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Synergy of Interleukin (IL)-5 and IL-18 in eosinophil mediated pathogenesis of allergic diseases

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Abstract

Eosinophils are circulating granulocytes that have pleiotropic effects in response to inflammatory signals in the body. In response to allergens or pathogens, exposure eosinophils are recruited in various organs that execute pathological immune responses. IL-5 plays a key role in the differentiation, development, and survival of eosinophils. Eosinophils are involved in a variety of allergic diseases including asthma, dermatitis and various gastrointestinal disorders (EGID). IL-5 signal transduction involves JAK-STAT-p38MAPK-NFkB activation and executes extracellular matrix remodeling, EMT transition and immune responses in allergic diseases. IL-18 is a classical cytokine also involved in immune responses and has a critical role in inflammasome pathway. We recently identified the IL-18 role in the generation, transformation, and maturation of (CD101+CD274+) pathogenic eosinophils. In, addition, several other cytokines like IL-2, IL-4, IL-13, IL-21, and IL-33 also contribute in advancing eosinophils associated immune responses in innate and adaptive immunity. This review discusses with a major focus (1) Eosinophils and its constituents, (2) Role of IL5 and IL18 in eosinophils development, transformation, maturation, signal transduction of IL5 and IL18, (3) the role of eosinophils in allergic disorders and (4) the role of several other associated cytokines in promoting eosinophils mediated allergic diseases.

Graphical Abstract

Fig. Synergy of Interleukin (IL)-5 and IL-18 in eosinophil mediated pathogenesis of allergic diseases

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Conflict of interest

No author has any conflict of interest.

Keywords

Allergy; Eosinophils; IL-5; IL-18; Immune responses; Th2 responses

1. Introduction

Eosinophils are likely first observed in 1846 by Wharton Jones and named by Paul Ehrlich in 1879, with time they have traditionally considered effector cells in the innate immune system. Eosinophils are granulocytic leukocytes that develop in the bone marrow from pluripotent progenitors and share many features with neutrophils, including growth and development in the bone marrow. In contrast with neutrophils, eosinophils exit the bone marrow and reside in the bloodstream with a half-life of ~18 hrs and migrate to the thymus, gastrointestinal tract under homeostatic conditions. Eosinophils arise from hematopoietic CD34+ progenitor cells in the bone marrow and are of 5 % of total white blood cells in healthy subjects. Eosinophils are distinguished phenotypically by their bilobed nuclei and cytoplasmic granules filled with cytotoxic and immunoregulatory proteins [1]. Mature eosinophils in the blood are less than 400 per $mm³$. Eosinophils respond to signals from cytokines, chemokines, and other proinflammatory mediators via cell-specific receptors most notably by cytokine interleukin-5 (IL-5) and the eotaxin chemokines. During eosinophilopoiesis, eosinophils acquire IL-5Rα and IL-18Rα on the cell surface and differentiate under the influence of both IL-5 and IL-18 [2]. Transcription factors play a crucial role in eosinophilic lineage commitment in the bone marrow to maturation and development of eosinophils. Eosinophilic progenitors express C/EBPα, C/EBPε, PU.1, and GATA-1 and GATA-2 [3]. The increased production of eosinophils is observed in various allergic conditions, skin diseases, parasitic infections, and reactions to drugs. Peripheral blood and tissue eosinophilia are hallmarks of parasitic helminth infection chronic asthma

and allergic diseases and play a role in host defense [4]. Eosinophils migrate from bone marrow to the specific tissues on exposure with allergens and execute immune responses [5]. Eosinophils display pleiotropic effects such as lymphocyte homeostasis, PMN activation, basophil degranulation, mastocytosis, macrophage phagocytosis, Th2/DC's stabilization, epithelial cell differentiation, neurite outgrowth, fat tissue homeostasis, glucose metabolism, tissue repair and fibrosis, cardiovascular morbidity, bronchospasm, pathogen defense, antigen presentation, extracellular pathogen trap, parasite antibody-dependent cell mediated cytotoxicity [6].

Eosinophils are key innate regulator cells. Under homeostasis, eosinophils are distributed in the blood, lung, thymus, uterus, adipose tissues, mammary gland, spleen and the lamina propria of the gastrointestinal (GI) tract [7]. However, under the allergic condition, the mature eosinophils under the influence of IL-5 and eotaxin are released from bone marrow to bloodstream and recruit target tissue and modulate immune reactions. Eosinophils also regulate adaptive immune reactions and modulate lymphocyte recruitment and homeostasis. Th2 cells produce the cytokine IL-5, which promotes eosinophil production, priming, and survival, and IL-13, which induces local cells to produce eotaxins, which attract circulating eosinophils into their niche eosinophils and are critical for T-cell homing in the lung. Eosinophil-deficient ΔdblGATA-1 mice reduced Peyer's patch development, and T helper cell cytokine production was observed, highlighting the promoting role of eosinophils on lymphocyte homeostasis [8-10]. Bone marrow eosinophils co-localize with plasma cells during their maturation, secrete the cytokines APRIL and IL-6 and contribute to the survival of bone marrow plasma cells. B cell process of IgA class switching was recently found to be positively regulated by GI eosinophils in the intestinal tissue[8, 10]. Eosinophils promote B cell proliferation upon eosinophil activation in mice [11].

Eosinophils act as antigen-presenting cells. Eosinophils can present antigen to T cells. GM-CSF–treated eosinophils induce antigen-specific T-cell clone proliferation [12]. Upon allergen exposure, eosinophils present antigens by costimulation of MHC class II [13, 14]. Eosinophils in the airway lumen migrate to the draining lymph node and promote antigenspecific T-cell proliferation ex vivo. Eosinophils, alone or via dendritic cells, drive Th2 polarization. Upon allergen stimulation, eosinophils produce Th2 cytokines (IL-4, IL-5, and IL-13). Eosinophils regulate dendritic cells and Th2 pulmonary immune responses [15]. Eosinophils suppress Th17 and Th1 responses via DC regulation. Eosinophil products can serve as Th2 adjuvants via DC regulation.

Eosinophils also regulate T-cell development in the thymus. Eosinophils recognize PAMPs and defense against viral, parasitic and bacterial infection. Eosinophils and mast cells mutually potentiate in asthma and eosinophilic esophagitis. Mast cells support the survival and activation of eosinophils by secreting IL-5. Eosinophil MBP activates mast cells and basophils, and release cytokines, histamine, and TNF-α [6]. Eosinophils play a key role in tissue destruction and repair. Epithelial cells of various tissues stimulate eosinophil secretion of TGF-β and fibroblast growth factors and play a role in tissue repair and also promote wound healing [16-19].

Adipose tissue residential eosinophils regulate local cytokine milieu and glucose homeostasis. Eosinophils play a role in adipose tissue macrophage polarization and glucose metabolism critical for maintaining adipose tissue M2 macrophages [20]. Human eosinophils promoted dorsal root ganglia neuron branching. Peripheral dorsal root ganglia and airway neurons produce eotaxins to chemoattract eosinophils [21, 22]. Eosinophils producing nerve growth factor and neurotrophins regulate neuronal activity [23]. Eosinophils in the GI tract play a role in homeostasis and disease contexts, such as in EGID promoting IgA class-switching, enhancing intestinal mucus secretions, determining intestinal microbiota and inducing the development of Peyer's patches [8, 10].

The current review summarizes the most recent advances in our understanding on the contributions of IL-5 and IL-18 to the growth, transformation, and maturation of eosinophils to the maintenance of innate and adaptive immune responses in various allergic diseases. The review also briefs the current pharmacotherapy available to treat various eosinophilmediated allergic diseases.

2. Eosinophilic constituents

Eosinophil constitutes a wide variety of molecules such as adhesion molecules, adhesion receptors, apoptosis and signaling factors, chemoattractant receptors, chemokines, cytokine and receptors, Fc receptors, growth factors and its receptors, lipid body contents, lipid mediators and its receptors, pattern recognition receptors, granule-derived receptors, eosinophil-derived granule proteins, enzymes and reactive oxygen intermediates. Eosinophils morphology, cell-surface receptors, and intracellular contents differ between species. Particularly interleukin-5 receptor subunit-α (IL-5Rα) and CC-chemokine receptor 3 (CCR3), as well as sialic acid binding immunoglobulin-like lectin 8 (SIGLEC-8) in humans, and SIGLEC-F (also known as SIGLEC-5) in mice. In addition, eosinophil also has CRTH2 receptor that helps in the recruitment of eosinophils in allergic condition via bindings its ligands [24]. Eosinophilic granular proteins comprises, major basic protein-1 and −2 (MBP-1, MBP-2), eosinophil peroxidase (EPX), eosinophil cationic protein (ECP), eosinophil-derived neurotoxin (EDN), Charcot-Leyden crystal protein/Galectin-10 (CLC/ Gal-10). These are routinely studied as markers of eosinophil activation, and we recently reported that IL-18 influence in the increase production of these toxic proteins during allergic condition [2].

2.1. Major basic protein

MBP comprised more than 50 % of the granule protein and localized in the core of the eosinophil secondary granules. MBP-1 damages cells by disrupting the lipid bilayer membrane or altering the activity of enzymes within tissues [25]. MBP-1 stimulates mediator (histamine) release from basophils and mast cells, activates neutrophils and platelets, and augments superoxide generation by alveolar macrophages. MBP-2 is expressed only in eosinophils and exhibits similar functions *in vitro* like MBP-1 [26-28].

2.2. Eosinophil peroxidase (EPX/EPO)

EPO is a heme peroxidase found in eosinophils, and its gene located on chromosome 17. Its activities include the oxidation of halide ions to bactericidal ROS, the cationic disruption of bacterial cell walls, and the post-translational modification of protein amino acid residues [29-31]. EPO also mediates protein nitration in allergic airway inflammation in mice. 3 nitrotyrosine (3NT) formation after allergen challenge is dependent on EPO activity, particularly in eosinophils [32].

2.3. Eosinophil cationic protein (ECP)

ECP is a basic cationic protein located in the eosinophil primary matrix and is released during degranulation of eosinophils. ECP possess helminth-toxic and ribonucleolytic activities. ECP is assayed as one of the predictive markers of eosinophil activation in biological fluids in diseases like asthma, bullous pemphigoid severity and outcome [32, 33]. ECP exerts cytotoxicity to parasites, bacteria, and virus and make pores on the cell membrane and kill the cell by osmotic lysis by the passage of water and other small molecules [34].

2.4. Eosinophil-derived neurotoxin (EDN)

EDN is released from eosinophil granules by cytokines and other proinflammatory mediators [35]. EDN is an active ribonuclease and capable of generating soluble ribonucleotides from insoluble tRNA [36]. Eosinophils upon activation with bacteria release EDN. Eosinophils in response to Staphylococcus aureus, release EDN [37]. EDN induces production of MMP-9 from the nasal epithelium and involves in the pathogenesis of chronic eosinophilic rhinosinusitis [38]. Eosinophil-derived neurotoxin correlated with the disease severity of AD and considered as a serum biomarker for disease severity in atopic dermatitis [39].

2.5. The Charcot-Leyden crystal protein/Galectin-10 (CLC/Gal-10)

CLC expression was increased in asthma, allergic reactions, and fungal and helminthic infections and related to the histological grade and with the number of eosinophils in the lesion of celiac disease patients and also higher in eosinophilic cystitis [40, 41]. CLCs upon their phagocytosis by primary human macrophages induce the release of the IL-1β and activate the NLRP3 Inflammasome. CLC functions as a T cell-suppressive molecule in eosinophils [42]. Neutralization of galectin-10 partially abrogated the suppressive function of the eosinophils, and recombinant galectin-10 was able to suppress T cell proliferation [43]. The components of eosinophils and their functions are described in Fig. 1 and Table 1.

3. IL5 mediated signal transduction and eosinophils development and

survival

Interleukin 5 (IL-5) plays a vital role in the activation, terminal differentiation, growth and survival of eosinophils at the early stage and the sites of /inflammation. Eosinophils highly express the interleukin-5Rα on their surface and also release significant amounts of IL-5 [67]. IL-5 signals via the IL-5 receptor (R) α chain and β chain (βc) complex [68]. Upon

activation, IL-5 signaling is initiated by the α -chain that binds IL-5 with low affinity, and dimerization with the βc-subunit forms high-affinity receptor [69]. IL-5 activation induces phosphorylation of a series of cellular proteins such as βc, HS1, Shc, Btk, JAK1, JAK2, STAT1, STAT3, STAT5, PI3K, MAPKs. Phosphorylation of these proteins activates downstream molecules such as Pim-1, c-fos, c-jun, NFκb, and thus induce cell survival and proliferation, immune responses and eosinophil number [70-76]. JAK2 is associated with hIL-5Rα, whereas JAK1 is associated with hIL-5Rα and forms. JAK1 and JAK2 are activated upon IL-5 stimulation. JAK2 and STAT5 activation are essential for IL-5 dependent signal transduction in eosinophils and B cells [77, 78]. IL-5 also activates, SHP2 and Raf-1 and Raf-1 regulate degranulation. Ras GTPase-ERK activation is vital for IL-5 mediated cell survival, and proliferation [79]. The inhibitors for kinases demonstrated the role of kinases in eosinophils activation [80]. In eosinophils p38 MAP kinases are activated, whereas its activation is inhibited in dying eosinophils [81]. The inhibitor for p38 MAP kinase SB239063 abolish lung eosinophilia in mice, another inhibitor SB203580 enhances apoptosis of eosinophils [82, 83]. However, there is a concern that these inhibitors might have side effects as they target signaling mechanisms in various cell types. In addition to eosinophil survival, IL-5 also inhibits eosinophil apoptosis, via NF-kB mediated induction of $BCL-X_L$ and play a role in the survival of eosinophils [84, 85] (Fig. 2.).

4. IL18 mediated signal transduction in inflammation

Interleukin-18 (IL-18) is an 18 kDa pro-inflammatory cytokine has been identified as an IFN-γ-inducing factor, and is a member of the IL-1 family of cytokines and display pleiotropic effects in inflammation and allergic diseases [86]. The pro-IL-18 expression observed in immune cells like monocytes, macrophages, dendritic cells, Kupffer cells and also in other cell types like keratinocytes, articular chondrocytes, synovial fibroblasts and osteoblasts [87]. Human eosinophils isolated from BALF and blood express IL-18 mRNA and protein [88]. Pro-IL-18 is cleaved by caspase1 and form IL-18. IL-18 recognizes heterodimeric IL18 receptors IL-18Rα and IL-18 Rβ, and signals via MyD88, IRAK, TRAF6 and degradation of IkB kinase and phosphorylates p50 and p65 complexes and induces NFkB signal transduction. On the other hand, IL18 mediated TRAF6 also phosphorylates MAPK, ERK, and JNK and induce AP-1 activation. These responses play a role in IL-18 mediated immune activation of T cells, B cells, NK cells, macrophages, mast cells and dendritic cells (Fig 4). IL-18 mediated Th1/Th2 response leads to B-cell activation via the release of cytokines IFN γ and IL-4 [89, 90]. Infection with pathogens like bacteria, virus, and fungi, and irritants exposure like silica, asbestos, and bleomycin, challenge activates NLRP3 inflammasome via PAMPS and DAMPs in immune cells and increase IL-1β secretion [91-94]. NLRP3 activation involves activation of adaptor protein ASC, and its sequestration in the nucleus to cytosol that interact with NLRP3 through its PYD and with caspase-1 through its CARD domains $(9, 57)$. Activated caspase-1 cleaves IL-1 β and IL-18, in the cytoplasm and these cytokines are released to the extracellular milieu and activate NLRP3-TLR-MYD88-NFkB signal transduction [95] (Fig 5). NLRP3 inflammasome responses drive steroid-resistant asthma [39]. NLRP3 inflammasome formation is observed in the development of COPD and many other inflammatory diseases [96].

5. Role of IL18 in eosinophils development and maturity

Previously, Ougura et al. [97] described that IL18 induces neutrophils, eosinophils, and lymphocytes with an elevation in IL-5 and GM-CSF levels. Most recently, we first time implicated the role of IL-18 in eosinophils differentiation and transformation of IL-5 differentiated naïve eosinophils into the CD101⁺CD274⁺ pathogenic eosinophils [98]. We showed that eosinophil subsets are not present in naïve IL-18 gene-deficient mice and an increase in the human blood and tissues in diseases state. Importantly, Il-18 is induced in most allergic disease and all eosinophils accumulated in the tissues of allergic patients are expressing CD101⁺CD274 [98].

In addition, Kumano et al. [99] earlier showed that Interleukin-18 enhances eosinophil deposition in the airways of mice immunized and sensitized with oval albumin. Further reports showed the role of IL-18 in eosinophil-mediated tumoricidal activity against a Colo-205 carcinoma cell line by upregulating LFA-1 and ICAM-1, however neutralization of IL-18 reduced eosinophil-mediated Colo-205 apoptosis and inhibited cell-cell adhesion [100]. IL-18 is critical cytokine involved in activation of iNKT cells and ICAM in promoting human EoE, additionally IL-18 in vitro also stimulates human iNKT cells and endothelial cells and induce eosinophil-active cytokines IL-5 and IL-13 [101]. We previously reported that IL-18 induce iNKT cell activation to release the eosinophil activating cytokine IL-5, as IL-5-deficient mice and iNKT cell-deficient (CD1d null) mice do not induce EoE in response to intranasal IL-18 challenge. Thus allergen-induced IL-18 has a significant role in promoting IL-5 and iNKT dependent EoE pathogenesis in mice [102]. IL-18 overexpression induces Th1, Th2 cytokine-mediated airway inflammation in mice with an increase in IFN- γ , IL-13, eotaxin levels and CD4⁺ T cells in ova sensitized mice [103].

6. Importance of IL5 and IL18 in eosinophils biology

IL-5 is a well established eosinophils differentiation, growth, and survival factor for eosinophils. Previous studies demonstrated that IL-5 levels were increased in asthma, allergy, and inflammation [104-106]. Thus IL-5 selectively affects eosinophil biology and is involved in a wide variety of allergic/inflammatory diseases mediated by eosinophils and is a therapeutic target for eosinophil-associated diseases.

Role of IL-18 in asthma development was studied in a model of airway hyperresponsiveness, and peribronchial eosinophilic inflammation of mice. Administration of mice with antihuman IL-18 antibodies protected against eosinophil-mediated airway inflammation and demonstrated the role of IL-18 in the development of asthmatic inflammation [107]. The current review signifies the critical synergetic role of IL-5 and IL-18 in eosinophils transformation from naïve eosinophils to pathogenic eosinophils. These understandings will provide an important input in eosinphils biology and future therapeutic interventions for eosinophils associated allergic diseases.

7. Eosinophils mediate immune responses

In general eosinophils defense helminth parasites by the release of cytotoxic granular proteins via degranulation mechanism [108]. Neutrophils infiltrate to the site of allergen

exposure or inflammation. Eosinophil major basic protein stimulates PI3K and PKCζ and releases superoxide via activation of NADPH oxidase [109]. MBP also enhance neutrophil adherence receptor CR3, thus enhance the inflammatory role of neutrophils in allergic diseases [110]. The transmembrane migration of eosinophils was observed upon coincubation of eosinophils with neutrophils and stimulation with IL-8, and this provides evidence that neutrophils migrate in response to IL-8 and leads to eosinophils accumulation in the airways in asthma [111]. Classical M1 type macrophages are proinflammatory and are activated by IFN-γ and LPS and release IL-12, IL-6, TNF-α, and CXCL10, CCL3. Alternatively, M2 macrophages exhibit anti-inflammatory activity and are activated by Th2 cytokines IL-4 and IL-13 via induction of STAT6 activation and release cytokines in asthma and allergic diseases [112, 113]. Depletion of alveolar macrophages increased eosinophils in bronchoalveolar lavage fluid of ovalbumin (OVA)-sensitized mice model of asthma, and transfer of unsensitized alveolar macrophages abrogated these changes [114]. IL-33 differentiates eosinophils from CD117⁺ progenitors in IL-5 dependent manner and enhances airway inflammation with an increase in eosinophils and macrophages [115].

Mast cells interact with eosinophils on IgE activation or dexamethasone challenge, and this interaction helps in eosinophils survival and communication and mediated by CD48, and 2B4 mediate mast cell–eosinophil interactions and increase eosinophil survival [116]. In another study of allergen-induced EOE model, Niranjan et al., [117] identified an increase in esophageal mast cell numbers in parallel with eosinophils and promoted muscle cell hyperplasia and hypertrophy. Mastocytosis and mast cell degranulation observed in the esophagi of patients with esophageal esophagitis [118]. Mast cell-mediated chemokine release leads to the recruitment of eosinophils and natural killer (NK) cells, through CCL11 and CXCL8 [119]. Mast cells alone are also able to mediate Th17 mediated allergic reactions independent of eosinophils via IL33 response in eosinophil-deficient ΔdblGATA mice model following OVA inhalation [120].

Basophils also play a key role in acute and chronic allergic reactions. Basophils secrete IL-4, and IL-13 and promote Th2 dependent immunity [121, 122]. Nakashima et al. [123] reported that basophils induce eosinophils upon interaction with mesenchymal fibroblasts in murine irritant contact dermatitis model via reactive oxygen species production. Upon allergen exposure, dendritic cells acquire the allergen and present to the draining lymph node, and the cells mature and induce T cell and B cell responses. Dendritic cell number increase with allergen exposure. Activation of TLR4 leads to DC migration and recruitment and induce Th2 responses [124]. On the other hand, eosinophil recruitment regulated by classical dendritic cells subsets. During memory stage of chronic asthma with allergen exposure in mouse cDC1s secrete CCL17 and CCL22 and recruit eosinophils and promote early eosinophil infiltration, as a part of type 2 immune response by lung CD24−CD11b+ DC2s by producing NO on day 1.5, and further this process is inhibited by releasing TGF-β1 on day 2.5 by CD24+ cDC2s [125].

Eosinophils also induce diverse immune responses upon allergen exposure and mediate cellular, humoral immune response and help defense the host immune system. Eosinophils induce Th2 immune response that involves the presentation of antigen to CD4+ T cells and Th2 differentiation, IL-4, IL-13 cytokine release within lymph nodes and tissues and

secretion of EDN and recruitment of Th2 to cells under inflammation. Previous studies also demonstrated the central role of Th2 cells and eosinophils in allergic responses, and mice lacking IL-13, a Th2 cytokine and pre eosinophilic cytokine IL-5 neutralization displayed Aspergillus fumigatus clearance [126]. Eosinophils induce B cell proliferation, survival, antibody secretion and plasma cell maintenance, proliferation, and survival (Fig.3.). IL-5 Tg hypereosinophilic mice with profound B cell expansion showed a decrease in B cell lymphocytosis upon genetic deletion of eosinophils. Also, patients with idiopathic hypereosinophilic syndrome (HES) revealed a correlation between eosinophils levels and circulating B cell numbers [11, 127-129]

8. Other cytokines associated involved in IL-5 and IL-18 mediated eosinophilic allergic diseases

Several cytokines play a critical role in allergic reactions, and contribute to disease pathology through the recruitment and activation of pro-inflammatory molecules and also mediated pro-fibrotic/remodeling events. T helper (Th) cells develop from naïve CD4+-T cells under lineage-specific culture conditions and release cytokines that play pleiotropic effects in allergic diseases. Th2 cells produce IL-4, IL-5, IL-9, and IL-13 and play a role in the protection against helminths. Th17 cells produce IL-17A, IL-17F, IL-22 and show activity on neutrophils, macrophages, B-cells and protect from bacteria, fungi and play a role in chronic inflammatory and autoimmune disorders [130, 131]. Th22 cells production of IL-22 plays a role in adaptive immune response [132]. IL-18 and IL-33 are involved in the activation of both Th1 and Th2 responses and have an important role in promoting adaptive immunity [133].

8.1. IL-4

IL-4 is produced by Th2 cells, basophils, mast cells, and eosinophils and regulates allergic conditions and the protective immune response against helminths and other extracellular parasites. IL-4+ TFH cell counts were increased in eosinophilic polyp tissues. IL-4 and IL-21 were involved in polyp TFH cell–induced IgE production from naive B cells, and nasal IL-4⁺ TFH cell counts correlated highly with local IgE levels in vivo, nasal IL-4+CXCR5+CD4+ T follicular helper cell counts correlate with local IgE production in eosinophilic nasal polyps [134]. ILC2s promote allergic inflammation, IL-4 production by IL-33-stimulated ILC2s blocks the generation of allergen-specific Treg cells and favors food allergy [135]. Thymic stromal lymphopoietin-elicited basophils promote epicutaneous sensitization to food antigens and subsequent IgE-mediated food allergy through Th2 polarization and IL-4 production [136]. Eosinophils were the major IL-4 expressing cell type in the heart during experimental autoimmune myocarditis. Eosinophils drive the progression of myocarditis to inflammatory dilated cardiomyopathy through IL-4 [137].

8.2. IL-9

IL-9 is a Th2-derived cytokine and an important mediator of allergic inflammation. IL-9 overexpressing mice show increased mast cell numbers in the intestine with inflammation, paneth cell hyperplasia and up-regulation of IL-13-dependent innate immunity mediators in the intestinal mucosa [138]. IL-9 governs allergen-induced mast cell activation, and anti-

IL-9 treatment protects from airway remodeling in asthma in mice [139]. Intestinal expression of IL-9 and mast cells are higher in patients with food allergy. In response to IL-33; IL-9-producing mucosal mast cells secrete increased amounts of IL-9 and IL-13 and play a key role in susceptibility to IgE-mediated food allergy [140].

8.3. IL-13

Overexpression of IL-13 in mice promotes asthma and EoE by an IL-5, eotaxin-1, and STAT6-dependent mechanism and increase eosinophils and epithelial cells hyperplasia [141]. However, it is also reorted that Il-5 has a critical role in promoting IL-13 induced disease pathogenesis [117], because IL-13 gene-deficient mice demonstrated development of allergen-induced EoE. Similarly, a report also indicates that esophageal eosinophilia is observedeven in IL-4/IL-13 double gene-deficient and STAT6 gene-deficient mice. Indicating, that anti-IL-4Rα therapy may not protect allergen or cytokine induced gastrointestinal disorders.

8.4. IL-17

IL-17A and IL-17F are Th17 cytokines produced by $CD4^+$ T cells and implicated in the pathogenesis of various allergic and chronic inflammatory diseases. IL-17+ cell population was found exclusively in CD3⁺CD4⁺ T cells, and the percentage of Th17 cells increased in peripheral blood of AD patients [142]. IL-17 levels elevated in mouse upon allergen exposure, and IL-17 knockout mice attenuated inflammation with a significant decrease in eosinophils [143]. IL-17 levels elevated in mice infected with the respiratory syncytial virus during OVA-induced allergic airway inflammation [144].

8.5. IL-21

IL-21 is a member of the type I cytokine family and is produced by $CD4⁺$ T cells, natural killer T cells, and Th17 cells and plays a major role in the control of innate and adaptive immune reactions [145]. An increase in serum IL-21 levels in adult atopic dermatitis patients with acute skin lesions was observed in patients [146]. rIL-21 nasal administration ameliorated allergic rhinitis by prevention of IgE production by B cells, eotaxin production by fibroblasts and attenuation of eosinophil infiltration into nasal mucosa [147]. Indicating anti-IL-21 may be effective in eosinophilic allergic patients.

8.6. IL-22

IL-22 play a role in inflammations, allergic diseases, and autoimmunity. IL-22 is mainly produced by innate lymphoid cells, not by Th cells in the inflamed lungs [148]. IL-22 in lesioned skin of chronic AD is higher than in psoriatic lesions [149], IL-22 elevated in asthma patients and the mouse model of asthma [150, 151]. Allergen-sensitized IL-22 deficient mice suffer from significantly higher airway hyperreactivity upon airway challenge with an increase in eosinophil infiltration, lymphocyte invasion and production of CCL17 (TARC), IL-5 and IL-13 in the lung. IL-22 play a role in pathogen elimination and innate lung defense against A. fumigatus mediated by Dectin-1-dependent IL-22 production [152].

8.7. IL-25

IL-25-high subset people had high Th2 cytokine levels serum IgE, with airway and blood eosinophils, airway hyperresponsiveness, and subepithelial thickening [153]. IL-25 neutralization in allergen-exposed mice exhibited a reduction in pulmonary eosinophilia and levels of Th2 associated cytokines, IL-5 and IL-13 and decreased fibrosis, airway smooth muscle hyperplasia and airway hyperreactivity [154].

8.8. IL-33

IL-33 is a nuclear cytokine that belongs to the IL-1 family and plays a crucial role in innate immunity. IL-33 expressed in epithelial cells, lymphoid organs and its levels increased in allergic inflammation and bronchial asthma [155]. Upon allergen challenge IL-33, levels were increased with IL-5 and IL-13 production and airway eosinophilia without T or B cells via CD25+CD44hi lymphoid cells, and this innate type-2 response abolished in IL-33 genedeficient mice [156].

9. Il-5 generated and Il-18 transformed pathogenic eosinophil mediated

allergic diseases

Allergy is a condition that the body responds abnormally to foreign substances. This is due to the hypersensitivity of a person's immune system to a particular or a variety of allergens in the environment. The common allergens include pollen, bee stings, certain foods, dust, mites, molds, drugs, latex, dander, exposure to animals, and seasonal climate change. The common symptoms associated with exposure to allergens include rashes on skins, eczema, hives, edema, inflammation, rhinitis, sneezing, itching, runny/stuffy nose, cough, chest tightness, wheezing or shortness of breath, watery, red or swollen eyes (conjunctivitis) and mucus formation.

Allergic diseases affect approximately 50 million Americans and several people worldwide. The common allergic diseases include asthma, atopic dermatitis or eczema, anaphylaxis, hay fever, sinusitis, urticaria or hives, and eosinophilic esophagitis. Eosinophil mediated allergic diseases listed in Figure.6.

9.1. Asthma

Asthma is narrowing of airways due to inflammation and an excessive mucous generation that results in coughing, wheezing, chest tightness and shortness of breath. Eosinophilic function in asthma is not clear, and all the asthmatic patients are not eosinophilic. Preclinical models of lung allergy delineated that mice lacking eosinophils still develop airway hyperreactivity [157-159]. Several animal and human studies are correlated with eosinophils mediated asthma development with an increase in IL-5 and IL-18 levels. Eosinophil mediated asthma in humans account 20 % with refractory asthma [160]. Increase in IL-5 levels observed in asthma patients with acute respiratory, viral, mixed viral/bacterial, infections, pneumonia, bronchitis [161]. Increased IL-18 levels were observed in asthma patients and correlated with asthma exacerbations [162]. We most recently showed that CD101+CD274+ pathogenic eosinophils accumulate in the nasal lavages of asthma and esophageal biopsies of eosinophilic esophagitis patients, respectively. Additionally, these

IL-18 responsive eosinophil subsets are also increased in the blood of asthma and eosinophilic esophagitis patients [2]. A similar observations are also reported in the mouse models of asthma and eosinophilic esophagitis.

9.2. Atopic dermatitis (AD) or eczema

Atopic dermatitis or eczema is one of the most prevalent skin disorder and often occurs in children and also may appear in adulthood. AD is genetical or occurs on exposure with allergens and or environmental factors that involve immune dysregulation and impaired skin barrier function and results in red itchy and dryness of the skin [163]. The allergens that enter the skin might be picked via IgE receptor by IgE specific antigen bound to Langerhans cells, and these cells migrate to lymph node and stimulate Th1 cells to differentiate to Th2 cells and promote differentiation of B cells to plasma cells and also induce chemokines, cytokines IL4, IL5 and IL-13 and IL-18 release and aid in eosinophil development and release [164]. Pathogenic eosinophilia has been shown to be present in patients with AD [165]. Activated eosinophils in turn also secrete cytokines and chemokines, such as IL-16, IL-12, TGF-β1, and IL-13 and induce immunological reactions. Eosinophil granular proteins deposition and its degranulation observed in AD patients [166]. Topical calcineurin inhibitors like tacrolimus, pimecrolimus, immunomodulatory agents prednisolone, systemic steroid-sparing agents azathioprine, methotrexate, cyclosporine, mycophenolate are available treatments for AD [167, 168].

9.3. Anaphylaxis

Anaphylaxis is a life-threatening hypersensitive reaction. The most common allergic reactions of anaphylaxis are exposure to wasp and bee venom, legumes, animal proteins, insect stings, and analgesic medications. The persons that come across anaphylaxis require immediate administration of epinephrine and medical attention. Mast cell signaling via FcεRI and IgE binds to FcεRI, and IgE and antigen-induced signaling events induce immunological reactions and mediate eosinophil-mediated allergic reactions [169]. Glucocorticosteroids, antihistamines, methylxanthines, venom immunotherapy considered for the management of anaphylaxis [170].

9.4. Hay fever or allergic rhinitis

Allergic rhinitis affects 400 million people worldwide [171]. On exposure with allergens such as pollens, grass, and dust mites result in a runny nose, and watery eyes with inflammation of the nose and mucous membranes, fever and the condition referred allergic rhinitis (AR). AR is part of a systemic inflammatory process and is associated with other inflammatory disorders of mucous membranes including asthma, rhinosinusitis, and allergic conjunctivitis. High prevalence of asthma is recorded in people with persistent and severe rhinitis [172]. Upon sensitization, to allergens, dendritic cells present allergens to T lymphocytes and promote Th2 responses and activate $CD4^+$ T cells and releases IL-4, IL-5, IL-10, and IL-13 cytokines. IL-4 induces IgE class switch in B lymphocyte and release IgE in the bloodstream that binds with mast cells and basophils. Mast cells release histamine and mediate nasal response. Histamine and TNF-α, leukotriene C4 and prostaglandin D2 contribute to the influx of eosinophils, $CD4⁺$ T lymphocytes, and basophils by stimulation of expression of adhesion molecules on endothelial cells and cause severe allergic responses

and nasal obstruction. CD4+ lymphocytes represent IL-5 that also play a role in allergic rhinitis by activation of eosinopoiesis, influx, and survival of eosinophils in nasal tissues [173-176].

Corticosteroids like fluticasone, mometasone, ciclesonide, triamcinolone, flunisolide, beclomethasone, betamethasone, antihistamines like azelastine, olopatadine, chromones like sodium cromoglicate, nedocromil sodium, anticholinergics like ipratropium bromide, decongestants like ephedrine, pseudoephedrine, xylometazoline; antihistamines like levocetirizine, cetirizine, desloratadine and loratadine, fexofenadine, acrivastine, rupatadine, carebastine and ebastine; corticosteroids like hydrocortisone, prednisolone; antileukotrienes like montelukast and zafirlukast and leukotriene synthesis inhibitors like zileuton; decongestants like pseudoephedrine are advised as a pharmacotherapy for allergic rhinitis. Apart from the medications immunotherapy is also a cure for people with allergic rhinitis [177].

9.5. Chronic hyperplastic eosinophilic sinusitis

Sinusitis is an inflammation of the sinuses that results in mucous formation. Acute sinusitis may be a result of allergens exposure and also results in fever, sore throat, headache and swelling of the face. Chronic sinuses are severe disease process associated with nasal polyposis and develop upon exposure with antigens, allergens, fungal, bacterial agents. Eosinophilic chronic rhinosinusitis is intractable and persistent disease and characterized by an inflammation of the nose and paranasal sinuses with nasal congestion and excessive mucus production and eosinophilic infiltration of nasal polyp tissue. IL-4, IL-5, or IL-1β may be critical for PDGFRα and play a pivotal role in the pathophysiology of eosinophilic and non-eosinophilic chronic rhinosinusitis with nasal polyps [178]. IL-16 may stimulate the migration and persistence of activated eosinophilic granulocytes in eosinophilic chronic rhinosinusitis [179]. Th2 mediated IL-25, IL-33, and IL-31, TGFα is up-regulated in eosinophilic chronic rhinosinusitis [180]. Higher leukotriene C4 levels can be an indicator of the risk of recurrence of nasal polyps in patients with recurrent sinonasal polyps after surgery. Leukotriene receptor antagonists like montelukast and zafirlukast and the leukotriene synthesis inhibitor zileuton are recommended therapeutic measures [180, 181]. Intranasal corticosteroids are also therapeutic strategies to modulate nasal mucosa, epithelial repair and regulating tissue remodeling markers, and to reduce tissue eosinophil infiltration [182].

9.6. Urticaria or hives

Body's reactions to allergens cause rashes on the skin that turn to swollen pale red bumps that are itchy and sometimes causes burning, and the condition also referred to as hives. Food allergens, medications, exposure to cold or hot environmental conditions and insect stings are triggers for hives. Eosinophils are recruited upon allergen exposure in urticaria and contribute to the pathogenesis by the release of proinflammatory cytokines, tissue-damaging protein, and immunoregulatory molecules. Eosinophilic granular proteins MBP and ECP observed in urticaria with IL-3, IL-4, IL-5, IL-2, IL-10, IL-12, and GM-CSF, IFN- γ , eosinophilic infiltration with deposits of cationic proteins MBP and EDN and high levels of leukotrienes C4 are observed in the lesioned skin [183]. Medications for urticaria include

epinephrine auto-injector, antihistamines, ultraviolet therapy, application of hydrophobic barrier creams, anti-IgE antibody Omalizumab administration [184-188].

9.7. Eosinophilia

The condition where eosinophils count increases <450 cells per μl of blood is eosinophilia [189]. Eosinophilia observed in a variety of allergic diseases that encountered as a result of increased eosinophil levels or cytokines such as IL-4, IL-5, and IL-13 that may also induce eosinophilia. Transgenic mice expressing IL-5 show eosinophilia [190]. The diagnosis of eosinophilia in patients characterized by an allergic condition like wheezing, rhinitis, or eczema [191]. The common factors that contribute to eosinophilia include exposure to helminth like schistosomiasis; the presence of a pet dog infected with *Toxocara canis*, and hypersensitivity to drugs. Eosinophilia observed in a variety of cancers [192, 193]. Parasitic eradication is the main cure for eosinophilia and albendazole is popularly used [194]. Therapeutics for eosinophilia with organ involvement include corticosteroids and IFN-α for the steroid-resistant disease. Other agents for the steroid-resistant disease, which are usually given as long-term maintenance regimens to control organ involvement, include the hydroxyurea, Chlorambucil, Vincristine, Cytarabine, 2-Chlorodeoxyadenosine (2-CdA) Etoposide, Cyclosporine [195].

9.8. Eosinophilic esophagitis (EoE)

EOE is one of the major cause of upper gastrointestinal morbidity with an estimated prevalence of 56.7/100,000 persons. Food sensitization is one of the main reasons for EOE and characterized by esophageal dysfunction with the formation of eosinophilic infiltrate in the esophageal mucosa [196]. Several eosinophils active Th2 cytokines, IL-4, IL-5, IL-13, and IL-18 shown to increase in EoE patients, and the overexpression of these mice promotes disease pathogenesis similar to the human. Our earlier study showed CD274 expressing and non-expressing eosinophils in pediatric and adult EoE patients. CD274 expression on blood eosinophils and blood mRNA expression levels were higher EoE patients and decreased following treatment [197]. Therapeutic approaches for EoE include chronic dietary elimination of allergic food, topical corticosteroid application, and esophageal dilation (Liacouras et al., 2011). Anti-IL-5, anti-IL-13 and anti-IL4 Ra clinical trials are in progress in most of the allergic diseases including EoE. Currently, the elimination diet and corticosteroids therapy are the best approaches to improve the EoE patients quality of life [198].

9.9. Eosinophilic cystitis (EC)

Eosinophilic cystitis characterized by infiltration of eosinophils and inflammation across the bladder wall. EC is characterized by injury of eosinophils to the bladder tissues resulting in gross hematuria, marked irritation of the lower urinary tract causing dysuria and frequency and fibrosis of the mucosa and muscularis necrosis of bladder wall [199]. Dysregulation of clonal eosinophilic proliferation, resulting in overproduction eosinophils in bladder mucosa with an elevation of IL-4, IL-5, IL-13, IFN γ , and TNFa observed in EC. Cyclophilin inhibitors and, cyclosporine identified as a promising strategy to treat EC [200].

EF characterized by edema and induration of the arms and legs and infiltration of eosinophils. EF diagnosed as inflammation and thickening of collagen bundles in the superficial muscle fascia with infiltration of immune cells. Laboratory diagnosis EF includes eosinophils accumulation in blood, hypergammaglobulinemia, and an increase in erythrocyte sedimentation rate [201]. An increase in T cell population and IL-5 levels observed in EF patients, and the increase in IL-5 levels correlated with enhanced production of eosinophils, their maturation, and recruitment to the inflamed regions [202-205]. Therapy for EF includes administration of corticosteroids and sulfasalazine, cyclosporine, methotrexate, hydroxychloroquine [206].

9.11. Eosinophilic granulomatosis with polyangiitis (EGPA), aka Churg-Strauss syndrome

EGPA is a systemic small- vessel eosinophilic vasculitis observed in association with asthma and eosinophilia. It is characterized by the presence of anti-myeloperoxidase (MPO), antineutrophil antibodies in the cytoplasm of 30–38% of patients and most frequently involves peripheral nerves and skin [207]. Apart from genetic background associated with HLA-DRB4 alleles the disease can be triggered by exposure to allergens, infections, vaccinations or drugs such as macrolide antibiotics, leukotriene receptor antagonists and exposure to particulate silica. EGPA is a Th2 mediated disease and the T cells produced Th2 cytokines such as IL-4, IL-13, IL-5, and CD294⁺ T cells observed with an increase in CCL17 and IL17A and a decrease in TREG cells. Clinical manifestation and laboratory findings demonstrate asthma and rhino- sinusitis, peripheral eosinophilia lung infiltrates due to bronchial plugging by mucus formation is also observed in patients [208]. Cyclophosphamide, Methotrexate, Rituximab, Mepolizumab suggested treatment strategy for EGPA [209, 210].

9.12. Eosinophilic pneumonia (EP)

Eosinophilic pneumonia is a pulmonary disease characterized by diffuse pulmonary infiltrates and an increase in eosinophils in the lungs and bronchoalveolar lavage fluid. EP's are due to lung infections, or drug-induced and off acute and chronic types. Several studies reported that cigarette smoking linked with the development of acute eosinophilic pneumonia. Chronic eosinophilic pneumonia characterized by radiographic evidence of bilateral peripherally located opacities with shortness of breath for several months. Peripheral blood eosinophilia with blood eosinophil count >1000/mL and increase in serum IgE level, erythrocyte sedimentation rate, C-reactive protein, and platelet counts [211, 212]. Corticosteroid treatment with intravenous methylprednisolone or oral prednisone and omalizumab were considered an effective strategy for EP [213, 214].

9.13. Hypereosinophilic syndrome (HES)

HES is characterized by a marked increase in eosinophils $(>1500 \text{ eosinophils/mm}^3)$ and its associated pathology with eosinophil-mediated end-organ damage. HES is also associated with cardiovascular complications with endomyocardial fibrosis and thrombosis [215]. HES patients showed clonal T-cell receptor rearrangements and IL-5 producing T cells characterized by CD3− CD4+ T cells [216]. HES also may be due to interstitial

chromosomal deletion on 4q12 leading to the creation of the FIP1L1-PDGFRA fusion gene (F/P+variant) [217]. Corticosteroids, hydroxyl urea, vincristine, prednisone, hydroxycarbamide, and interferon-α, mepolizumab (anti–IL-5 mAb), imatinib mesylate, cyclophosphamide, 6-thioguanine, methotrexate, cytarabine, 2-CDA, alemtuzumab, cyclosporine, intravenous immunoglobulin are suggested therapy for HES [218, 219],

9.14. Eosinophilic COPD

Chronic obstructive pulmonary disease is a lethal disease and is the 4th leading cause of deaths worldwide (WHO 2015). IL-18 overexpression is linked with emphysematous lesions and mediates Th1, Th2, and Th17 cytokine responses [220], Increased expression of IL18 was observed in COPD patients and correlated with disease severity [221-223]. Due to the pathogenic role of eosinophils in COPD clinical results demonstrated the benefits of monoclonal antibodies targeting IL-5 in severe eosinophilic COPD. Mepolizumab, a humanized monoclonal antibody, reduces eosinophil counts in blood and tissues by blocking IL-5, through binding to eosinophil surface receptors [67, 224]. Mepolizumab at a dose of 100 mg was associated with a lower annual rate of moderate or severe exacerbations than placebo among patients with COPD and an eosinophilic phenotype and suggested that eosinophilic airway inflammation contributes to COPD exacerbations[225].

9.15. Eosinophilic myocarditis (EM)

EM is an acute life-threatening cardiac inflammatory disease that is characterized by eosinophilic infiltration and has a poor prognosis and requires endomyocardial biopsy for definitive diagnosis. EM often accompanied by eosinophilia and is associated with immune dysfunctions. The clinical observation of patients with EM is with abrupt impairment of left ventricular ejection fraction and high risk of malignant arrhythmias. Thus, corticosteroid therapy, some form of inotropes and mechanical circulatory support for recovery in patients with EM was supportive and beneficial. Elevated IL-5 levels were observed in patients with EM and anti-IL5 therapy by administration of Mepolizumab improved cardiac function [226]. Despite the progress in understanding the EM further clinical trials and awareness of disease progress and treatment are required for treating patients with EM [227, 228]. Induction of EM in hypereosinophilic IL-5Tg mice resulted in eosinophilic myocarditis with ventricular and atrial inflammation and progress to dilated cardiomyopathy [137].

9.16. Eosinophilic cellulitis (ECE)/ Wells syndrome

ECE also was known as Wells syndrome is a hypersensitivity of the dermis against a variety of exogenous and endogenous antigenic stimuli and is usually seen in adults and rarely in children. ECE represents acute pruritic dermatosis, with scarring and flame figures of the skin with an accumulation of inflammatory cells and eosinophilic debris. IL-5 overproduction might contribute to eosinophil accumulation [229]. CD4+CD7−T cells may be implicated in IL-5 production and recruitment of dysregulated eosinophils in ECE [230]. IL-4 and IL-13 mediated activation of CD163+ M2 macrophages might also contribute to the recruitment of eosinophils in ECE [231]. IL-2 mediated platelet activation stimulated activation CD25 expressing eosinophils degranulation and release of eosinophilic cationic protein and tissue damage documented in ECE [232]. Treatment measures of ECE include

corticosteroids, dapsone, antihistamines, colchicine, interferon- α, antimalarial drugs, and immunosuppressive agents like cyclosporine and azathioprine, and tacrolimus [233-236].

9.17. Pulmonary eosinophilia (PE)

Following allergen exposure in pulmonary eosinophilia characterized by eosinophilia, goblet cell metaplasia, and increased Th1, Th2, Th17 mediated cytokine production. Particularly, Th17 mediated IL-17 play a key role in immune reactions in pulmonary eosinophilia following Aspergillus fumigatus exposure in mice. Inflammation attenuated in IL-17 knockout mice challenged with A. fumigatus with a significant decrease in eosinophils, conidial clearance, and the early transient peak of $CD4^+$ $CD25^+$ FoxP3⁺ cells was blunted explaining the role for IL-17 in driving Th2-type inflammation to repeated inhalation of fungal conidia [143]. Therapy for PE includes prednisolone, methylprednisolone, and oral glucocorticoids, inhaled glucocorticoids [237]. The JAK-3 inhibitor CP-690550 acts as an anti-inflammatory agent for pulmonary eosinophilia in a mouse model [238].

9.18. Eosinophilic gastrointestinal disorders (EGID)

EGID's selectively affect the gastrointestinal tract with inflammation and accumulation of eosinophils in the absence of known causes for eosinophilia like drug reactions, parasitic infections, and malignancy. The common factors for EGID's are genetic, food sensitization and persons with EGID often have an atopic disease. EGID symptoms include difficulty in swallowing, vomiting, reflux, abdominal and chest pain. IL-5, IL-13, and IL-18 implicated in the EGID pathogenesis. The diagnosis for EGID is through biopsies from endoscopy and/or colonoscopy [239, 240]. The common EGID's described below.

9.18.1. Eosinophilic colitis (ECO)—It's a rare form of the eosinophilic gastrointestinal disease and observed in neonates and also in young adults. The pathophysiology associated with ECO is altered hypersensitivity and associated with food allergy in infants, and in adults, it is T lymphocyte-associated. Common symptoms include abdominal pain, weight loss, and diarrhea, mucosal injury and malabsorption [241]. ECO characterized by eosinophilic infiltration in the colon, which is also associated with high serum IgE, and IL-5 levels. Treatment for ECO includes administration with glucocorticoids, antihistamines, leukotriene receptors antagonists and drugs that target IL-5 and IgE levels [242].

9.18.2. Eosinophilic gastritis (EG)—EG is commonly confined to stomach eosinophilic inflammation; however it also represents coexisting eosinophilic accumulation in the esophagus. Blood eosinophil counts, and accumulation of $MIB-1^+$, CD117⁺ mast cells, and FOXP3+ T-reg cells, activated T cells, are increased in patients with EG. Cytokinelike IL-4, IL-5, IL-13, IL-17, IL-18 and CCL26 was also significantly increased in EG patients that may correlate with elevated eosinophil levels or vice versa [243]. Therapies for EG include a dietary restriction in patients allergic to milk, egg, soy, wheat, nuts, seafood, red meats and other foods [244].

9.18.3. Eosinophilic gastroenteritis—EGE is commonly associated with food allergy and characterized by abdominal pain, dysphagia, nausea, vomiting, and diarrhea. EGE

characterized by the eosinophilic accumulation in the gastrointestinal tract, serum IgE levels are also observed in EGE patients [245]. IL-3, IL-5 and GM-CSF accumulation observed along with eosinophil levels in EGE. Eotoxin associated eosinophil accumulation seen in lamina propria of stomach and intestine [246]. Treatment for EGE includes administration of corticosteroids like fluticasone, prednisolone and budesonide and mast cell stabilizer cromolyn, leukotriene receptor antagonists like montelukast [247].

10. Conclusion and future perspectives

Eosinophils are normal constituents of the GI tract, mammary glands, and bone marrow, and play roles in development and homeostasis of these organs. In the present review, we discussed the evolution of the eosinophil as a multi-factorial leukocyte with pleiotropic functions, and the significance of IL-5, IL-18 and other cytokines in eosinophil associated allergic diseases. Although historically eosinophils are considered as cells that exhibit defense against parasites, the vast majority of data supports that eosinophils are pleiotropic multifunctional leukocytes that promote allergy and allergic diseases and elicit innate and adaptive immune responses upon hyperactivity to allergens. In Th2 type inflammation/ allergy IL-4, IL-5, IL-9, IL-13, IL-18, IL-25, and IL-31 cytokines play a critical role. Several studies carried out in humans, and animal models of eosinophil allergic diseases revealed that among these cytokines IL-5 and IL-18 play a major role in eosinophil differentiation, maturation, development, transformation, migration, accumulation and elicit immune reactions. Small molecule antagonists of CCR3, CRTH2 or eosinophil inhibitory receptors might also be an effective strategy to treat eosinophils in allergic diseases. Therapies with molecules that block IL-5 or and IL-18 signaling might be promising to treat eosinophilmediated allergic diseases. Although anti-IL-5 antibody therapy for allergic diseases remains elusive, a potential of humanized anti-human-IL-5Rα mAb treatment or anti-hIL-18Rα mAb treatment or in combination for asthmatic and allergic disease patients may be beneficial as a therapeutic strategy in the treatment to eliminate and control eosinophils and subsets of eosinophilic phenotypes that mediate allergic diseases. Blockade of eosinophils production in bone marrow, transformation into pathogenic eosinophils, inhibition of eosinophil activation and factors that contribute to eosinophil apoptosis is also interesting strategies for eosinophil depletion in allergic diseases. In summary, we propose the initiation of the anti-IL-18 clinical trial to block the transformation of naïve eosinophils into the pathogenic eosinophils. Anti-IL-18 therapy may maintain the eosinophils associated innate immunity and restrict disease pathogenesis by restricting the maturation of pathogenic eosinophils. Thus, we need additional studies using genetically engineered IL-5 and IL-18 double gene-deficient mice for *in vivo*, characterization of the immunobiology eosinophils associated disease pathogenesis.

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Abbreviations:

References

- [1]. Fulkerson PC, Rothenberg ME, Eosinophil Development, Disease Involvement, and Therapeutic Suppression, Adv Immunol 138 (2018) 1–34. [PubMed: 29731004]
- [2]. Venkateshaiah SU, Mishra A, Manohar M, Verma AK, Rajavelu P, Niranjan R, Wild LG, Parada NA, Blecker U, Lasky JA, Mishra A, A critical role for IL-18 in transformation and maturation of naive eosinophils to pathogenic eosinophils, J Allergy Clin Immunol 142(1) (2018) 301–305. [PubMed: 29499224]
- [3]. Fulkerson PC, Transcription Factors in Eosinophil Development and As Therapeutic Targets, Front Med (Lausanne) 4 (2017) 115. [PubMed: 28791289]
- [4]. Rosenberg HF, Phipps S, Foster PS, Eosinophil trafficking in allergy and asthma, J Allergy Clin Immunol 119(6) (2007) 1303–10; quiz 1311–2. [PubMed: 17481712]
- [5]. Uhm TG, Kim BS, Chung IY, Eosinophil development, regulation of eosinophil-specific genes, and role of eosinophils in the pathogenesis of asthma, Allergy Asthma Immunol Res 4(2) (2012) 68–79. [PubMed: 22379601]
- [6]. Wen T, Rothenberg ME, The Regulatory Function of Eosinophils, Microbiol Spectr 4(5) (2016).
- [7]. Rothenberg ME, Hogan SP, The eosinophil, Annu Rev Immunol 24 (2006) 147–74. [PubMed: 16551246]
- [8]. Chu VT, Beller A, Rausch S, Strandmark J, Zanker M, Arbach O, Kruglov A, Berek C, Eosinophils promote generation and maintenance of immunoglobulin-A-expressing plasma cells and contribute to gut immune homeostasis, Immunity 40(4) (2014) 582–93. [PubMed: 24745334]
- [9]. Jacobsen EA, Ochkur SI, Pero RS, Taranova AG, Protheroe CA, Colbert DC, Lee NA, Lee JJ, Allergic pulmonary inflammation in mice is dependent on eosinophil-induced recruitment of effector T cells, J Exp Med 205(3) (2008) 699–710. [PubMed: 18316417]
- [10]. Jung Y, Wen T, Mingler MK, Caldwell JM, Wang YH, Chaplin DD, Lee EH, Jang MH, Woo SY, Seoh JY, Miyasaka M, Rothenberg ME, IL-1beta in eosinophil-mediated small intestinal homeostasis and IgA production, Mucosal Immunol 8(4) (2015) 930–42. [PubMed: 25563499]
- [11]. Wong TW, Doyle AD, Lee JJ, Jelinek DF, Eosinophils regulate peripheral B cell numbers in both mice and humans, J Immunol 192(8) (2014) 3548–58. [PubMed: 24616476]
- [12]. Del Pozo V, De Andres B, Martin E, Cardaba B, Fernandez JC, Gallardo S, Tramon P, Leyva-Cobian F, Palomino P, Lahoz C, Eosinophil as antigen-presenting cell: activation of T cell clones and T cell hybridoma by eosinophils after antigen processing, Eur J Immunol 22(7) (1992) 1919– 25. [PubMed: 1623930]
- [13]. Akuthota P, Melo RC, Spencer LA, Weller PF, MHC Class II and CD9 in human eosinophils localize to detergent-resistant membrane microdomains, Am J Respir Cell Mol Biol 46(2) (2012) 188–95. [PubMed: 21885678]
- [14]. Shi HZ, Humbles A, Gerard C, Jin Z, Weller PF, Lymph node trafficking and antigen presentation by endobronchial eosinophils, J Clin Invest 105(7) (2000) 945–53. [PubMed: 10749574]
- [15]. Jacobsen EA, Zellner KR, Colbert D, Lee NA, Lee JJ, Eosinophils regulate dendritic cells and Th2 pulmonary immune responses following allergen provocation, J Immunol 187(11) (2011) 6059–68. [PubMed: 22048766]
- [16]. Horiuchi T, Weller PF, Expression of vascular endothelial growth factor by human eosinophils: upregulation by granulocyte macrophage colony-stimulating factor and interleukin-5, Am J Respir Cell Mol Biol 17(1) (1997) 70–7. [PubMed: 9224211]
- [17]. Ohno I, Nitta Y, Yamauchi K, Hoshi H, Honma M, Woolley K, O'Byrne P, Dolovich J, Jordana M, Tamura G, et al., Eosinophils as a potential source of platelet-derived growth factor B-chain

(PDGF-B) in nasal polyposis and bronchial asthma, Am J Respir Cell Mol Biol 13(6) (1995) 639–47. [PubMed: 7576701]

- [18]. Stenfeldt AL, Wenneras C, Danger signals derived from stressed and necrotic epithelial cells activate human eosinophils, Immunology 112(4) (2004) 605–14. [PubMed: 15270732]
- [19]. Todd R, Donoff BR, Chiang T, Chou MY, Elovic A, Gallagher GT, Wong DT, The eosinophil as a cellular source of transforming growth factor alpha in healing cutaneous wounds, Am J Pathol 138(6) (1991) 1307–13. [PubMed: 2053590]
- [20]. Wu D, Molofsky AB, Liang HE, Ricardo-Gonzalez RR, Jouihan HA, Bando JK, Chawla A, Locksley RM, Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis, Science 332(6026) (2011) 243–7. [PubMed: 21436399]
- [21]. Foster EL, Simpson EL, Fredrikson LJ, Lee JJ, Lee NA, Fryer AD, Jacoby DB, Eosinophils increase neuron branching in human and murine skin and in vitro, PLoS One 6(7) (2011) e22029. [PubMed: 21811556]
- [22]. Fryer AD, Stein LH, Nie Z, Curtis DE, Evans CM, Hodgson ST, Jose PJ, Belmonte KE, Fitch E, Jacoby DB, Neuronal eotaxin and the effects of CCR3 antagonist on airway hyperreactivity and M2 receptor dysfunction, J Clin Invest 116(1) (2006) 228–36. [PubMed: 16374515]
- [23]. Kobayashi H, Gleich GJ, Butterfield JH, Kita H, Human eosinophils produce neurotrophins and secrete nerve growth factor on immunologic stimuli, Blood 99(6) (2002) 2214–20. [PubMed: 11877300]
- [24]. Verma AK, Manohar M, Venkateshaiah SU, Blecker U, Collins MH, Mishra A, Role of Vasoactive Intestinal Peptide in Promoting the Pathogenesis of Eosinophilic Esophagitis (EoE), Cell Mol Gastroenterol Hepatol 5(1) (2018) 99–100 e7. [PubMed: 29276755]
- [25]. Gleich GJ, Adolphson CR, Leiferman KM, The biology of the eosinophilic leukocyte, Annu Rev Med 44 (1993) 85–101. [PubMed: 8476270]
- [26]. Acharya KR, Ackerman SJ, Eosinophil granule proteins: form and function, J Biol Chem 289(25) (2014) 17406–15. [PubMed: 24802755]
- [27]. Puxeddu I, Berkman N, Nissim Ben Efraim AH, Davies DE, Ribatti D, Gleich GJ, Levi-Schaffer F, The role of eosinophil major basic protein in angiogenesis, Allergy 64(3) (2009) 368–74. [PubMed: 19120069]
- [28]. Xue A, Wang J, Sieck GC, Wylam ME, Distribution of major basic protein on human airway following in vitro eosinophil incubation, Mediators Inflamm 2010 (2010) 824362. [PubMed: 20339471]
- [29]. Kandikattu HK, Mishra A, Immunomodulatory effects of tacrolimus (FK506) for the treatment of allergic diseases, Int. J Cell Biol. Physiol 1 (2018) 5–13.
- [30]. O'Brien PJ, Peroxidases, Chem Biol Interact 129(1–2) (2000) 113–39. [PubMed: 11154738]
- [31]. Wang J, Slungaard A, Role of eosinophil peroxidase in host defense and disease pathology, Arch Biochem Biophys 445(2) (2006) 256–60. [PubMed: 16297853]
- [32]. Duguet A, Iijima H, Eum SY, Hamid Q, Eidelman DH, Eosinophil peroxidase mediates protein nitration in allergic airway inflammation in mice, Am J Respir Crit Care Med 164(7) (2001) 1119–26. [PubMed: 11673196]
- [33]. Koh GC, Shek LP, Goh DY, Van Bever H, Koh DS, Eosinophil cationic protein: is it useful in asthma? A systematic review, Respir Med 101(4) (2007) 696–705. [PubMed: 17034998]
- [34]. Venge P, Bystrom J, Carlson M, Hakansson L, Karawacjzyk M, Peterson C, Seveus L, Trulson A, Eosinophil cationic protein (ECP): molecular and biological properties and the use of ECP as a marker of eosinophil activation in disease, Clin Exp Allergy 29(9) (1999) 1172–86. [PubMed: 10469025]
- [35]. Rosenberg HF, Dyer KD, Foster PS, Eosinophils: changing perspectives in health and disease, Nat Rev Immunol 13(1) (2013) 9–22. [PubMed: 23154224]
- [36]. Slifman NR, Loegering DA, Mckean DJ, Gleich GJ, Ribonuclease-Activity Associated with Human Eosinophil-Derived Neurotoxin and Eosinophil Cationic Protein, Journal of Immunology 137(9) (1986) 2913–2917.
- [37]. Hosoki K, Nakamura A, Kainuma K, Sugimoto M, Nagao M, Hiraguchi Y, Tanida H, Tokuda R, Wada H, Nobori T, Suga S, Fujisawa T, Differential activation of eosinophils by bacteria

associated with asthma, Int Arch Allergy Immunol 161 Suppl 2 (2013) 16–22. [PubMed: 23711849]

- [38]. Tsuda T, Maeda Y, Nishide M, Koyama S, Hayama Y, Nojima S, Takamatsu H, Okuzaki D, Kinehara Y, Kato Y, Nakatani T, Obata S, Akazawa H, Shikina T, Takeda K, Hayama M, Inohara H, Kumanogoh A, Eosinophil-derived neurotoxin enhances airway remodeling in eosinophilic chronic rhinosinusitis and correlates with disease severity, Int Immunol (2018).
- [39]. Kim HS, Kim JH, Seo YM, Chun YH, Yoon JS, Kim HH, Lee JS, Kim JT, Eosinophil-derived neurotoxin as a biomarker for disease severity and relapse in recalcitrant atopic dermatitis, Ann Allergy Asthma Immunol 119(5) (2017) 441–445. [PubMed: 28866306]
- [40]. Staribratova D, Belovejdov V, Staikov D, Dikov D, Demonstration of Charcot-Leyden crystals in eosinophilic cystitis, Arch Pathol Lab Med 134(10) (2010) 1420. [PubMed: 20923292]
- [41]. De Re V, Simula MP, Cannizzaro R, Pavan A, De Zorzi MA, Toffoli G, Canzonieri V, Galectin-10, eosinophils, and celiac disease, Ann N Y Acad Sci 1173 (2009) 357–64. [PubMed: 19758173]
- [42]. Rodriguez-Alcazar JF, Ataide MA, Engels G, Schmitt-Mabmunyo C, Garbi N, Kastenmuller W, Latz E, Franklin BS, Charcot-Leyden Crystals Activate the NLRP3 Inflammasome and Cause IL-1beta Inflammation in Human Macrophages, J Immunol 202(2) (2019) 550–558. [PubMed: 30559319]
- [43]. Lingblom C, Andersson J, Andersson K, Wenneras C, Regulatory Eosinophils Suppress T Cells Partly through Galectin-10, J Immunol 198(12) (2017) 4672–4681. [PubMed: 28515279]
- [44]. Wang H, Rudd CE, SKAP-55, SKAP-55-related and ADAP adaptors modulate integrin-mediated immune-cell adhesion, Trends Cell Biol 18(10) (2008) 486–93. [PubMed: 18760924]
- [45]. Mishra A, Hogan SP, Brandt EB, Wagner N, Crossman MW, Foster PS, Rothenberg ME, Enterocyte expression of the eotaxin and interleukin-5 transgenes induces compartmentalized dysregulation of eosinophil trafficking, J Biol Chem 277(6) (2002) 4406–12. [PubMed: 11733500]
- [46]. Liao W, Long H, Chang CC, Lu Q, The Eosinophil in Health and Disease: from Bench to Bedside and Back, Clin Rev Allergy Immunol 50(2) (2016) 125–39. [PubMed: 26410377]
- [47]. Shen ZJ, Malter JS, Determinants of eosinophil survival and apoptotic cell death, Apoptosis 20(2) (2015) 224–34. [PubMed: 25563855]
- [48]. Goldsmith KC, Gross M, Peirce S, Luyindula D, Liu X, Vu A, Sliozberg M, Guo R, Zhao H, Reynolds CP, Hogarty MD, Mitochondrial Bcl-2 family dynamics define therapy response and resistance in neuroblastoma, Cancer Res 72(10) (2012) 2565–77. [PubMed: 22589275]
- [49]. Leiferman KM, Beck LA Gleich GJ, Regulation of the production and activation of eosinophils, Rev Immunol 24 (2006) 147–74.
- [50]. Lee SC, Brummet ME, Shahabuddin S, Woodworth TG, Georas SN, Leiferman KM, Gilman SC, Stellato C, Gladue RP, Schleimer RP, Beck LA, Cutaneous injection of human subjects with macrophage inflammatory protein-1 alpha induces significant recruitment of neutrophils and monocytes, J Immunol 164(6) (2000) 3392–401. [PubMed: 10706735]
- [51]. Michael BD, Griffiths MJ, Granerod J, Brown D, Davies NW, Borrow R, Solomon T, Characteristic Cytokine and Chemokine Profiles in Encephalitis of Infectious, Immune-Mediated, and Unknown Aetiology, PLoS One 11(1) (2016) e0146288. [PubMed: 26808276]
- [52]. Giembycz MA, Lindsay MA, Pharmacology of the eosinophil, Pharmacol Rev 51(2) (1999) 213– 340. [PubMed: 10353986]
- [53]. Bonini S, Lambiase A, Bonini S, Angelucci F, Magrini L, Manni L, Aloe L, Circulating nerve growth factor levels are increased in humans with allergic diseases and asthma, Proc Natl Acad Sci U S A 93(20) (1996) 10955–60. [PubMed: 8855290]
- [54]. Broudy VC, Stem cell factor and hematopoiesis, Blood 90(4) (1997) 1345–64. [PubMed: 9269751]
- [55]. Davoine F, Lacy P, Eosinophil cytokines, chemokines, and growth factors: emerging roles in immunity, Front Immunol 5 (2014) 570. [PubMed: 25426119]
- [56]. Hartman M, Piliponsky AM, Temkin V, Levi-Schaffer F, Human peripheral blood eosinophils express stem cell factor, Blood 97(4) (2001) 1086–91. [PubMed: 11159541]

- [57]. Levi-Montalcini R, Skaper SD, Dal Toso R, Petrelli L, Leon A, Nerve growth factor: from neurotrophin to neurokine, Trends Neurosci 19(11) (1996) 514–20. [PubMed: 8931279]
- [58]. Matsuda H, Coughlin MD, Bienenstock J, Denburg JA, Nerve growth factor promotes human hemopoietic colony growth and differentiation, Proc Natl Acad Sci U S A 85(17) (1988) 6508– 12. [PubMed: 3413109]
- [59]. Minshall EM, Leung DY, Martin RJ, Song YL, Cameron L, Ernst P, Hamid Q, Eosinophilassociated TGF-beta1 mRNA expression and airways fibrosis in bronchial asthma, Am J Respir Cell Mol Biol 17(3) (1997) 326–33. [PubMed: 9308919]
- [60]. Powell PP, Klagsbrun M, Abraham JA, Jones RC, Eosinophils expressing heparin-binding EGFlike growth factor mRNA localize around lung microvessels in pulmonary hypertension, Am J Pathol 143(3) (1993) 784–93. [PubMed: 8362977]
- [61]. Solomon A, Aloe L, Pe'er J, Frucht-Pery J, Bonini S, Bonini S, Levi-Schaffer F, Nerve growth factor is preformed in and activates human peripheral blood eosinophils, Journal of Allergy and Clinical Immunology 102(3) (1998) 454–460. [PubMed: 9768588]
- [62]. Wong DTW, Weller PF, Galli SJ, Elovic A, Rand TH, Gallagher GT, Chiang T, Chou MY, Matossian K, Mcbride J, Todd R, Human Eosinophils Express Transforming Growth Factor-Alpha, Journal of Experimental Medicine 172(3) (1990) 673–681. [PubMed: 1696954]
- [63]. Feltenmark S, Gautam N, Brunnstrom A, Griffiths W, Backman L, Edenius C, Lindbom L, Bjorkholm M, Claesson HE, Eoxins are proinflammatory arachidonic acid metabolites produced via the 15-lipoxygenase-1 pathway in human eosinophils and mast cells, Proc Natl Acad Sci U S A 105(2) (2008) 680–5. [PubMed: 18184802]
- [64]. Kvarnhammar AM, Cardell LO, Pattern-recognition receptors in human eosinophils, Immunology 136(1) (2012) 11–20. [PubMed: 22242941]
- [65]. Stahle-Backdahl M, Parks WC, 92-kd gelatinase is actively expressed by eosinophils and stored by neutrophils in squamous cell carcinoma, Am J Pathol 142(4) (1993) 995–1000. [PubMed: 7682768]
- [66]. Yagisawa M, Yuo A, Yonemaru M, ImajohOhmi S, Kanegasaki S, Yazaki Y, Takaku F, Superoxide release and NADPH oxidase components in mature human phagocytes: Correlation between functional capacity and amount of functional proteins, Biochemical and Biophysical Research Communications 228(2) (1996) 510–516. [PubMed: 8920944]
- [67]. Varricchi G, Bagnasco D, Borriello F, Heffler E, Canonica GW, Interleukin-5 pathway inhibition in the treatment of eosinophilic respiratory disorders: evidence and unmet needs, Curr Opin Allergy Clin Immunol 16(2) (2016) 186–200. [PubMed: 26859368]
- [68]. Adachi T, Alam R, The mechanism of IL-5 signal transduction, Am J Physiol 275(3 Pt 1) (1998) C623–33. [PubMed: 9730944]
- [69]. Stein ML, Munitz A, Targeting interleukin (IL) 5 for asthma and hypereosinophilic diseases, Recent Pat Inflamm Allergy Drug Discov 4(3) (2010) 201–9. [PubMed: 20807192]
- [70]. Kouro T, Kikuchi Y, Kanazawa H, Hirokawa K, Harada N, Shiiba M, Wakao H, Takaki S, Takatsu K, Critical proline residues of the cytoplasmic domain of the IL-5 receptor alpha chain and its function in IL-5-mediated activation of JAK kinase and STAT5, International Immunology 8(2) (1996) 237–245. [PubMed: 8671609]
- [71]. Ogata N, Kouro T, Yamada A, Koike M, Hanai N, Ishikawa T, Takatsu K, JAK2 and JAK1 constitutively associate with an interleukin-5 (IL-5) receptor alpha and beta c subunit, respectively, and are activated upon IL-5 stimulation, Blood 91(7) (1998) 2264–2271. [PubMed: 9516124]
- [72]. Pazdrak K, Adachi T, Alam R, Src homology 2 protein tyrosine phosphatase (SHPTP2)/Src homology 2 phosphatase 2 (SHP2) tyrosine phosphatase is a positive regulator of the interleukin 5 receptor signal transduction pathways leading to the prolongation of eosinophil survival, J Exp Med 186(4) (1997) 561–8. [PubMed: 9254654]
- [73]. Pazdrak K, Olszewska-Pazdrak B, Stafford S, Garofalo RP, Alam R, Lyn, Jak2, and Raf-1 kinases are critical for the antiapoptotic effect of interleukin 5, whereas only Raf-1 kinase is essential for eosinophil activation and degranulation, Journal of Experimental Medicine 188(3) (1998) 421– 429. [PubMed: 9687520]

- [74]. Sato N, Sakamaki K, Terada N, Arai K, Miyajima A, Signal transduction by the high-affinity GM-CSF receptor: two distinct cytoplasmic regions of the common beta subunit responsible for different signaling, EMBO J 12(11) (1993) 4181–9. [PubMed: 8223433]
- [75]. Sato S, Katagiri T, Takaki S, Kikuchi Y, Hitoshi Y, Yonehara S, Tsukada S, Kitamura D, Watanabe T, Witte O, Takatsu K, Il-5 Receptor-Mediated Tyrosine Phosphorylation of Sh2/Shs-Containing Proteins and Activation of Bruton Tyrosine and Janus-2 Kinases, Journal of Experimental Medicine 180(6) (1994) 2101–2111. [PubMed: 7525847]
- [76]. Takaki S, Kanazawa H, Shiiba M, Takatsu K, A Critical Cytoplasmic Domain of the Interleukin-5 (Il-5) Receptor-Alpha Chain and Its Function in Il-5-Mediated Growth Signal-Transduction, Molecular and Cellular Biology 14(11) (1994) 7404–7413. [PubMed: 7935454]
- [77]. Horikawa K, Kaku H, Nakajima H, Davey HW, Hennighausen L, Iwamoto I, Yasue T, Kariyone A, Takatsu K, Essential role of Stat5 for IL-5-dependent IgH switch recombination in mouse B cells, J Immunol 167(9) (2001) 5018–26. [PubMed: 11673510]
- [78]. Kagami S, Nakajima H, Suzuki K, Saito Y, Takatsu K, Leonard W, Iwamoto I, Both Stat5a and Stat5b are required for antigen-induced eosinophil and T cell recruitment into the tissue, Journal of Allergy and Clinical Immunology 105(1) (2000) S355–S355.
- [79]. Hall DJ, Cui J, Bates ME, Stout BA, Koenderman L, Coffer PJ, Bertics PJ, Transduction of a dominant-negative H-Ras into human eosinophils attenuates extracellular signal-regulated kinase activation and interleukin-5-mediated cell viability, Blood 98(7) (2001) 2014–21. [PubMed: 11567984]
- [80]. Chung KF, p38 mitogen-activated protein kinase pathways in asthma and COPD, Chest 139(6) (2011) 1470–1479. [PubMed: 21652557]
- [81]. Park YM, Bochner BS, Eosinophil survival and apoptosis in health and disease, Allergy Asthma Immunol Res 2(2) (2010) 87–101. [PubMed: 20358022]
- [82]. Kankaanranta H, De Souza PM, Barnes PJ, Salmon M, Giembycz MA, Lindsay MA, SB 203580, an inhibitor of p38 mitogen-activated protein kinase, enhances constitutive apoptosis of cytokinedeprived human eosinophils, J Pharmacol Exp Ther 290(2) (1999) 621–8. [PubMed: 10411570]
- [83]. Underwood DC, Osborn RR, Kotzer CJ, Adams JL, Lee JC, Webb EF, Carpenter DC, Bochnowicz S, Thomas HC, Hay DW, Griswold DE, SB 239063, a potent p38 MAP kinase inhibitor, reduces inflammatory cytokine production, airways eosinophil infiltration, and persistence, J Pharmacol Exp Ther 293(1) (2000) 281–8. [PubMed: 10734180]
- [84]. Amruta N, Kandikattu HK, Apoptosis of inflammatory cells in Asthma, Int J Cell Biol Physiol 1 (2018) 1–6.
- [85]. Schwartz C, Willebrand R, Huber S, Rupec RA, Wu D, Locksley R, Voehringer D, Eosinophilspecific deletion of IkappaBalpha in mice reveals a critical role of NF-kappaB-induced Bcl-xL for inhibition of apoptosis, Blood 125(25) (2015) 3896–904. [PubMed: 25862560]
- [86]. Dinarello CA, Novick D, Kim S, Kaplanski G, Interleukin-18 and IL-18 binding protein, Front Immunol 4 (2013)289. [PubMed: 24115947]
- [87]. Dinarello CA, Interleukin-18, Methods 19(1) (1999) 121–32. [PubMed: 10525448]
- [88]. Mathur SK, Sedgwick JB, Vrtis RF et al.,, Human eosinophils express IL-18 mRNA and protein, J. Allergy Clin. Immunol 113 (2004) S166–7.
- [89]. Nakanishi K, Unique Action of Interleukin-18 on T Cells and Other Immune Cells, Front Immunol 9 (2018) 763. [PubMed: 29731751]
- [90]. Sandersa NL, Venkateshaiah SU, Manohar M, Verma AK, Kandikattu HK, Mishra A, Interleukin-18 has an Important Role in Differentiation and Maturation of Mucosal Mast Cells, J Mucosal Immunol Res 2(1) (2018).
- [91]. Dostert C, Petrilli V, Van Bruggen R, Steele C, Mossman BT, Tschopp J, Innate immune activation through Nalp3 inflammasome sensing of asbestos and silica, Science 320(5876) (2008) 674–7. [PubMed: 18403674]
- [92]. Gasse P, Mary C, Guenon I, Noulin N, Charron S, Schnyder-Candrian S, Schnyder B, Akira S, Quesniaux VF, Lagente V, Ryffel B, Couillin I, IL-1R1/MyD88 signaling and the inflammasome are essential in pulmonary inflammation and fibrosis in mice, J Clin Invest 117(12) (2007) 3786– 99. [PubMed: 17992263]

- [93]. Hornung V, Bauernfeind F, Halle A, Samstad EO, Kono H, Rock KL, Fitzgerald KA, Latz E, Silica crystals and aluminum salts activate the NALP3 inflammasome through phagosomal destabilization, Nat Immunol 9(8) (2008) 847–56. [PubMed: 18604214]
- [94]. Xu JF, Washko GR, Nakahira K, Hatabu H, Patel AS, Fernandez IE, Nishino M, Okajima Y, Yamashiro T, Ross JC, Estepar RS, Diaz AA, Li HP, Qu JM, Himes BE, Come CE, D'Aco K, Martinez FJ, Han MK, Lynch DA, Crapo JD, Morse D, Ryter SW, Silverman EK, Rosas IO, Choi AM, Hunninghake GM, C.O. Investigators, Statins and pulmonary fibrosis: the potential role of NLRP3 inflammasome activation, Am J Respir Crit Care Med 185(5) (2012) 547–56. [PubMed: 22246178]
- [95]. dos Santos G, Kutuzov MA, Ridge KM, The inflammasome in lung diseases, Am J Physiol Lung Cell Mol Physiol 303(8) (2012) L627–33. [PubMed: 22904168]
- [96]. Yang W, Ni H, Wang H, Gu H, NLRP3 inflammasome is essential for the development of chronic obstructive pulmonary disease, Int J Clin Exp Pathol 8(10) (2015) 13209–16. [PubMed: 26722520]
- [97]. Ogura T, Ueda H, Hosohara K, Tsuji R, Nagata Y, Kashiwamura S, Okamura H, Interleukin-18 stimulates hematopoietic cytokine and growth factor formation and augments circulating granulocytes in mice, Blood 98(7) (2001) 2101–7. [PubMed: 11567996]
- [98]. Mesnil C, Raulier S, Paulissen G, Xiao X, Birrell MA, Pirottin D, Janss T, Starkl P, Ramery E, Henket M, Schleich FN, Radermecker M, Thielemans K, Gillet L, Thiry M, Belvisi MG, Louis R, Desmet C, Marichal T, Bureau F, Lung-resident eosinophils represent a distinct regulatory eosinophil subset, J Clin Invest 126(9) (2016) 3279–95. [PubMed: 27548519]
- [99]. Kumano K, Nakao A, Nakajima H, Hayashi F, Kurimoto M, Okamura H, Saito Y, Iwamoto I, Interleukin-18 enhances antigen-induced eosinophil recruitment into the mouse airways, Am J Respir Crit Care Med 160(3) (1999) 873–8. [PubMed: 10471611]
- [100]. Gatault S, Delbeke M, Driss V, Sarazin A, Dendooven A, Kahn JE, Lefevre G, Capron M, IL-18 Is Involved in Eosinophil-Mediated Tumoricidal Activity against a Colon Carcinoma Cell Line by Upregulating LFA-1 and ICAM-1, J Immunol 195(5) (2015) 2483–92. [PubMed: 26216891]
- [101]. Niranjan R, Rajavelu P, Ventateshaiah SU, Shukla JS, Zaidi A, Mariswamy SJ, Mattner J, Fortgang I, Kowalczyk M, Balart L, Shukla A, Mishra A, Involvement of interleukin-18 in the pathogenesis of human eosinophilic esophagitis, Clin Immunol 157(2) (2015) 103–13. [PubMed: 25638412]
- [102]. Dutt P, Shukla JS, Ventateshaiah SU, Mariswamy SJ, Mattner J, Shukla A, Mishra A, Allergeninduced interleukin-18 promotes experimental eosinophilic oesophagitis in mice, Immunol Cell Biol 93(10) (2015) 849–57. [PubMed: 25801352]
- [103]. Sawada M, Kawayama T, Imaoka H, Sakazaki Y, Oda H, Takenaka S, Kaku Y, Azuma K, Tajiri M, Edakuni N, Okamoto M, Kato S, Hoshino T, IL-18 induces airway hyperresponsiveness and pulmonary inflammation via CD4+ T cell and IL-13, PLoS One 8(1) (2013) e54623. [PubMed: 23382928]
- [104]. Brusselle GG, Maes T, Bracke KR, Eosinophils in the spotlight: Eosinophilic airway inflammation in nonallergic asthma, Nat Med 19(8) (2013) 977–9. [PubMed: 23921745]
- [105]. Tanaka H, Komai M, Nagao K, Ishizaki M, Kajiwara D, Takatsu K, Delespesse G, Nagai H, Role of interleukin-5 and eosinophils in allergen-induced airway remodeling in mice, Am J Respir Cell Mol Biol 31(1) (2004) 62–8. [PubMed: 14975941]
- [106]. Turkeli A, Yilmaz O, Taneli F, Horasan GD, Kanik ET, Kizilkaya M, Gozukara C, Yuksel H, IL-5, IL-8 and MMP −9 levels in exhaled breath condensate of atopic and nonatopic asthmatic children, Respir Med 109(6) (2015) 680–8. [PubMed: 25937050]
- [107]. Kuroda-Morimoto M, Tanaka H, Hayashi N, Nakahira M, Imai Y, Imamura M, Yasuda K, Yumikura-Futatsugi S, Matsui K, Nakashima T, Sugimura K, Tsutsui H, Sano H, Nakanishi K, Contribution of IL-18 to eosinophilic airway inflammation induced by immunization and challenge with Staphylococcus aureus proteins, Int Immunol 22(7) (2010) 561–70. [PubMed: 20497957]
- [108]. Shamri R, Xenakis JJ, Spencer LA, Eosinophils in innate immunity: an evolving story, Cell and Tissue Research 343(1) (2011) 57–83. [PubMed: 21042920]

- [109]. Shenoy NG, Gleich GJ, Thomas LL, Eosinophil major basic protein stimulates neutrophil superoxide production by a class IA phosphoinositide 3-kinase and protein kinase C-zetadependent pathway, J Immunol 171(7) (2003) 3734–41. [PubMed: 14500673]
- [110]. Moy JN, Thomas LL, Whisler LC, Eosinophil major basic protein enhances the expression of neutrophil CR3 and p150,95, J Allergy Clin Immunol 92(4) (1993) 598–606. [PubMed: 8409119]
- [111]. Kikuchi I, Kikuchi S, Kobayashi T, Hagiwara K, Sakamoto Y, Kanazawa M, Nagata M, Eosinophil trans-basement membrane migration induced by interleukin-8 and neutrophils, Am J Respir Cell Mol Biol 34(6) (2006) 760–5. [PubMed: 16456187]
- [112]. Moreira AP, Hogaboam CM, Macrophages in allergic asthma: fine-tuning their pro- and antiinflammatory actions for disease resolution, J Interferon Cytokine Res 31(6) (2011) 485–91. [PubMed: 21631355]
- [113]. Murray PJ, Wynn TA, Protective and pathogenic functions of macrophage subsets, Nat Rev Immunol 11(11) (2011) 723–37. [PubMed: 21997792]
- [114]. Bang BR, Chun E, Shim EJ, Lee HS, Lee SY, Cho SH, Min KU, Kim YY, Park HW, Alveolar macrophages modulate allergic inflammation in a murine model of asthma, Exp Mol Med 43(5) (2011) 275–80. [PubMed: 21415590]
- [115]. Stolarski B, Kurowska-Stolarska M, Kewin P, Xu D, Liew FY, IL-33 exacerbates eosinophilmediated airway inflammation, J Immunol 185(6) (2010) 3472–80. [PubMed: 20693421]
- [116]. Elishmereni M, Alenius HT, Bradding P, Mizrahi S, Shikotra A, Minai-Fleminger Y, Mankuta D, Eliashar R, Zabucchi G, Levi-Schaffer F, Physical interactions between mast cells and eosinophils: a novel mechanism enhancing eosinophil survival in vitro, Allergy 66(3) (2011) 376–85. [PubMed: 20977491]
- [117]. Niranjan R, Mavi P, Rayapudi M, Dynda S, Mishra A, Pathogenic role of mast cells in experimental eosinophilic esophagitis, Am J Physiol Gastrointest Liver Physiol 304(12) (2013) G1087–94. [PubMed: 23599040]
- [118]. Abonia JP, Blanchard C, Butz BB, Rainey HF, Collins MH, Stringer K, Putnam PE, Rothenberg ME, Involvement of mast cells in eosinophilic esophagitis, J Allergy Clin Immunol 126(1) (2010) 140–9. [PubMed: 20538331]
- [119]. Burke SM, Issekutz TB, Mohan K, Lee PW, Shmulevitz M, Marshall JS, Human mast cell activation with virus-associated stimuli leads to the selective chemotaxis of natural killer cells by a CXCL8-dependent mechanism, Blood 111(12) (2008) 5467–76. [PubMed: 18424663]
- [120]. Cho KA, Suh JW, Sohn JH, Park JW, Lee H, Kang JL, Woo SY, Cho YJ, IL-33 induces Th17 mediated airway inflammation via mast cells in ovalbumin-challenged mice, Am J Physiol Lung Cell Mol Physiol 302(4) (2012) L429–40. [PubMed: 22180658]
- [121]. Otsuka A, Nakajima S, Kubo M, Egawa G, Honda T, Kitoh A, Nomura T, Hanakawa S, Sagita Moniaga C, Kim B, Matsuoka S, Watanabe T, Miyachi Y, Kabashima K, Basophils are required for the induction of Th2 immunity to haptens and peptide antigens, Nat Commun 4 (2013) 1739.
- [122]. Perrigoue JG, Saenz SA, Siracusa MC, Allenspach EJ, Taylor BC, Giacomin PR, Nair MG, Du Y, Zaph C, van Rooijen N, Comeau MR, Pearce EJ, Laufer TM, Artis D, MHC class IIdependent basophil-CD4+ T cell interactions promote T(H)2 cytokine-dependent immunity, Nat Immunol 10(7) (2009) 697–705. [PubMed: 19465906]
- [123]. Nakashima C, Otsuka A, Kitoh A, Honda T, Egawa G, Nakajima S, Nakamizo S, Arita M, Kubo M, Miyachi Y, Kabashima K, Basophils regulate the recruitment of eosinophils in a murine model of irritant contact dermatitis, J Allergy Clin Immunol 134(1) (2014) 100–7. [PubMed: 24713170]
- [124]. Gill MA, The role of dendritic cells in asthma, J Allergy Clin Immunol 129(4) (2012) 889–901. [PubMed: 22464668]
- [125]. Yi S, Zhai J, Niu R, Zhu G, Wang M, Liu J, Huang H, Wang Y, Jing X, Kang L, Song W, Shi Y, Tang H, Eosinophil recruitment is dynamically regulated by interplay among lung dendritic cell subsets after allergen challenge, Nat Commun 9(1) (2018) 3879. [PubMed: 30250029]
- [126]. Porter P, Susarla SC, Polikepahad S, Qian Y, Hampton J, Kiss A, Vaidya S, Sur S, Ongeri V, Yang T, Delclos GL, Abramson S, Kheradmand F, Corry DB, Link between allergic asthma and

airway mucosal infection suggested by proteinase-secreting household fungi, Mucosal Immunol 2(6) (2009) 504–17. [PubMed: 19710638]

- [127]. Chu VT, Berek C, Immunization induces activation of bone marrow eosinophils required for plasma cell survival, Eur J Immunol 42(1) (2012) 130–7. [PubMed: 22057654]
- [128]. Chu VT, Frohlich A, Steinhauser G, Scheel T, Roch T, Fillatreau S, Lee JJ, Lohning M, Berek C, Eosinophils are required for the maintenance of plasma cells in the bone marrow, Nat Immunol 12(2) (2011) 151–9. [PubMed: 21217761]
- [129]. Wong TW, Kita H, Hanson CA, Walters DK, Arendt BK, Jelinek DF, Induction of malignant plasma cell proliferation by eosinophils, PLoS One 8(7) (2013) e70554. [PubMed: 23894671]
- [130]. Cosmi L, Liotta F, Maggi E, Romagnani S, Annunziato F, Th17 cells: new players in asthma pathogenesis, Allergy 66(8) (2011) 989–98. [PubMed: 21375540]
- [131]. Kandikattu HK, Rachitha P, Jayashree GV, Krupashree K, Sukhith M, Majid A, Amruta N, Khanum F, Anti-inflammatory and anti-oxidant effects of Cardamom (Elettaria repens (Sonn.) Baill) and its phytochemical analysis by 4D GCXGC TOF-MS, Biomed Pharmacother 91 (2017) 191–201. [PubMed: 28458157]
- [132]. Jia L, Wu C, The biology and functions of Th22 cells, Adv Exp Med Biol 841 (2014) 209–30. [PubMed: 25261209]
- [133]. Arend WP, Palmer G, Gabay C, IL-1, IL-18, and IL-33 families of cytokines, Immunol Rev 223 (2008) 20–38. [PubMed: 18613828]
- [134]. Zhang YN, Song J, Wang H, Wang H, Zeng M, Zhai GT, Ma J, Li ZY, Liao B, Wang BF, Zhen Z, Wang N, Cao PP, Lin P, Ning Q, Liu Z, Nasal IL-4(+)CXCR5(+)CD4(+) T follicular helper cell counts correlate with local IgE production in eosinophilic nasal polyps, J Allergy Clin Immunol 137(2) (2016) 462–73. [PubMed: 26329514]
- [135]. Noval Rivas M, Burton OT, Oettgen HC, Chatila T, IL-4 production by group 2 innate lymphoid cells promotes food allergy by blocking regulatory T-cell function, J Allergy Clin Immunol 138(3) (2016) 801–811 e9. [PubMed: 27177780]
- [136]. Hussain M, Borcard L, Walsh KP, Pena Rodriguez M, Mueller C, Kim BS, Kubo M, Artis D, Noti M, Basophil-derived IL-4 promotes epicutaneous antigen sensitization concomitant with the development of food allergy, J Allergy Clin Immunol 141(1) (2018) 223–234 e5. [PubMed: 28390860]
- [137]. Diny NL, Rose NR, Cihakova D, Eosinophils in Autoimmune Diseases, Frontiers in Immunology 8 (2017).
- [138]. Steenwinckel V, Louahed J, Lemaire MM, Sommereyns C, Warnier G, McKenzie A, Brombacher F, Van Snick J, Renauld JC, IL-9 promotes IL-13-dependent paneth cell hyperplasia and up-regulation of innate immunity mediators in intestinal mucosa, J Immunol 182(8) (2009) 4737–43. [PubMed: 19342650]
- [139]. Kearley J, Erjefalt JS, Andersson C, Benjamin E, Jones CP, Robichaud A, Pegorier S, Brewah Y, Burwell TJ, Bjermer L, Kiener PA, Kolbeck R, Lloyd CM, Coyle AJ, Humbles AA, IL-9 governs allergen-induced mast cell numbers in the lung and chronic remodeling of the airways, Am J Respir Crit Care Med 183(7) (2011) 865–75. [PubMed: 20971830]
- [140]. Chen CY, Lee JB, Liu B, Ohta S, Wang PY, Kartashov AV, Mugge L, Abonia JP, Barski A, Izuhara K, Rothenberg ME, Finkelman FD, Hogan SP, Wang YH, Induction of Interleukin-9- Producing Mucosal Mast Cells Promotes Susceptibility to IgE-Mediated Experimental Food Allergy, Immunity 43(4) (2015) 788–802. [PubMed: 26410628]
- [141]. Liu J, Harberts E, Tammaro A, Girardi N, Filler RB, Fishelevich R, Temann A, Licona-Limon P, Girardi M, Flavell RA, Gaspari AA, IL-9 regulates allergen-specific Th1 responses in allergic contact dermatitis, J Invest Dermatol 134(7) (2014) 1903–1911. [PubMed: 24487305]
- [142]. Koga C, Kabashima K, Shiraishi N, Kobayashi M, Tokura Y, Possible pathogenic role of Th17 cells for atopic dermatitis, J Invest Dermatol 128(11) (2008) 2625–30. [PubMed: 18432274]
- [143]. Murdock BJ, Falkowski NR, Shreiner AB, Sadighi Akha AA, McDonald RA, White ES, Toews GB, Huffnagle GB, Interleukin-17 drives pulmonary eosinophilia following repeated exposure to Aspergillus fumigatus conidia, Infect Immun 80(4) (2012) 1424–36. [PubMed: 22252873]
- [144]. Newcomb DC, Boswell MG, Reiss S, Zhou W, Goleniewska K, Toki S, Harintho MT, Lukacs NW, Kolls JK, Peebles RS Jr., IL-17A inhibits airway reactivity induced by respiratory syncytial

virus infection during allergic airway inflammation, Thorax 68(8) (2013) 717–23. [PubMed: 23422214]

- [145]. Akdis M, Burgler S, Crameri R, Eiwegger T, Fujita H, Gomez E, Klunker S, Meyer N, O'Mahony L, Palomares O, Rhyner C, Ouaked N, Schaffartzik A, Van De Veen W, Zeller S, Zimmermann M, Akdis CA, Interleukins, from 1 to 37, and interferon-gamma: receptors, functions, and roles in diseases, J Allergy Clin Immunol 127(3) (2011) 701–21 e1–70. [PubMed: 21377040]
- [146]. Mizutani H, Tamagawa-Mineoka R, Nakamura N, Masuda K, Katoh N, Serum IL-21 levels are elevated in atopic dermatitis patients with acute skin lesions, Allergol Int 66(3) (2017) 440–444. [PubMed: 27884624]
- [147]. Hiromura Y, Kishida T, Nakano H, Hama T, Imanishi J, Hisa Y, Mazda O, IL-21 administration into the nostril alleviates murine allergic rhinitis, J Immunol 179(10) (2007) 7157–65. [PubMed: 17982108]
- [148]. Taube C, Tertilt C, Gyulveszi G, Dehzad N, Kreymborg K, Schneeweiss K, Michel E, Reuter S, Renauld JC, Arnold-Schild D, Schild H, Buhl R, Becher B, IL-22 is produced by innate lymphoid cells and limits inflammation in allergic airway disease, PLoS One 6(7) (2011) e21799. [PubMed: 21789181]
- [149]. Nograles KE, Zaba LC, Shemer A, Fuentes-Duculan J, Cardinale I, Kikuchi T, Ramon M, Bergman R, Krueger JG, Guttman-Yassky E, IL-22-producing "T22" T cells account for upregulated IL-22 in atopic dermatitis despite reduced IL-17-producing TH17 T cells, J Allergy Clin Immunol 123(6) (2009) 1244–52 e2. [PubMed: 19439349]
- [150]. Besnard AG, Sabat R, Dumoutier L, Renauld JC, Willart M, Lambrecht B, Teixeira MM, Charron S, Fick L, Erard F, Warszawska K, Wolk K, Quesniaux V, Ryffel B, Togbe D, Dual Role of IL-22 in allergic airway inflammation and its cross-talk with IL-17A, Am J Respir Crit Care Med 183(9) (2011) 1153–63. [PubMed: 21297073]
- [151]. Farfariello V, Amantini C, Nabissi M, Morelli MB, Aperio C, Caprodossi S, Carlucci A, Bianchi AM, Santoni G, IL-22 mRNA in peripheral blood mononuclear cells from allergic rhinitic and asthmatic pediatric patients, Pediatr Allergy Immunol 22(4) (2011) 419–23. [PubMed: 21535180]
- [152]. Gessner MA, Werner JL, Lilly LM, Nelson MP, Metz AE, Dunaway CW, Chan YR, Ouyang W, Brown GD, Weaver CT, Steele C, Dectin-1-dependent interleukin-22 contributes to early innate lung defense against Aspergillus fumigatus, Infect Immun 80(1) (2012) 410–7. [PubMed: 22038916]
- [153]. Cheng D, Xue Z, Yi L, Shi H, Zhang K, Huo X, Bonser LR, Zhao J, Xu Y, Erle DJ, Zhen G, Epithelial interleukin-25 is a key mediator in Th2-high, corticosteroid-responsive asthma, Am J Respir Crit Care Med 190(6) (2014) 639–48. [PubMed: 25133876]
- [154]. Gregory LG, Jones CP, Walker SA, Sawant D, Gowers KH, Campbell GA, McKenzie AN, Lloyd CM, IL-25 drives remodelling in allergic airways disease induced by house dust mite, Thorax 68(1) (2013) 82–90. [PubMed: 23093652]
- [155]. Cayrol C, Girard JP, IL-33: an alarmin cytokine with crucial roles in innate immunity, inflammation and allergy, Curr Opin Immunol 31 (2014) 31–7. [PubMed: 25278425]
- [156]. Bartemes KR, Iijima K, Kobayashi T, Kephart GM, McKenzie AN, Kita H, IL-33-responsive lineage- CD25+ CD44(hi) lymphoid cells mediate innate type 2 immunity and allergic inflammation in the lungs, J Immunol 188(3) (2012) 1503–13. [PubMed: 22198948]
- [157]. Fattouh R, Al-Garawi A, Fattouh M, Arias K, Walker TD, Goncharova S, Coyle AJ, Humbles AA, Jordana M, Eosinophils are dispensable for allergic remodeling and immunity in a model of house dust mite-induced airway disease, Am J Respir Crit Care Med 183(2) (2011) 179–88. [PubMed: 20732990]
- [158]. Humbles AA, Lloyd CM, McMillan SJ, Friend DS, Xanthou G, McKenna EE, Ghiran S, Gerard NP, Yu C, Orkin SH, Gerard C, A critical role for eosinophils in allergic airways remodeling, Science 305(5691) (2004) 1776–9. [PubMed: 15375268]
- [159]. Lloyd CM, Saglani S, Eosinophils in the spotlight: Finding the link between obesity and asthma, Nat Med 19(8) (2013) 976–7. [PubMed: 23921744]

- [160]. Haldar P, Pavord ID, Shaw DE, Berry MA, Thomas M, Brightling CE, Wardlaw AJ, Green RH, Cluster analysis and clinical asthma phenotypes, Am J Respir Crit Care Med 178(3) (2008) 218– 224. [PubMed: 18480428]
- [161]. Giuffrida MJ, Valero N, Mosquera J, Duran A, Arocha F, Chacin B, Espina LM, Gotera J, Bermudez J, Mavarez A, Alvarez-Mon M, Increased Systemic Cytokine/Chemokine Expression in Asthmatic and Non-asthmatic Patients with Bacterial, Viral or Mixed Lung Infection, Scand J Immunol 85(4) (2017) 280–290. [PubMed: 28168862]
- [162]. Tanaka H, Miyazaki N, Oashi K, Teramoto S, Shiratori M, Hashimoto M, Ohmichi M, Abe S, IL-18 might reflect disease activity in mild and moderate asthma exacerbation, J Allergy Clin Immunol 107(2) (2001) 331–6. [PubMed: 11174201]
- [163]. Wallach D, Taieb A, Atopic dermatitis/atopic eczema, Chem Immunol Allergy 100 (2014) 81– 96. [PubMed: 24925387]
- [164]. Werfel T, Allam JP, Biedermann T, Eyerich K, Gilles S, Guttman-Yassky E, Hoetzenecker W, Knol E, Simon HU, Wollenberg A, Bieber T, Lauener R, Schmid-Grendelmeier P, Traidl-Hoffmann C, Akdis CA, Cellular and molecular immunologic mechanisms in patients with atopic dermatitis, J Allergy Clin Immunol 138(2) (2016) 336–49. [PubMed: 27497276]
- [165]. Celakovska J, Bukac J, Ettler K, Vaneckova J, Krcmova I, Ettlerova K, Krejsek J, Evaluation of Peripheral Blood Eosinophilia in Adolescent and Adult Patients Suffering from Atopic Dermatitis and the Relation to the Occurrence of Allergy to Aeroallergens, Indian Journal of Dermatology 64(1) (2019) 34–40. [PubMed: 30745633]
- [166]. Rasheed Z, Zedan K, Saif GB, Salama RH, Salem T, Ahmed AA, El-Moniem AA, Elkholy M, Al Robaee AA, Alzolibani AA, Markers of atopic dermatitis, allergic rhinitis and bronchial asthma in pediatric patients: correlation with filaggrin, eosinophil major basic protein and immunoglobulin E, Clin Mol Allergy 16 (2018)23. [PubMed: 30473631]
- [167]. Hall A, Atopic Dermatitis (Atopic Eczema), Atlas of Male Genital Dermatology (2019) 41–43.
- [168]. Kandikattu HKM, A., Oxido-nitrosative stress and antioxidants in asthma, Int J Basic Clin Immonol 1(9–12) (2018).
- [169]. Kalesnikoff J, Galli SJ, Anaphylaxis: mechanisms of mast cell activation, Chem Immunol Allergy 95 (2010)45–66. [PubMed: 20519881]
- [170]. Dhami S, Panesar SS, Roberts G, Muraro A, Worm M, Bilo MB, Cardona V, Dubois AE, DunnGalvin A, Eigenmann P, Fernandez-Rivas M, Halken S, Lack G, Niggemann B, Rueff F, Santos AF, Vlieg-Boerstra B, Zolkipli ZQ, Sheikh A, Allergy EF, Anaphylaxis Guidelines G, Management of anaphylaxis: a systematic review, Allergy 69(2) (2014) 168–75. [PubMed: 24251536]
- [171]. Bousquet J, Khaltaev N, Cruz AA, Denburg J, Fokkens WJ, Togias A, Zuberbier T, Baena-Cagnani CE, Canonica GW, van Weel C, Agache I, Ait-Khaled N, Bachert C, Blaiss MS, Bonini S, Boulet LP, Bousquet PJ, Camargos P, Carlsen KH, Chen Y, Custovic A, Dahl R, Demoly P, Douagui H, Durham SR, van Wijk RG, Kalayci O, Kaliner MA, Kim YY, Kowalski ML, Kuna P, Le LT, Lemiere C, Li J, Lockey RF, Mavale-Manuel S, Meltzer EO, Mohammad Y, Mullol J, Naclerio R, O'Hehir RE, Ohta K, Ouedraogo S, Palkonen S, Papadopoulos N, Passalacqua G, Pawankar R, Popov TA, Rabe KF, Rosado-Pinto J, Scadding GK, Simons FE, Toskala E, Valovirta E, van Cauwenberge P, Wang DY, Wickman M, Yawn BP, Yorgancioglu A, Yusuf OM, Zar H, Annesi-Maesano I, Bateman ED, Ben Kheder A, Boakye DA, Bouchard J, Burney P, Busse WW, Chan-Yeung M, Chavannes NH, Chuchalin A, Dolen WK, Emuzyte R, Grouse L, Humbert M, Jackson C, Johnston SL, Keith PK, Kemp JP, Klossek JM, Larenas-Linnemann D, Lipworth B, Malo JL, Marshall GD, Naspitz C, Nekam K, Niggemann B, Nizankowska-Mogilnicka E, Okamoto Y, Orru MP, Potter P, Price D, Stoloff SW, Vandenplas O, Viegi G, Williams D, O. World Health, Galen, AllerGen, Allergic Rhinitis and its Impact on Asthma (ARIA) 2008 update (in collaboration with the World Health Organization, GA(2)LEN and AllerGen), Allergy 63 Suppl 86 (2008) 8–160. [PubMed: 18331513]
- [172]. Bousquet J, Van Cauwenberge P, Khaltaev N, Aria Workshop G, O. World Health, Allergic rhinitis and its impact on asthma, J Allergy Clin Immunol 108(5 Suppl) (2001) S147–334. [PubMed: 11707753]
- [173]. Kato A, Schleimer RP, Beyond inflammation: airway epithelial cells are at the interface of innate and adaptive immunity, Curr Opin Immunol 19(6) (2007) 711–20. [PubMed: 17928212]

- [174]. Miyata M, Hatsushika K, Ando T, Shimokawa N, Ohnuma Y, Katoh R, Suto H, Ogawa H, Masuyama K, Nakao A, Mast cell regulation of epithelial TSLP expression plays an important role in the development of allergic rhinitis, Eur J Immunol 38(6) (2008) 1487–92. [PubMed: 18461563]
- [175]. Nakamura Y, Miyata M, Ohba T, Ando T, Hatsushika K, Suenaga F, Shimokawa N, Ohnuma Y, Katoh R, Ogawa H, Nakao A, Cigarette smoke extract induces thymic stromal lymphopoietin expression, leading to T(H)2-type immune responses and airway inflammation, J Allergy Clin Immunol 122(6) (2008) 1208–14. [PubMed: 18926564]
- [176]. van den Oord RA, Sheikh A, Filaggrin gene defects and risk of developing allergic sensitisation and allergic disorders: systematic review and meta-analysis, BMJ 339 (2009) b2433. [PubMed: 19589816]
- [177]. Greiner AN, Hellings PW, Rotiroti G, Scadding GK, Allergic rhinitis, Lancet 378(9809) (2011) 2112–22. [PubMed: 21783242]
- [178]. Lin H, Lin D, Xiong XS, Dai XX, Lin T, Role of platelet-derived growth factor-alpha in eosinophilic and non-eosinophilic chronic rhinosinusitis with nasal polyps, Int Forum Allergy Rhinol 4(11) (2014) 909–14. [PubMed: 25256824]
- [179]. Lackner A, Raggam RB, Stammberger H, Beham A, Braun H, Kleinhappl B, Buzina W, Kittinger C, Reinisch S, Berghold A, Freudenschuss K, Barth S, Marth E, The role of interleukin-16 in eosinophilic chronic rhinosinusitis, Eur Arch Otorhinolaryngol 264(8) (2007) 887–93. [PubMed: 17431659]
- [180]. Shah SA, Ishinaga H, Takeuchi K, Pathogenesis of eosinophilic chronic rhinosinusitis, J Inflamm (Lond) 13 (2016) 11. [PubMed: 27053925]
- [181]. Steinke JW, Kennedy JL, Leukotriene Inhibitors in Sinusitis, Curr Infect Dis Rep (2012).
- [182]. de Borja Callejas F, Martinez-Anton A, Picado C, Alobid I, Pujols L, Valero A, Roca-Ferrer J, Mullol J, Corticosteroid treatment regulates mucosal remodeling in chronic rhinosinusitis with nasal polyps, Laryngoscope 125(5) (2015) E158–67. [PubMed: 25641502]
- [183]. Antia C, Baquerizo K, Korman A, Bernstein JA, Alikhan A, Urticaria: A comprehensive review: Epidemiology, diagnosis, and work-up, J Am Acad Dermatol 79(4) (2018) 599–614. [PubMed: 30241623]
- [184]. Kreft B, Wohlrab J, Marsch WC, [Aquagenic urticaria. A case report], Hautarzt 65(2) (2014) 130–2. [PubMed: 24141429]
- [185]. Kursbaum A, Spalter L, Ben-Assuly S, [A dramatic presentation of primary idiopathic cold urticaria], Harefuah 104(9) (1983) 393–4. [PubMed: 6674051]
- [186]. Mazarakis A, Bardousis K, Almpanis G, Mazaraki I, Markou S, Kounis NG, Kounis syndrome following cold urticaria: the swimmer's death, Int J Cardiol 176(2) (2014) e52–3. [PubMed: 25064198]
- [187]. Trevisonno J, Balram B, Netchiporouk E, Ben-Shoshan M, Physical urticaria: Review on classification, triggers and management with special focus on prevalence including a metaanalysis, Postgrad Med 127(6) (2015) 565–70. [PubMed: 25959894]
- [188]. Zuberbier T, Bindslev-Jensen C, Canonica W, Grattan CE, Greaves MW, Henz BM, Kapp A, Kozel MM, Maurer M, Merk HF, Schafer T, Simon D, Vena GA, Wedi B, Eaaci/Ga2Len/Edf, EAACI/GA2LEN/EDF guideline: definition, classification and diagnosis of urticaria, Allergy 61(3) (2006) 316–20. [PubMed: 16436140]
- [189]. Fulkerson PC, Rothenberg ME, Targeting eosinophils in allergy, inflammation and beyond, Nat Rev Drug Discov 12(2) (2013) 117–29. [PubMed: 23334207]
- [190]. Dent LA, Strath M, Mellor AL, Sanderson CJ, Eosinophilia in transgenic mice expressing interleukin 5, J Exp Med 172(5) (1990) 1425–31. [PubMed: 2230651]
- [191]. Rothenberg ME, Eosinophilia N Engl J Med 338(22) (1998) 1592–600.
- [192]. Davis BP, Rothenberg ME, Eosinophils and cancer, Cancer Immunol Res 2(1) (2014) 1–8. [PubMed: 24778159]
- [193]. Manohar M, Verma AK, Venkateshaiah SU, Mishra A, Significance of Eosinophils in Promoting Pancreatic malignancy, J Gastroenterol Pancreatol Liver Disord 5(1) (2017).
- [194]. Insiripong S, Siriyakorn N, Treatment of eosinophilia with albendazole, Southeast Asian J Trop Med Public Health 39(3) (2008) 517–20. [PubMed: 18564693]

- [195]. Michaelann L, Erik Zeger L, Palaniandy K, Eosinophilia Treatment & Management, Hematology (2018).
- [196]. Dellon ES, Jensen ET, Martin CF, Shaheen NJ, Kappelman MD, Prevalence of eosinophilic esophagitis in the United States, Clin Gastroenterol Hepatol 12(4) (2014) 589–96 e1. [PubMed: 24035773]
- [197]. Venkateshaiah SU, Manohar M, Verma AK, Blecker U, Mishra A, Possible Noninvasive Biomarker of Eosinophilic Esophagitis: Clinical and Experimental Evidence, Case Rep Gastroenterol 10(3) (2016) 685–692.
- [198]. Sgouros SN, Bergele C, Mantides A, Eosinophilic esophagitis in adults: a systematic review, Eur J Gastroenterol Hepatol 18(2) (2006) 211–7. [PubMed: 16394804]
- [199]. Li G, Cai B, Song H, Yang Z, Clinical and radiological character of eosinophilic cystitis, Int J Clin Exp Med 8(1) (2015) 533–9. [PubMed: 25785027]
- [200]. Aleem S, Kumar B, Fasano MB, Takacs E, Azar AE, Successful use of cyclosporine as treatment for eosinophilic cystitis: a case report, World Allergy Organ J 9 (2016) 22. [PubMed: 27458500]
- [201]. Pinal-Fernandez I, Selva-O' Callaghan A, Grau JM, Diagnosis and classification of eosinophilic fasciitis, Autoimmun Rev 13(4–5) (2014) 379–82. [PubMed: 24424187]
- [202]. Antic M, Lautenschlager S, Itin PH, Eosinophilic fasciitis 30 years after what do we really know? Report of 11 patients and review of the literature, Dermatology 213(2) (2006) 93–101. [PubMed: 16902285]
- [203]. Dziadzio L, Kelly EA, Panzer SE, Jarjour N, Huttenlocher A, Cytokine abnormalities in a patient with eosinophilic fasciitis, Ann Allergy Asthma Immunol 90(4) (2003) 452–5. [PubMed: 12722970]
- [204]. Enokihara H, Furusawa S, Nakakubo H, Kajitani H, Nagashima S, Saito K, Shishido H, Hitoshi Y, Takatsu K, Noma T, et al., T cells from eosinophilic patients produce interleukin-5 with interleukin-2 stimulation, Blood 73(7) (1989) 1809–13. [PubMed: 2785414]
- [205]. French LE, Shapiro M, Junkins-Hopkins JM, Wolfe JT, Rook AH, Eosinophilic fasciitis and eosinophilic cellulitis in a patient with abnormal circulating clonal T cells: increased production of interleukin 5 and inhibition by interferon alfa, J Am Acad Dermatol 49(6) (2003) 1170–4. [PubMed: 14639411]
- [206]. Bischoff L, Derk CT, Eosinophilic fasciitis: demographics, disease pattern and response to treatment: report of 12 cases and review of the literature, Int J Dermatol 47(1) (2008) 29–35.
- [207]. Mouthon L, Dunogue B, Guillevin L, Diagnosis and classification of eosinophilic granulomatosis with polyangiitis (formerly named Churg-Strauss syndrome), J Autoimmun 48– 49 (2014) 99–103.
- [208]. Vaglio A, Buzio C, Zwerina J, Eosinophilic granulomatosis with polyangiitis (Churg-Strauss): state of the art, Allergy 68(3) (2013) 261–73. [PubMed: 23330816]
- [209]. Mohammad AJ, Hot A, Arndt F, Moosig F, Guerry MJ, Amudala N, Smith R, Sivasothy P, Guillevin L, Merkel PA, Jayne DR, Rituximab for the treatment of eosinophilic granulomatosis with polyangiitis (Churg-Strauss), Ann Rheum Dis 75(2) (2016) 396–401. [PubMed: 25467294]
- [210]. Wechsler ME, Akuthota P, Jayne D, Khoury P, Klion A, Langford CA, Merkel PA, Moosig F, Specks U, Cid MC, Luqmani R, Brown J, Mallett S, Philipson R, Yancey SW, Steinfeld J, Weller PF, Gleich GJ, Team EMS, Mepolizumab or Placebo for Eosinophilic Granulomatosis with Polyangiitis, N Engl J Med 376(20) (2017) 1921–1932. [PubMed: 28514601]
- [211]. Allen J, Wert M, Eosinophilic Pneumonias J Allergy Clin Immunol Pract 6(5) (2018) 1455– 1461.
- [212]. Marchand E, Cordier JF, Idiopathic chronic eosinophilic pneumonia, Orphanet J Rare Dis 1 (2006) 11. [PubMed: 16722612]
- [213]. Kaya H, Gumus S, Ucar E, Aydogan M, Musabak U, Tozkoparan E, Bilgic H, Omalizumab as a steroid-sparing agent in chronic eosinophilic pneumonia, Chest 142(2) (2012) 513–516. [PubMed: 22871762]
- [214]. Rhee CK, Min KH, Yim NY, Lee JE, Lee NR, Chung MP, Jeon K, Clinical characteristics and corticosteroid treatment of acute eosinophilic pneumonia, Eur Respir J 41(2) (2013) 402–9. [PubMed: 22599359]

- [215]. Ogbogu PU, Rosing DR, Horne MK 3rd, Cardiovascular manifestations of hypereosinophilic syndromes, Immunol Allergy Clin North Am 27(3) (2007) 457–75. [PubMed: 17868859]
- [216]. Helbig G, Wieczorkiewicz A, Dziaczkowska-Suszek J, Majewski M, Kyrcz-Krzemien S, T-cell abnormalities are present at high frequencies in patients with hypereosinophilic syndrome, Haematologica 94(9) (2009) 1236–41. [PubMed: 19734416]
- [217]. Roufosse FE, Goldman M, Cogan E, Hypereosinophilic syndromes, Orphanet J Rare Dis 2 (2007) 37. [PubMed: 17848188]
- [218]. Gleich GJ, Leiferman KM, Pardanani A, Tefferi A, Butterfield JH, Treatment of hypereosinophilic syndrome with imatinib mesilate, Lancet 359(9317) (2002) 1577–8. [PubMed: 12047970]
- [219]. Klion AD, Bochner BS, Gleich GJ, Nutman TB, Rothenberg ME, Simon HU, Wechsler ME, Weller PF, G. The Hypereosinophilic Syndromes Working, Approaches to the treatment of hypereosinophilic syndromes: a workshop summary report, J Allergy Clin Immunol 117(6) (2006) 1292–302. [PubMed: 16750989]
- [220]. Dima E, Koltsida O, Katsaounou P, Vakali S, Koutsoukou A, Koulouris NG, Rovina N, Implication of Interleukin (IL)-18 in the pathogenesis of chronic obstructive pulmonary disease (COPD), Cytokine 74(2) (2015)313–7. [PubMed: 25922275]
- [221]. Imaoka H, Hoshino T, Takei S, Kinoshita T, Okamoto M, Kawayama T, Kato S, Iwasaki H, Watanabe K, Aizawa H, Interleukin-18 production and pulmonary function in COPD, Eur Respir J 31(2) (2008) 287–97. [PubMed: 17989120]
- [222]. Nakajima T, Owen CA, Interleukin-18: the master regulator driving destructive and remodeling processes in the lungs of patients with chronic obstructive pulmonary disease?, Am J Respir Crit Care Med 185(11) (2012) 1137–9. [PubMed: 22661518]
- [223]. Wang J, Liu X, Xie M, Xie J, Xiong W, Xu Y, Increased expression of interleukin-18 and its receptor in peripheral blood of patients with chronic obstructive pulmonary disease, COPD 9(4) (2012) 375–81. [PubMed: 22489883]
- [224]. Hart TK, Cook RM, Zia-Amirhosseini P, Minthorn E, Sellers TS, Maleeff BE, Eustis S, Schwartz LW, Tsui P, Appelbaum ER, Martin EC, Bugelski PJ, Herzyk DJ, Preclinical efficacy and safety of mepolizumab (SB-240563), a humanized monoclonal antibody to IL-5, in cynomolgus monkeys, J Allergy Clin Immunol 108(2) (2001) 250–7. [PubMed: 11496242]
- [225]. Pavord ID, Chanez P, Criner GJ, Kerstjens HAM, Korn S, Lugogo N, Martinot JB, Sagara H, Albers FC, Bradford ES, Harris SS, Mayer B, Rubin DB, Yancey SW, Sciurba FC, Mepolizumab for Eosinophilic Chronic Obstructive Pulmonary Disease, N Engl J Med 377(17) (2017) 1613– 1629. [PubMed: 28893134]
- [226]. Song T, Jones DM, Homsi Y, Therapeutic effect of anti-IL-5 on eosinophilic myocarditis with large pericardial effusion, BMJ Case Rep 2017 (2017).
- [227]. Brambatti M, Matassini MV, Adler ED, Klingel K, Camici PG, Ammirati E, Eosinophilic Myocarditis: Characteristics, Treatment, and Outcomes, J Am Coll Cardiol 70(19) (2017) 2363– 2375. [PubMed: 29096807]
- [228]. Cooper LT Jr., Eosinophilic Myocarditis as a Cause of Acute Cardiac Syndromes: The Importance of Awareness, J Am Coll Cardiol 70(19) (2017) 2376–2377. [PubMed: 29096808]
- [229]. Espana A, Sanz ML, Sola J, Gil P, Wells' syndrome (eosinophilic cellulitis): correlation between clinical activity, eosinophil levels, eosinophil cation protein and interleukin-5, Br J Dermatol 140(1) (1999) 127–30. [PubMed: 10215782]
- [230]. Yagi H, Tokura Y, Matsushita K, Hanaoka K, Furukawa F, Takigawa M, Wells' syndrome: a pathogenic role for circulating CD4+CD7− T cells expressing interleukin-5 mRNA, Br J Dermatol 136(6) (1997) 918–23. [PubMed: 9217826]
- [231]. Fujimura T, Kambayashi Y, Furudate S, Kakizaki A, Aiba S, A possible mechanism in the recruitment of eosinophils and Th2 cells through CD163(+) M2 macrophages in the lesional skin of eosinophilic cellulitis, Eur J Dermatol 24(2) (2014) 180–5. [PubMed: 24721436]
- [232]. Simon HU, Plotz S, Simon D, Seitzer U, Braathen LR, Menz G, Straumann A, Dummer R, Levi-Schaffer F, Interleukin-2 primes eosinophil degranulation in hypereosinophilia and Wells' syndrome, Eur J Immunol 33(4) (2003) 834–9. [PubMed: 12672048]

- [233]. Caputo R, Marzano AV, Vezzoli P, Lunardon L, Wells syndrome in adults and children: a report of 19 cases, Arch Dermatol 142(9) (2006) 1157–61. [PubMed: 16983003]
- [234]. Cherng E, McClung AA, Rosenthal HM, Hicks J, Levy ML, Wells' syndrome associated with parvovirus in a 5-year old boy, Pediatr Dermatol 29(6) (2012) 762–4. [PubMed: 22150362]
- [235]. Gilliam AE, Bruckner AL, Howard RM, Lee BP, Wu S, Frieden IJ, Bullous "cellulitis" with eosinophilia: case report and review of Wells' syndrome in childhood, Pediatrics 116(1) (2005) e149–55. [PubMed: 15995016]
- [236]. Ohtsuka T, Oral tacrolimus treatment for refractory eosinophilic cellulitis, Clin Exp Dermatol 34(8) (2009)e597–8. [PubMed: 19489865]
- [237]. Katz U, Shoenfeld Y, Pulmonary eosinophilia, Clin Rev Allergy Immunol 34(3) (2008) 367–71. [PubMed: 18197481]
- [238]. Kudlacz E, Conklyn M, Andresen C, Whitney-Pickett C, Changelian P, The JAK-3 inhibitor CP-690550 is a potent anti-inflammatory agent in a murine model of pulmonary eosinophilia, Eur J Pharmacol 582(1–3) (2008) 154–61. [PubMed: 18242596]
- [239]. Rothenberg ME, Eosinophilic gastrointestinal disorders (EGID), J Allergy Clin Immunol 113(1) (2004) 11–28; quiz 29. [PubMed: 14713902]
- [240]. Straumann A, Eosinophil-Associated Gastrointestinal Disorders, In Clinical Immunology (2019) 633–640.
- [241]. Alfadda AA, Storr MA, Shaffer EA, Eosinophilic colitis: epidemiology, clinical features, and current management, Therap Adv Gastroenterol 4(5) (2011) 301–9.
- [242]. Uppal V, Kreiger P, Kutsch E, Eosinophilic Gastroenteritis and Colitis: a Comprehensive Review, Clin Rev Allergy Immunol 50(2) (2016) 175–88. [PubMed: 26054822]
- [243]. Caldwell JM, Collins MH, Stucke EM, Putnam PE, Franciosi JP, Kushner JP, Abonia JP, Rothenberg ME, Histologic eosinophilic gastritis is a systemic disorder associated with blood and extragastric eosinophilia, TH2 immunity, and a unique gastric transcriptome, J Allergy Clin Immunol 134(5) (2014) 1114–24. [PubMed: 25234644]
- [244]. Ko HM, Morotti RA, Yershov O, Chehade M, Eosinophilic gastritis in children: clinicopathological correlation, disease course, and response to therapy, Am J Gastroenterol 109(8) (2014) 1277–85. [PubMed: 24957155]
- [245]. Baig MA, Qadir A, Rasheed J, A review of eosinophilic gastroenteritis, J Natl Med Assoc 98(10) (2006) 1616–9. [PubMed: 17052051]
- [246]. Desreumaux P, Bloget F, Seguy D, Capron M, Cortot A, Colombel JF, Janin A, Interleukin 3, granulocyte-macrophage colony-stimulating factor, and interleukin 5 in eosinophilic gastroenteritis, Gastroenterology 110(3) (1996) 768–74. [PubMed: 8608886]
- [247]. Ingle SB, Hinge Ingle CR, Eosinophilic gastroenteritis: an unusual type of gastroenteritis, World J Gastroenterol 19(31) (2013) 5061–6. [PubMed: 23964139]

Highlights

- **•** Eosinophils and its constituents
- **•** Role of IL5 and IL18 in eosinophils development, transformation, maturation
- **•** Signal transduction of IL5 and IL18
- **•** The role of eosinophils in allergic disorders
- **•** The role of several other associated cytokines in promoting eosinophils mediated allergic diseases

Fig.2.

IL5 mediated cell signaling in eosinophils.

Fig.3.

Production, migration, and recruitment of eosinophils from bone marrow to targeted tissues in eosinophil mediated allergic diseases and activation of innate and adaptive immune responses.

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Fig.6.

Eosinophils associated allergic diseases

Table.1

Eosinophil constituents and function.

