

High Corticosterone Levels in Prenatally Stressed Rats Predict Persistent Paradoxical Sleep Alterations

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Prenatal stress predisposes rats to long-lasting disturbances that persist throughout adulthood (e.g., high anxiety, dysfunction of the hypothalamo–pituitary–adrenal axis, and abnormal circadian timing). These disturbances parallel to a large extent those found in depressed patients, in which hypercortisolemia and sleep alterations may be related to stress-inducing events. We studied sleep–wake parameters in control and prenatally stressed adult rats (3–4 months old) and examined possible relationships with their corticosterone levels (determined at 2 months of age). Under baseline conditions, prenatally stressed rats showed increased amounts of paradoxical sleep, positively correlated to plasma corticosterone levels. Other changes include increased sleep fragmentation, total light slow-wave

sleep time, and a slight decrease in the percentage of deep slow-wave sleep relative to total sleep time. During recovery sleep from acute restraint stress, all sleep changes persisted and were correlated with stress-induced corticosterone secretion. High corticosterone levels under baseline conditions as well as an acute stress challenge may thus predict long-term sleep–wake alterations in rats. Taken together with other behavioral and hormonal abnormalities in prenatally stressed animals, the pronounced changes in sleep–wake parameters that are similar to those found in depressed patients suggest that prenatal stress may be a useful animal model of depression.

Key words: prenatal stress; sleep–wake parameters; paradoxical sleep; corticosterone; restraint stress; rat

Prenatal stress in rats can lead to profound behavioral and endocrine abnormalities in adult animals. Many of these abnormalities parallel those found in human depression, suggesting that the prenatally stressed (PNS) rat may be an animal model for depression. For example, PNS rats exhibit “behavioral despair” (Alonso et al., 1997) in the forced swimming test, a proposed measure of “learned helplessness” in rodents (Porsolt et al., 1977). They also show increased reactivity to novelty (open field and Y maze) and a higher propensity to self-administer amphetamine (Deminière et al., 1992), indicating an enhanced emotional reactivity (Joffe, 1978; Vallée et al., 1997; Weinstock, 1997) and anxiety (Vallée et al., 1997). In addition, feedback inhibition of the hypothalamo–pituitary–adrenal (HPA) axis by circulating glucocorticoids is impaired in both PNS rats and depressed patients (Holsboer et al., 1984; Maccari et al., 1995; Barbazanges et al., 1996; Modell et al., 1997; Vallée et al., 1997; Plotsky et al., 1998). Abnormalities in a variety of overt circadian rhythms, including the cortisol rhythm, have been documented in depressed patients (Wirz-Justice, 1995; Rosenwasser and Wirz-Justice, 1997), and we have recently discovered that PNS adult rats also show disturbances in circadian rhythms (locomotor activity, corticosterone secretion) that are consistent with those observed in depressed humans (Koehl et al., 1997, 1999; Van Reeth et al., 1998).

One of the hallmarks of human depression is an alteration in the sleep–wake cycle, including a shortened rapid eye movement (REM) sleep latency, an increase in the amount and frequency of REM sleep during the first part of the night, increased sleep fragmentation, and a decrease in the amount of slow-wave sleep (Kupfer and Reynolds, 1992). In the present study we sought to determine whether prenatal stress could have long-term effects on the sleep–wake cycle in adult rats. On the basis of previous findings on the influence of stress on sleep regulation (Cespuglio et al., 1995), the sleep patterns of PNS rats in response to acute stress were assessed and compared to controls. In view of the established correlation between HPA axis disturbances and some behavioral abnormalities in adult PNS rats (Vallée et al., 1997), we evaluated possible correlations between sleep–wake parameters and plasma corticosterone levels under both baseline and acute stress conditions.

Our results demonstrate pronounced effects of prenatal stress on sleep in adult rats that parallel, to some extent, changes in sleep architecture found in depressed patients. These data add further support for the use of the PNS rat as an animal model for depression. The persistence of sleep abnormalities and other depressive-like symptoms even after termination of the adverse stimulus in the prenatal stress model makes it a particularly attractive model for the design and testing of new therapeutical strategies in mood and sleep disorders.

Received Feb. 23, 1999; revised June 21, 1999; accepted July 20, 1999.

This work was supported by the Belgian Fonds National de la Recherche Scientifique and Fonds de la Recherche Scientifique Médicale, a NATO collaborative research grant (960771), and a Commissariat Général aux Relations Internationales–Institut National de la Santé et de la Recherche Médicale grant to O.V.R. and S.M. We thank R. Biermans, A. Heylen, and W. Van Den Broeck for their skillful technical assistance.

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MATERIALS AND METHODS

Animals. Adult virgin Sprague Dawley female rats (IFFA Credo, Brussels, Belgium) weighing ~240 gm were first group-housed (10 per cage, size ≈ 60 × 80 cm) for 10 d in order to coordinate their estrous cycle, and then individually housed for a full estrous cycle (4 d) in the presence of a sexually experienced male Sprague Dawley rat weighing ~400 gm. Mating was performed in a separate and very quiet animal room. Males

were then removed from the cages, and pregnant females were randomly assigned to PNS or control (CONT) groups, individually housed in plastic cages, allowed *ad libitum* access to food and water, and maintained on a 12 hr light/dark cycle (LD; lights on, 8:30 A.M. to 8:30 P.M.; light intensity, ~100 lux) at constant room temperature ($22 \pm 2^\circ\text{C}$) and humidity (60%). After birth, pups were raised with their mother in a separate room. All animal procedures were approved by the institutional animal care and use committee.

Prenatal stress procedure. Prenatal stress was performed daily during the last week of pregnancy until delivery. Pregnant females were individually restrained three times a day (at 9:00 A.M., 12:00 A.M., and 5:00 P.M.) for 45 min in transparent plastic cylinders (7 cm diameter; 19 cm long) and exposed to bright light (1500 lux). Control pregnant females were left undisturbed in their home cages. Offspring were weaned 21 d after birth and housed in same-sex groups of four until the age of 2 months. The animals tested were derived from a total of five litters per condition (five CONT, five PNS). Only litters of 8–13 pups with similar numbers of males and females were used for the study. Only one or two male pups per litter were studied as adults to eliminate any possible “litter effects” (Chapman and Stern, 1979) on the measured variables. Experiments were performed on eight male CONT and eight PNS rats. No female rats were included to avoid the influence of the estrous cycle on spontaneous sleep patterns (Schwierin et al., 1998).

Blood sampling and corticosterone measurements. At 2 months of age blood samples were collected (via the tail vein) three times between 9:30 and 11:30 A.M. Rats were moved to an adjacent room and were placed individually in a restraint transparent tube, and blood was collected quickly (<2 min) to determine basal corticosterone levels. The second sampling was performed 20 min after restraint stress was initiated. The rats were then returned to their home cages with their usual partners, until the last blood sampling was performed 60 min after the initiation of the stress procedure. Restraint was performed in plastic cylinders identical to those used for the prenatal stress procedure. Blood corticosterone levels were determined by radioimmunoassay using a highly specific corticosterone antiserum (ICN Biomedicals, Cleveland, OH) with a detection threshold of 0.1 $\mu\text{g}/100$ ml. The interassay and intra-assay variations were, respectively, 6 and 3.5% at a mean value of 1.5 ng/tube and 6.8 and 4% at a mean value of 10 ng/tube.

Electrode implantation and EEG sleep recording. One month after the restraint stress procedure, the rats were moved to a sleep recording room. The animals were implanted under deep anesthesia (Thalamonal; 1 ml, s.c.) with chronic electrodes for polygraphic recordings of frontoparietal electroencephalogram (EEG), electrooculogram (EOG), and nuchal electromyogram (EMG). All electrodes were attached to a microconnector and fixed to the skull with dental cement. After surgery, the rats were individually housed in Plexiglas cages (30 cm diameter, 40 cm high), maintained under similar environmental conditions as before and left undisturbed for 2 weeks. The animals were then habituated to the sleep recording procedure for the next 14 d. In their home cages and in the same room, the rats were connected with a cable to a rotating swivel allowing free movements, and EEG, EOG, and EMG activities were recorded on a polygraph (EEG-4414 A/K; Nihon-Khodon) with an output connected to a computer for on-line spectral analysis of the EEG. Habituation consisted of two recording sessions for 8 hr and two sessions of 24 hr.

At the end of the habituation period, sleep was recorded for a period of 24 hr, beginning at the onset of the light phase. Over the next month, when the rats were 4 months of age, sleep was recorded during two additional sessions. In the first of these sessions, sleep was recorded for 22 hr starting 2 hr after light onset, and served as a baseline for the effects of a restraint stress procedure on sleep 2 d later. Restraint, for 1 hr beginning 1 hr after lights on, was performed on a table next to the home cage, under bright light (1500 lux) in plastic cylinders identical to those used for the prenatal stress procedure. Polygraphic recordings were again obtained for the following 22 hr time interval.

Data analysis. Polygraphic recordings were visually scored by 30 sec epochs. Those epochs were classified as being either wake (W), light slow-wave sleep (SWS1), deep slow-wave sleep (SWS2), or paradoxical sleep (PS), as described earlier (Dugovic et al., 1989). Briefly, the different vigilance states were characterized as follows: W, low-voltage fast EEG activity, high EMG activity, and numerous eye movements; SWS1, high-voltage slow cortical waves interrupted by low-voltage fast waves, and reduced EMG activity; SWS2, continuous high-amplitude slow-wave activity in EEG, very low EMG activity, and no EOG activity; and PS, low-voltage fast cortical waves with a regular theta rhythm,

absence of muscular tone, and presence of rapid eye movements. Scores were entered into a computer that calculated various sleep–wake parameters: amount of time spent in the four vigilance states and number and duration of episodes for each state. Sleep–wake parameters were analyzed over 2 and 4 hr time intervals, as well as over the total light and dark phases. The duration of time spent in the different states of vigilance was expressed in minutes as well as a percentage of recording time.

All software routines for EEG spectral analysis were written using a VAX lab 1.4 Scientific Library. The EEG signal was high-pass frequency filtered at 1 Hz and low-pass frequency filtered at 35 Hz. The analog signal was digitized at a sampling rate of 102.4 per sec with a 12 bit resolution, to yield epochs of 5 sec duration composed of 512 samples. A spectral window filter (Welch) was applied to the digitized signal. Every 5 sec, Fast Fourier Transformations were performed on these 512 data points within the frequency range of 1–32 Hz and with a frequency resolution of 0.2 Hz. EEG power spectra in the different states of vigilance were calculated by computing average power density values in each frequency bin per 2 hr time interval.

Parameters of the sleep–wake cycle of CONT animals were compared with those of the PNS animals under baseline conditions. The effects of restraint on sleep in the CONT and PNS animals were also analyzed to determine whether the two groups responded differentially to the stress procedure. Correlations between any sleep–wake differences at the age of 3–4 months and basal and stress-induced corticosterone secretion obtained at 2 months of age were assessed.

Sleep data were analyzed with factorial ANOVA at different times across the entire recording period. Subsequently, differences between groups were assessed by means of the unpaired two-tailed Student's *t* test. Within each group, differences between baseline and recovery from 1 hr restraint stress were assessed by means of paired two-tailed Student's *t* tests for the sleep–wake parameters and by the Wilcoxon signed-rank test for the EEG power spectral values. ANOVA was used to compare values of plasma corticosterone levels in response to the restraint procedure between the two groups of rats (factorial ANOVA, treatment effect) and whether there was an interaction between treatment (prenatal stress) and the time of blood sampling. Relationships between the different sleep–wake parameters and corticosterone levels were assessed by Pearson's correlation analyses. The area under the curve (AUC) was calculated by the use of the trapezoidal rule. All values are expressed as mean \pm SEM.

RESULTS

Basal and stress-induced corticosterone secretion

Mean (\pm SEM) basal corticosterone levels in CONT (0.38 ± 0.06 $\mu\text{g}/100$ ml) and PNS (1.70 ± 0.83 $\mu\text{g}/100$ ml) animals were not significantly different (ANOVA, $F_{(1,14)} = 2.434$; $p = 0.1411$). Stress-induced corticosterone secretion differed between the two groups (ANOVA, interaction treatment \times time, $F_{(2,28)} = 5.591$; $p = 0.0091$). Higher corticosterone values were observed in PNS rats, both 20 min (CONT, 20.00 ± 3.46 $\mu\text{g}/100$ ml; PNS, 39.13 ± 3.40 $\mu\text{g}/100$ ml; group effect, $F_{(1,14)} = 15.208$; $p = 0.0016$) and 60 min after the initiation of the stress procedure (CONT, 6.38 ± 0.79 $\mu\text{g}/100$ ml; PNS, 19.38 ± 4.59 $\mu\text{g}/100$ ml; group effect, $F_{(1,14)} = 7.759$; $p = 0.0147$). Furthermore, the AUC (micrograms per 100 ml \times 1 hr) for stress-induced plasma corticosterone levels was significantly larger in PNS (1578 ± 133 $\mu\text{g}/100$ ml \times 1 hr) compared to CONT (731 ± 112 $\mu\text{g}/100$ ml \times 1 hr) rats ($t = 4.841$; $p = 0.0003$) (Fig. 1).

Baseline EEG sleep and correlations with basal and stress-induced corticosterone secretion

Prenatal stress induced substantial changes in both the structure and the continuity of sleep in adult rats. A plot of the distribution of vigilance states in 4 hr bins over the 24 hr LD cycle revealed that PS was the most altered state (Fig. 2). Compared to CONT rats, PNS rats showed increased total PS time, as well as an increase in the percentage of total sleep time in PS, during both the light and dark phases (Table 1). The increase in time spent in PS over the 24 hr recording period (+33%; $p < 0.0001$) was caused by an increase in the number of PS episodes (+31% over

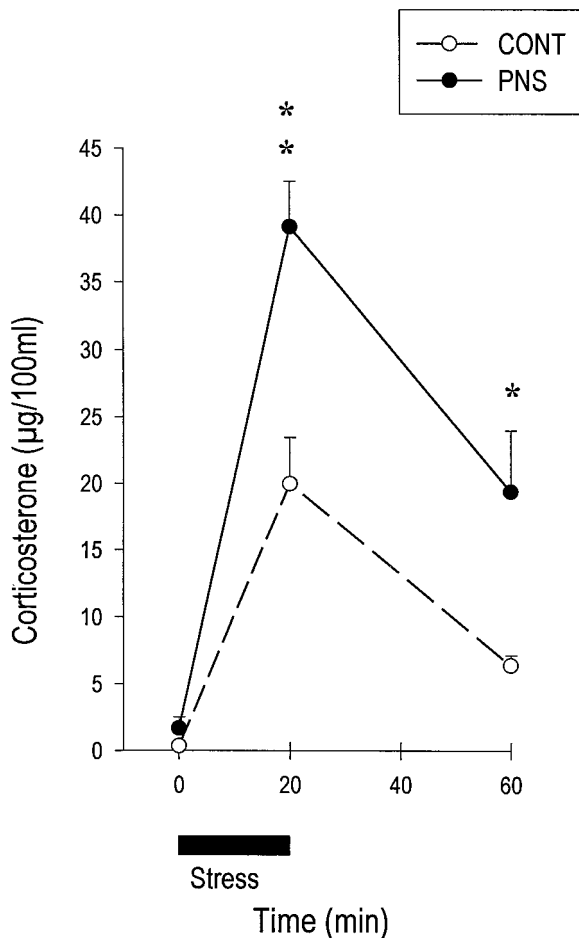


Figure 1. Corticosterone secretion induced by 20 min restraint stress (black line of abscissa) in control (CONT) and prenatally stressed (PNS) rats. At 2 months of age, PNS male rats showed higher corticosterone levels both 20 and 60 min after initiation of the restraint procedure. No change was observed in basal corticosterone levels between the two groups. * $p < 0.05$; ** $p < 0.01$ (ANOVA).

the 24 hr interval; $p = 0.0014$) (Fig. 3), whereas the mean duration of PS episodes remained the same in the two groups.

In addition to the clear effects on PS, prenatal stress also induced an increase in total SWS1 time that was restricted to the dark phase (+33%; $p = 0.0007$). Although the total SWS2 time was not significantly changed, the percentage of total sleep time in SWS2 was significantly decreased in both the light and dark phases in PNS animals (Fig. 2, Table 1). In addition, sleep was more fragmented during the dark phase in PNS animals, as indexed by a larger number of episodes in each vigilance state (Fig. 3) and a shorter duration of W episodes (CONT, 4.4 ± 0.2 ; PNS, 3.5 ± 0.1 ; $p = 0.0007$). This resulted in less time spent in W (–8%; $p = 0.0011$) during the dark phase (Table 1).

Quantitative analysis of EEG activity during specific vigilance states over the 12 hr light phase revealed minor differences in power spectral values (1–32 Hz) between the two groups of rats. EEG slow-wave activity (SWA; EEG power in the 1–5 Hz range), an indicator of sleep intensity (Borbély and Neuhaus, 1979), was evaluated during SWS2 across the light phase. SWA progressively decreased over consecutive 2 hr intervals during the light phase, and the time course for this decrease was comparable in the two groups (Fig. 4). No differences in SWA levels or in power densi-

ties within the higher frequency ranges (5–32 Hz) in any of the 2 hr intervals were observed between CONT and PNS rats (data not shown). During PS, similar power spectral values were obtained in PNS rats and CONT rats in the entire frequency range studied (1–32 Hz).

The only significant correlation between basal plasma corticosterone levels at 2 months of age and the measured sleep parameters at 3–4 months of age was a positive correlation with the total amount of PS during the 12 hr light phase ($r = 0.53$; $p = 0.0365$) as well as over the entire 24 hr period of baseline sleep recording ($r = 0.53$; $p = 0.0356$) (data not shown). The AUC for the stress-induced increase in corticosterone levels was also positively correlated with the 24 hr amount of baseline PS ($r = 0.80$; $p = 0.0002$), as was sleep fragmentation during the 12 hr dark phase ($r = 0.57$; $p = 0.0224$) (Fig. 5).

Effects of 1 hr restraint stress on sleep EEG and correlations with stress-induced corticosterone secretion

During the second baseline sleep recording period (22 hr instead of 24 hr), the sleep–wake patterns of all rats were similar to those obtained in the first recording session. Again, compared to CONT rats, PNS rats exhibited enhanced total PS time over the total 22 hr period (+32%; $p = 0.0001$), which was caused by an increased number of PS episodes (+28%; $p = 0.0131$). The percentage of PS relative to total sleep time was increased as well (+30%; $p < 0.0001$). Similarly, PNS rats spent significantly more time in SWS1 (+66%; $p = 0.0001$) and showed increased sleep fragmentation during the dark phase when compared to CONT animals. Whereas in the initial baseline sleep recording session the total SW2 time was not different between the two groups, in the second baseline session there was a significant decrease in the total SWS2 time (–13%; $p = 0.004$) during the light phase of PNS animals. The decrease of SWS2 was also observed during both the light and the dark phases when expressed as a percentage of total sleep time (data not shown).

The 1 hr restraint stress resulted in long-lasting effects on PS in both groups of rats. Compared to their baseline values, CONT and PNS rats showed significantly increased amounts of PS during the remaining 10 hr of light (CONT, +16%, $p = 0.0006$; PNS, +9%, $p = 0.0031$) and the subsequent 12 hr of dark (CONT, +30%, $p = 0.0196$; PNS, +27%, $p = 0.0018$). Although PS values were significantly higher in PNS rats compared to CONT rats during the light phase (+21%; $p = 0.0052$) (Fig. 6, top), the relative stress-induced PS increase during the light phase was significantly less in PNS than in CONT rats ($p = 0.0394$) (Fig. 6, bottom). In both groups, EEG power spectral activities during PS were not affected by restraint stress.

The 1 hr restraint stress resulted, in CONT rats only, in a moderate rebound increase of total SWS2 time limited to the first 2 hr of recovery (+20%; $p = 0.0123$) (Fig. 7, top). In both groups, SWS2 intensity (indexed by SWA) was more affected than its duration. Indeed, EEG power density in the 1–5 Hz range was significantly increased above baseline during the three 2 hr intervals after restraint, with a comparable magnitude in the two groups (Fig. 7, bottom). In response to the 1 hr restraint, other abnormalities that characterized sleep in PNS rats under baseline conditions were still present during sleep recovery, i.e., total SWS2 time decreased in the light phase (–15%; $p = 0.0001$) whereas total SWS1 time (+60%; $p < 0.0001$) and sleep fragmentation (+20%; $p = 0.0265$) increased during the dark phase.

A number of significant correlations were found between the

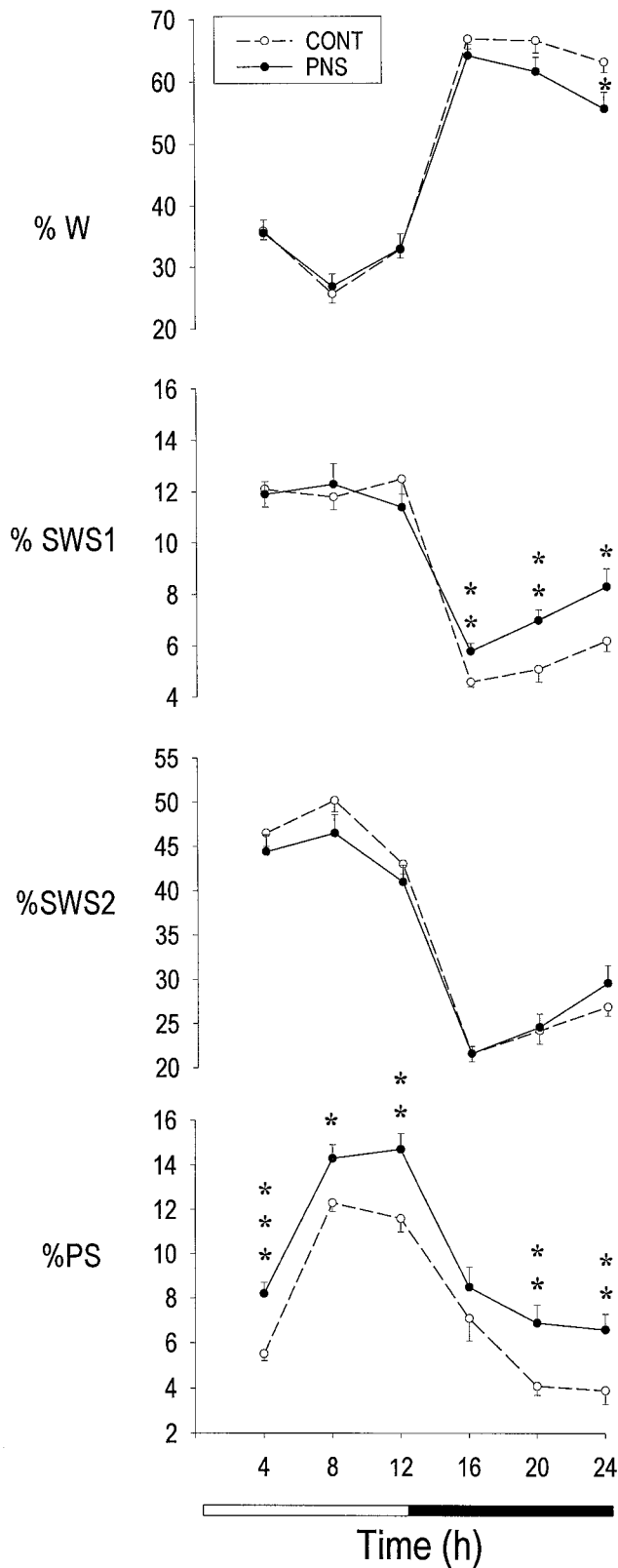


Figure 2. Distribution per 4 hr intervals of vigilance states in eight control (CONT) and eight prenatally stressed (PNS) rats under baseline conditions. Mean (\pm SEM) values of wake (W), light slow-wave sleep (SWS1), deep slow-wave sleep (SWS2), and paradoxical sleep (PS) are expressed as percentage of recording time. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$ (two-tailed unpaired Student's *t* test) for between-groups comparisons.

Table 1. Total duration of each vigilance state during the light (0–12 hr) and dark phase (12–24 hr) in control and prenatally stressed rats

Baseline vigilance states		Control	Prenatal stress
W (min)	0–12 hr	226.8 \pm 6.9	229.2 \pm 13.5
	12–24 hr	471.3 \pm 5.4	434.7 \pm 7.1**
SWS1 (min)	0–12 hr	87.3 \pm 2.2	85.5 \pm 3.4
	12–24 hr	38.2 \pm 1.8	50.8 \pm 2.4***
SWS2 (min)	0–12 hr	335.3 \pm 6.2	316.4 \pm 12.7
	12–24 hr	174.5 \pm 5.0	181.9 \pm 5.6
PS (min)	0–12 hr	70.6 \pm 2.0	89.0 \pm 3.1***
	12–24 hr	36.1 \pm 2.4	52.6 \pm 3.5**
% SWS1/TST	0–12 hr	17.7 \pm 0.5	17.5 \pm 0.9
	12–24 hr	15.4 \pm 0.7	17.8 \pm 0.9
% SWS2/TST	0–12 hr	67.9 \pm 0.4	64.3 \pm 0.9**
	12–24 hr	70.2 \pm 1.0	64.0 \pm 1.2**
% PS/TST	0–12 hr	14.3 \pm 0.4	18.2 \pm 0.6***
	12–24 hr	14.4 \pm 0.9	18.3 \pm 1.0*

Mean (\pm SEM) values ($n = 8$ rats in each group) of W, SWS1, SWS2, and PS are expressed in minutes. SWS1, SWS2, and PS are also expressed as a percentage of total sleep time (TST).

* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$ (two-tailed unpaired Student's *t* test) for between-groups comparisons.

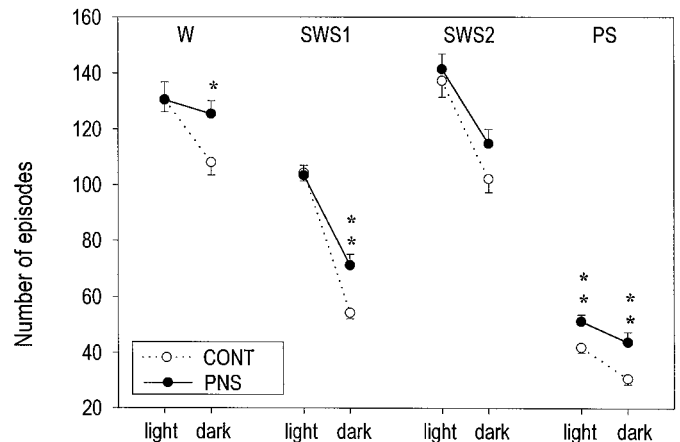


Figure 3. Mean (\pm SEM) number of episodes for each vigilance state (W, SWS1, SWS2, PS) during light or dark phases, in CONT and PNS rats under baseline conditions. * $p < 0.05$; ** $p < 0.01$ (two-tailed unpaired Student's *t* test) for between-groups comparisons.

AUC for stress-induced plasma corticosterone determined at 2 months of age and sleep–wake parameters at 3–4 months after an acute restraint stress. The amount of PS time over the 22 hr recording period ($r = 0.63$; $p = 0.0096$), as well as the amount of SWS1 time during the dark phase ($r = 0.69$; $p = 0.0030$) and the number of episodes in each vigilance state during the dark phase ($r = 0.58$; $p = 0.0197$) were all positively correlated with the AUC for corticosterone levels after acute restraint stress. In addition, stress-induced corticosterone levels were negatively correlated to the amount of SWS2 time ($r = -0.79$; $p = 0.0003$) during the light phase (Fig. 8). Thus, the stress-induced increase in corticosterone secretion revealed significant correlations with each sleep–wake parameter that was already altered under baseline conditions by prenatal stress.

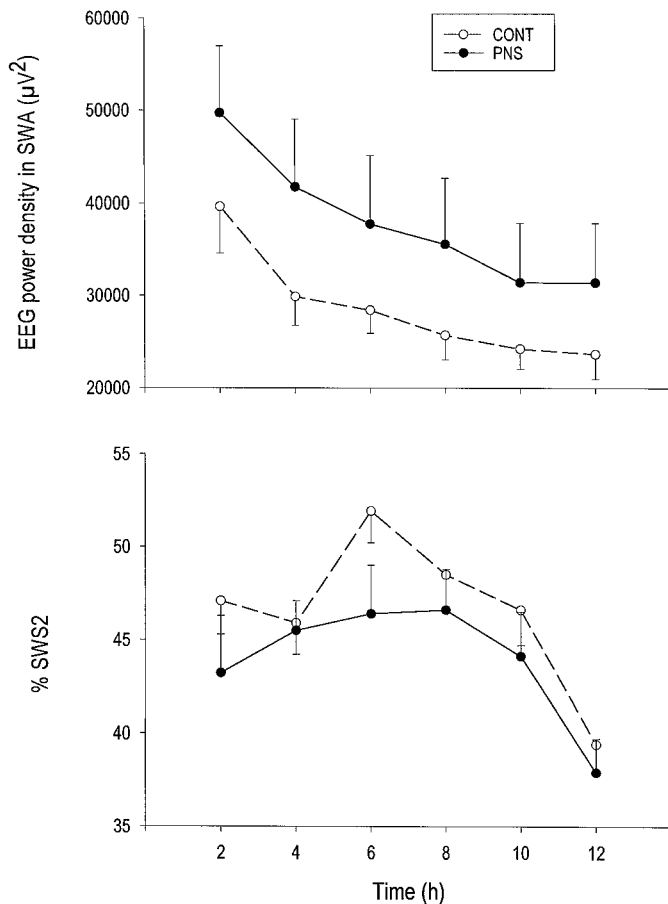


Figure 4. *Top*, Time course of mean (\pm SEM) slow-wave activity (SWA) per 2 hr intervals in SWS2 during light phase, in CONT and PNS rats under baseline conditions. SWA values are EEG power densities (square microvolts) averaged in the 1–5 Hz frequency band. *Bottom*, Corresponding mean (\pm SEM) values of SWS2 duration expressed as percentage of recording time.

DISCUSSION

The present results demonstrate that exposure to prenatal stress can produce long-term and selective changes in both the structure and the continuity of sleep. Although there are reports of abnormal “sleep-like behaviors” in PNS monkeys (Schneider, 1992) and PNS humans (Stott, 1973), our data provide the first polygraphic demonstration of long-term effects of prenatal stress on the sleep–wake cycle when the animals reach adulthood.

Baseline conditions

Under baseline conditions, sleep–wake alterations measured at the age of 3–4 months consisted primarily in an increase in PS duration at all phases of the light–dark cycle that was positively correlated to basal and stress-induced plasma corticosterone levels. Other sleep changes in PNS rats (i.e., increase in nighttime SWS1 and sleep fragmentation) were not correlated with basal corticosterone levels at the age of 2 months, suggesting that PS changes might represent a specific response to prenatal stress-induced changes in the HPA axis. The observation of larger PS amounts and a lower percentage of SWS2 relative to total sleep time in rats showing higher stress-induced corticosterone responses, further suggests an important role for the HPA axis in sleep regulation and its changes in response to prenatal stress. Interestingly, studies in humans have indeed suggested a role for

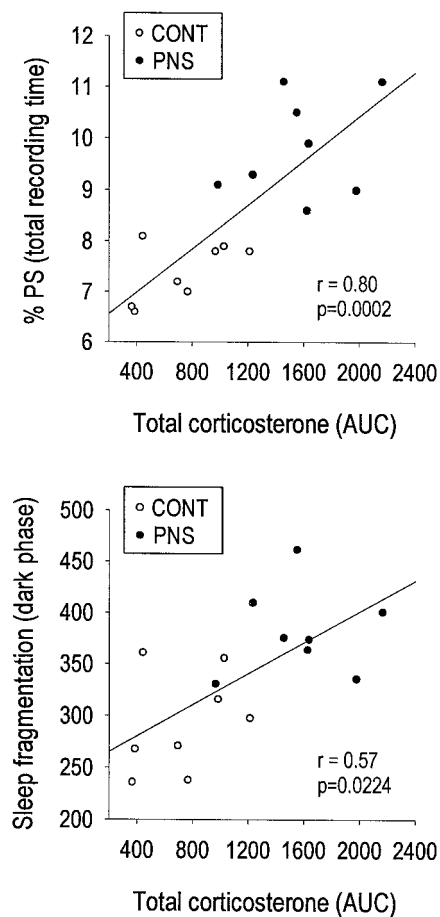


Figure 5. Positive correlations between individual stress-induced plasma corticosterone AUC values and amounts of PS expressed as percentage of total recording time (*top*) or sleep fragmentation in the dark phase (i.e., the number of episodes in each vigilance state) (*bottom*), in eight CONT and eight PNS rats. r = coefficient of Pearson’s correlation analysis.

glucocorticoids in the modulation of PS and a role for mineralocorticoids in the modulation of SWS2 (Born et al., 1991).

The finding that stress is often a predisposing factor in the development of depression (Sapolsky, 1996) has led to the hypothesis that stress-induced models of depression have construct validity (Rosenwasser and Wirz-Justice, 1997; Willner, 1997). Exposure of laboratory animals to chronic stressors at adult age can lead to PS abnormalities (i.e., PS increase, shorter PS latency). In all these models, only transient PS changes were found during or after stress. In both the learned helplessness (Adrien et al., 1991) and the intermittent foot shock paradigms (Kant et al., 1995), PS was increased only during the first day of recovery. In rats exposed to chronic mild stress, an increase in PS was only observed during the first day of stress recovery (Cheeta et al., 1997). Although an increase in PS was found after 2 weeks of chronic stress, the increase disappeared soon after stress termination (Moreau et al., 1995). Thus, persistence of PS alterations in rats exposed to stress *in utero* dramatically contrasts with the reversibility of those in the above-mentioned models. Interestingly, we have noted that even at 6 months of age, sleep–wake differences between PNS and CONT rats were still present (our unpublished data).

In addition to the major effects on PS, prenatal stress produced an increase in sleep fragmentation and in total SWS1 time re-

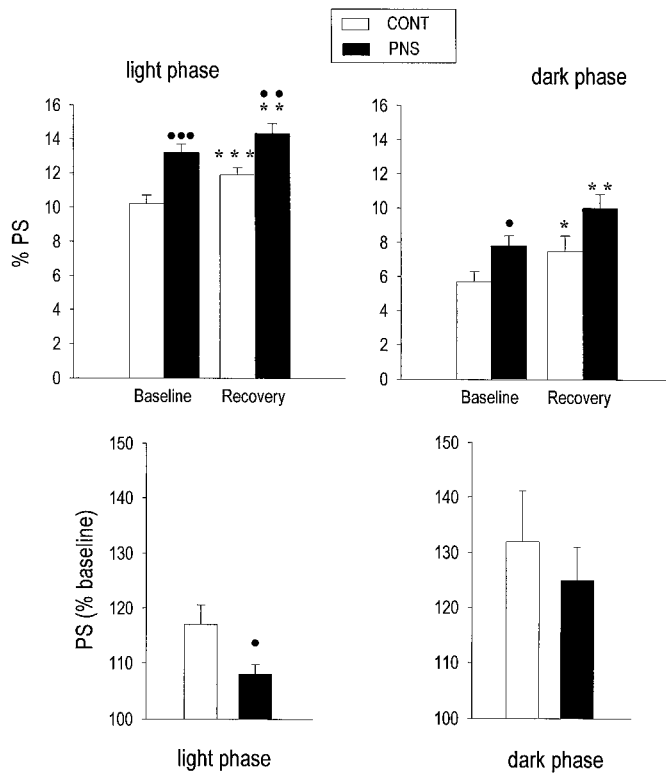


Figure 6. Amounts of PS during recovery from the 1 hr restraint stress in CONT and PNS rats. *Top*, Mean (\pm SEM) PS values (percentage of total recording time) under baseline and recovery conditions in the light (*left panel*) and the dark (*right panel*) phases. Under both baseline and recovery conditions, PNS rats had higher PS levels during the light phase compared to CONT rats ($*p < 0.05$; $**p < 0.01$; $***p < 0.001$, two-tailed unpaired Student's *t* test). During recovery sleep, a significant PS increase was observed in both groups during the light and the dark phase, as compared to their own baseline values ($*p < 0.05$; $**p < 0.01$; $***p < 0.001$; two-tailed paired Student's *t* test). *Bottom*, Mean (\pm SEM) values of PS increase during recovery in the light and the dark phase are expressed as percentage of baseline values in each group. The magnitude of PS increase in the light phase was significantly lower in PNS compared to CONT rats ($p = 0.0234$; unpaired two-tailed Student's *t* test).

stricted to the dark phase. SWS2 was decreased only when expressed as a percentage relative to total sleep time. A transient decrease in SWS2 has been reported in rats after 3 weeks of chronic mild stress (Cheeta et al., 1997).

Acute restraint stress

In response to an acute stress, there was an increase in total PS time in both groups of animals. A 1 hr immobilization stress was previously found to be followed by an increase of sleep duration (mainly PS) when the acute stress was applied either at the beginning of the dark phase (Rampin et al., 1991) or during the light phase (Bouyer et al., 1997). It is tempting to speculate that the smaller percent increase over baseline during the light phase in the PNS group could be caused by an alteration of the PS response to stress in PNS rats. However, it could also reflect a ceiling effect because the baseline levels were already elevated in the PNS animals.

The 1 hr restraint stress induced a compensatory effect on SWS2 duration only in CONT animals. Its magnitude was similar to that observed in other studies in response to either a 1 hr restraint stress or sleep deprivation (Cespuglio et al., 1995; Bonnet et al., 1997). The failure of PNS animals to show a similar

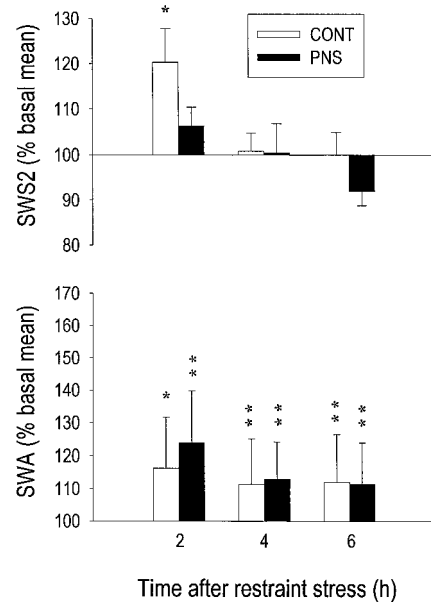


Figure 7. Effect of 1 hr restraint stress on the amount of SWS2 and on SWA (EEG power in 1–5 Hz band) during SWS2, as measured during three consecutive 2 hr intervals of recovery sleep in eight CONT and eight PNS rats. Mean values (\pm SEM) of SWS2 or SWA are presented as a percentage of their corresponding baseline mean values. Both groups showed elevated SWA during the 6 hr of analysis, whereas a significant increase in the amount of SWS2 was observed only in CONT rats during the first 2 hr interval ($*p < 0.05$; $**p < 0.01$; paired two-tailed Student's *t* test).

increase in SWS2 indicates a possible alteration in the SWS response, as does the decrease in SWS2 observed under the corresponding baseline conditions. In contrast, an increase in SWA, an index of sleep intensity, in response to restraint was observed in both groups. It should be noted that no increase in SWA was reported by others in response to a 3 hr sleep deprivation (Tobler and Borbély, 1990), indicating that the increase observed in the present study was caused by the stress of the restraint procedure and not by any possible associated sleep loss. In support of this hypothesis is the finding of a similar increase in SWA in rats exposed to a 1 hr social stress (Meerlo et al., 1997).

In response to acute stress, PNS rats show decreased binding capacity of hippocampal type I (mineralocorticoid) and type II (glucocorticoid) corticosteroid receptors (Henry et al., 1994; Maccari et al., 1995; Barbazanges et al., 1996). In view of the involvement of glucocorticoid and mineralocorticoid receptors in both SWS and PS (Friess et al., 1995), this desensitization could be related to the diminished response on PS and SWS2 after the acute stress challenge.

As shown previously, restraint stress leads to a rapid increase in plasma corticosterone (Henry et al., 1994), which in the present study was magnified at both the 20 and 60 min sampling time after the initiation of restraint. We also confirm here the previous finding that restraint stress can induce abnormally prolonged corticosterone secretion in PNS rats (Maccari et al., 1995; Barbazanges et al., 1996; Vallée et al., 1997). In the present study, significantly higher plasma corticosterone levels were found in PNS rats after 20 min of restraint, which were not observed in our previous studies in which rats were restrained for 30 min before blood sampling. These higher corticosterone levels might be explained by a different time course in plasma corticosterone levels,

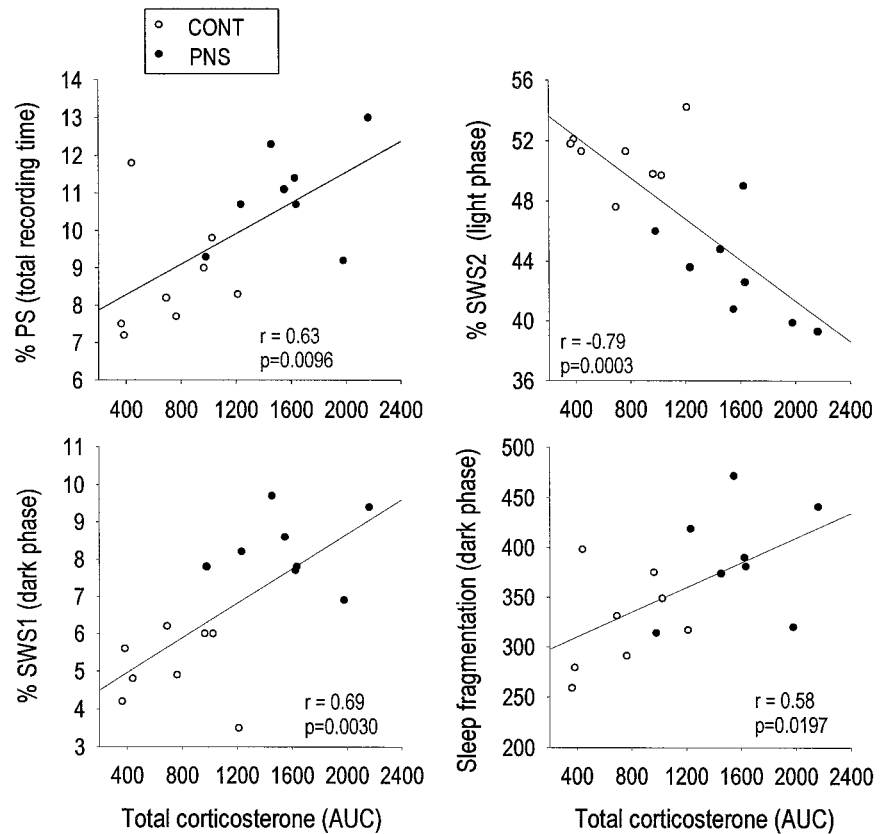


Figure 8. Correlations between stress-induced plasma corticosterone levels and sleep parameters during recovery from the 1 hr restraint stress in eight CONT and eight PNS rats. Individual corticosterone AUC values (from 0 to 60 min after stress) were positively correlated to the amount of PS (*top left panel*), the amount of SWS1 in the dark phase (*bottom left panel*), and sleep fragmentation in the dark phase (*bottom right panel*). Individual corticosterone values were negatively correlated to the amount of SWS2 in the light phase (*top right panel*). Individual amounts of PS, SWS1, and SWS2 are expressed as percentage of recording time, and sleep fragmentation is expressed as number of episodes for each vigilance state. r = coefficient of Pearson's correlation analysis.

characterized by a steeper slope of corticosterone rise in response to stress. All sleep–wake differences between the two groups found under baseline conditions were still present during the 22 hr of sleep recording after restraint stress. Interestingly, after the 1 hr restraint stress experiment, there were more robust correlations between plasma corticosterone levels (at the age of 2 months) and the various sleep parameters (at the age of 3–4 months) than under baseline conditions. In addition to the positive correlation between plasma corticosterone levels and total PS time or sleep fragmentation found under baseline conditions, in response to restraint, a positive correlation was also found with total SWS1 time in the dark phase and a negative one with total SWS2 time in the light phase. Taken together, these results indicate that high corticosterone levels can predict long-term and selective sleep–wake alterations in PNS rats.

In addition to glucocorticoids, other factors may be involved in the long-term effects of prenatal stress on sleep. Corticotropin-releasing hormone (CRH) is involved in the regulation of physiological waking (Opp, 1995) and in sleep–wake modifications induced by acute stress exposure (Gonzalez and Valatx, 1997). Under stress conditions, CRH acting as a neurotransmitter in the locus coeruleus (LC) induces an increase in activity of noradrenergic neurons, which leads to an increase in PS (Gonzalez et al., 1996; Gonzalez and Valatx, 1997). Long-term alterations of CRH neurotransmission in PNS rats (Cratty et al., 1995) and developmental changes in their LC (Peters, 1984; Johnson et al., 1992) may explain the altered PS under baseline conditions. The serotonergic (5-HT) system may also be involved in the effects of prenatal stress on the adult sleep–wake cycle because exposure to high glucocorticoid levels or to acute stressors results in significant alterations in 5-HT turnover in the midbrain–pons area in

PNS rats, which also show long-term behavioral abnormalities and/or an altered response to 5-HT receptor agonists (Peters, 1988; Muneoka et al., 1997). In view of the permissive role played by the 5-HT system on PS regulation (Jouvet, 1984) and by a number of 5-HT receptors on sleep–wake modulation (for review, see Adrien, 1995; Boutrel et al., 1999), developmental alterations in brain 5-HT metabolism may contribute to PS modifications in PNS rats. In addition, during an acute restraint stress, adrenocorticotropin hormone and its derivatives, α -melanocyte-stimulating hormone and corticotropin-like intermediate lobe peptide, released under the influence of the stress-induced 5-HT release might participate in the PS increase (Bonnet et al., 1997). A cholinergic component may be involved as well, as suggested by increased acetylcholine release after acute stress (Imperato et al., 1991; Kaufer et al., 1998). The cholinergic system is involved in the executive mechanisms of PS (Hobson et al., 1986), and recent data indicate that PNS rats have an acetylcholine hypersensitivity to CRH challenges (Day et al., 1998). Thus, permanent neurochemical changes in both monoaminergic (NA; 5-HT) and cholinergic systems might all participate in the PS modifications found in PNS rats.

Abnormalities in a variety of overt circadian rhythms have been documented in depressed patients, including rhythms of body temperature, various peripheral hormone concentrations, and urinary levels of neurotransmitter metabolites (Wehr and Goodwin, 1983). Circadian abnormalities include changes in free-running period, amplitude, cohesion, and entrainment to photic and/or social “zeitgebers” (Wehr and Goodwin, 1983; Rosenwasser and Wirz-Justice, 1997). Interestingly, circadian timing disturbances have recently been reported in PNS rats that include an advanced acrophase in the rhythm of corticosterone secretion

(Koehl et al., 1997), a phase advance in the rhythm of wheel-running behavior (Koehl et al., 1999), a shortened free-running period of locomotor activity (Maccari et al., 1997), and an abnormal rate of resynchronization of the activity rhythm after abrupt shifts in the light–dark cycle (Van Reeth et al., 1998).

Alterations in sleep–wake patterns, a common biological trait of human depression, usually include a shortened latency of the first REM sleep episode, increased REM sleep amount and frequency during the first part of the night, decreased SWS amounts, and increased sleep fragmentation (Kupfer and Reynolds, 1992). Significant correlations between sleep abnormalities and dysfunctions of the HPA axis have been shown in depressed patients (Poland et al., 1992; Hubain et al., 1998) and may result from a stress component (Sapolsky, 1996; Rosenwasser and Wirz-Justice, 1997). In the present study in adult rats, sleep–wake changes resulting from prenatal stress were consistent with those described in depressed patients. Similarly, significant correlations between these sleep changes and corticosterone levels parallel those found in depression. Added to our previous findings in PNS rats of high anxiety and emotionality, dysfunction of the HPA axis, and circadian timing abnormalities, the observation of long-term changes in their sleep structure supports the validity of the “prenatal stress” model as a valid animal model of depression. The persistence of all induced abnormalities after stressor removal could be seen as being particularly advantageous for the design and testing of new therapeutical strategies in mood and sleep disorders.

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