



# A nomogram based on bi-regional radiomics features from multimodal magnetic resonance imaging for preoperative prediction of microvascular invasion in hepatocellular carcinoma

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**Background:** We aimed to develop and validate a nomogram combining bi-regional radiomics features from multimodal magnetic resonance imaging (MRI) and clinicoradiological characteristics to preoperatively predict microvascular invasion (MVI) of hepatocellular carcinoma (HCC).

**Methods:** A total of 267 HCC patients were divided into training (n=194) and validation (n=73) cohorts according to MRI data. Bi-regional features were extracted from whole tumors and peritumoral regions in multimodal MRI. The minimum redundancy maximum relevance (mRMR) algorithm was applied to select features and build signatures. The predictive performance of the optimal radiomics signature was further evaluated within subgroups defined by tumor size and alpha fetoprotein (AFP) level. Then, a radiomics nomogram including the optimal radiomics signature, radiographic descriptors, and clinical variables was developed using multivariable regression. The nomogram performance was evaluated based on its discrimination, calibration, and clinical utility.

**Results:** The fusion radiomics signature derived from triphasic dynamic contrast-enhanced (DCE) MR images can effectively classify MVI and non-MVI HCC patients, with an AUC of 0.784 (95% CI: 0.719–0.840) in the training cohort and 0.820 (95% CI: 0.713–0.900) in the validation cohort. The fusion radiomics signature also performed well in the subgroups defined by the two risk factors, respectively. The nomogram, consisting of the fusion radiomics signature, arterial peritumoral enhancement, and AFP level, outperformed the clinicoradiological prediction model in the validation cohort (AUCs: 0.858 *vs.* 0.729; *P*=0.022), fitting well in the calibration curves (*P*>0.05). Decision curves confirmed the clinical utility of the nomogram.

**Conclusions:** The radiomics nomogram can serve as a visual predictive tool for MVI in HCCs, and thus assist clinicians in selecting optimal treatment strategies to improve clinical outcomes.

**Keywords:** Microvascular invasion (MVI); hepatocellular carcinoma (HCC); magnetic resonance imaging (MRI); radiomics; nomogram

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## Introduction

Hepatocellular carcinoma (HCC) is the sixth most prevalent cancer worldwide and one of the leading causes of cancer-related death (1). Hepatectomy and liver transplantation (LT) are potential curative modalities for HCC patients. Although several clinically based classification schemes have been used to guide the selection of therapeutic strategies, the 5-year recurrence rates can be as high as 70% after hepatectomy and 35% after LT (2-4). Studies have demonstrated that microvascular invasion (MVI) is the strongest independent predictor of early tumor recurrence (5). Alternative treatment strategies and preoperative adjuvant therapy should be considered for HCC patients who are at high risk of MVI (6,7). Unfortunately, MVI diagnosis can only reliably be made by histopathology of surgical or biopsy specimens, which is when its clinical utility is marginal. To minimize the risk of recurrence, a non-invasive method capable of accurately identifying MVI preoperatively is urgently needed to better stratify HCC patients for treatment decisions.

Over the past decade, many efforts have been made to preoperatively predict MVI. Current practice uses two clinical surrogate biomarkers, the Milan criteria, and serum alpha fetoprotein (AFP) levels, as predictors (8). LT is generally recommended in patients with tumors that meet the Milan criteria (a single tumor  $\leq 5$  cm or up to 3 tumors, each  $\leq 3$  cm). However, 30% of HCCs selected by the Milan criteria for LT has MVI and, conversely, 50% of tumors outside the Milan criteria have no MVI and would potentially benefit from LT (9,10). Serum AFP lacks sensitivity for the prediction of MVI, and even the model that combines both the Milan criteria and serum AFP has a sensitivity of only 50% (11). In addition to clinical predictors of MVI, several studies have reported that certain imaging findings, including the gross type of HCC (12), arterial rim enhancement or peritumoral enhancement (13,14), internal arteries (15), incomplete capsule (16), washout (17), and peritumoral hypointensity during the hepatobiliary phase (HBP) (18), are useful for predicting MVI of HCC. However, such qualitative radiographic descriptors suffer from limitations, including inter-reader variability and lack of external validation (19).

Recently, the emergence of radiomics has allowed the automated, high-throughput extraction of quantitative imaging features from regions of interest (ROIs) in routinely acquired radiologic images, which provide crucial insights into tumor heterogeneity, phenotype, and

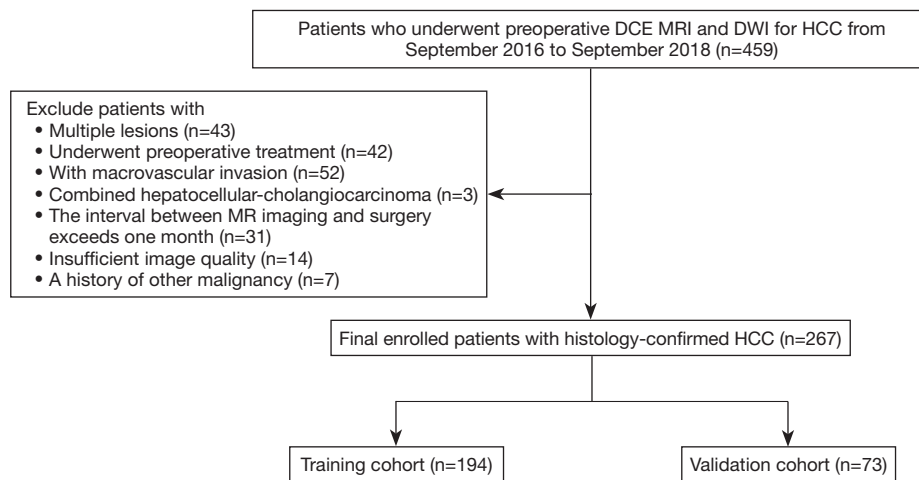
microenvironment (20). Few investigations have been published on the utility of radiomics for MVI prediction in HCC patients (8,19,21-24), and most of them are based on CT images. However, magnetic resonance imaging (MRI) carries an advantage of depicting more soft-tissue characteristics, and dynamic contrast-enhanced (DCE) MRI and diffusion-weighted imaging (DWI) has the potential to assess the tumor's metabolism and proliferation with higher accuracy (25). Consequently, multimodal MRI may allow more effective radiomics. In addition, due to the multiregional and microenvironmental heterogeneity in HCC, it is reasonable to hypothesize that quantitative features from the whole tumor and peritumoral region could have the best accuracy in predicting MVI than those features from a single region.

Therefore, in this study, we aimed to investigate the use of bi-regional radiomics parameters derived from multimodal MR images to predict MVI of HCC, as well as to establish and validate a combined nomogram model that incorporates the developed radiomics signature with radiologic features and clinical risk factors for the preoperative prediction of MVI in HCC patients.

## Methods

### Patients

This study was approved by our Institutional Review Board, and the requirement for informed consent was waived. We retrospectively searched our institutional database between September 2016 and September 2018 and identified 459 patients who had undergone preoperative multiparameter MRI for HCC. The inclusion criteria were as follows: (I) histologically confirmed HCC; (II) single tumor with or without satellite nodules, which are lesions having a tumor diameter  $\leq 2$  cm and a distance from the main tumor  $\leq 2$  cm; (III) no preoperative cancer-related treatments, including transarterial chemoembolization and radiofrequency ablation; (IV) no macrovascular invasion on MRI; (V) an MR scan received within 1 month before curative hepatic resection or transplantation; and (VI) MR images with sufficient image quality. Patients who met the following criteria were excluded: (I) combined hepatocellular-cholangiocarcinoma, (II) recurrent HCC and (III) a history of other malignancies. The patient recruitment process is shown in *Figure 1*. The final cohort consisted of 267 consecutive patients (229 males and 38 females; range, 31–83 years; mean age,  $57.93 \pm 10.65$  years). The cohort



**Figure 1** Flowchart of the enrolled study patients. HCC, hepatocellular carcinoma; DCE MRI, dynamic contrast-enhanced MRI; DWI, diffusion-weighted imaging.

was divided into a training set (n=194; 165 males and 29 females; range, 31–83 years; mean age, 58.35±10.85 years; from September 2016 to February 2018) and a validation set (n=73; 64 males and 9 females; range, 40–77 years; mean age, 56.82±9.40 years; from March 2018 to September 2018) according to the MR examination date.

### Clinical and pathological variables

The demographic characteristics, including sex, age, and hepatitis status (presence or absence), were gathered from the electronic medical record system. Data on cirrhosis, MVI, and Edmondson-Steiner (E-S) grade were obtained from pathology reports. MVI was defined as a tumor within a vascular space lined by endothelium that was visible only by microscopy (5). Specimens from hepatectomy or LT were sampled by using the “7 points” baseline sampling method (26). Laboratory data, including albumin, gamma-glutamyl transferase (GGT), and AFP were obtained via preoperative routine blood tests. Patients were classified by the Barcelona clinic liver cancer (BCLC) staging system and Child-Pugh score based on MRI findings and medical history. Clinical and pathological descriptions of these two cohorts are fully detailed in *Table 1*.

### MR image acquisition

MR examinations were performed using a 3.0 T scanner (GE Medical Systems, Milwaukee, WI, USA). Our liver MRI protocol consisted of axial T2-weighted imaging

with fat suppression, dual-echo (in-phase and opposed-phase) T1-weighted imaging, DWI, and pre-contrast and post-contrast dynamic three-dimensional fast-spoiled gradient-recalled echo sequence (liver acceleration volume acquisition, LAVA). DWI was obtained using a respiratory triggering, a single-shot echo-planar imaging pulse sequence with b values of 0 and 1,000 mm<sup>2</sup>/s. The acquisition parameters for LAVA were as follows: repetition time (TR) of 3.2 ms; echo time (TE) of 1.5 ms; reverse time of 5 ms; field of view of 380×304 mm<sup>2</sup>; flip angle of 10°; bandwidth of 100 kHz; and image resolution of 1.19×1.19×5 mm<sup>3</sup>. Post-contrast dynamic LAVA was performed at the arterial phase (20 s), portal venous phase (60 s), and equilibrium phase (180 s) after a rapid bolus injection of 0.1 mmol/kg of gadopentetate dimeglumine (magnevist®, Bayer Schering Pharma, Berlin, Germany) into the cubital vein at a rate of 2.5 mL/s, followed by a 15-mL saline flush.

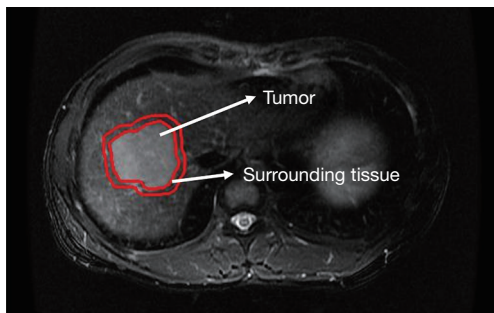
### Qualitative radiographic descriptors

Image analysis was performed by two abdominal radiologists (R. Z. and J. Z., with 10 and 15 years of experience in liver MRI, respectively) who were blinded to clinical, laboratory, and pathologic information. The two radiologists independently evaluated the following eight MR imaging features for each HCC including tumor size, the gross type of HCC (including nodule and non-nodule types), arterial rim enhancement, arterial peritumoral parenchymal enhancement, washout, and peritumoral hypointensity in the later phase, radiological capsule, and internal arteries, as

**Table 1** Comparisons of clinicoradiological characteristics in training and validation cohorts

Characteristics	Training cohort (N=194)		P	Validation cohort (N=73)		P
	MVI (+) (N=64)	MVI (-) (N=130)		MVI (+) (N=26)	MVI (-) (N=47)	
Clinicoradiological characteristics						
Age (years), mean [range]	57.84 [32–83]	58.60 [31–81]	0.6508	56.38 [40–77]	57.06 [37–74]	0.7675
Gender (male/female)	55/9	110/20	0.8081	24/2	40/7	0.5999
Primary liver disease (hepatitis B/hepatitis C/ non-alcoholic steatohepatitis/none or other)	57/0/1/6	112/1/0/17	0.3823	22/0/0/4	43/0/2/2	NA
Cirrhosis (present/absent)	52/12	100/30	0.4914	21/5	35/22	0.5419
Albumin (normal/abnormal)	61/3	118/12	0.4076	25/1	44/3	0.9355
GGT (normal/abnormal)	33/31	70/60	0.7644	12/14	25/22	0.5647
AFP (normal/abnormal)	25/39	71/59	0.0416	8/18	27/20	0.0289
Child-Pugh class (A/B)	62/2	121/9	0.4561	25/1	45/2	0.5952
BCLC staging system (0/A)	3/61	17/113	0.1198	1/25	6/41	0.4097
E-S grade (I+II/III+IV)	24/40	77/53	0.0044	8/18	20/27	0.3214
MR imaging features						
Tumor size (cm), mean (range)	5.16 (1.5–14.2)	4.14 (0.8–13.0)	0.0062	5.13 (1.4–10.2)	4.003 (0.8–9.7)	0.0344
Gross type			0.0022			0.0238
Non-nodular type	56	87		23	30	
Nodular type	8	43		3	17	
Arterial peritumoral enhancement			0.0015			0.0885
Absence	40	108		17	39	
Presence	24	22		9	8	
Arterial rim enhancement			0.0030			0.6897
Absence	46	115		23	40	
Presence	18	15		3	7	
Peritumoral hypointensity			0.0770			0.1549
Absence	48	111		20	42	
Presence	16	19		6	5	
Radiological capsule			0.0147			0.2066
Absence or incomplete	57	96		22	32	
Complete	7	34		4	15	
Wash out			0.2127			0.0673
Absence	9	28		1	11	
Presence	55	102		25	36	
Internal arteries			<0.0001			0.0234
Absence	21	84		10	31	
Presence	43	46		16	16	

MVI, microvascular invasion; GGT, gamma-glutamyl transferase; AFP,  $\alpha$ -fetoprotein; BCLC staging system, Barcelona clinic liver cancer staging system; E-S grade, Edmondson-Steiner grade.



**Figure 2** Lesion segmentation for radiomics analysis. First, radiologists manually drew a seed region that enclosed the contour of tumor; then, a region with 10 mm distance to tumor contour was automatically created.

previously described (12-18).

### *Image segmentation and radiomics feature extraction*

The whole tumor was manually depicted along with the lesion outline on each axial slice of the T1-weighted images, T2-weighted images, DWI images, and three-phase DCE images by a radiologist with ten years of professional experience using ITK-SNAP (<http://www.nitrc.org/projects/itk-snap/>). The delineated tumor boundary was independently validated by another radiologist to reduce possible bias.

The pre-processing procedure was undertaken before feature extraction, including image resampling and grey level normalization. The voxel resample was performed using three-dimensional Lagrangian polygon interpolation on MATLAB software (27). All voxel sizes of all images were resampled with the same size of  $1 \times 1 \times 1 \text{ mm}^3$ . The image grey level was normalized to a scale of 1 to 64. To capture quantitative features from the area around the tumor, a surrounding tissue area of approximately 10 mm was created using MATLAB 2017b (MathWorks Inc., Natick, MA, USA) (<http://www.mathworks.com/products/matlab/>), the lesion segmentation is shown in *Figure 2*. A total of 484 radiomics features were extracted from each of the 2 ROIs (tumor and surrounding tissue), which included three types: (I) 7 intensity features; (II) 53 texture features; and (III) 424 wavelet features. For triphasic DCE MR images, 14 inside-outside co-occurrence features were further calculated for each of the 2 ROIs, including the intensity changes and the corresponding intensity change ratios. The intensity changes were calculated by subtracting the signal intensity of the lesion in the first/third phase from that

in the second phase. The corresponding intensity change ratios were calculated by dividing the signal intensity of the lesion in the second phase by the intensity changes of the second phase. During the liver DCE MR scanning, the diaphragm was hardly assured to be in the same position for each scan. Thus, the intensities of the lesion in the three phases were calculated using the mean value of the intensities within ROIs. The detailed descriptions of these features are presented in *Figure S1*. By performing stability and reproducibility test for radiomics features, a number of 2,516 features showed desirable reproducibility. The detailed results of reproducibility test were provided in Supplementary I.

### *Radiomics feature selection and signature construction*

The extracted radiomics features were normalized to eliminate the value scales of the data both in the training and validation cohorts. The training cohort was used to build a radiomics signature as the MVI classifier. More features may improve the classifiers performance; however, more features may cause the model overfitting. The minimum redundancy maximum relevance (mRMR) algorithm was applied to identify the optimal radiomics features. The mRMR ranks the input radiomics features by maximizing the mutual information (MI) with respect to the class labels and minimizing the average MI of selected features (28). After the mRMR feature selection, the optimal feature subset was generated. Then, a radiomics signature was built with multivariable logistic regression analysis using the selected features. The performances of the radiomics signatures from single MR sequences and fusion radiomics signatures from multiple sequences were evaluated using the area under the receiver operating characteristic (ROC) curve (AUC) both in the training and validation cohort. Multiple comparisons of the curves were performed using the Delong test to identify the optimal signature.

To minimize the impact of established predictors of MVI in HCC (tumor size and AFP level) (29,30), all HCC patients were divided into two subgroups defined according to two risk factors. The predictive performance of the optimal radiomics signature was evaluated within subgroups using the AUCs from both the training and validation cohorts.

### *Construction and evaluation of MVI prediction models*

Clinicoradiological risk factors were analyzed for significant

**Table 2** Predictive performance of the clinicoradiological model, fusion radiomics signature, and nomogram

Model	Training cohort (N=194)				Validation cohort (N=73)			
	AUC	95% CI	Sensitivity	Specificity	AUC	95% CI	Sensitivity	Specificity
Clinicoradiological model	0.753	0.686–0.812	46.88%	93.08%	0.729	0.612–0.827	26.92%	93.62%
Fusion radiomics signature	0.784	0.719–0.840	65.65%	80.00%	0.820	0.713–0.900	69.23%	80.85%
Nomogram	0.825	0.764–0.875	82.71%	70.77%	0.858	0.756–0.928	80.77%	68.09%

AUC, area under ROC curve; CI, confidence interval. The AUC was reported in 95% confidence interval.

differences between the MVI and non-MVI groups using the Mann-Whitney U test, and Chi-square test, as appropriate. In the training cohort, a clinicoradiological model was developed with stepwise multivariable logistic regression using a backward search method.

In the training cohort, a combination model incorporating the optimal radiomics and clinicoradiological characteristics were developed using multivariable logistic regression analysis. The backward step-wise selection was applied using the likelihood ratio test with Akaike's information criterion (AIC) as the stopping rule. The model with the minimum AIC value was selected as the final model.

Comparisons of the predictive performances among the optimal radiomics signature, the clinicoradiological model, and the combined radiomics model were performed using the Delong test.

### Construction and validation of the radiomics nomogram

A radiomics nomogram was generated on the prediction model as a graphical presentation (31). The performance of the nomogram was evaluated based on its capabilities in discrimination, calibration, and clinical utility in both the training and validation cohorts. The discrimination capability was measured using the AUC. Calibration curves were plotted to intuitively evaluate the predictive accuracy of the nomogram (32). The calibration curve takes the nomogram-predicted probability of MVI as the abscissa and the actual rate obtained by the bootstrapping method as the ordinate. The closer the calibration curve is to the diagonal line, the higher the prediction accuracy of the model. The Hosmer-Lemeshow test was used to test the goodness-of-fit of the nomogram. Decision curve analysis was conducted to evaluate the clinical utility of the nomogram by quantifying the probabilities of net benefits at a threshold across 0.0 to 1.0 (33). The decision curve takes the threshold of nomogram-predicted probability for MVI as the abscissa

and clinical decision net benefit as the ordinate. The two extreme curves of the treat-all strategy and the treat-none strategy are used as references; the farther the decision curve is from the two extreme curves, the higher the clinical decision net benefit of the model.

### Statistical analysis

All statistical analyses were performed in R software (version 3.4.1; <http://www.Rproject.org>) and MedCalc software (Version 15.2.2; <https://www.medcalc.org>). The mRMR feature selection was performed using the "mRMRe" package. Nomogram and calibration curves were plotted using the "rms" package. The Hosmer-Lemeshow test was performed using the "generalhoslem" package. The ROC curves were plotted using MedCalc software. A two-sided P value of 0.05 was used to determine the statistical significance.

## Results

### Clinicoradiological characteristics

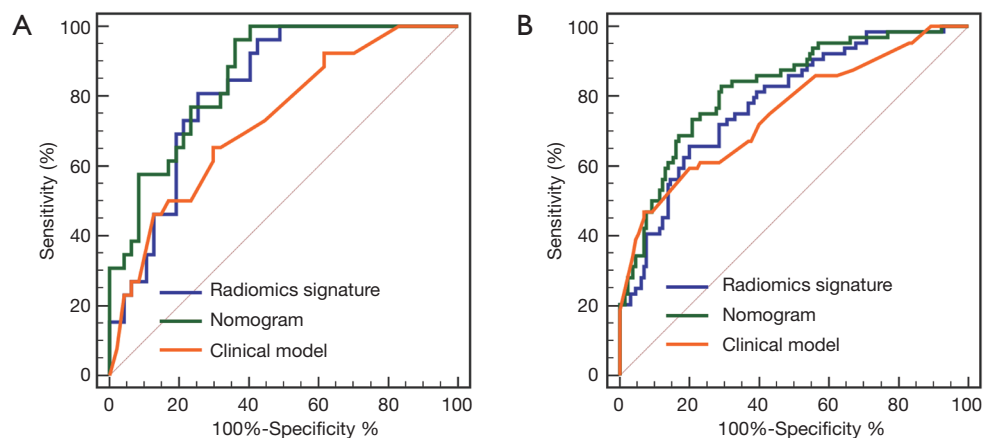
Clinicoradiological characteristics for patients in the training and validation cohorts are listed in *Table 1*. The univariate analyses showed that four clinicoradiological characteristics including AFP level, tumor size, gross tumor type, and presence of internal arteries had significant differences between the patients with and without MVI in both the training and validation cohorts.

Based on the multivariate analysis that used the minimum Akaike information criterion, an optimal clinicoradiological model was constructed using gross tumor type, arterial peritumoral enhancement, arterial rim enhancement, internal arteries, and AFP level. The formula of the clinicoradiological model is provided in *Figure S2*. The AUCs of the clinicoradiological model was 0.753 (95% CI: 0.686–0.812) for the training cohort and 0.729 (95% CI: 0.612–0.827) for the validation cohort (*Table 2*).

**Table 3** Subgroup analyses using the fusion radiomics signature

Risk factors	Subgroups	MVI (+)	MVI (-)	P	AUC (95% CI)
Tumor size	≤5 cm	-0.5327±0.9536	-2.0223±2.3438	<0.0001	0.808 (0.743–0.863)
	>5 cm	0.7277±2.2340	-0.2168±0.5376	0.0079	0.743 (0.637–0.832)
AFP level	≤20 ng/mL	0.3912±2.5910	-1.5549±2.6558	0.0004	0.793 (0.714–0.859)
	>20 ng/mL	-0.1610±2.5910	-1.5965±1.4372	<0.0001	0.807 (0.731–0.870)

The radiomics score obtained from the fusion radiomics signature are reported using mean ± standard deviation. P values were obtained from the Mann-Whitney U test between MVI positive and negative groups. The predictive performance of the fusion radiomics signature was evaluated using the area under ROC curve (AUC); the AUC values are reported with 95% confidence interval. MVI, microvascular invasion.



**Figure 3** Comparison of receiver operating characteristic (ROC) curves for the prediction of microvascular invasion. ROC curves of the clinicoradiological model, the fusion radiomics signature and the nomogram, which combines the fusion radiomics signature and clinicoradiological factors in the training (A) and validation (B) datasets.

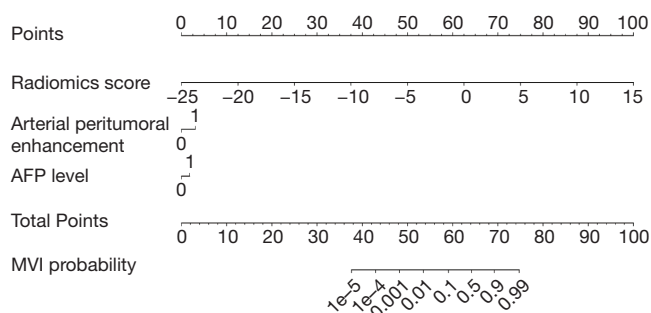
### Construction and validation of the radiomics signatures

The performances of radiomics signatures using a single MR sequence and the fusion radiomics signatures using multi-sequence are shown in *Table S2*. Obviously, the fusion radiomics signature based on triphasic DCE MR images showed the best predictive performance, with an AUC of 0.784 (95% CI: 0.719–0.840) in the training cohort and 0.820 (95% CI: 0.713–0.900) in the validation cohort. The formula for the fusion radiomics signature is provided as a *Figure S2*.

According to tumor size, all HCC patients were divided into two sub-groups: tumor size ≤5 cm (n=182) and tumor size >5 cm (n=85). According to the AFP level, all patients were divided into two groups: AFP level ≤20 ng/mL (n=131) and AFP level >20 ng/mL (n=136). The predictive performance of the fusion radiomics signature within subgroups defined by tumor size and AFP level is summarized in *Table 3*.

### Construction and comparison of MVI prediction models

The combined radiomics model that added the fusion radiomics signature into the clinicoradiological factors exhibited an AUC of 0.825 (95% CI: 0.764–0.875) with a sensitivity of 82.71% and a specificity of 70.77% in the training cohort. The nomogram yielded an AUC of 0.858 (95% CI: 0.756–0.928) with a sensitivity of 80.77% and a specificity of 68.09% in the validation cohort (*Table 2*). For MVI prediction, the combined radiomics model showed the best performance compared with the clinicoradiological model (AUCs: 0.858 *vs.* 0.729; P=0.022) in the validation cohort, although the training did not have statistical significance (AUCs: 0.825 *vs.* 0.753; P=0.055). ROC curves for the prediction for MVI were compared among the optimal radiomics signatures, and the best clinicoradiological model and the combined radiomics model are shown in *Figure 3A,B*, respectively.



**Figure 4** Radiomics nomogram combining the fusion radiomics signature derived from triphasic dynamic contrast-enhanced (DCE) MR images and clinicoradiological factors including serum alpha fetoprotein (AFP) levels and arterial peritumoral enhancement for predicting microvascular invasion (MVI).

#### Development and validation of the radiomics nomogram

The nomogram based on the combined radiomics model is presented in *Figure 4*. Calibration curves (*Figure 5A,B*) showed good consistency between the nomogram-predicted probability of MVI and the actual MVI rate. The Hosmer-Lemeshow test yielded nonsignificant statistics for both the training cohort ( $P=0.7541$ ) and the validation cohort ( $P=0.7916$ ). The decision curve (*Figure 5C,D*) demonstrated a higher net benefit of the nomogram than that of the treat-all strategy and the treat-none strategy. This result suggests that a therapy strategy based on the nomogram will improve the clinical outcome. An example of representative multimodal MR and relevant pathological images of two patients is given in *Figure 6*.

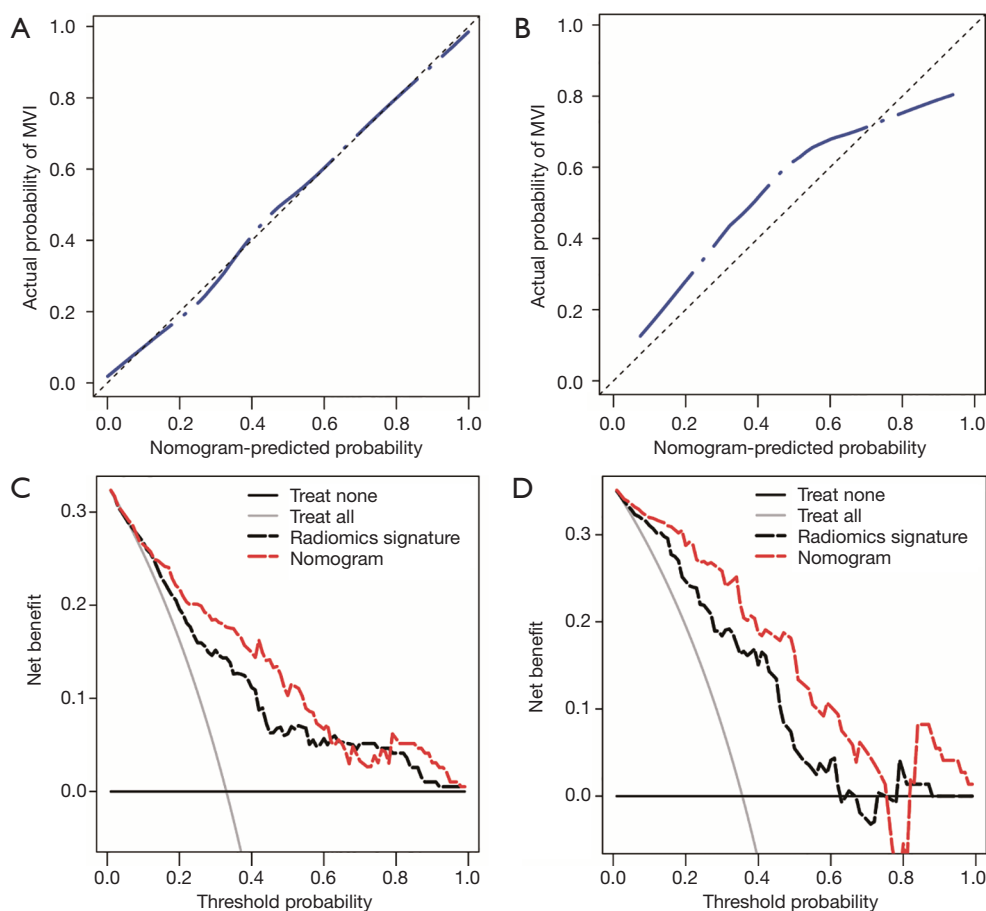
#### Discussion

In this study, we developed and validated a radiomics signature derived from MR images for preoperatively predicting MVI in HCC patients, which showed good discrimination both in the training and validation cohorts. Encouragingly, the diagnostic performance of the radiomics signature did not vary with tumor size or AFP level, suggesting that it is a robust predictor of MVI. The radiomics nomogram that incorporates fusion radiomics signature and clinicoradiological characteristics achieved satisfactory preoperative prediction of MVI, which may aid in clinical decision-making.

Recently few studies about the utility of radiomics in MVI prediction of HCC have been reported. Bakr *et al.* (8)

explored noninvasive biomarkers of MVI in HCC using quantitative and semantic image features extracted from triphasic contrast-enhanced CT images, showing that the radiomics signature better-identified MVI (AUC=0.76), compared to the semantic feature signature. However, the case number was 28 only, and the data lacked further validation. Zheng *et al.* (19) created two multivariate models for the prediction of MVI using quantitative image analysis, where the tumor was stratified by size, one model for patients with  $\leq 5$  cm tumors predicted MVI with an AUC of 0.80, and the other model for patients with  $>5$  cm tumors with an AUC of 0.88. The study similarly lacked further validation, and quantitative image analysis was only performed on a tumor-liver interface; however, the internal region of a tumor is also very informative and may reflect the tumor's underlying biological properties. Thus, our study used bi-regional radiomics parameters to predict the MVI of HCC. Peng *et al.* (21) reported that a nomogram that included portal venous and arterial phase CT-based radiomics features, radiologic features, and clinical factors could potentially distinguish MVI+ and MVI- in HBV-related HCC, with an AUC of 0.846 in the training cohort and 0.844 in the validation cohort, which was in accord with the results of our study. However, unlike our study, their study used only the largest cross-sectional area rather than the entire tumor, which might have caused information loss (34), and only extract portal venous and arterial phase CT data. Because our study fully extracted and utilized the space and timing characteristics contained in the triphasic DCE-MR images, the results of our study were more repeatable. Xu *et al.* (22) developed a computational approach integrating large-scale clinical and imaging modalities, especially radiomic features from contrast-enhanced CT, to predict MVI and clinical outcomes in HCCs, but the study did not include an equilibrium phase that would have been useful for the prediction of MVI (8). Ma *et al.* (23) were first to use 3D contrast-enhanced CT images to build a radiomics signature for predicting the MVI status of HCC, with an AUC of 0.727 in the training dataset. Our radiomics signature exhibited better performance; in addition to bi-regional radiomic features, the additional information carried by MR imaging might be one of the reasons for this improved performance. In addition to the above studies based on CT images, recently, Feng *et al.* (24) reported radiomics analysis of MR images for MVI prediction; however, the above studies were both based on liver-specific agent-enhanced MR images, and the costs of liver-specific agents are too high to be suitable for



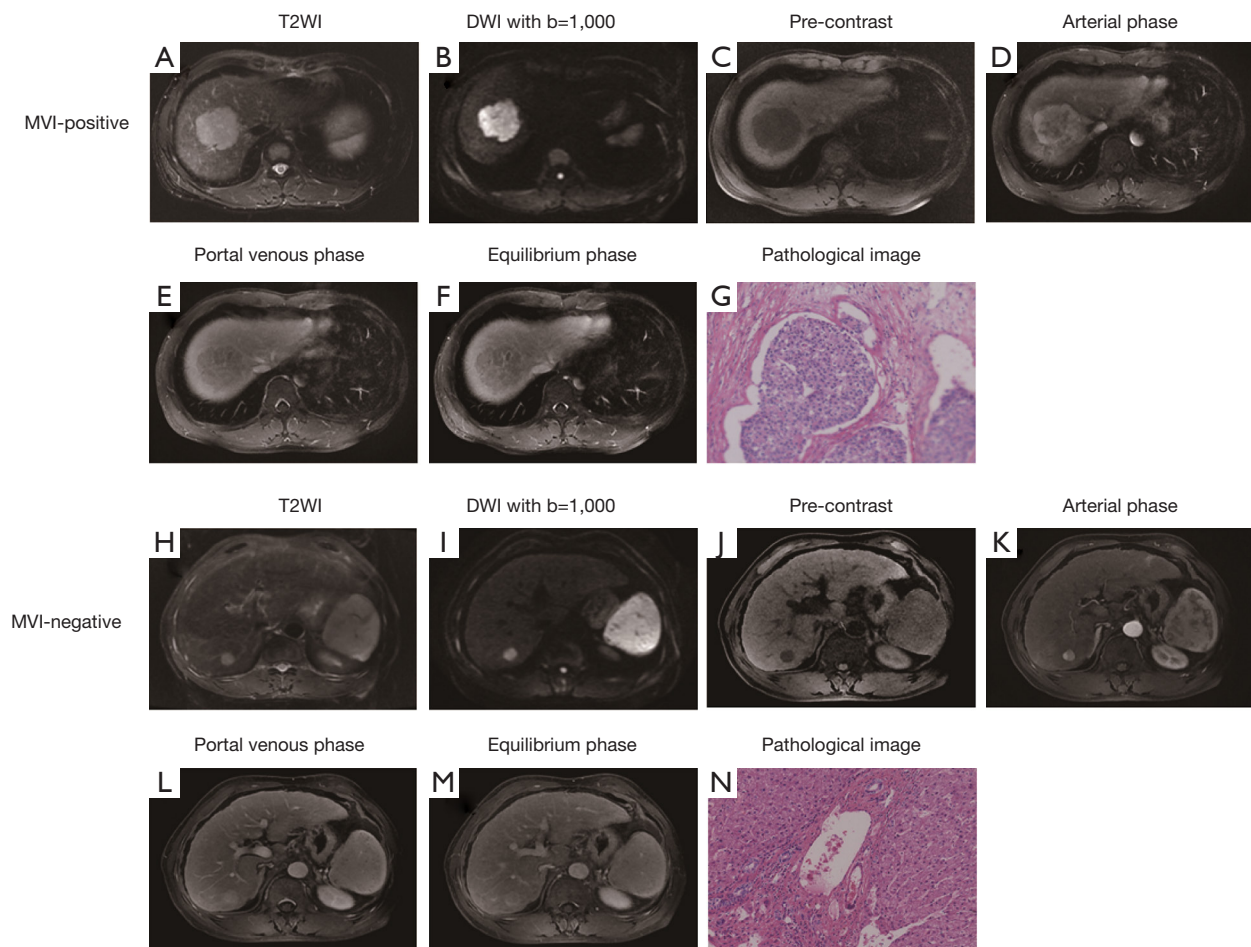


**Figure 5** The performance of the nomogram was assessed by calibration curves in the training cohort (A) and the validation cohort (B). In the calibration curves, the diagonal dashed line represents the ideal prediction, and the closer the blue dashed line is to the diagonal line, the higher the prediction accuracy of the model. The clinical utility of the nomogram was evaluated by decision curves in the training cohort (C) and the validation cohort (D). In the decision curves, the black line indicates the net benefit of assuming that there are no patients with microvascular invasion (MVI), and the grey line indicates the net benefit of assuming all patients with MVI. The red dashed line indicates the expected net benefit per patient based on the nomogram, and the black dashed line indicates the expected net benefit per patient based on the fusion radiomics signature. The farther the decision curve is from the two extreme curves, the higher the clinical decision net benefit of the model.

most HCC patients.

Our results revealed that the fusion radiomics signature derived from triphasic DCE MR images achieved optimal performance. This finding was not unexpected given that MVI can change the tumor perfusion by minute portal venous invasion (35,36), leading to detectable differences in contrast-enhancement between HCCs with and without MVI. However, the AUC of the radiomics signature derived from DW images for prediction of MVI in the present study was not very high (up to 0.732). The possible explanations may be due to the lower imaging resolution of DW images, which might be insufficient to reveal radiomics feature differences. Some predictive features came from the

peritumoral region in our study. Zheng *et al.* (19) showed that texture analysis in CT enhancement at the liver-tumor interface could predict MVI status in HCCs. The peritumoral region is a very informative region that may reflect underlying tumor biological properties, including MVI, extracellular matrix remodeling, and associated inflammatory response (15,36,37). During construction of the optimal radiomics signature, nearly all of the predictive features were wavelet features. This is in accord with previous studies that incorporated wavelet features in radiomics signature construction (38-42). The three-dimensional wavelet transformation decomposed image data into different frequency components, which may be useful



**Figure 6** Representative multimodal MR and relevant pathological images of microvascular invasion (MVI)-positive and MVI-negative hepatocellular carcinoma (HCC). (A,B,C,D,E,F,G) show a case of MVI-positive HCC. T2-weighted (A) and pre-contrast (B) images display a non-nodular lesion. Diffusion-weighted image with  $b=1,000 \text{ mm}^2/\text{s}$  (C) shows restricted diffusion. Arterial phase image (D) shows the presence of rim enhancement and peritumoral enhancement. Portal venous phase (E) and equilibrium phase (F) images display “washout” and incomplete radiological capsule. Pathological image (G) displays a tumor embolus in vascular channel (H&E,  $\times 100$ ). Figures h-n show a case of MVI-negative HCC. T2-weighted (H) and pre-contrast (I) images display a nodular lesion. Diffusion-weighted image with  $b=1,000 \text{ mm}^2/\text{s}$  (J) shows restricted diffusion. Arterial phase image (K) shows the absence of peritumoral enhancement. Portal venous phase (L) and equilibrium phase (M) images display complete radiological capsule. Pathological image (N) shows no tumor embolus in vascular channel (H&E,  $\times 100$ ).

for further exploration of the spatial tumor heterogeneity on multiple scales (39).

The fusion radiomics signature still performed well in the subgroups defined by tumor size and AFP level, which has very important clinical applicability. The fusion radiomics signature could contribute to the improved stratification of HCC patients into subgroups for individualized treatment. For example, the Milan criteria for LT is clearly imperfect, as described above. In this regard, the use of the radiomics signature for MVI prediction could broaden the criteria and further optimize the selection of LT patients. Moreover, we

found that the radiomics signature had more discriminating power in tumors  $\leq 5 \text{ cm}$ , compared to those that were  $> 5 \text{ cm}$ , and it also showed a strong discriminating power in tumors with normal serum AFP levels, suggesting that the biomarker could be used as an early detector for the aggressive behavior of HCC.

In this study, we also evaluated the clinicoradiological factors as surrogate biomarkers of MVI in HCC. Serum AFP levels and arterial peritumoral enhancement, which were included in our combined radiomics model, were also independent risk factors for MVI. These results agreed

with those from prior studies (8,11,13,14). Surprisingly, in this study, tumor size was not an independent risk factor for MVI, even though it was a significant predictor in the univariate analysis. Large tumor size has long been considered to increase the risk of MVI (43), but it has not always been proven to be an independent predictor (44). One possible reason was the selection bias: in this study, nearly 34% of HCCs exhibited MVI positivity, having a mean tumor size of 4.6 cm.

The combined model incorporating the significant clinicoradiological characteristics with the radiomics signature performed better than the radiomics signature alone did in the preoperative prediction of MVI for different individuals with HCCs. Perhaps a combination of more factors such as genomic datasets might obtain better efficiency of MVI prediction in HCCs. However, using multiple factors to predict MVI might also increase the burden on patients due to additional tests and exorbitant costs. Our developed combined model based on routinely available radiologic images and clinical laboratory data was easy to implement in routine clinical practice. Furthermore, of all the available prediction tools, a nomogram can provide a personalized, risk score-based, and highly accurate estimation along with ease of use (31). Our nomogram, based on the combined radiomics model, performed well, and demonstrated clinical utility for individualized therapeutic decision making.

This study has several limitations. First, this is a retrospective single-center study and requires further prospective multicentre validation with larger cohorts. Second, we did not examine tumor histopathology to correlate radiomics features with the exact locations of microscopic tumor thrombi, but promising results in this study may guide future studies on radiopathological correlations. Third, a follow-up of postoperative patients was not performed in this study. Next, we will follow up these patients and explore the potential of the combined nomogram in a survival prediction.

In conclusion, the fusion radiomics signature derived from triphasic DCE MR images can effectively predict MVI in HCC patients. The diagnostic performance of the fusion radiomics signature did not vary with tumor size or AFP level, suggesting that it is a robust predictor of MVI. The combined radiomics nomogram that incorporates clinicoradiological characteristics and the fusion radiomics signature achieved satisfactory preoperative prediction of MVI and thus will be able to assist clinicians in selecting optimal treatment strategies to improve clinical outcomes.

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## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

*Ethical Statement:* This study was approved by our Institutional Review Board, and the requirement for informed consent was waived.

## References

1. Forner A, Llovet JM, Bruix J. Hepatocellular carcinoma. *Lancet* (London, England) 2012;379:1245-55.
2. Fujiwara N, Friedman SL, Goossens N, Hoshida Y. Risk factors and prevention of hepatocellular carcinoma in the era of precision medicine. *J Hepatol* 2018;68:526-49.
3. Llovet JM, Schwartz M, Mazzaferro V. Resection and liver transplantation for hepatocellular carcinoma. *Semin Liver Dis* 2005;25:181-200.
4. De Angelis R, Sant M, Coleman MP, Francisci S, Baili P, Pierannunzio D, Trama A, Visser O, Brenner H, Ardanaz E, Bielska-Lasota M, Engholm G, Nennecke A, Siesling S, Berrino F, Capocaccia R; EUROCARE-5 Working Group. Cancer survival in Europe 1999-2007 by country and age: results of EUROCARE--5-a population-based study. *Lancet Oncol* 2014;15:23-34.
5. Roayaie S, Blume IN, Thung SN, Guido M, Fiel MI, Hiotis S, Labow DM, Llovet JM, Schwartz ME. A system of classifying microvascular invasion to predict outcome after resection in patients with hepatocellular carcinoma. *Gastroenterology* 2009;137:850-5.
6. Shi M, Guo RP, Lin XJ, Zhang YQ, Chen MS, Zhang CQ, Lau WY, Li JQ. Partial hepatectomy with wide versus narrow resection margin for solitary hepatocellular carcinoma: a prospective randomized trial. *Ann Surg* 2007;245:36-43.
7. Omata M, Cheng AL, Kokudo N, Kudo M, Lee JM, Jia J, Tateishi R, Han KH, Chawla YK, Shiina S, Jafri W, Payawal DA, Ohki T, Ogasawara S, Chen PJ, Lesmana

- CRA, Lesmana LA, Gani RA, Obi S, Dokmeci AK, Sarin SK. Asia-Pacific clinical practice guidelines on the management of hepatocellular carcinoma: a 2017 update. *Hepatol Int* 2017;11:317-370.
8. Bakr S, Echegaray S, Shah R, Kamaya A, Louie J, Napel S, Kothary N, Gevaert O. Noninvasive radiomics signature based on quantitative analysis of computed tomography images as a surrogate for microvascular invasion in hepatocellular carcinoma: a pilot study. *J Med Imaging (Bellingham)* 2017;4:041303.
  9. Mazzaferro V, Regalia E, Doci R, Andreola S, Pulvirenti A, Bozzetti F, Montalto F, Ammatuna M, Morabito A, Gennari L. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. *N Engl J Med* 1996;334:693-9.
  10. Grasso A, Stigliano R, Morisco F, Martines H, Quaglia A, Dhillon AP, Patch D, Davidson BR, Rolles K, Burroughs AK. Liver transplantation and recurrent hepatocellular carcinoma: predictive value of nodule size in a retrospective and explant study. *Transplantation* 2006;81:1532-41.
  11. Duvoux C, Roudot-Thoraval F, Decaens T, Pessione F, Badran H, Piardi T, Francoz C, Compagnon P, Vanlemmens C, Dumortier J, Dharancy S, Gugenheim J, Bernard PH, Adam R, Radenne S, Muscari F, Conti F, Hardwigsen J, Pageaux GP, Chazouillères O, Salame E, Hilleret MN, Lebray P, Abergel A, Debette-Gratien M, Kluger MD, Mallat A, Azoulay D, Cherqui D; Liver Transplantation French Study Group. Liver transplantation for hepatocellular carcinoma: a model including  $\alpha$ -fetoprotein improves the performance of Milan criteria. *Gastroenterology* 2012;143:986-94.e3.
  12. He J, Shi J, Fu X, Mao L, Zhou T, Qiu Y, Zhu B. The Clinicopathologic and Prognostic Significance of Gross Classification on Solitary Hepatocellular Carcinoma After Hepatectomy. *Medicine (Baltimore)* 2015;94:e1331.
  13. An C, Kim DW, Park YN, Chung YE, Rhee H, Kim MJ. Single hepatocellular carcinoma: preoperative MR imaging to predict early recurrence after curative resection. *Radiology* 2015;276:433-43.
  14. Renzulli M, Brocchi S, Cucchetti A, Mazzotti F, Mosconi C, Sportoletti C, Brandi G, Pinna AD, Golfieri R. Can current preoperative imaging be used to detect microvascular invasion of hepatocellular carcinoma? *Radiology* 2016;279:432-42.
  15. Banerjee S, Wang DS, Kim HJ, Sirlin CB, Chan MG, Korn RL, Rutman AM, Siripongsakun S, Lu D, Imanbayev G, Kuo MD. A computed tomography radiogenomic biomarker predicts microvascular invasion and clinical outcomes in hepatocellular carcinoma. *Hepatology* 2015;62:792-800.
  16. Reginelli A, Vanzulli A, Sgrazutti C, Caschera L, Serra N, Raucci A, Urraro F, Cappabianca S. Vascular microinvasion from hepatocellular carcinoma: CT findings and pathologic correlation for the best therapeutic strategies. *Med Oncol* 2017;34:93.
  17. Witjes CD, Willemsen FE, Verheij J, van der Veer SJ, Hansen BE, Verhoef C, de Man RA, Ijzermans JN. Histological differentiation grade and microvascular invasion of hepatocellular carcinoma predicted by dynamic contrast-enhanced MRI. *J Magn Reson Imaging* 2012;36:641-7.
  18. Kim KA, Kim MJ, Jeon HM, Kim KS, Choi JS, Ahn SH, Cha SJ, Chung YE. Prediction of microvascular invasion of hepatocellular carcinoma: usefulness of peritumoral hypointensity seen on gadoxetate disodium-enhanced hepatobiliary phase images. *J Magn Reson Imaging* 2012;35:629-34.
  19. Zheng J, Chakraborty J, Chapman WC, Gerst S, Gonen M, Pak LM, Jarnagin WR, DeMatteo RP, Do RKG, Simpson AL; Hepatopancreatobiliary Service in the Department of Surgery of the Memorial Sloan Kettering Cancer Center; Research Staff in the Department of Surgery at Washington University School of Medicine. Preoperative Prediction of Microvascular Invasion in Hepatocellular Carcinoma Using Quantitative Image Analysis. *J Am Coll Surg* 2017;225:778-788.e1.
  20. Lambin P, Leijenaar RTH, Deist TM, Peerlings J, de Jong EEC, van Timmeren J, Sanduleanu S, Larue RTHM, Even AJG, Jochems A, van Wijk Y, Woodruff H, van Soest J, Lustberg T, Roelofs E, van Elmpt W, Dekker A, Mottaghy FM, Wildberger JE, Walsh S. Radiomics: the bridge between medical imaging and personalized medicine. *Nat Rev Clin Oncol* 2017;14:749-62.
  21. Peng J, Zhang J, Zhang Q, Xu Y, Zhou J, Liu L. A radiomics nomogram for preoperative prediction of microvascular invasion risk in hepatitis B virus-related hepatocellular carcinoma. *Diagn Interv Radiol* 2018;24:121-7.
  22. Xu X, Zhang HL, Liu QP, Sun SW, Zhang J, Zhu FP, Yang G, Yan X, Zhang YD, Liu XS. Radiomic analysis of contrast-enhanced CT predicts microvascular invasion and outcome in hepatocellular carcinoma. *J Hepatol* 2019;70:1133-44.
  23. Ma X, Wei J, Gu D, Zhu Y, Feng B, Liang M, Wang S, Zhao X, Tian J. Preoperative radiomics nomogram for microvascular invasion prediction in hepatocellular carcinoma using contrast-enhanced CT. *Eur Radiol*

- 2019;29:3595-605.
24. Feng ST, Jia Y, Liao B, Huang B, Zhou Q, Li X, Wei K, Chen L, Li B, Wang W, Chen S, He X, Wang H, Peng S, Chen ZB, Tang M, Chen Z, Hou Y, Peng Z, Kuang M. Preoperative prediction of microvascular invasion in hepatocellular cancer: a radiomics model using Gd-EOB-DTPA-enhanced MRI. *Eur Radiol* 2019;29:4648-59.
  25. Colen RR, Wang J, Singh SK, Gutman DA, Zinn PO. Glioblastoma: imaging genomic mapping reveals sexspecific oncogenic associations of cell death. *Radiology* 2015;275:215-27.
  26. Cong WM, Bu H, Chen J, Dong H, Zhu YY, Feng LH, Chen J; Guideline Committee. Practice guidelines for the pathological diagnosis of primary liver cancer: 2015 update. *World J Gastroenterol* 2016;22:9279-87.
  27. Wu J, Aguilera T, Shultz D, Gudur M, Rubin DL, Loo BW Jr, Diehn M, Li R. Early-Stage Non-Small Cell Lung Cancer: Quantitative Imaging Characteristics of (18)F Fluorodeoxyglucose PET/CT Allow Prediction of Distant Metastasis. *Radiology* 2016;281:270-8.
  28. Prasanna P, Patel J, Partovi S, Madabhushi A, Tiwari P. Radiomic features from the peritumoral brain parenchyma on treatment-naïve multi-parametric MR imaging predict long versus short-term survival in glioblastoma multiforme: Preliminary findings. *Eur Radiol* 2017;27:4188-97.
  29. Duffy JP, Vardanian A, Benjamin E, Watson M, Farmer DG, Ghobrial RM, Lipshutz G, Yersiz H, Lu DS, Lassman C, Tong MJ, Hiatt JR, Busuttil RW. Liver transplantation criteria for hepatocellular carcinoma should be expanded: a 22-year experience with 467 patients at UCLA. *Ann Surg* 2007;246:502-9.
  30. Shim JH, Han S, Lee YJ, Lee SG, Kim KM, Lim YS, Chung YH, Lee YS, Lee HC. Half-life of serum alpha-fetoprotein: an early prognostic index of recurrence and survival after hepatic resection for hepatocellular carcinoma. *Ann Surg* 2013;257:708-17.
  31. Balachandran VP, Gonen M, Smith JJ, DeMatteo RP. Nomograms in oncology: more than meets the eye. *Lancet Oncol* 2015;16:e173-80.
  32. Kramer AA, Zimmerman JE. Assessing the calibration of mortality benchmarks in critical care: The Hosmer-Lemeshow test revisited. *Crit Care Med* 2007;35:2052-6.
  33. Vickers AJ, Elkin EB. Decision curve analysis: a novel method for evaluating prediction models. *Med Decis Making* 2006;26:565-74.
  34. Ng F, Kozarski R, Ganeshan B, Goh V. Assessment of tumor heterogeneity by CT texture analysis: can the largest crosssectional area be used as an alternative to whole tumor analysis? *Eur J Radiol* 2013;82:342-8.
  35. Simpson-Herren L, Noker PE, Wagoner SD. Variability of tumor response to chemotherapy. II. Contribution of tumor heterogeneity. *Cancer Chemother Pharmacol* 1988;22:131-6.
  36. Eccles SA, Welch DR. Metastasis: recent discoveries and novel treatment strategies. *Lancet* 2007;369:1742-57.
  37. Ünal E, İdilman İS, Akata D, Özmen MN, Karçaaltınçaba M. Microvascular invasion in hepatocellular carcinoma. *Diagn Interv Radiol* 2016;22:125-32.
  38. Liang W, Yang P, Huang R, Xu L, Wang J, Liu W, Zhang L, Wan D, Huang Q, Lu Y, Kuang Y, Niu T. A Combined Nomogram Model to Preoperatively Predict Histologic Grade in Pancreatic Neuroendocrine Tumors. *Clin Cancer Res* 2019;25:584-94.
  39. Wilson R, Devaraj A. Radiomics of pulmonary nodules and lung cancer. *Transl Lung Cancer Res* 2017;6:86-91.
  40. Huang Y, Liu Z, He L, Chen X, Pan D, Ma Z, Liang C, Tian J, Liang C. Radiomics Signature: A Potential Biomarker for the Prediction of Disease-Free Survival in Early-Stage (I or II) Non-Small Cell Lung Cancer. *Radiology* 2016;281:947-57.
  41. Wu S, Zheng J, Li Y, Yu H, Shi S, Xie W, Liu H, Su Y, Huang J, Lin T. A Radiomics Nomogram for the Preoperative Prediction of Lymph Node Metastasis in Bladder Cancer. *Clin Cancer Res* 2017;23:6904-11.
  42. Wu S, Zheng J, Li Y, Wu Z, Shi S, Huang M, Yu H, Dong W, Huang J, Lin T. Development and Validation of an MRI-Based Radiomics Signature for the Preoperative Prediction of Lymph Node Metastasis in Bladder Cancer. *EBioMedicine* 2018;34:76-84.
  43. Kaibori M, Ishizaki M, Matsui K, Kwon AH. Predictors of microvascular invasion before hepatectomy for hepatocellular carcinoma. *J Surg Oncol* 2010;102:462-8.
  44. Shirabe K, Kajiyama K, Abe T, Sakamoto S, Fukuya T, Akazawa K, Morita K, Maehara Y. Predictors of microscopic portal vein invasion by hepatocellular carcinoma: measurement of portal perfusion defect area ratio. *J Gastroenterol Hepatol* 2009;24:1431-6.
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### Supplementary I Stability and reproducibility test for radiomics features

We performed the stability and reproducibility test for image features. We randomly chose 60 patients from the overall patient dataset. A radiologist (J. Z.) repeated the tumor segmentation process for these 60 patients in DCE images. Then, the quantitative image features were extracted again. We defined the feature set used in the signature construction process as feature-set 1, the feature set extracted to test the stability and reproducibility as the feature-set 2. We used the Mann-Whitney U test to assess

the difference for each image feature between the feature-set 1 and feature-set 2. Then, the correlation coefficient for each image feature was calculated using the Spearman's rank correlation test between the feature set-1 and feature set-2.

A total of 2,561 features (the overall number: 2,932) showed no statistical significant difference between the feature-set 1 and feature-set 2. By using the correlation test, 2,463 image features showed statistical correlation between feature-set 1 and feature-set 2. The correlation coefficients and P values for features used in the radiomics signature are shown in *Table S1*.

**Table S1** The correlation coefficients and P values for features used in the radiomics signature

Feature name	Correlation coefficient	P value	P value of significant analysis
LRLGE_LHH_DCE_2_Outside	0.414822	0.000215	0.478613
LRHGE_LHH_DCE_3_Inside	0.497639	0.000006	0.971515
senh_DCE_1_Inside	0.945519	<0.000001	0.820107
SZE_HLL_DCE_2_Inside	0.532603	0.000001	0.231237
LZLGE_DCE_1_Inside	0.955420	0.000000	0.950547
LRLGE_LLH_DCE_1_Outside	0.800199	<0.000001	0.873079
LZHGE_DCE_2_Outside	0.512290	0.000003	0.881970
maxpr_LHH_DCE_2_Inside	0.386117	0.000623	0.497477
Ratio_Kurtosis_DCE_23	0.354794	0.001789	0.342571
SZE_HHL_DCE_2_Inside	0.363272	0.001358	0.116578
LRLGE_LLL_DCE_1_Inside	0.796358	<0.000001	0.750774
Skewness_DCE_1_Inside	0.845206	<0.000001	0.492725

T1-weighted images (968)	Inside feature	Intensity features	Texture features	Wavelet features
	Outside feature	Intensity features	Texture features	Wavelet features
T2-weighted images (968)	Inside feature:	Intensity features	Texture features	Wavelet features
	Outside feature:	Intensity features	Texture features	Wavelet features
DCE image (2,932)	Inside feature:	Intensity features	Texture features	Wavelet features
	Outside feature:	Intensity features	Texture features	Wavelet features
	Inside-outside co-occurrence feature	Delta-intensity features	Intensity ratio features	
DWI images (968)	Inside feature	Intensity features	Texture features	Wavelet features
	Outside feature	Intensity features	Texture features	Wavelet features

**Figure S1** Radiomics features used in this study.

**Table S2** Performances of the signatures from different modalities

Modality	Training group				Validation group			
	Accuracy	AUC (95% CI)	Sensitivity	Specificity	Accuracy	AUC (95% CI)	Sensitivity	Specificity
T1-weighted imaging	65.46%	0.705 (0.636–0.768)	76.56%	60.00%	54.79%	0.647 (0.527–0.756)	76.92%	46.81%
T2-weighted imaging	56.70%	0.699 (0.629–0.763)	92.19%	39.23%	41.10%	0.635 (0.514–0.745)	80.77%	40.43%
DCE MRI	75.26%	0.784 (0.719–0.840)	65.65%	80.00%	75.34%	0.820 (0.713–0.900)	69.23%	80.85%
DWI	66.49%	0.732 (0.615–0.829)	73.08%	68.09%	69.86%	0.706 (0.636–0.769)	64.06%	67.69%
All modality	71.13%	0.778 (0.712–0.834)	76.56%	68.46%	71.23%	0.803 (0.693–0.887)	76.92%	74.74%

AUC, area under ROC curve; CI, confidence interval; DCE, dynamic contrast-enhanced MRI; DWI, diffusion-weighted imaging. The AUC was reported in 95% confidence interval.

#### Radiomics signature

$$\begin{aligned}
 &= -1.0442 - 0.2045 \times \text{LRLGE\_LHH\_DCE\_2\_Outside} \\
 &- 0.0359 \times \text{LRHGE\_LHH\_DCE\_3\_Inside} - 0.1586 \times \text{se nth\_DCE\_1\_Inside} \\
 &- 0.1364 \times \text{SZE\_HLL\_DCE\_2\_Inside} - 0.0924 \times \text{LZLGE\_DCE\_1\_Inside} \\
 &- 0.0486 \times \text{LRLGE\_LLH\_DCE\_1\_Outside} + 0.1091 \times \text{LZHGE\_DCE\_2\_Outside} \\
 &- 0.1498 \times \text{maxpr\_LHH\_DCE\_2\_Inside} - 1.8547 \times \text{Ratio\_Kurtosis\_DCE\_23} \\
 &- 0.5099 \times \text{SZE\_HHL\_DCE\_2\_Inside} - 0.5536 \times \text{LRLGE\_LLL\_DCE\_1\_Inside} \\
 &+ 0.5819 \times \text{Skewness\_DCE\_1\_Inside}
 \end{aligned}$$

#### Clinical model

$$\begin{aligned}
 &= -2.8480 + 0.9843 \times \text{Gross type} \\
 &+ 0.8765 \times \text{Arterial peritumoral enhancement} \\
 &+ 1.1694 \times \text{Arterial rim enhancement} + 1.1340 \times \text{Internal arteries} \\
 &+ 0.6243 \times \text{AFP level}
 \end{aligned}$$

**Figure S2** Formulas for the radiomics signature and clinical model.