

Scavenger receptor class a, member 3 is associated with severity of hand, foot, and mouth disease in a case-control study

Ye Tian, MD, Kai Zhou, MD, Jing Hu, MD, Ming-Feng Shan, MD, Hong-Jian Chen, MD, Shan Cheng, MD, Li-Fei Liu, MD, Xiao-Li Mei, MD*

Abstract

Hand, foot, and mouth disease (HFMD) spreads rapidly and has been recognized as a public health problem in recent years in China. Unfortunately, there is no effective vaccine or antiviral drug currently for EV71 infection. In this study, we aim to identify biomarker which are associated with for severity of EV71 infection cases using high-throughput RNA sequencing approach.

RNA sequencing of samples from severe HFMD (S) patients group (n=10) and control HFMD (C) patients group (n=10) were performed and the results were verified by qPCR. mRNA with the highest expression level was selected to be validated in an independent cohort comprising of 45 severe EV71 infected patients and 45 control by qPCR assay.

Seventeen significant differentially expressed genes were identified. Scavenger receptor class A, member 3 (SCARA3) was one of the significantly upregulated genes with the highest expression level and was selected for validation. The mean relative expression levels in severe HFMD and control HFMD patients were 10.1-fold and 5.0-fold, respectively, *P* value <.001.

We found that SCARA3 is associated with severity of HFMD, and it may be a potential prognostic marker to predict the HFMD progression in EV71 infected patients.

Abbreviations: CSF = cerebrospinal fluid, CVA16 = coxsackievirus A16, EV71 = enterovirus 71, HFMD = hand foot mouth disease, SCARA3 = Scavenger receptor class A, member 3, WBC = white blood cell.

Keywords: biomarker, foot and mouth disease, hand, SCARA3

1. Introduction

Hand, foot, and mouth disease (HFMD) is a common infectious diseases in infants and young children and characterized by ulcers on the hand and foot and in the mouth.^[1] HFMD can spread and cause outbreaks all over the world which leads to serious public health threat and heavy economic burden.^[2-4] In China, the epidemic of HFMD has become a serious public health problem^[5-9] and recent outbreaks in Shanghai and Zunyi had

over 2,000,000 and 6,000 cases, respectively occurred within 1 year.^[10,11]

HFMD is mainly caused by enterovirus 71 (EV71) and coxsackievirus A16 (CVA16).^[12,13] Most of HFMD cases caused by CVA16 are mild with symptoms limited to fever, malaise, rashes on the volar regions of the hands and feet, herpangina and difficulty eating and drinking. EV71 associated patients appear to be more severe with high mortality and can cause neurological complications, such as aseptic meningitis, encephalitis, brain stem encephalitis, neurogenic pulmonary edema, and hemorrhage.^[1,14,15] Although only 1% of HFMD patients developed cardiopulmonary or neurological complications, the fatality rate was as high as 3% in those severe cases.^[16] Therefore, large scale EV71 infection outbreak has huge impact on the society and healthcare system. In recent years, CVA6 has been frequently identified in HFMD outbreaks in many Asian countries including China, Thailand, Japan, and Singapore.^[17-21] CVA6 infection is characterized with high fever and abnormal white blood cell (WBC) count in blood.^[22,23]

EV71 vaccines had been developed and some of them had completed phase 3 clinical trials.^[24] However, there is no single vaccine for all EV71 infections due to the wide genetic diversity of HFMD.^[25-27] Therefore it is essential to develop other preventive or interventional methods for better outcome. One of them is to use noninvasive blood markers to identify the severe cases or predict the progression so early intervention could be implemented to reduce the mortality in severe EV71 cases.

The current study aims to investigate biomarkers which are associated with severity of EV71 infection in HFMD cases.

Editor: Leyi Wang.

This study was supported by grants from the Administration of Traditional Chinese Medicine of Jiangsu Province, China (No. LZ13117).

The authors have no conflicts of interests to disclose.

Children's Hospital of Nanjing Medical University, Nanjing, People's Republic of China.

* Correspondence: Xiao-Li Mei, Department of Infectious Disease, Children's Hospital of Nanjing Medical University, 72 Guangzhou Rd, Nanjing, People's Republic of China (e-mail: himeiji@126.com).

Copyright © 2019 the Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial License 4.0 (CCBY-NC), where it is permissible to download, share, remix, transform, and buildup the work provided it is properly cited. The work cannot be used commercially without permission from the journal.

How to cite this article: Tian Y, Zhou K, Hu J, Shan MF, Chen HJ, Cheng S, Liu LF, Mei XL. Scavenger receptor class a, member 3 is associated with severity of hand, foot, and mouth disease in a case-control study. *Medicine* 2019;98:40 (e17471).

Received: 12 February 2019 / Received in final form: 2 August 2019 / Accepted: 11 September 2019

<http://dx.doi.org/10.1097/MD.00000000000017471>

2. Methods

2.1. Study subjects and sample

All HFMD patients were recruited from the Nanjing Children's Hospital Affiliated to Nanjing Medical University in 2014 to 2015. HFMD patients were diagnosed according to the China Ministry of Health, 2010 version of the "Hand, Foot and Mouth Disease Diagnosis and Treatment Guidelines".^[28] Throat swab and blood samples were collected upon diagnosis of HFMD. The types of enterovirus were tested by polymerase chain reaction (PCR) and was verified by presence of EV71 IgM antibody in blood sample. Only those who were EV71 positive were included in the current study. Patients were divided into severe HFMD (S) and control HFMD (C) based on the progression of the disease. Severe HFMD patients were those who presented with one or more conditions of the following central nervous system complications:

- (1) aseptic meningitis,
- (2) encephalitis.

Aseptic meningitis was confirmed based on examination of cerebrospinal fluid (CSF), WBC $>20 \times 10^6/L$ with normal glucose level, normal or increased protein level (based on *Zhu Futang Practice of Pediatrics 8th Edition Chinese Edition*), and negative for Gram stain smear in addition to fever, vomiting, irritability, and other meningeal signs, without disturbed consciousness. Encephalitis was confirmed based on presence of WBC of $>20 \times 10^6/L$ in CSF, disturbed consciousness and neurological signs. Control HFMD patients were defined as those who did not have central nervous system complications. Both groups had been ruled out merger other common infectious diseases, such as measles, cytomegalovirus infection, and mumps. This study was reviewed and approved by Institutional Ethics Committees of the Nanjing Children's Hospital Affiliated to Nanjing Medical University. Consents were obtained from parents of young patients.

2.2. Study design

All HFMD patients who were positive for EV71 were recruited for the study and categorized as severe HFMD or control HFMD based on presence or absence of central nervous system complications. In the discovery phase, genome-wide transcriptional analysis was performed in 10 severe HFMD patients and 10 control HFMD patients. The mRNA expression was analyzed using High-throughput RNA sequencing and the result was further verified by q-PCR. The gene with the highest expression level was selected as potential marker and was validated in an independent cohort.

2.3. Blood sample processing

Lithium heparin anticoagulant tubes were used to collect 5 ml peripheral blood from each volunteer. Samples were allowed to clot for 15 minutes at room temperature and then put on ice in a cooler and transported to the laboratory within 2 hours after collection. Tubes were centrifuged ($1000 \times g$ for 10 minutes) in a refrigerated centrifuge and serum was apportioned into 0.5 ml aliquots and stored at $-80^\circ C$ until further processing.

2.4. RNA extraction and quality control

RNA was extracted from serum using RNeasy Mini Kit (Qiagen, Germany) according to the kit instructions. The quality and

concentration of RNA was detected using NanoDrop 2000 (Thermo, USA). Each of the RNA samples showed an A260:A230 ratio above 2.0 and an A260:A280 ratio above 1.8. RNA integrity was assessed using the Agilent 2200 Tape Station, a standard denaturing agarose gel electrophoresis and each sample had a RIN^c above 7.0.

2.5. High-throughput RNA sequencing and data analysis

High-throughput RNA sequencing was performed by Ribobio (Guangzhou, China). Briefly, total RNA was isolated from serums of the patients, then purified mRNA was fragmented in fragmentation buffer. We used these short fragments of mRNA as templates to synthesize the first-strand cDNA, using random hexamer primers. The second-strand cDNA was synthesized using buffer, dNTPs, RNase H, and DNA polymerase I, respectively. Following the purify double-stranded cDNAs and addition of a single A base, Illumina sequencing adaptors were ligated onto the short fragments. Constructed libraries were sequenced using Illumina HiSeq 2500, according to the manufacturer's instructions. The samples expression levels were presented as RPKM (expected number of Reads Per Kilobase of transcript sequence per Million base pairs sequenced). The gene-level RPKM values were normalized using the log₂ values for further analyses. Differential expression was determined with log₂ fold change more than 1 or less than -1, and *P* value $\leq .05$. The data is available upon request.

2.6. Quantitative real-time PCR

The mRNA expression was validated with SYBR Green qPCR assay and β -actin was considered as a house keeping gene. RNA was reversed transcribed into cDNA using PrimeScriptTM RT reagent Kit (Takara companies, Japan). The qPCR Primers used was shown in Table 2. Reaction conditions as follows: $95^\circ C$, 30 s; $95^\circ C$, 5 s, $60^\circ C$, 34 s, 40 cycles. The data were expressed as mRNA copy number relative to the β -actin.

2.7. Statistical analysis

The data was analyzed using the SPSS version 19. Differential expression levels of mRNAs were compared by independent-samples *t* test among different groups. Relative expression values of SCARA3 were expressed as the mean \pm standard deviation, and a *P* value of $<.05$ was considered statistically significant.

3. Results

A total of 110 HFMD patients who were positive for EV71 and had blood samples available were included in this study. Out of 110 patients, 55 who presented with central nervous system complications were classified as severe HFMD and 55 who did not have any central nervous system complications were classified as control HFMD (Table 1.). In the discovery phase, we performed genome-wide transcriptional analysis using the serum RNA isolated from 10 severe HFMD patients (S group) and 10 control HFMD patients (C group). Clustering analysis showed that 549 genes were differently expressed in S and C groups. There were 392 upregulated mRNAs and 157 downregulated mRNAs in S group compared to C group. A heat map of the 17 genes with the highest fold change of at least 3-fold was created

Table 1
Characteristics of HFMD patients.

	Severe group (n = 55)	Control group (n = 55)	P value
EV71 infection	100%	100%	-
Gender (male/female)	32:23	30:25	.848
Age (year)	1.97 ± 1.05	2.17 ± 1.24	.367
Duration of illness at sampling (hour)	36.56 ± 9.03	33.47 ± 10.06	.09
Fever	100% (55/55)	90.90% (50/55)	.057
Rash	96.36% (53/55)	100% (55/55)	.495
Oral ulcer	90.90% (50/55)	94.54% (52/55)	.716
Vomiting	14.54% (8/55)	5.45% (3/55)	.202
Convulsion	10.90% (6/55)	3.63% (2/55)	.271
Lethargy	54.54% (30/55)	0% (0/55)	<.001
Mean white blood cell (/mm ³) ± SD	17.80 ± 1.79	7.03 ± 1.70	<.001
Blood glucose level (mmol/L) ± SD	9.64 ± 0.90	5.32 ± 1.14	<.001

HFMD = hand foot mouth disease.

(Fig. 1). Among 17 significant differentially expressed genes, 16 of them were upregulated while 1 mRNA was downregulated.

To verify the data from RNA sequencing, the 15 mRNA were validated using qPCR. The results showed the expression patterns of those mRNA were consistent with sequencing data. SCARA3 was the highest upregulated gene in the severe patients compared to controls (Fig. 2).

SCARA3 was selected as a potential biomarker for validation because of the highest expression level among all significantly expressed mRNA. The expression level of SCARA3 were assessed in independent cohort comprising 45 severe HFMD patients and 45 control HFMD patients using qPCR assay. The mean relative expression levels in severe HFMD and control HFMD patients were 10.1-fold and 5.0-fold, respectively, P value <.001 (Fig. 3).

4. Discussion

In this study, we first compared the different mRNA expression of severe patients group with control group in EV71 patients by High-throughput RNA sequencing. Sixteen genes were upregulated and 1 gene was downregulated in severe HFMD patients compared with control group. We further verified this result by q-PCR and found that the expression of SCARA3 was the most significantly increased in severe HFMD patients. SCARA3 was selected as potential prognostic marker and was validated in an independent cohort. The mean value of SCARA3 expression was 2 times higher in severe EV71 patients than that in control group. It suggested the upregulation of SCARA3 may be an indication of

Table 2
List of primers used in the study.

Primer name	Forward	Reverse
CASQ2	5'- CTGAGCATCCTGTGGATCG-3'	5'- TCTTCTCCCAGTAGGCAACG-3'
TFPI2	5'-TTTCGTATAAAGCGGGTATT-3'	5'-ACGACCCGCTAAACAAAAC-3'
USP46	5'-GAAACCTTCCCTAGAGGGAATAC-3'	5'-CTGTCTTGTCAAGCATTCTAGG-3'
SLC46A3	5'-CTAACAGGACTGTCATCTGGC-3'	5'-CATAGTAACATTCTGAGATG-3'
GUSB	5'-CTGTACACGACCCACCAC-3'	5'-ATTGCGACGACTTGT-3'
SAMD4B	5'-TCGGAGTACATGAGGCTACTGC-3'	5'-AGGGCCAGTGCATTGCGGTC-3'
ZNF44	5'-TCATCCCTGAATTGCTACAT-3'	5'-GCGATAACTTAAGCCTT-3'
ANTXR1	5'TGCAACACAGAAATGCTCTGCCTG-3'	5'-TTTATCCCTGGGTGATGAAGCCCA-3'
SCARA3	5'- TGCAGCTGGATAACATCTCG-3'	5'- CTTGGTCATCCTGGGCTTTA-3'
TTN	5'-GGCCTTTCATTCCTGGT-3'	5'-TCGGCTTCTATTGGTGAAGG-3'
RWDD1	5'- GGAGTCCATCTACCTGACTCC-3'	5'-CTGATAATACTTAGCAGGACCT-3'
HELLS	5'-GGTGAAGTCCAGGATGCC-3'	5'-CCAAGAGTCTGCATCAAAC-3'
GABRB2	5'-TCTGAAGTCAATATGGATTA-3'	5'-GGTCTTAACAGTCACTCCG-3'
FARP2	5'-GATGGCCAGGTATTACTGAC-3'	5'-GAAGCACCACATCTTTGGC-3'
FAM181B	5'-TGCGCGAGGCCACCCGCGAT-3'	5'-CGGCCGTTTGGCTGGCGTGT-3'
β-actin	5'- AGATGTGGATCAGCAAGCAG-3'	5'-GCGCAAGTTAGGTTTTGTCA-3'

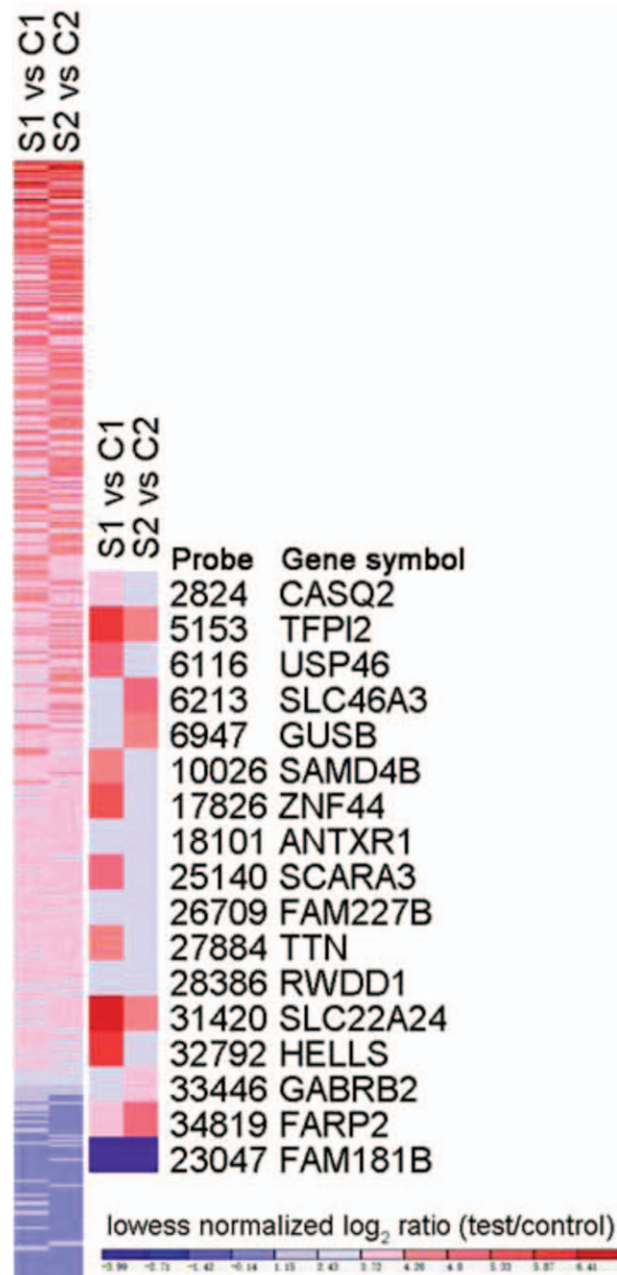


Figure 1. High-throughput RNA sequencing of severe HFMD patients and control HFMD patients. Heat map of the high-throughput RNA sequencing data obtained for severe patients and control patients. Blue color refers to downregulation; red color refers to upregulation. HFMD = hand foot mouth disease.

progression of EV71 infection in HFMD. Our report is the first to show the association between SCARA3 and the severity of EV71 infection.

The class A scavenger receptors (SCARAs) are a family of cell surface glycoproteins and involved in the uptake of nucleic acids,

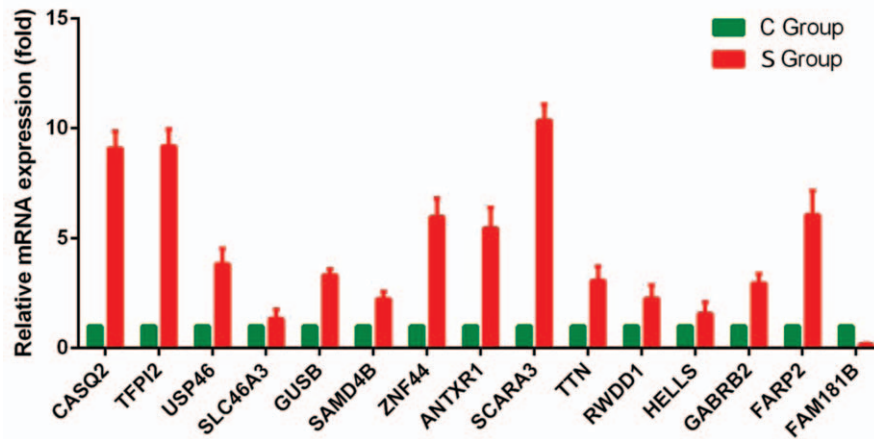


Figure 2. Validation of the expression of significantly upregulated and downregulated genes between S group and C group. The genes mRNA levels were verified using qPCR in serum from severe patients (n=10) and control patients (n=10). The expression of mRNA was $2^{-\Delta\Delta Ct}$ relative to β -actin.

double-stranded RNA, and oligonucleotide. There are 5 members have been identified in scavenger receptor family: SR-AI/II/III, MARCO, SCARA3, SCARA4, and SCARA5.^[29–31] SCARAs have a vital roles in the immune response against bacterial and viral pathogens,^[32–34] such as the increased expression of SCARA5 involved in the cancer metastatic progression, and MARCO plays an important role in bacteria-binding innate immune response.^[31,35] As for SCARA3, its expression could be altered by oxidative stress, also functioned as a cellular stress response gene to scavenge reactive oxygen species (ROS) and other harmful products of oxidation. Moreover, SCARA3 can acquire fatty acids by binding to polyanionic ligands and representing a major route through dendritic cells and macrophages in cancers.^[36,37] It seemed the progression of EV71 infection had stimulated the immune system and caused the overexpression of SCARA3. Animal studies on how the EV71 infection activates secretion of SCARA3 will be helpful.

There is no specific antiviral medications or effective vaccine for EV71. The severe cases can progress quickly and the current

treatment is largely supportive. SCARA3 could be used as prognostic marker to identify severe cases. Continuous electrocardiogram monitoring and early intervention such as treatment with vasoactive drugs, Gamma globulin or Glucocorticoid in those severe EV71 infection cases could help reduce the mortality. Although the level of SCARA3 expression in severe EV71 patients was statistically significantly higher than that in control group ($P < .001$, Fig. 3), there were some overlaps between severe group and control group. This suggested SCARA3 should be used carefully together with observation of patient symptoms.

The study has some limitations. The sample size of validation cohort was small. Although there were 17 significant differentially expressed genes, only SCARA3 was selected for validation because of budget constrain. Lastly, the SCARA3 expression level was not measured after patient with severe EV71 infection recovered to verify its use as prognostic effect.

In conclusion, we found the expression of SCARA3 was increased in severe HFMD patients compared with the control patients, indicating that SCARA3 was associated with the severity of HFMD. Our study suggested that SCARA3 may participate in the progression of HFMD and could serve as a potential prognostic indicator in HFMD. The result is required to be validated in a prospective study before it can be used as prognostic marker in clinic. More work is needed to understand the molecular mechanism of SCARA3 in EV71 infection.

Acknowledgments

We appreciate staffs, doctors and nurses of Nanjing Children's Hospital Affiliated to Nanjing Medical University for patient management, sample collection, and analysis in this work.

Author contributions

Conceptualization: Ye Tian, Kai Zhou, Jing Hu, Ming-Feng Shan, Hong-Jian Chen, Shan Cheng, Li-Fei Liu, Xiao-Li Mei.
Data curation: Ye Tian, Kai Zhou, Hong-Jian Chen, Shan Cheng, Li-Fei Liu.

Formal analysis: Ye Tian.

Funding acquisition: Xiao-Li Mei.

Investigation: Ye Tian, Kai Zhou, Jing Hu, Ming-Feng Shan, Hong-Jian Chen, Shan Cheng, Li-Fei Liu, Xiao-Li Mei.

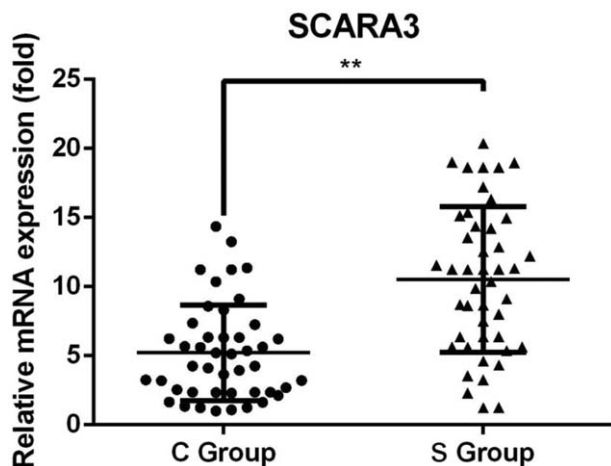


Figure 3. SCARA3 is associated with severity of hand foot mouth disease. Comparing differences in the mRNA expression levels of SCARA3 between severe patients (n=45) and control patients (n=45). β -actin was used as an internal control. $P < 0.001$. SCARA3=Scavenger receptor class A, member 3.

Methodology: Ye Tian, Kai Zhou, Jing Hu, Ming-Feng Shan, Hong-Jian Chen, Shan Cheng, Li-Fei Liu, Xiao-Li Mei.

Project administration: Ye Tian, Xiao-Li Mei.

Resources: Xiao-Li Mei.

Software: Ye Tian.

Supervision: Xiao-Li Mei.

Validation: Ye Tian.

Writing – original draft: Ye Tian, Xiao-Li Mei.

Writing – review & editing: Ye Tian, Kai Zhou, Jing Hu, Ming-Feng Shan, Hong-Jian Chen, Shan Cheng, Li-Fei Liu, Xiao-Li Mei.

References

- [1] Solomon T, Lewthwaite P, Perera D, et al. Virology, epidemiology, pathogenesis, and control of enterovirus 71. *Lancet Infect Dis* 2010;10:778–90.
- [2] Nassef C, Ziemer C, Morrell DS. Hand-foot-and-mouth disease: a new look at a classic viral rash. *Curr Opin Pediatr* 2015;27:486–91.
- [3] Geoghegan JL, Tan le V, Kuhnt D, et al. Phylogenetics of enterovirus A71-associated hand, foot, and mouth disease in Viet Nam. *J Virol* 2015;89:8871–9.
- [4] Machain-Williams C, Dzul-Rosado AR, Yeh-Gorocica AB, et al. Detection of hand, foot and mouth disease in the Yucatan Peninsula of Mexico. *Infect Dis Rep* 2014;6:5627.
- [5] Zhou HT, Guo YH, Chen MJ, et al. Changes in enterovirus serotype constituent ratios altered the clinical features of infected children in Guangdong Province, China, from 2010 to 2013. *BMC Infect Dis* 2016;16:399.
- [6] Mao YJ, Sun L, Xie JG, et al. Epidemiological features and spatio-temporal clusters of hand-foot-mouth disease at town level in Fuyang, Anhui Province, China (2008–2013). *Epidemiol Infect* 2016;144:3184–97.
- [7] Zhu J, Luo Z, Wang J, et al. Phylogenetic analysis of enterovirus 71 circulating in Beijing, China from 2007 to 2009. *PLoS One* 2013;8:e56318.
- [8] Xia Y, Shan J, Ji H, et al. Study of the epidemiology and etiological characteristics of hand, foot, and mouth disease in Suzhou City, East China, 2011–2014. *Arch Virol* 2016;161:1933–43.
- [9] Lee MK, Chan PK, Ho II, et al. Enterovirus infection among patients admitted to hospital in Hong Kong in 2010: epidemiology, clinical characteristics, and importance of molecular diagnosis. *J Med Virol* 2013;85:1811–7.
- [10] Wang Y, Zou G, Xia A, et al. Enterovirus 71 infection in children with hand, foot, and mouth disease in Shanghai, China: epidemiology, clinical feature and diagnosis. *Virol J* 2015;12:83.
- [11] Zhang W, Huang B, She C, et al. An epidemic analysis of hand, foot, and mouth disease in Zunyi, China between 2012 and 2014. *Saudi Med J* 2015;36:593–8.
- [12] Yong W, Qiao M, Shi L, et al. Genetic characteristics of coxsackievirus A16 associated with hand, foot, and mouth disease in Nanjing, China. *J Infect Dev Ctries* 2016;10:168–75.
- [13] Tan X, Huang X, Zhu S, et al. The persistent circulation of enterovirus 71 in People's Republic of China: causing emerging nationwide epidemics since 2008. *PLoS One* 2011;6:e25662.
- [14] Liu CC, Hwang CS, Yang WS, et al. Long-term immunogenicity studies of formalin-inactivated enterovirus 71 whole-virion vaccine in macaques. *PLoS One* 2014;9:e106756.
- [15] Chong CY, Chan KP, Shah VA, et al. Hand, foot and mouth disease in Singapore: a comparison of fatal and non-fatal cases. *Acta Paediatr* 2003;92:1163–9.
- [16] Xing W, Liao Q, Viboud C, et al. Hand, foot, and mouth disease in China, 2008–12: an epidemiological study. *Lancet Infect Dis* 2014;14:308–18.
- [17] Fujimoto T, Iizuka S, Enomoto M, et al. Hand, foot, and mouth disease caused by coxsackievirus A6, Japan, 2011. *Emerg Infect Dis* 2012;18:337–9.
- [18] Wu Y, Yeo A, Phoon MC, et al. The largest outbreak of hand; foot and mouth disease in Singapore in 2008: the role of enterovirus 71 and coxsackievirus A strains. *Int J Infect Dis* 2010;14:e1076–81.
- [19] Puenpa J, Chieochansin T, Linsuwanon P, et al. Hand, foot, and mouth disease caused by coxsackievirus A6, Thailand, 2012. *Emerg Infect Dis* 2013;19:641–3.
- [20] Zeng H, Lu J, Zheng H, et al. The epidemiological study of coxsackievirus A6 revealing hand, foot and mouth disease epidemic patterns in Guangdong, China. *Sci Rep* 2015;5:10550.
- [21] Tan X, Li L, Zhang B, et al. Molecular epidemiology of coxsackievirus A6 associated with outbreaks of hand, foot, and mouth disease in Tianjin, China, in 2013. *Arch Virol* 2015;160:1097–104.
- [22] Li J, Sun Y, Du Y, et al. Characterization of coxsackievirus A6- and enterovirus 71-associated hand foot and mouth disease in Beijing, China, from 2013 to 2015. *Front Microbiol* 2016;7:391.
- [23] Hu YQ, Xie GC, Li DD, et al. Prevalence of coxsackievirus A6 and enterovirus 71 in hand, foot and mouth disease in Nanjing, China in 2013. *Pediatr Infect Dis J* 2015;34:951–7.
- [24] Li JX, Mao QY, Liang ZL, et al. Development of enterovirus 71 vaccines: from the lab bench to Phase III clinical trials. *Expert Rev Vaccines* 2014;13:609–18.
- [25] Aswathyrj S, Arunkumar G, Alidjinou EK, et al. Hand, foot and mouth disease (HFMD): emerging epidemiology and the need for a vaccine strategy. *Med Microbiol Immunol* 2016;205:397–407.
- [26] Yee PT, Poh CL. Development of novel miRNA-based vaccines and antivirals against enterovirus 71. *Curr Pharm Des* 2016;22:6694–700.
- [27] Liu L, Mo Z, Liang Z, et al. Immunity and clinical efficacy of an inactivated enterovirus 71 vaccine in healthy Chinese children: a report of further observations. *BMC Med* 2015;13:226.
- [28] The Ministry of Health of the People's Republic of China. Guideline for the Diagnosis and Treatment of Hand Foot and Mouth Disease. Beijing, China, 2010.
- [29] Helmfors H, Lindberg S, Langel U. SCARA involvement in the uptake of nanoparticles formed by cell-penetrating peptides. *Methods Mol Biol* 2015;1324:163–74.
- [30] Ezzat K, Helmfors H, Tudoran O, et al. Scavenger receptor-mediated uptake of cell-penetrating peptide nanocomplexes with oligonucleotides. *FASEB J* 2012;26:1172–80.
- [31] He J, Liu H, Wu C. Identification of SCARA3, SCARA5 and MARCO of class A scavenger receptor-like family in *Pseudosciaena crocea*. *Fish Shellfish Immunol* 2014;41:238–49.
- [32] Bonocchi R, Garlanda C, Mantovani A, et al. Cytokine decoy and scavenger receptors as key regulators of immunity and inflammation. *Cytokine* 2016;87:37–45.
- [33] Yu X, Guo C, Fisher PB, et al. Scavenger receptors: emerging roles in cancer biology and immunology. *Adv Cancer Res* 2015;128:309–64.
- [34] Areschoug T, Gordon S. Scavenger receptors: role in innate immunity and microbial pathogenesis. *Cell Microbiol* 2009;11:1160–9.
- [35] Armengol C, Bartoli R, Sanjurjo L, et al. Role of scavenger receptors in the pathophysiology of chronic liver diseases. *Crit Rev Immunol* 2013;33:57–96.
- [36] Brown CO, Schibler J, Fitzgerald MP, et al. Scavenger receptor class A member 3 (SCARA3) in disease progression and therapy resistance in multiple myeloma. *Leuk Res* 2013;37:963–9.
- [37] Bock AJ, Nymoen DA, Brenne K, et al. SCARA3 mRNA is overexpressed in ovarian carcinoma compared with breast carcinoma effusions. *Hum Pathol* 2012;43:669–74.