OH-DPAT-stimulated <sup>35</sup>S-GTP- $\gamma$ -S binding in the hippocampus of normal mice. *Top*, <sup>35</sup>S-GTP- $\gamma$ -S binding in the presence of 8-OH-DPAT. *Middle*, <sup>35</sup>S-GTP- $\gamma$ -S binding in the absence of 8-OH-DPAT. *Bottom*, Nonspecific binding defined by the presence of 10<sup>-5</sup> M GTP- $\gamma$ -S.



**Figure 5.** Autoradiography of 8-OH-DPAT-stimulated <sup>35</sup>S-GTP- $\gamma$ -S binding in the midbrain sections of 5-HTT mutant mice. *Left*, <sup>35</sup>S-GTP- $\gamma$ -S binding in the presence of 8-OH-DPAT. *Right*, <sup>35</sup>S-GTP- $\gamma$ -S binding in the absence of 8-OH-DPAT.

change in the hypothermic response to 8-OH-DPAT as compared with 5-HTT *+/+* littermates (Li et al., 1999). However, our present data indicated that this hypothermic response in female 5-HTT *+/-* mice is virtually identical to 5-HTT *-/-* mice. The reduction in the density of 5-HT<sub>1A</sub> receptors in the dorsal raphe of 5-HTT *+/-* and *-/-* mice may be, at least partly, attributable to a decrease in 5-HT<sub>1A</sub> gene expression. On the other hand, no decrease in 5-HT<sub>1A</sub> receptor mRNA was observed in the hypothalamus of 5-HTT *-/-* mice. These results suggest that more than one mechanism is involved in the regulation of the density of 5-HT<sub>1A</sub> receptors in different brain regions. In addition, our studies of G-protein coupling to 5-HT<sub>1A</sub> receptors and G-protein levels indicated that these elements in the 5-HT<sub>1A</sub> signal transduction pathway do not play a major role in the desensitization of 5-HT<sub>1A</sub> receptors in the 5-HTT mutant mice.

Among different brain regions the most marked reduction of 5-HT<sub>1A</sub> receptors was observed in the dorsal raphe nucleus of both male and female 5-HTT *-/-* mice. These data could account for eliminated electrophysiological and hypothermic responses to 8-OH-DPAT in 5-HTT *-/-* mice (Gobbi et al., 1999; Li et al., 1999), functional tests for 5-HT<sub>1A</sub> receptors in the dorsal raphe of mice (Bill et al., 1991). Also, these data are consistent with the observations from other investigators who also reported a reduction of <sup>3</sup>H-WAY-100635 (a 5-HT<sub>1A</sub> antagonist) binding sites in the dorsal raphe nucleus, but not the CA1 region of hippocampus, in 5-HTT mutant mice (Fabre et al., 1998). Besides the dorsal raphe, a reduction in the density of 5-HT<sub>1A</sub> receptors also was demonstrated in the median raphe, most nuclei of hypothalamus, as well as some nuclei of the amygdala and septum of female 5-HTT *-/-* mice. Although the density of 5-HT<sub>1A</sub> receptors in the nuclei of hypothalamus of male 5-HTT *-/-* mice was not statistically different, a tendency toward a reduction was observed in all of the hypothalamic nuclei. In fact, a homogenate ligand-binding assay showed that the density of 5-HT<sub>1A</sub> receptors was reduced significantly in the hypothalamus of male 5-HTT knock-out mice (Li et al., 1999). It has been known that the hypothalamus and amygdala are related to the regulation of emotion. It is possible that the reduction in the 5-HT<sub>1A</sub> receptors in these regions contributes to the increase of anxiety and decrease in depression-like behaviors that recently were observed in the 5-HTT *-/-* mice (C. Wichems, unpublished data). On the other hand, no significant changes of 5-HT<sub>1A</sub> receptors have been found in the hippocampus and frontal cortex.

All together, these results suggest that 5-HT<sub>1A</sub> receptors in 5-HTT mutant mice are regulated differently in the different brain regions. In the dorsal raphe of 5-HTT mutant mice the 5-HT<sub>1A</sub> receptors are reduced extensively, which might be attributable to a decrease in gene expression, i.e., regulation of gene transcription. On the other hand, the density, but not mRNA, of 5-HT<sub>1A</sub> receptors is reduced in the hypothalamus, amygdala, septum, and median raphe of 5-HTT mutant mice, suggesting that translational regulation and/or post-translational modifications may be involved. The mechanism underlying the regulation of 5-HT<sub>1A</sub> receptors in

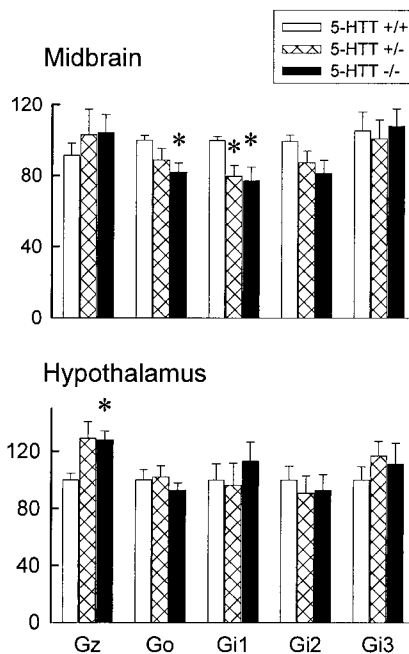
**Table 4.** 8-OH-DPAT-stimulated <sup>35</sup>S-GTP- $\gamma$ -S binding in 5-HTT knock-out mice

	<sup>35</sup> S-GTP- $\gamma$ -S binding (fmol/mg tissue)			Ratio of <sup>125</sup> I-MPPI/ <sup>35</sup> S-GTP- $\gamma$ -S binding		
	5-HTT <i>+/+</i>	5-HTT <i>+/-</i>	5-HTT <i>-/-</i>	5-HTT <i>+/+</i>	5-HTT <i>+/-</i>	5-HTT <i>-/-</i>
<b>Female</b>						
DRN	128 ± 15	104 ± 14	78 ± 14*	5.6 ± 0.6	5.3 ± 1.1	3.6 ± 0.5
CA1	198 ± 15	180 ± 5	174 ± 11	1.6 ± 0.1	1.6 ± 0.1	1.8 ± 0.3
<b>Male</b>						
DRN	89 ± 16	124 ± 13	81 ± 22	8.3 ± 1	4.2 ± 0.6	6.9 ± 2.4
CA1	228 ± 14	227 ± 11	214 ± 17	1.7 ± 0.1	1.6 ± 0.1	1.7 ± 0.1

Data are represented as mean ± SEM of five to seven mice.

\*Significant difference from 5-HTT *+/+* mice; *p* < 0.05.

The abbreviations are as same as listed in Figure 1.

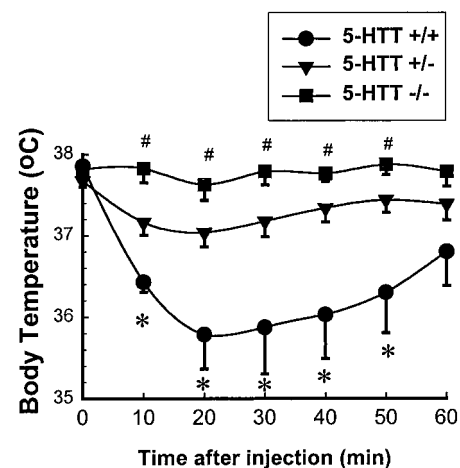


**Figure 6.** G-protein concentrations in the midbrain (*top*) and hypothalamus (*bottom*) of female 5-HTT mutant mice. The data represent the mean  $\pm$  SEM of eight mice per group. \*Significant difference from 5-HTT +/+ mice;  $p < 0.05$ .

the different brain regions of 5-HTT mutant mice is still unknown. 5-HT<sub>1A</sub> receptors in the dorsal and median raphe nuclei are autoreceptors. The reduction in the density of 5-HT<sub>1A</sub> receptors could result from a feedback regulation induced by sustained high extracellular concentrations of 5-HT because of the lack of the 5-HTT (Mathews et al., 2000). The fact that the reduction of the 5-HT<sub>1A</sub> receptors in the dorsal raphe is more extensive than in other brain regions supports the hypothesis that the 5-HT<sub>1A</sub> autoreceptors are more sensitive to an increase of 5-HT than postsynaptic 5-HT<sub>1A</sub> receptors (Artigas et al., 1996; Blier et al., 1998).

An important finding in the present studies is that the reduction of 5-HT<sub>1A</sub> receptors is substantially greater in the female than in the male 5-HTT mutant mice. Furthermore, in comparison to our previous results in male mice (Li et al., 1999), the hypothermic response to 8-OH-DPAT in female 5-HTT +/- mice is more marked than that in male 5-HTT mutant mice. This result is important because it may provide an insight into why women more frequently develop affective and some anxiety disorders. It has been known that female gonadal hormones, especially estrogen, can regulate 5-HT<sub>1A</sub> receptors (Lakoski, 1989; Maswood et al., 1995; Trevino et al., 1999; Osterlund et al., 2000). For example, estrogen treatment attenuated 5-HT<sub>1A</sub> receptor functions (Lakoski, 1989; Maswood et al., 1995) and decreased 5-HT<sub>1A</sub> receptor mRNA levels in ovariectomized monkey and rats (Pecins-Thompson and Bethea, 1999; Osterlund et al., 2000). On the other hand, estrogen also reduces 5-HTT mRNA concentrations (Pecins-Thompson et al., 1998). Therefore, the remarkable decrease of the 5-HT<sub>1A</sub> receptors in female 5-HTT mutant mice could result from synergistic effects of estrogen and the 5-HTT mutation on 5-HT<sub>1A</sub> receptors and/or interaction between estrogen and 5-HTT.

Another possible cause for the desensitization of 5-HT<sub>1A</sub> receptors is alteration of their G-protein coupling. Although 8-OH-DPAT-stimulated GTP- $\gamma$ -S binding in the dorsal raphe was reduced significantly in the 5-HTT -/- mice, the ratio of <sup>125</sup>I MPPI and 8-OH-DPAT-stimulated GTP- $\gamma$ -S binding was not altered significantly. This suggests that the reduction in the 8-OH-DPAT-stimulated GTP- $\gamma$ -S binding is attributable to a decrease of the number of 5-HT<sub>1A</sub> receptors, but not any attenuation of G-protein coupling to 5-HT<sub>1A</sub> receptors. Furthermore, although G<sub>o</sub> and G<sub>i1</sub> proteins in the midbrain are reduced significantly, but modestly, in



**Figure 7.** Hypothermic response to 8-OH-DPAT in female 5-HTT knock-out mice. The data represent the mean  $\pm$  SEM of 10 mice per group. Two-way ANOVA: main effect of genotype,  $F_{(2,147)} = 31.07$ ,  $p < 0.0001$ ; main effect of time,  $F_{(6,147)} = 8.77$ ,  $p < 0.0001$ ; interaction between genotype and time,  $F_{(12,147)} = 1.49$ ,  $p = 0.132$ . \*Significant difference from those in the 0 time points of the same genotype of 5-HTT,  $p < 0.05$  (Student–Newman–Keuls' multiple range test). #, Significant difference from the 5-HTT +/+ mice (at same time point),  $p < 0.05$  (Student–Newman–Keuls' multiple range test).

the female 5-HTT -/- mice, these and other G-proteins in the hypothalamus, hippocampus, and frontal cortex were not reduced significantly. It is interesting that G<sub>z</sub> proteins in the hypothalamus are increased even modestly in the 5-HTT mutant mice. G<sub>z</sub> proteins are coupled to 5-HT<sub>1A</sub> receptors in the hypothalamus that regulate oxytocin release (Serres et al., 2000). The elevation of the G<sub>z</sub> proteins in the 5-HTT mutant mice could be a compensatory effect for reduction in the 5-HT<sub>1A</sub> receptors in the hypothalamus. All together, these data suggest that alteration in the G-protein coupling to the 5-HT<sub>1A</sub> receptors is not a major mechanism for the desensitization of 5-HT<sub>1A</sub> receptors in 5-HTT mutant mice.

Similar to these findings in 5-HTT mutant mice, the desensitization of 5-HT<sub>1A</sub> receptors also has been observed after chronic administration of SSRIs in human and rodents (Li et al., 1997a; Lerer et al., 1999; Raap et al., 1999). In both situations the desensitization of 5-HT<sub>1A</sub> receptors occurs in the hypothalamus and dorsal raphe, but not in the hippocampus and cortex (Le Poul et al., 1995, 2000; Li et al., 1997a). However, the mechanisms underlying the desensitization may be different between 5-HTT mutant mice and that found after chronic treatment with SSRIs. In 5-HTT mutant mice the density of 5-HT<sub>1A</sub> receptors, but not their signal transduction, is reduced. On the other hand, chronic treatment with SSRIs does not reduce the density of 5-HT<sub>1A</sub> receptors in any brain regions (Li et al., 1997b; Le Poul et al., 2000). Instead, some decreases in several G-protein concentrations have been observed (Li et al., 1996, 1997a). These differences may be attributable to the much longer and more marked lack or reduction of 5-HTT function in the 5-HTT mutant mice than that in the SSRI-treated animals. An alternative explanation could be that the reduction of 5-HT<sub>1A</sub> receptors in 5-HTT mutant mice results from more complex changes during neuron development.

In conclusion, the present studies demonstrate that 5-HT<sub>1A</sub> receptors in the 5-HTT mutant mice are brain region-specifically reduced. This result could account for our previous results indicating that 5-HT<sub>1A</sub> receptor-mediated temperature and neuroendocrine responses are desensitized in 5-HTT mutant mice (Li et al., 1999). The reduction in the density of 5-HT<sub>1A</sub> receptors in the dorsal raphe, but not in hypothalamus, may be attributable to an attenuation of 5-HT<sub>1A</sub> gene expression. More interestingly, the reduction of 5-HT<sub>1A</sub> receptors is more extensive in the female than in the male 5-HTT mutant mice. This is consistent with our finding that female 5-HTT mutant mice have a more prominent and consistent increase in anxiety than males when compared with

their normal littermates. The significance of these observations is that it may provide a greater insight for the frequently reported higher incidence of affective and some anxiety disorders in women and also the premenstrual syndrome symptomatology.

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