

Levels of Serotonin in the Hemolymph of *Aplysia* Are Modulated by Light/Dark Cycles and Sensitization Training

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Serotonin (5-hydroxytryptamine, 5-HT) modulates the behavior and physiology of both vertebrate and invertebrate animals. Effects of injections of 5-HT and the morphology of the serotonergic system of *Aplysia* indicate that 5-HT may have a humoral, in addition to a neurotransmitter, role. To study possible humoral roles of 5-HT, we measured 5-HT in the hemolymph. The concentration of 5-HT in the hemolymph was ~18 nM, a value close to previously reported thresholds for eliciting physiological responses. The concentration of 5-HT in the hemolymph expressed a diurnal rhythm. In addition, electrical stimulation that leads to long-term sensitization significantly increased levels of 5-HT in the hemolymph during training, 1.5 hr after training, and 24 hr after training. Moreover, levels of 5-HT in the hemolymph were significantly correlated with the magnitude of sensitization. The half-life of an increase in 5-HT in the hemolymph was ~0.5 hr. Therefore, the persistent in-

crease of 5-HT in the hemolymph 24 hr after sensitization training indicates that training caused a long-lasting increase in the release of 5-HT. This long-lasting increase in 5-HT in the hemolymph was blocked by treatment with an inhibitor of protein synthesis during training. Based on the levels of 5-HT in the hemolymph and its regulation by environmental events, we propose that 5-HT has a humoral role in regulation of the behavioral state of *Aplysia*. In support of this hypothesis, we found that increasing levels of 5-HT in the hemolymph led to significant alterations in feeding behavior. Increasing levels of 5-HT during the daytime when they were normally low increased the latency to assume feeding posture from daytime to nighttime values.

Key words: *Aplysia*; mollusk; serotonin; hemolymph; sensitization; diurnal rhythms; learning; feeding; emetine

Serotonin influences behavioral state in a number of vertebrates and invertebrates (for review, see Edwards and Kravitz, 1997). In *Aplysia*, 5-HT is involved in mediating sensitization of defensive withdrawal reflexes (Brunelli et al., 1976; Bernier et al., 1982; Klein et al., 1982; Walters et al., 1983; Glanzman et al., 1989; Hammer et al., 1989; Schacher et al., 1990; Trudeau and Castellucci, 1993; Emptage et al., 1996) and regulating the phase of the ocular circadian rhythm (Corrent and Eskin, 1982), cardiac output (Liebeswar et al., 1975; Sawada et al., 1984), muscle tone (Weiss et al., 1975; Cooper et al., 1989; McPherson and Blankenship, 1992), bag cell afterdischarge (Kaczmarek et al., 1978), locomotor activity (Mackey and Carew, 1983), swimming behavior (Parsons and Pinsker, 1989; McPherson and Blankenship, 1991; Laurienti and Blankenship, 1997), and the sensitivity of photoreceptors and mechanoreceptors (Eskin and Maresh, 1982; Billy and Walters, 1989). Feeding behavior is also modulated by 5-HT released by the metacerebral cell (Weiss et al., 1975; Kupfermann et al., 1979; Rosen et al., 1983). Moreover, activity of the serotonergic metacerebral cell directly correlates with central arousal in *Aplysia* (Kupfermann and Weiss, 1982).

Several lines of evidence indicate that 5-HT may have a humoral role in *Aplysia*. 5-HT has widespread physiological effects,

and injections of 5-HT into the hemocoel increase the number and frequency of a variety of spontaneous behaviors for a short period of time (Palovcik et al., 1982; Mackey and Carew, 1983; Parsons and Pinsker, 1989). Also, the projections of serotonergic fibers indicates that 5-HT may be released into the hemocoel. For example, serotonergic fibers innervate the aorta and other arteries, and also form a basket around the eye (Takahashi et al., 1988; Alevizos et al., 1989; Skelton and Koester, 1992).

As a first step to further examine the possible humoral role of 5-HT in *Aplysia*, we investigated whether 5-HT was present in the hemolymph and, if so, whether its levels were regulated. In this study, we demonstrate that 5-HT is present in the hemolymph of *Aplysia* and that its concentration is near threshold for a number of physiological effects of 5-HT. We show that levels of 5-HT in the hemolymph are regulated by light/dark cycles and by electrical stimulation that leads to long-term sensitization. Furthermore, we show that perturbations of 5-HT in the hemolymph affect aspects of appetitive feeding behavior.

MATERIALS AND METHODS

Aplysia californica (100–150 gm) were obtained from Marinus (Long Beach, CA). Animals were maintained in Instant Ocean (Aquarium Systems, Mentor, OH) at 15°C under a 12 hr light/dark (LD) cycle. Animals were allowed to adapt to laboratory conditions for 3 d before use except in the experiment measuring the 5-HT levels of animals just after their arrival in the laboratory. Animals used for sensitization training were kept in individual plastic cages in recirculating seawater tanks. They were fed dried seaweed every 2–3 d. All other animals, except those used in feeding latency experiments (described below), were placed directly in recirculating seawater tanks (100 gal) and fed romaine lettuce every 2–3 d. Animals were housed in constant darkness (DD) by transfer to a 30 gal recirculating seawater tank kept in light-tight housing.

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Measurements of levels of 5-HT in hemolymph of *Aplysia* living in the field were performed on animals (100–150 gm) captured from tidal pools along the coast of Southern California and immediately sampled (see below). All animals were captured and sampled during the day in mid-October. Conditions at the site of capture were calm, and water temperature was ~19°C.

To measure levels of 5-HT in the hemolymph of animals immediately after arrival into the laboratory, animals were removed from original shipping containers and equilibrated for 15–30 min. During this equilibration, animals remained in individually sealed plastic bags, which were placed in seawater tanks maintained at 15°C. After equilibration, bags were opened, and hemolymph was immediately sampled (see below). Animals were then fed (lettuce), transferred to individual plastic cages, and maintained under standard environmental conditions (see above).

Long-term sensitization produced by electrical stimulation was performed as previously described (Goldsmith and Byrne, 1993; Cleary et al., 1998). Briefly, animals were administered 4 blocks of 10 shocks over 1.5 hr to one randomly selected side of the animal. Each shock consisted of a 500 msec pulse of 60 mA applied to the body wall. Release of ink and opaline, defensive secretions caused by noxious stimuli, were reliably elicited with this training protocol (Goldsmith and Byrne, 1993). Baseline hemolymph samples were withdrawn 0.5 hr before the first stimulus. Other samples were withdrawn at times indicated in the text. Single animals were sampled multiple times during training (as noted in Results), but in all other cases hemolymph was sampled only once from an animal. Control animals did not receive electrical stimulation, but were handled identically to sensitized animals in all other respects.

Siphon withdrawal duration was measured, and hemolymph was subsequently sampled to correlate changes in duration of siphon withdrawal with changes in levels of 5-HT in the hemolymph. Teflon-coated silver wire electrodes were implanted into the posterior region of the tail, as previously described (Scholz and Byrne, 1987; Goldsmith and Byrne, 1993), for delivery of electrical stimuli (20 msec, AC shock). Baseline hemolymph samples were withdrawn 30 min before threshold determination. Several test stimuli were administered to determine threshold current required for eliciting siphon-withdrawal. Siphon withdrawal was assessed at two-times the threshold current required to elicit the reflex. Immediately after determining the baseline siphon withdrawal duration for each side, electrical stimulation was administered over one side of the body wall as above. Siphon withdrawal duration was measured 24 hr after the end of electrical stimulation, and then hemolymph was withdrawn immediately afterward. Hemolymph was withdrawn from the side of the animal that did not receive electrical stimulation. Withdrawal of hemolymph did not induce long-term sensitization. Control animals did not receive electrical stimulation, but were handled identically to sensitized animals in all other respects. The experimenter who measured siphon withdrawals was blind to whether the animals received electrical stimulation and the side from which hemolymph was withdrawn.

To measure the half-life of 5-HT in the hemolymph of animals that were electrically stimulated once 5-HT was elevated, individual *Aplysia* were sensitized by the standard long-term training protocol and then placed in 3 l of seawater that contained 250 μ M 5-HT (creatine-sulfate complex; Sigma, St. Louis, MO) for 0.5 hr (Alberini et al., 1994). Hemolymph was sampled before, immediately after, and then 0.5, 1, and 6 hr after treatment. Animals were only sampled once during this experiment. Sensitized animals were used in this experiment because the turnover of 5-HT or its diffusion through the skin might be altered by the training procedure.

Collection of hemolymph from animals used in this study was performed using the following protocol. Hemolymph samples (0.5 ml) were withdrawn from animals using a sterile 16 gauge needle and 1 cc syringe. Hemolymph was withdrawn from the animal at a level between the rhinophores and parapodia near the foot. Care was taken to minimize trauma to the animal. Withdrawal of hemolymph only required ~10 sec of handling. In addition, removal of hemolymph only elicited inking in 10% of the animals used in this study. Subsequent samples of hemolymph from animals indicates that withdrawal of hemolymph did not produce elevation of 5-HT in the hemolymph.

Hemolymph was centrifuged at 4°C for 2 min to pellet hemocytes. Previous studies have shown that the hemocytes of various invertebrates, including *Aplysia*, contain 5-HT (Stefano et al., 1989; Ottaviani and Cossarizza, 1990; Ottaviani et al., 1992; Levenson and Eskin, unpublished observations). Centrifugation did not elicit release of 5-HT from *Aplysia* hemocytes, because the level of 5-HT measured from centrifuged hemolymph was nearly identical to the level measured from hemolymph

diluted with isotonic MgCl₂ (180 mM final concentration) or hemolymph that was passed through a 0.22 μ m filter (data not shown). Cell-free hemolymph was aliquoted, immediately frozen in liquid N₂, and stored at -80°C. Assays were always performed within 5 d of collection of hemolymph. 5-HT was measured using a commercially available ELISA (ICN Biochemicals, Costa Mesa, CA). Hemolymph was processed as plasma according to the manufacturer's protocol. After acylation, the resulting solution was diluted with 200 μ l of assay buffer. During processing, hemolymph was exposed to 14% acetone to precipitate proteins. This treatment should denature any proteins that may bind 5-HT. Thus, total 5-HT in the hemolymph was measured in all of our assays.

Several control experiments were done to verify that the ELISA was valid, because the primary antiserum used actually binds to *N*-acyl-5-HT. Hemolymph that was not exposed to the acylation reagent did not have detectable levels of 5-HT. No 5-HT was present in seawater that had been exposed to acylation. Moreover, the ELISA could accurately measure known amounts of 5-HT added to seawater or hemolymph (data not shown).

The role of protein synthesis in the long-lasting increase in hemolymph 5-HT caused by electrical stimulation was assessed via injection of emetine into the intact animal. Baseline hemolymph samples were withdrawn from animals 0.5 hr before receiving an injection of emetine. Animals were injected with 1 ml, 9 mM emetine [dihydrochloride; Sigma; dissolved in buffered-filtered seawater (BFSW) with 3% ethanol] per 140 gm body weight 30 min before electrical stimulation. Injections were estimated to yield ~100 μ M emetine in the hemolymph, assuming 65% of the weight of the animal is hemolymph. This concentration was sufficient to significantly inhibit protein synthesis (see Results; Schwartz et al., 1971; Castellucci et al., 1986; Montarolo et al., 1986). Animals injected with emetine responded to electrical stimulation similar to control animals; copious amounts of ink and opaline were secreted, and animals withdrew from the site of shock. Animals injected with emetine became flaccid near the site of injection, but appeared to recover ~3–4 hr after injection. Control animals received a vehicle injection (3% ethanol in BFSW) in place of emetine. The vehicle did not appear to affect behavior and/or muscle tone. Electrical stimulation was performed as described above. Hemolymph was collected 24 hr after the end of electrical stimulation.

The effect of emetine injections on protein synthesis was assayed by measuring incorporation of [³H]leucine into protein in both pleural-pedal and abdominal ganglia as previously described (Raju et al., 1991). In each experiment, two experimental animals were injected with emetine, and two control animals were injected with the vehicle (as above). This experiment was repeated three times. Thirty minutes after injection, pleural-pedal and abdominal ganglia were removed and exposed to 40 μ Ci/ml [³H]leucine (1 mCi/ml; ICN Biochemicals) in BFSW for 1 hr at 15°C. After exposure to leucine, ganglia were rinsed 6 times with ice-cold BFSW, blotted dry, and frozen in liquid N₂. Ganglia were homogenized, and incorporation of leucine into protein was measured as TCA-precipitable counts. This value was normalized to total counts in the homogenate to adjust for differences in uptake of leucine by the ganglia.

Latency to feeding posture was measured following the technique of Kupfermann (1974). After adaptation to the laboratory, animals were fed to satiation with dried seaweed (Kupfermann, 1974), housed in individual plastic cages, and fasted for 5 d. On the fifth day, latency to feeding posture was tested with a piece of seaweed ~2.5 mm² in area and 0.25 gm in weight. Animals were only included in this study that had an initial behavioral state of 0 or 1, as defined by Kupfermann (1974), just before experimentation. The 0 and 1 states are characterized by an immobile animal. Rhinophores were brushed with a piece of seaweed, and the seaweed was then kept in the seawater near its surface. Latency to feeding posture was designated as the time elapsed between brushing the rhinophores with seaweed and the display of characteristic "heads-up feeding posture" (Kupfermann, 1974). Latencies measured at night, zeitgeber time 18 (ZT18), were performed under dim red light. Experimental animals received an injection of 1 ml, 3 μ M 5-HT (hydrochloride; Sigma) per 150 gm body weight, 20 min before testing feeding latency. These injections increased levels of 5-HT in the hemolymph within 5 min to 25.8 \pm 4.9 nM, a 5-HT level normally seen at night. Control animals received an equivalent injection of seawater. All feeding experiments were performed blind. Specifically, the investigator responsible for measuring feeding latency was different from the investigator responsible for injection of 5-HT.

Analysis of 5-HT from animals versus time spent in the laboratory was

Table 1. Thresholds of physiological responses to 5-HT

Phenomenon	Threshold (nM)	Reference
Cardioacceleration	0.1–1	Liebeswar et al., 1975
Potentialiation of ARC contraction	1	Weiss et al., 1975
Potentialiation of ERG	10–100	Eskin and Maresh, 1982
Phase-shifting of ocular circadian rhythm	10–100	Corrent and Eskin, 1982
Inhibition of spontaneous activity in the eye	10–100	Corrent et al., 1982
Body wall relaxation	10–100	Cooper et al., 1989
Inhibition of R15 bursting	20–100	Drummond et al., 1980a
Inhibition of afterdischarge	125	Kaczmarek et al., 1978
Facilitation of sensory-motor synapse	1000	Brunelli et al., 1976
Induction of swimming behavior	10,000	Parsons and Pinsker, 1989

performed with a paired, Student's *t* test. Time-dependent changes, after electrical stimulation and the effect of emetine on the 24 hr increase in 5-HT were analyzed via two-way ANOVA. Elevation of 5-HT in the hemolymph and rhythms in levels of 5-HT were analyzed for significance using one-way ANOVA. The relationship between duration of siphon withdrawal and the magnitude of increase in hemolymph 5-HT was analyzed via product-moment correlation. Feeding latency at midday and midnight, and after 5-HT injection was analyzed using a Wilcoxon two-sample test. All *post hoc* analyses were performed using the T-method of unplanned comparisons among means (Sokal and Rohlf, 1995). Statistical significance for all tests was set at $p \leq 0.05$.

RESULTS

The mean level of 5-HT in the hemolymph during the day taken from *Aplysia* ≥ 3 d after their arrival to the laboratory was 18.0 ± 0.6 nM (SEM; $n = 117$), and is comparable to the mean level of 5-HT in the hemolymph measured from animals in the field (16.7 ± 1.3 nM; $n = 17$). Interestingly, this value is also close to the level of 5-HT in the hemolymph of another mollusc, *Helix* (Cardot, 1971). This level of 5-HT in the hemolymph of *Aplysia* is close to threshold values for a number of physiological responses (Table 1).

Initially, animals were sampled immediately (15–30 min, between 10:00 A.M. and 12:00 P.M.) and 3 d after arrival into the laboratory, a time at which animals are normally used in experiments. The levels of 5-HT in the hemolymph of animals that had just arrived were significantly elevated relative to levels observed after animals had been in the laboratory for 3 d (Fig. 1; $t = 5.32$; $df = 7$; $p < 0.001$). Levels of 5-HT in the hemolymph did not appear to decrease further after animals had been in the laboratory for >3 d (data not shown). The decrease in 5-HT with time in the laboratory suggested that levels of 5-HT in the hemolymph may be regulated by environmental stimuli and be indicative of the state of the animal. Moreover, this observation indicates that animals should be equilibrated in the laboratory before performing behavioral or cellular studies. Several factors may have contributed to the increase in hemolymph 5-HT during shipment of the animals. Animals arrived in the laboratory after approximately one half day of travel, during which they were maintained in cold seawater ($\sim 4^\circ\text{C}$) in dark, styrofoam boxes. Some possible factors that might contribute to the elevated 5-HT include stress caused by water temperature, handling, or being held in darkness for a long period of time (see Fig. 3). Further experiments are required to determine which of these factors, or others, might be responsible for this change.

Diurnal rhythm of 5-HT

Aplysia have circadian locomotor rhythms, and 5-HT plays an important role in regulation of ocular circadian pacemakers in

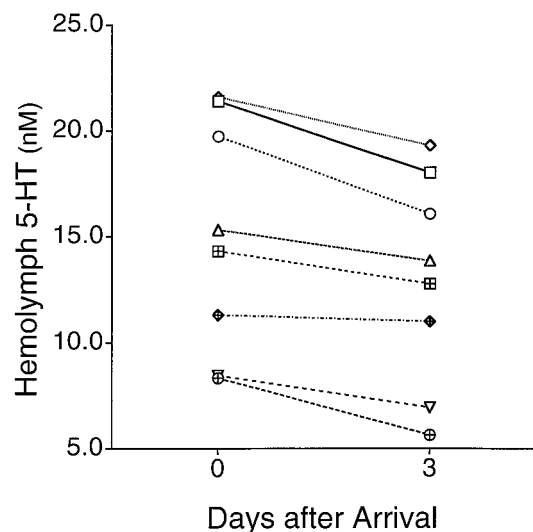


Figure 1. Levels of 5-HT in the hemolymph decrease with time after arrival in the laboratory. Concentration of 5-HT in the hemolymph is plotted as a function of time spent in the laboratory for individual animals. Hemolymph was sampled immediately after arrival at the lab and 3 d afterward. There was a significant decrease in 5-HT during this period.

Aplysia (Jacklet, 1969; Strumwasser, 1973; Kupfermann, 1974; Lickey et al., 1977; Corrent and Eskin, 1982; Corrent et al., 1982; Eskin and Maresh, 1982; Colwell, 1990). Therefore, we investigated whether the concentration of 5-HT in the hemolymph varied with respect to time of day. Hemolymph was sampled from individual animals once at various times throughout a complete photoperiod. Levels of 5-HT in the hemolymph were significantly affected by light/dark cycles (Fig. 2; $F = 3.54$; $df = 47$; $p < 0.01$). The average concentration of 5-HT (18.0 ± 1.2 nM; average of samples from ZT1–ZT12) in the hemolymph during the day was significantly ($t = 4.53$; $df = 6$; $p < 0.0001$) lower than the average value at night (24.4 ± 0.7 nM; average of samples from ZT15–ZT24).

To determine whether changes in the levels of 5-HT observed in a light/dark cycle were under control of a circadian pacemaker, hemolymph was sampled in another experiment from individual animals placed in constant darkness. In these animals, no significant temporal variation of 5-HT in the hemolymph was observed (Fig. 3; $F = 0.38$; $df = 35$; $p > 0.9$). The mean concentration of 5-HT in the hemolymph from animals in DD (25.1 ± 0.7 nM) was similar to levels seen at night in the LD experiment (24.4 ± 0.7

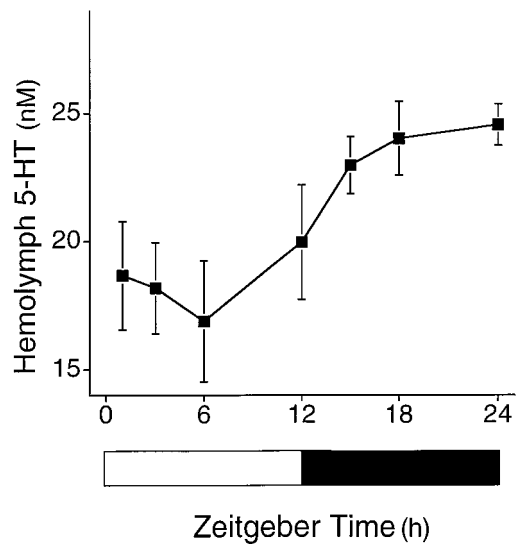


Figure 2. Concentration of 5-HT in the hemolymph is regulated by light/dark cycles. Concentration of 5-HT in the hemolymph is plotted as a function of time of day. Hemolymph was sampled one time from each animal maintained in 12 hr LD cycle. Five to eight animals were sampled at each time point. ZT0 is dawn, ZT12 is dusk. The concentration of 5-HT shows significant variation as a function of time of day. Error bars indicate SEM.

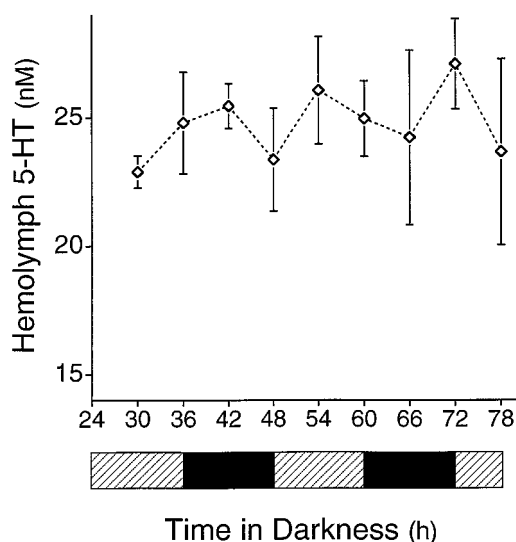


Figure 3. Concentration of 5-HT in the hemolymph does not change when animals are kept in constant darkness. Each animal was sampled only once. No significant variation was seen in the concentration of 5-HT in the hemolymph. In this experiment, 5-HT remained near the night-time levels that were observed when animals were maintained under a light/dark cycle (Fig. 2). Four animals were sampled at each time point.

nm). This finding indicates that the rhythm of 5-HT observed in LD is caused by light decreasing levels of 5-HT in the hemolymph. Light could lower levels of 5-HT in the hemolymph by affecting levels of enzymes involved in 5-HT biosynthesis. A similar mechanism explains the inhibition of melatonin production by light in the mammalian pineal (Deguchi and Axelrod, 1972; Klein and Weller, 1972; Moore and Klein, 1974). Alternatively, light could modulate the activity of serotonergic neurons to decrease their release of 5-HT, light could inhibit enzymes in-

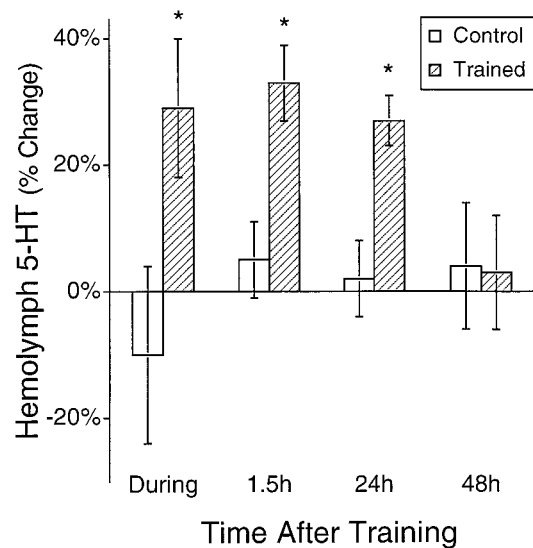


Figure 4. Electrical stimulation elevated 5-HT in the hemolymph during, 1.5, and 24 hr after stimulation. Concentrations of 5-HT in the hemolymph of experimental animals sampled during, 1.5, 24, or 48 hr after the end of electrical stimulation are plotted relative to the value of 5-HT for each animal 0.5 hr before initiation of stimulation. Control animals were handled exactly as animals that were trained, with the exception that control animals did not receive any electrical stimulation. Four hemolymph samples were taken during training; individual values were averaged to yield a concentration of 5-HT in the hemolymph during training. Individual hemolymph samples were taken 1.5, 24, and 48 hr after training from individual animals that had not been previously sampled. In this and subsequent illustrations, an asterisk indicates a significant ($p \leq 0.05$) difference relative to control values.

involved in 5-HT degradation, or light could increase renal clearance of 5-HT.

Effect of electrical stimulation on 5-HT in the hemolymph

Sensitization in *Aplysia* appears to be mediated, at least in part, via 5-HT. Application of 5-HT mimics many of the effects produced by electrical stimulation that produces sensitization (Brunelli et al., 1976; Bernier et al., 1982; Klein et al., 1982; Walters et al., 1983; Hammer et al., 1989; Schacher et al., 1990; Emptage and Carew, 1993; Trudeau and Castellucci, 1993; Emptage et al., 1996; Zhang et al., 1997), and serotonergic neurons appear to be necessary for sensitization (Glanzman et al., 1989). We investigated whether electrical stimulation would release 5-HT and subsequently increase levels of 5-HT in the hemolymph. Electrical stimulation consisted of 4 blocks of 10 shocks to one side of the animal, with 0.5 hr between each block of shocks. Hemolymph was collected 0.5 hr before stimulation and 1 min after each block of stimulation. Changes in levels of 5-HT in the hemolymph are expressed relative to the level of 5-HT measured 0.5 hr before training. Electrical stimulation significantly elevated levels of 5-HT in the hemolymph (Fig. 4; $F = 21$; $df = 65$; $p < 0.0001$). *Post hoc* analysis indicated that 5-HT was significantly elevated during training [Fig. 4; minimum significant difference (MSD; Sokal and Rohlf, 1995) = 15%; $p < 0.05$]. In addition, an increase in the level of 5-HT in the hemolymph was seen 1.5 and 24 hr after electrical stimulation, however no significant increase of 5-HT in the hemolymph was seen 48 hr after electrical stimulation (Fig. 4).

If the increase in 5-HT in the hemolymph was related to the behavioral sensitization caused by noxious stimulation of the

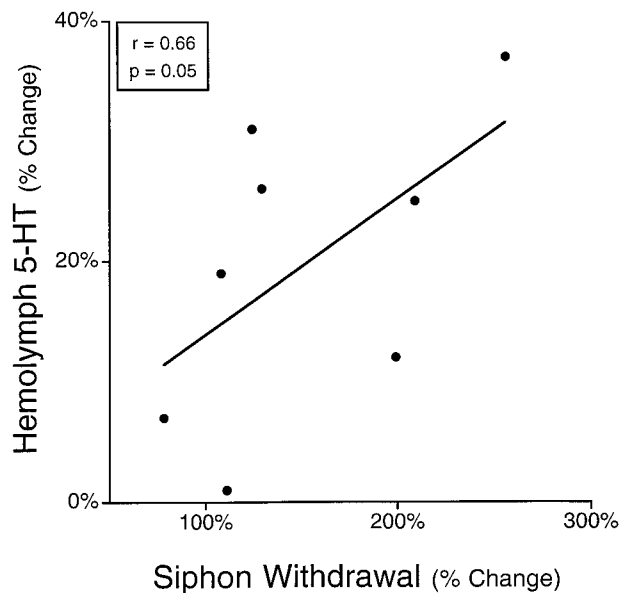


Figure 5. Increases in the duration of siphon withdrawal produced by electrical stimulation were correlated with increases in 5-HT in the hemolymph. Animals were electrically stimulated, and 24 hr later, 5-HT in the hemolymph was measured after determination of the duration of siphon withdrawal. Behavioral changes were significantly correlated with changes in levels of 5-HT in the hemolymph. Measurement of siphon withdrawal did not lead to the elevation of 5-HT in the hemolymph because electrical stimulation alone without behavioral measurement also led to elevation of 5-HT in the hemolymph (see Fig. 4).

body wall, then increases in 5-HT in the hemolymph should occur in animals that were sensitized by noxious stimulation. Therefore, siphon withdrawal and changes in levels of 5-HT in the hemolymph were measured in the same animal. Unilateral sensitization of the siphon withdrawal reflex was observed in animals 24 hr after electrical stimulation. The side of the animals that was trained showed a significant $152 \pm 22\%$ ($n = 8$; SEM) increase in the duration of siphon withdrawal compared to the control side, which showed no sensitization ($2 \pm 3\%$ increase; $t = 6.96$; $df = 7$; $p < 0.001$). In these same animals, 5-HT in the hemolymph significantly increased $20 \pm 4\%$ 24 hr after electrical stimulation ($t = 4.6$; $df = 7$; $p < 0.005$). Furthermore, control animals that received no electrical stimulation showed no long-term behavioral sensitization as measured by siphon withdrawal duration ($9 \pm 6\%$ increase, $n = 5$) and no change in levels of 5-HT in the hemolymph ($0 \pm 7\%$). The magnitude of the change in duration of siphon withdrawal was significantly correlated with the magnitude of the change in 5-HT in the hemolymph (Fig. 5; $r = 0.66$; $df = 7$; $p = 0.05$). This result indicates that changes in the levels of 5-HT in the hemolymph are related to changes in the siphon-withdrawal reflex, and thus may be involved in behavioral sensitization. In these experiments in which siphon withdrawal and 5-HT were both determined, it is unlikely that the behavioral testing itself led to the increase in 5-HT in the hemolymph since levels of 5-HT in the hemolymph did not increase in control animals which received no electrical stimulation (Fig. 5), and electrical stimulation alone led to a similar increase in 5-HT when no behavioral measurements were made (Fig. 4).

The increase we observed in 5-HT in the hemolymph 24 hr after training could be caused by several factors. An initial increase of 5-HT could lead to a long-lasting elevation if 5-HT in the hemolymph was stable and the skin of the animal was imper-

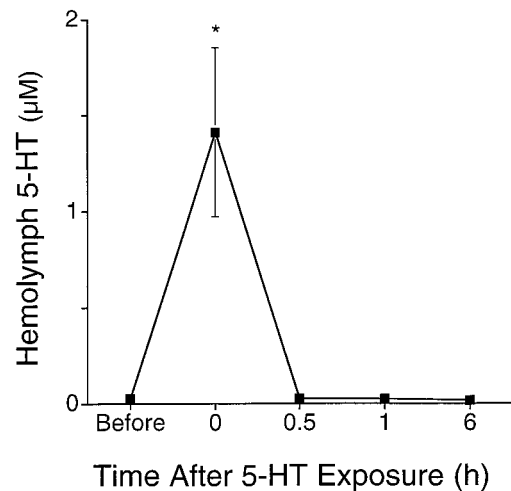


Figure 6. Disappearance of 5-HT from the hemolymph. Concentration of 5-HT in the hemolymph is plotted as a function of different times before and after an *in vivo* exposure to 5-HT (Alberini et al., 1994). Hemolymph was sampled from sensitized animals just before, immediately after, and then 0.5, 1, and 6 hr after a 0.5 hr treatment of $250 \mu\text{M}$ 5-HT. The *in vivo* treatment increased the concentration of 5-HT to $1.4 \pm 0.4 \mu\text{M}$. By 0.5 hr after the end of the treatment, levels of 5-HT returned to baseline values. Three to four animals were sampled at each time point. Animals were only sampled once. No significant differences were observed for samples collected 0.5 hr after treatment or later when compared to baseline values.

meable to 5-HT. To ascertain whether long-term increases in 5-HT could be caused by its persistence in the hemolymph once it was elevated in sensitized animals, we investigated how long an increase in 5-HT in the hemolymph would persist. Sensitized animals were used in this experiment because the turnover of 5-HT or its diffusion through the skin might be altered by the training procedure. Animals were sensitized and then bathed in $250 \mu\text{M}$ 5-HT for 0.5 hr to elevate 5-HT in the hemolymph (Alberini et al., 1994). Hemolymph was sampled from individual animals before and at various times after exposure to 5-HT. Immediately after this treatment, 5-HT in the hemolymph was $\sim 1.4 \pm 0.4 \mu\text{M}$ (Fig. 6; $F = 36.9$; $df = 16$; $p < 0.0001$; MSD = 188 μM ; $p < 0.05$). By 0.5 hr after the end of treatment, 5-HT had returned to baseline values (Fig. 6). Similar results were obtained when this experiment was performed in nonsensitized animals and when smaller changes in hemolymph 5-HT were produced (data not shown). It is possible that the clearance mechanisms used for micromolar concentrations of 5-HT differ from those used at the nanomolar concentration. If this is so, there should be a rapid decrease in levels of 5-HT in the hemolymph from micromolar to nanomolar, followed by a slower decay back to baseline. This pattern was not seen in our data. Levels of 5-HT returned to baseline within 0.5 hr. This suggests that there is no “slow” decay for 5-HT. Therefore, since increases in 5-HT in the hemolymph produced by *in vivo* exposure to 5-HT lasted relatively brief periods of time, long-term sensitization training appears to lead to a prolonged release of 5-HT from serotonergic neurons or other sources that lasts at least for 24 hr.

The long-lasting increase in 5-HT in the hemolymph could be produced by mechanisms similar to those responsible for producing long-term memory in *Aplysia* and other organisms. New protein synthesis appears to be a general requirement during the induction phase of long-term memory (Flexner et al., 1963; Agranoff, 1967; Barondes, 1975; Agranoff et al., 1978; Davis and

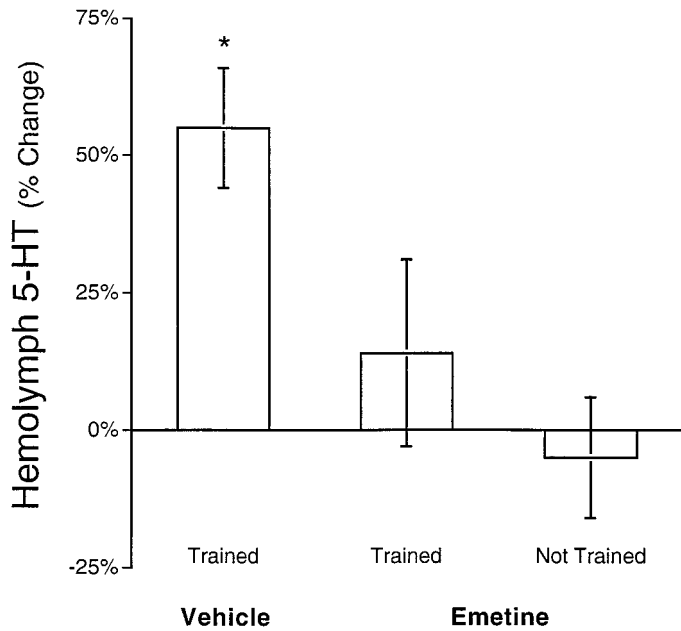


Figure 7. Inhibition of protein synthesis blocks the 24 hr increase in hemolymph 5-HT caused by sensitization training. The level of 5-HT in the hemolymph was measured 24 hr after sensitization training and compared to baseline values measured 1 hr before training. Animals were injected with 1 ml, 9 mM emetine (in BFSW with 3% ethanol) per 140 gm body weight 0.5 hr before sensitization training. The control group received an injection of vehicle (BFSW with 3% ethanol) 0.5 hr before sensitization training. There was a significant increase in 5-HT in the hemolymph of the control group 24 hr after the end of sensitization training ($p < 0.05$). However, animals that received an injection of emetine before sensitization training did not have increased levels of hemolymph 5-HT. Also, injection of emetine alone (*Not Trained* group) did not alter levels of 5-HT 24 hr later. The asterisk indicates that the control group was significantly different from the two groups that received emetine. Each group consisted of seven different animals.

Squire, 1984; Castellucci et al., 1986; Goelet et al., 1986; Milner et al., 1998). Therefore, we investigated whether the 24 hr increase in 5-HT in the hemolymph was dependent on protein synthesis. The protein synthesis inhibitor emetine, which has been used for studies both *in vivo* and *in vitro* in *Aplysia* (Schwartz et al., 1971; Castellucci et al., 1986; Montarolo et al., 1986; Hegde et al., 1997; Martin et al., 1997), was injected into animals 0.5 hr before electrical stimulation. Inhibition of protein synthesis does not appear to have effects on the neurophysiological properties of *Aplysia* neurons (Schwartz et al., 1971) or synaptic transmission (Castellucci et al., 1986; Montarolo et al., 1986). The concentration of emetine in the hemolymph after injection (1 ml, 9 mM per 150 gm body weight) was estimated to be $\sim 100 \mu\text{M}$. The effect of emetine on protein synthesis was assayed by measuring [^3H]leucine incorporation into protein in ganglia removed from *Aplysia* 0.5 hr after they were injected with emetine (see Materials and Methods). Emetine injections inhibited protein synthesis by 93% ($\pm 2\%$; $n = 3$) in the pleural-pedal ganglia and 91% ($\pm 2\%$; $n = 3$) in the abdominal ganglion. Emetine blocked the increase in 5-HT in the hemolymph 24 hr after electrical stimulation (Fig. 7; $F = 5.42$; $df = 20$; $p < 0.02$; $\text{MSD} = 24\%$; $p < 0.05$). Furthermore, emetine injection alone did not change levels of 5-HT in the hemolymph (Fig. 7). Although other neural effects cannot be ruled out, the results suggest that blockage of the long-term increase in hemolymph 5-HT by emetine was caused by inhibition of protein synthesis.

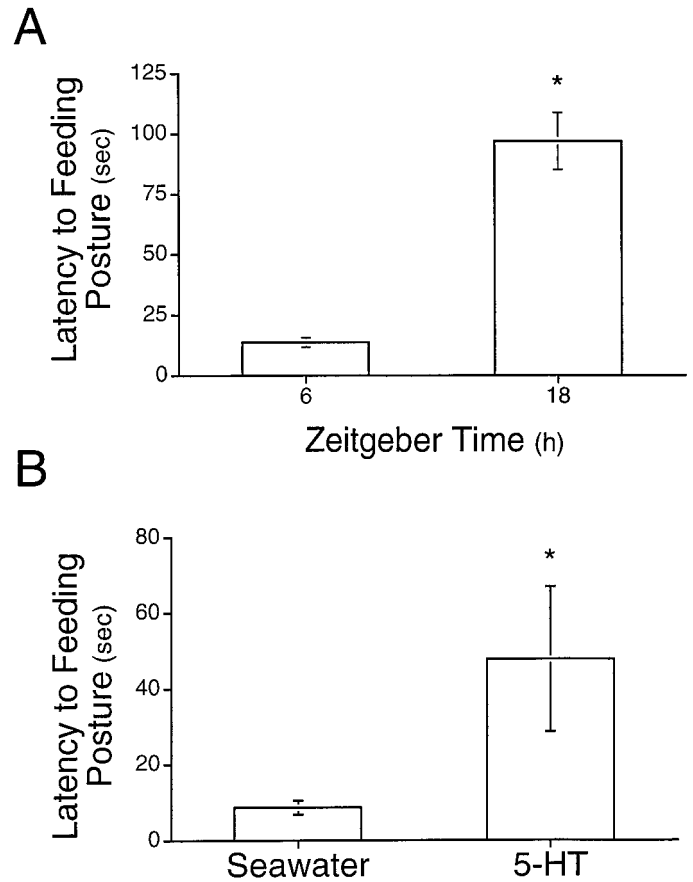


Figure 8. 5-HT in the hemolymph regulates feeding. *A*, Latency to assumption of feeding posture expressed a significant diurnal rhythm (Kupfermann, 1974). *B*, Injection of 5-HT into the hemocoel of *Aplysia* in the middle of the day (ZT6) significantly lengthened the latency to assume feeding posture. Injections were designed to elevate 5-HT in the hemolymph to levels normally seen at night. Asterisks indicate a significant ($p < 0.05$) difference relative to values at ZT6.

Influence of hemolymph 5-HT on feeding

There are many possible functional consequences of changes in the levels of 5-HT in the hemolymph (for example, see Table 1). To begin investigating functional consequences of elevations of 5-HT in the hemolymph, we examined an aspect of appetitive feeding behavior, since 5-HT has been implicated in regulation of feeding behavior (Weiss et al., 1975; Weiss et al., 1978; Kupfermann et al., 1979). *Aplysia* were exposed to dried seaweed, and the time required to elicit the feeding posture was measured (see Materials and Methods). Confirming Kupfermann (1974), we found that *Aplysia* expressed a diurnal rhythm of latency to assume feeding posture (Fig. 8*A*). *Aplysia* responded much more quickly during the day (13.7 ± 1.9 sec) than they did at night (97.0 ± 11.8 sec). This rhythm was specific for food, because stimulation of animals with filter paper failed to elicit the feeding posture at either time ($n = 5$; data not shown). To investigate whether 5-HT in the hemolymph might be involved in modulating feeding latency, 5-HT was increased in the hemolymph at a time during the day (ZT6) when 5-HT was normally low. We predicted that increasing 5-HT during the day would lead to an increase in the latency to assume feeding posture, similar to what is normally observed at night when 5-HT is elevated. Experimental animals received an injection of 5-HT during the day that increased levels

of 5-HT in the hemolymph within 5 min to 25.8 ± 4.9 nM, a level normally seen at night (see Materials and Methods; Fig. 2). As predicted, animals that were injected with 5-HT had significantly longer latencies to feed than animals injected with saline (Fig. 7B; $U = 60$; $df = 16$; $p < 0.05$).

DISCUSSION

This study demonstrates that environmental events that affect the state of the animal (LD cycles, sensitization training) also regulate levels of 5-HT in the hemolymph. Furthermore, levels of 5-HT in the hemolymph were significantly elevated after arrival of the animals in the laboratory relative to levels measured 3 d later (Fig. 1). The level of 5-HT in the hemolymph is close to threshold values reported for several physiological responses (Table 1). Therefore, the changes in 5-HT we observed, given their magnitudes and persistent nature, are probably sufficient to produce significant effects on the properties of neurons. In support of this idea, we found that elevating 5-HT in the hemolymph from a “day” level to a “night” level led to an increase in latency to assume feeding posture (Fig. 8). Our findings, taken together with those of others showing effects of injections of 5-HT on behavior (Palovcik et al., 1982; Mackey and Carew, 1983; Parsons and Pinsker, 1989), indicate that 5-HT in the hemolymph may play a humoral role and modulate the behavior of *Aplysia*.

Levels of 5-HT in the hemolymph could be regulated via several processes. A number of possible sites of release of 5-HT into the hemolymph exist. Electrical stimulation or training of the animal may acutely release 5-HT from synapses in the CNS or peripheral nervous system and this 5-HT may “spill over” into the hemolymph (Peretz and Estes, 1974; Tritt et al., 1983; Goldstein et al., 1984; Ono and McCaman, 1984; Kistler et al., 1985; Longley and Longley, 1986; Rathouz and Kirk, 1988; Zhang et al., 1991; Hernádi et al., 1992). Also, the eyes are surrounded by a basket of serotonergic fibers originating from the accessory optic nerve, and the blood vessels of *Aplysia* are highly innervated by serotonergic processes (Liebeswar et al., 1975; Takahashi et al., 1988; Alevizos et al., 1989; Skelton and Koester, 1992). Finally, hemocytes may also release 5-HT into the hemolymph (Stefano et al., 1989; Ottaviani and Cossarizza, 1990; Ottaviani et al., 1992; Levenson and Eskin, unpublished observations). Clearance of 5-HT by the kidneys or hemocytes (Goldman and Schwartz, 1977) may also contribute to the regulation of 5-HT in the hemolymph.

The possible sites of action of 5-HT in the hemolymph are diverse. In the nervous system, humoral 5-HT may exert its effects through synaptic 5-HT receptors, or perhaps through extrasynaptic 5-HT receptors located on cell bodies (Drummond et al., 1980b; Li et al., 1995; Angers et al., 1998). Humoral 5-HT would reach these sites through delivery via the open circulatory system and by penetration of the connective tissue sheath. Humoral 5-HT appears to have ready access to cell bodies, and perhaps slightly less access to the neuropil (Brunelli et al., 1976; Kaczmarek et al., 1978; Drummond et al., 1980a; Byrne et al., 1991; Emptage et al., 1996; Bunge et al., 1997).

Diurnal variation of 5-HT in the hemolymph

The diurnal rhythm of 5-HT in the hemolymph may play a role in regulating or modulating daily behaviors. In several invertebrates, 5-HT and other neurohormones have been shown to be important modulators of behavior or the neuronal circuitry underlying those behaviors (Poon, 1980; Livingstone et al., 1980; Willard, 1981; Schwarz et al., 1984; Claassen and Kammer, 1985; Marder et al., 1986; Mulloney et al., 1987; Tuersley and McCrohan, 1987; Nus-

baum and Marder, 1988; Mangan et al., 1994; Malyshev et al., 1997). Feeding behavior in *Aplysia* undergoes daily changes (Kupfermann, 1974; Fig. 8A), and we have found that those changes correlate with the level of 5-HT in the hemolymph (Fig. 8A). Specifically, when levels of 5-HT in the hemolymph were low, latency to feed was low. When levels of 5-HT in the hemolymph were high, feeding latency was high. Moreover, when levels of 5-HT in the hemolymph were elevated to nighttime levels by injection of 5-HT during the day, latency to feed increased (Fig. 8B). These results suggest that one function of the diurnal rhythm of 5-HT in the hemolymph may be to regulate feeding behavior. Further tests of this specific idea will require examining feeding latency when levels of 5-HT in the hemolymph are lowered or 5-HT receptors are blocked *in vivo* during the night.

Another daily behavior that could be regulated by 5-HT in the hemolymph is locomotor activity. Injections of 5-HT increase locomotor activity (Palovcik et al., 1982; Mackey and Carew, 1983; Parsons and Pinsker, 1989; see also Adamo and Chase, 1991), and the diurnal pattern of locomotor activity peaks during the day (Lickey et al., 1977). However, the levels of 5-HT in the hemolymph are lowest during the day, when activity is highest. In the studies that examined the effects of injection of 5-HT into *Aplysia*, behavior was measured no more than 15 min after injection, and the effective concentration of 5-HT after injection was much higher than levels observed in this study. Thus, lower levels of 5-HT might have opposite effects than high doses. In addition, acute treatments may produce different effects than longer term treatments. Indeed, modulation of pituitary lactotroph prolactin secretion by endothelin-1 is either inhibitory or stimulatory depending on whether the cells are exposed to an acute or chronic treatment of dopamine, respectively (Kanyicska et al., 1995, 1997).

Elevation of 5-HT caused by electrical stimulation

A significant increase in levels of 5-HT in the hemolymph was observed in our studies during electrical stimulation that leads to sensitization (Fig. 4). This finding is the first evidence that release of 5-HT actually accompanies sensitization training in *Aplysia* and, as such, further suggests that 5-HT plays a role in mediating the effects of sensitization training. Moreover, the increase in siphon withdrawal duration was correlated with the magnitude of the increase in 5-HT in the hemolymph (Fig. 5). This further indicates a relationship between elevation of 5-HT in the hemolymph and sensitization.

A significant increase in 5-HT in the hemolymph also occurred 1.5 and 24 hr after electrical stimulation (Fig. 4). The long-lasting increase in 5-HT in the hemolymph appeared to be caused by a persistent release of 5-HT from serotonergic neurons or other sources, because the half-life of 5-HT in the hemolymph was relatively short (Fig. 6). These results demonstrated that a single 1.5 hr block of electrical stimulation led to an increase in the levels of 5-HT in the hemolymph that persisted for 24 hr. Multiple blocks of such stimulation performed over several (2–4) days leads to sensitization that persists for weeks (Pinsker et al., 1973). It would be interesting to examine the effect of multiple blocks of electrical stimulation on the duration of the increase in 5-HT to determine whether the increase in 5-HT in the hemolymph might persist even longer than a day.

Formation of long-term memory in *Aplysia* and many other organisms requires protein synthesis during the induction phase of learning (Davis and Squire, 1984; Golet et al., 1986; Milner et al., 1998). Similarly, many of the long-lasting changes in the

Aplysia nervous system believed to be responsible for memory have been shown to be dependent on protein synthesis (Castellucci et al., 1986; Montarolo et al., 1986; Manseau et al., 1998; Bailey et al., 1992; O'Leary et al., 1995). Like these other processes, the 24 hr increase in 5-HT in the hemolymph induced by sensitization training was blocked via inhibition of protein synthesis (Fig. 7). The similarities in time course and requirement of new protein synthesis between changes in levels of 5-HT in the hemolymph and long-term changes responsible for memory indicate that changes in 5-HT in the hemolymph might be one of the constellation of factors that contribute to various forms of long-term memory in *Aplysia*. Moreover, some of the same molecular mechanisms (Abel et al., 1998) involved in the formation of long-term memory produced by sensitization training may also be involved in producing the long-lasting increase in 5-HT in the hemolymph.

What is the role of the elevated levels of 5-HT in the hemolymph or the proposed persistent increase in activity of serotonergic neurons produced by sensitization training? An increase in circulating 5-HT produced by training may increase the sensitivity of peripheral mechanoreceptors and photoreceptors (Eskin and Maresh, 1982; Billy and Walters, 1989). Elevations of 5-HT may also elevate heart rate and increase vascular tone (Liebeswar et al., 1975; Krontiris-Litowitz et al., 1989). Nonessential behaviors may be inhibited by increases in circulating 5-HT. Humoral effects of 5-HT could contribute to, or even cause in some cases the behavioral sensitization seen immediately after sensitization training and 24 hr later. It is important to note that in some instances, including the present study, electrical stimulation leads to long-term unilateral sensitization (Scholz and Byrne, 1987; Goldsmith and Byrne, 1993; Cleary et al., 1998). In these cases, humoral 5-HT could not contribute to the induction of sensitization unless the unilateral sensitization was produced by an associative mechanism involving humoral 5-HT, and neural activity on the side of the animal that was trained.

The increase in humoral 5-HT during training also could strengthen memory formation by helping to induce facilitation of the sensorimotor synapse and interneuronal connections (Bailey et al., 1981; Trudeau and Castellucci, 1993; Xu et al., 1995). This possible role of an increase in humoral 5-HT would be similar to the proposed enhancing role of catecholamines in memory formation of mammals (for review, see McGaugh et al., 1996). In the rat, elevation of epinephrine in the plasma during aversive training enhances memory acquisition (McCarty and Gold, 1981; Galvez et al., 1996).

5-HT was also elevated in the hemolymph 1.5 and 24 hr after sensitization training of *Aplysia*. The prolonged elevation of 5-HT or the activation of neurons causing the prolonged elevation of 5-HT may aid in the induction of facilitation and participate in the mechanism maintaining facilitation for long periods of time. Thus, the prolonged release of 5-HT might be part of the "memory" for long-term sensitization. Moreover, the prolonged release of 5-HT might potentiate future training as epinephrine does in the rat, and be relevant for additional training that lasts for several days. Subsequent training sessions of *Aplysia* on following days would be done in the presence of heightened activity of serotonergic neurons or elevated 5-HT in the hemolymph. Prolonged increases in neuronal activity and levels of humoral factors present a possible mechanism for strengthening formation of memories when learning takes place over a relatively long time scale, such as days or weeks.

Our findings raise the possibility that 5-HT may act as a

neuroendocrine molecule in the hemolymph to regulate the behavioral state of *Aplysia*. Humoral 5-HT may directly mediate some events, whereas it may act as a modulator in other cases. The acute changes in levels of 5-HT in the hemolymph may have different roles than the chronic presence of 5-HT in the hemolymph (Kanyicska et al., 1995, 1997). It is also possible that 5-HT released at synapses because of persistent neural activity induced by sensitization training may be more or less important than humoral 5-HT. Ultimately, it will be important to determine the role of 5-HT in the hemolymph and distinguish its humoral role from its neurotransmitter role. It will also be important to determine the site of release of 5-HT into the hemolymph and the mechanisms responsible for persistence of 5-HT in the hemolymph after sensitization training.

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