

Performance Evaluation of Three URiSCAN Devices for Routine Urinalysis

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Background: This study compares the diagnostic performance (in routine urinalysis) of three URiSCAN devices and three Roche analyzers to manual microscopy and quantitative assays. **Methods:** We analyzed eight dipstick tests using three URiSCAN devices. The results were compared to those of the tests performed using three Roche analyzers. The results of leukocyte and erythrocyte screens were compared to those obtained using manual microscopy. Protein, glucose, pH, and specific gravity (SG) assays performed on the URiSCAN devices were compared with the results of corresponding quantitative assays. **Results:** The rates of agreement within one grade difference were found to be more than 94.3%. When compared with manual microscopy,

Key words: urinalysis; urine dipstick analysis; microscopic examination; semiquantitative; quantitative

the Optima provided better diagnostic performance for the detection of leukocytes compared with the Urisys 1100. Compared to the Urisys 2400, the Super plus provided better diagnostic performance with regard to both leukocytes and erythrocytes. There was good correlation between the three URiSCAN devices and each quantitative assay, except for SG detection. **Conclusion:** There were well correlated results between those of the three URiSCAN devices and those obtained using the corresponding Roche analyzers, quantitative assays, and manual microscopy. URiSCAN series devices are therefore suitable for routine urinalysis in clinical laboratories. *J. Clin. Lab. Anal.* **30**:424–430, 2016. © 2015 Wiley Periodicals, Inc.

Abbreviations

AUC = area under the curve
ROC = receiver-operating characteristic
SG = specific gravity

INTRODUCTION

Urinalysis is an important clinical tool for screening, diagnosis, and follow-up. Urinalysis also allows for the detection of urogenital or systemic disorders that give rise to chemical and physical abnormalities (1). Urine is evaluated by most urinalysis procedures using diagnostic reagent strips (dipsticks) for urine chemistry and microscopy for cell counts. The traditional microscopic techniques that involve urinary sediment are labor-intensive, time-consuming, imprecise, and potentially influenced by interobserver variability. Given these limitations, routine

urine testing with multiparameter dipsticks is considered the optimal initial step in analysis (2, 3). When used in combination with a urine analyzer, urinalysis with dipstick analysis is valuable for its convenience, speed, and reproducibility. These microchemistry systems have been available for many years and allow for qualitative and semiquantitative analyses in routine urinalysis. Using semiautomatic or fully automatic instruments to read a dipstick may eliminate interobserver variability and

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time-sensitive errors associated with visual interpretation (4).

URiSCAN devices (YD Diagnostics, Yongin-si, Republic of Korea) are some of the most commonly used semi-automatic and/or automatic urine analyzers in Korea. The Korean Association of Quality Assurance for Clinical Laboratory (2009 KEQAS-UA) conducted a quality assessment trial and found that 45.8% of the 692 analyzers used by participating institutions were URiSCAN devices (5). Although a prior performance evaluation of the URiSCAN Pro II and a comparative device has been conducted (6), there are no data available comparing the performance of the URiSCAN Optima and the URiSCAN Super plus with comparable devices, or with quantitative assays and microscopic examination.

Therefore, the aim of this study was to compare the diagnostic performances of three URiSCAN devices (the Optima, Pro II, and Super plus) and three Roche urine analyzers (the Urisys 1100, Cobas u411, and Urisys 2400; Roche Diagnostics, Mannheim, Germany). The analyzers in routine urinalysis were also compared with results obtained by manual microscopy and quantitative assays.

MATERIALS AND METHODS

Specimens

Fresh urine samples collected from 1,273 inpatients and outpatients between June 2013 and November 2013 were used in this study. The samples did not contain any preservatives. The urine specimens were transferred to two different test tubes. At least 10 ml was allocated for dipstick analysis, along with sequential microscopic examination. More than 2 ml were allocated for quantitative analysis. The samples were analyzed within 2 hr of arrival to the laboratory. This study was approved by our institutional review board (KBC13073D).

Methods

Dipstick analysis was performed, and analyzed using the six analyzers from URiSCAN and Roche. Eight measurements were made, including protein, blood, glucose, ketone, urobilinogen, bilirubin, nitrite, and leukocytes. Each URiSCAN device was compared to a similar Roche analyzer with regard to size, weight, test velocity, test cycle, and memory capacity. Each specimen was well mixed before it was tested with the dipstick analysis. All tests were performed according to each manufacturer's instructions. Three independent sets of comparison studies were performed sequentially for each specimen. A total of 1,273 noncentrifuged urine samples were analyzed with dipstick analyses and microscopic examination. These

included 438 specimens for the Optima versus Urisys 1100 comparison, 437 specimens for the URiSCAN Pro II versus Cobas u411 comparison, and 398 specimens for the URiSCAN Super plus versus Urisys 2400 comparison.

The results obtained in each pairwise comparison were considered concordant if they were within one grade difference of each other. The pairwise concordance rates between each set of analyzers were defined by the percent agreement rate. The differences in grading systems between URiSCAN and Roche analyzers precluded the direct application of statistical evaluation. In order to compensate for the differences in the grading systems used for different instruments, the grading system levels (of each test parameter) were converted to comparable scales. For example, the Pro II has five grades with regard to urobilinogen detection [\pm (0.1 mg/dl), + (1 mg/dl), ++ (4 mg/dl), +++ (8 mg/dl), ++++ (12 mg/dl)], while the Cobas u411 has five different grades [- , \pm (1 mg/dl), + (4 mg/dl), ++ (8 mg/dl), +++ (12 mg/dl)]. In this case (Pro II and Cobas u411), the following result pairs were considered to be comparable: (+/- and -), (+ and +/-), (++) and +), (+++ and ++), (++++ and +++).

Following dipstick analysis, urine specimens were centrifuged at 1800 rpm for 3 min. The remaining 200 μ l of sediment was reserved for microscopic examination. A qualified medical technologist performed all of the microscopic examinations with a single microscope under 400 \times magnification (DMLS2; Leica, Lockbourne, OH). In each sample, erythrocytes and leukocytes were counted in high power fields, and the average numbers (from ten fields) were recorded. A positive finding was defined as the presence of at least three erythrocytes and five leukocytes per high power field (7).

The URiSCAN device results for protein, glucose, pH, and specific gravity (SG) were compared to those of corresponding quantitative assays. These quantitative assays included use of the benzethonium chloride method for protein determination on the Cobas Integra 800 (Roche, Indianapolis, IN), the hexokinase method for glucose measurement on the Advia 1800 (Siemens, Tokyo, Japan), a pH meter (Mettler Toledo S220; Mettler Toledo, Zurich, Switzerland), and a refractometer (UG- α ; Atago, Tokyo, Japan). Other test parameters were not considered, as we were unable to perform the requisite quantitative assays due to laboratory limitations. The principle of the URiSCAN device is based on light reflected from the surface of the urine strip. The light is transferred through an optical fiber to a charge-coupled device sensor, where the reflected light is analyzed to determine the ratio of the three primary colors. The ratio of the three primary colors and compensation colors is used to calculate the change of reflectance rate (%R). An analog to digital converter is then used to convert this %R to an equivalent grade

TABLE 1. Correlation Between URiSCAN Devices and Roche Urine Analyzers Represented as Rate (%) of Results Within One Grading Difference

	Blood	Bil	Urobil	Ket	Pro	NT	Glu	Leu
Optima versus Urisys 1100	97.5	99.8	100	99.5	94.3	100	96.8	98.6
Pro II versus Cobas u411	97.5	100	100	99.8	95.7	100	98.2	97.5
Super plus versus Urisys 2400	99.5	100	99.7	98.5	98.2	100	94.7	99.0

Bil, bilirubin; Urobil, urobilinogen; Ket, ketones; Pro, protein; NT, nitrite; Glu, glucose; Leu, leukocytes.

or concentration (8). The average %R was calculated for each test parameter based on duplicate tests performed on the same dipstick. The average %R values were used in each comparison.

Statistical and Analytical Methods

The agreement rates and within-one-grade differences were calculated between the classified grades of the simultaneously evaluated URiSCAN and corresponding Roche devices. The area under the curve (AUC) was calculated from the receiver-operating characteristic (ROC) according to the trace or positive dipstick grading results using manual microscopic examination ($P < 0.05$). The correlation coefficient and regression line were obtained to verify the correlations between the change %R (of the URiSCAN devices) and the corresponding assays ($P < 0.001$). IBM SPSS version 18.0 (IBM, Armonk, New York, NY) and STATA version 13.0 (Stata Corp., College Station, TX) were used for statistical analyses. The correlation coefficients and regression lines were calculated using Pearson correlation and linear regression or Spearman's correlation, respectively.

RESULTS

Table 1 summarizes the agreements between the URiSCAN devices and the Roche analyzers for the same urine specimens. The concordance levels of greater than 94.3% to 100% were obtained for the eight parameters

investigated (considering the results within one grade of difference).

The results of leukocyte and erythrocyte screens were compared to those obtained by manual microscopy. The sensitivities and specificities of the URiSCAN devices and the Roche analyzers were then calculated (Table 2). The Super plus device offered better diagnostic performance (AUC 0.883) in erythrocyte detection than did the Urisys 2400 device (AUC 0.870). In the leukocyte screen, the Optima and Super plus units had better diagnostic performance (AUC 0.790 and 0.850, respectively) than did the Urisys 1100 and Urisys 2400 devices (AUC 0.740 and 0.770, respectively; $P < 0.05$). There were no statistical differences in the performances of the Optima and Urisys 1100 erythrocyte screens, or between the Pro II and Cobas u411 combined screens.

Correlation coefficients and regression lines were obtained by comparing the change %R of the URiSCAN devices and the corresponding quantitative methods (Figs. 1, 2, and 3). The quantitative protein results for each quantitative assay were adjusted by applying logarithmic values. There were good correlations using linear regression between protein and SG, both of which exceeded 0.904 ($P < 0.001$). In contrast, there was a poor correlation between the SG detection of the Optima and Pro II. Nonparametric and nonlinear scatter plots were observed for both glucose and pH. Spearman's correlation revealed good correlations, with glucose exceeding 0.712 ($P < 0.001$) and pH exceeding 0.934 ($P < 0.001$).

TABLE 2. Diagnostic Accuracy of Erythrocyte and Leukocyte Detection Using URiSCAN Devices and Roche Urine Analyzers With Manual Microscopy as the Standard

	Erythrocytes				Leukocytes			
	Sensitivity	Specificity	AUC	<i>P</i>	Sensitivity	Specificity	AUC	<i>P</i>
URiSCAN Optima	91.4	78.1	0.848	0.931	76.6	93.1	0.790	0.003 ^a
Urisys 1100	86.7	82.6	0.846		64.9	97.8	0.740	
URiSCAN Pro II	79.6	85.8	0.827	0.400	72.1	93.7	0.795	0.746
Cobas u411	76.8	86.4	0.816		70.9	94.6	0.790	
URiSCAN Super plus	92.9	83.8	0.883	0.018 ^a	87.5	84.8	0.850	0.008 ^a
Urisys 2400	92.9	81.1	0.870		95.8	61.5	0.770	

^a $P < 0.05$.

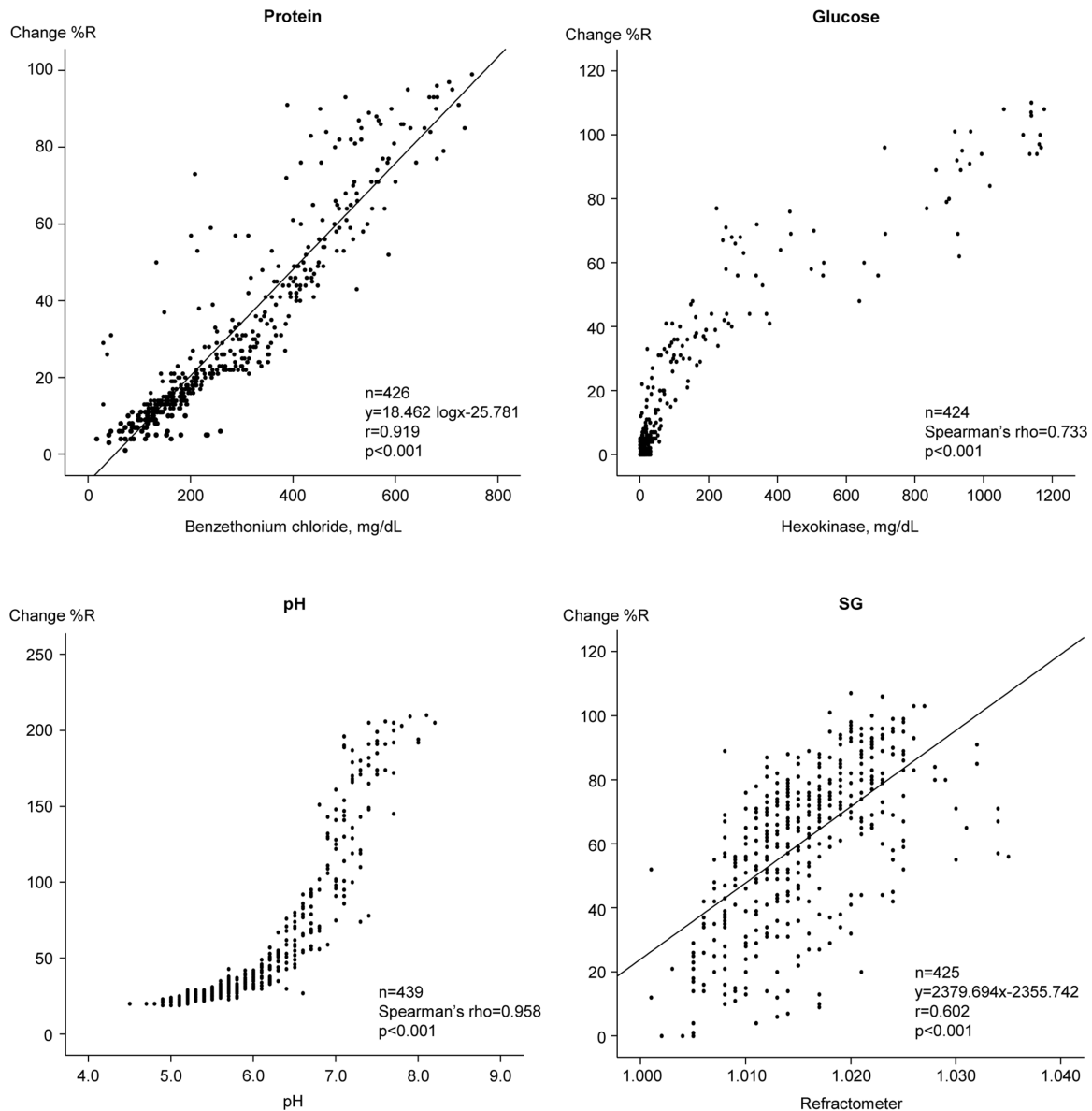


Fig. 1. Correlation coefficients between the quantitative assays and URiSCAN Optima results.

DISCUSSION

As shown in Table 1, there was good overall agreement between the three URiSCAN devices and their corresponding Roche analyzers. The agreement rates between Optima versus Urisys 1100 for protein and glucose were 94.3% and 96.8%, respectively. Among the discrepancy results, 24 of 25 results for protein were only detected by Optima. In contrast, Urisys 1100 demonstrated all 14 glucose results above one-grading scale. We did not investigate these analyzer discrepancies any further. However, we advise that urinalyses are carefully interpreted to avoid false-positives readings for protein and/or falsely elevated glucose measurements.

Microscopic urine examination is recommended to clarify false-negative or false-positive results on urinalysis. In this study, we compared the URiSCAN and Roche devices based on microscopic examination. In addition, AUCs were calculated according to the devices' sensitivities and specificities (Table 2). The URiSCAN Optima showed superior diagnostic performance for the detection of leukocytes, whereas we observed no difference in performance for erythrocyte detection. The URiSCAN Pro II showed equivalent performance levels for the detection of both leukocytes and erythrocytes. The URiSCAN Super plus showed the best diagnostic performance for the detection of both leukocytes and erythrocytes. One limitation to this

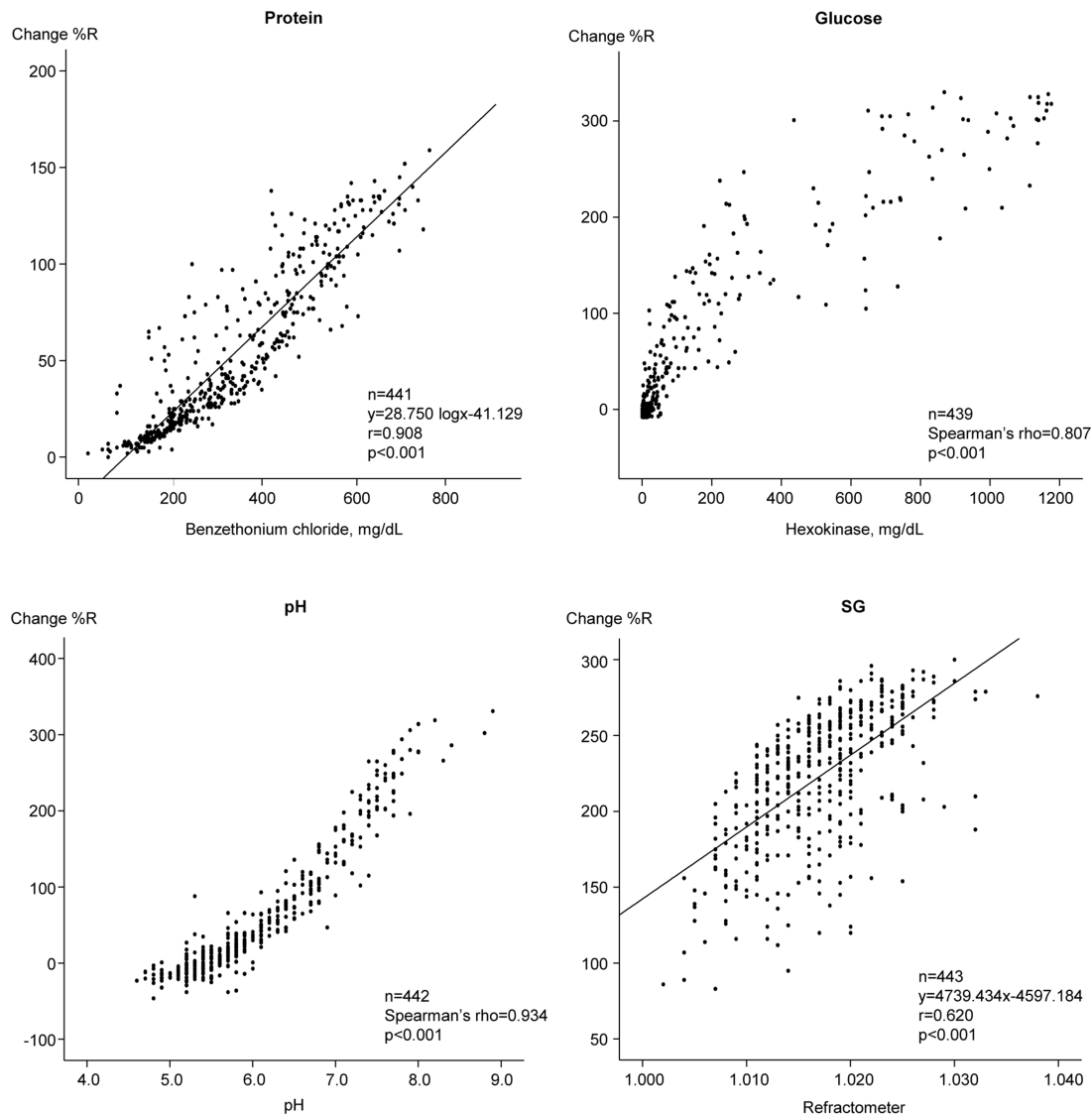


Fig. 2. Correlation coefficients between the quantitative assays and URiSCAN Pro II results.

study, however, is that we did not investigate factors that may have influenced false-negative and/or false-positive results. According to previous studies, there are numerous factors that influence dipstick-based urinalysis and can cause false-negative/false-positive results. When dipstick analysis is used for leukocyte detection, false-positive results may be caused by urine contamination by vaginal discharge and bacteriuria. False-negative results, in contrast, may occur in the setting of elevated SG, glycosuria, ketonuria, proteinuria, oxidizing drugs (cephalexin, nitrofurantoin, tetracycline, and gentamicin), and ascorbic acid. False-positive erythrocyte detection may occur in the presence of hemoglobinuria, myoglobinuria, menstrual blood, or dehydration. False-negative erythrocyte detection, in contrast, may occur if the patient takes

captopril, or has high SG, acidemia, proteinuria, or ascorbic acid in the urine (1, 9–11). The sensitivity of leukocyte detection ranged between 64.9% and 95.8%. Previous studies have suggested that leukocyte sensitivity varies according to each urinalysis device. For example, that of the Combostick Reader 720 is 82.4% (6), the URiSCAN Gen 10SGL is 63.6% (8), UriSed is 82% (12), Fus100 is 68% (12), iQ200 is 87% (13), UF-100 is 71.8% (14), and URiSCAN Super is 87% (15). The differences may be due to varying methodologies used to determine these parameters, such as the strip test analyzers, which measure enzymatic activity rather than counting cells.

The three URiSCAN devices were well correlated with the quantitative assays with regard to four parameters tested (protein, glucose, pH, and SG; Figs. 1–3. We

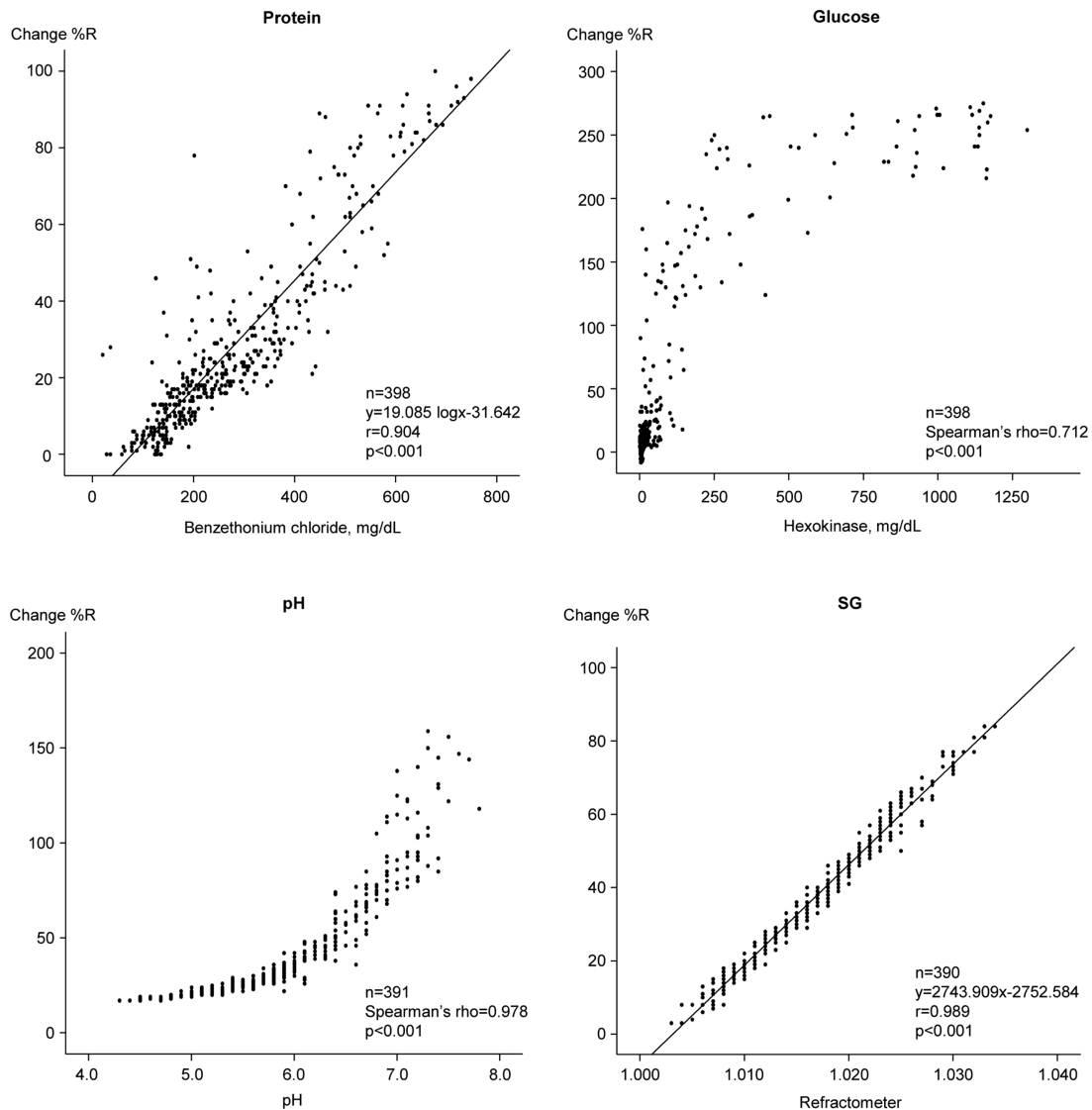


Fig. 3. Correlation coefficients between the quantitative assays and URiSCAN Super plus results.

observed parametric and linear scatters for protein and SG; therefore, linear regression was used to evaluate these two variables. In contrast, we observed nonparametric and nonlinear scatters for glucose and pH. We evaluated these variables by adjusting Spearman's correlation. There was poor correlation between the dipstick analyses with Optima and Pro II instruments and the quantitative assay (with a refractometer) for urine SG. This finding may have resulted from differences in the measurement principles for this assay. Most dipstick analyzers use similar principles for measuring SG. The strip relies on the correlation between the ionic solute concentration and the urine SG to provide an indirect measurement (16). In contrast, the Optima and the Pro II SG tests are based on the apparent pKa change of pretreated polyelectrolytes in relation to the ionic concentration. The presence of cations in urine

causes proton release from complex substances, which leads to changes in pH and thus changes the indicator colors of the stick. This method is not affected by nonionic substances. However, the presence of moderate quantities of protein may cause false elevation, while highly buffered alkaline urine samples may cause false-negative results (16–18). Refractometry is based on light refraction and can be affected by nonionic substances in the urine. When light passes from one medium into another, the light beam changes its direction at the surface boundary if its speed in the second medium is different from that in the first. The angle created by the light bending is called the critical angle and the ability of a substance to bend light is its refractivity. Therefore, the refractivity of a solution is an indirect measurement of the total solute concentration (7). In previous studies that compared the dipstick

method, which uses ionic environmental alteration as a measure, and the refractometer method for detection of urine SG, low correlations were reported (19–21). Urine SG measurements by the dipstick method has been used as an easy, rapid, noninvasive, and inexpensive way to interpret a patient's hydration status. However, these results should be carefully interpreted given the discrepancy that may exist between the dipstick and refractometer results (22–24). Ideally, the dipstick SG results should be confirmed using a refractometer before a final diagnosis is made. The URiSCAN Super plus contains a built-in refractometer unlike the Optima or Pro II. Therefore, the URiSCAN Super plus may be more useful for measuring urine SG than are the Optima or Pro II (25). As might be expected, therefore, the URiSCAN Super plus results were more highly correlated with those of the quantitative assay ($r = 0.989$, $P < 0.001$) than were the results obtained using the Optima ($r = 0.602$) and Pro II ($r = 0.620$).

In conclusion, the three URiSCAN devices (Optima, Pro II, and Super plus) are well correlated with the corresponding Roche analyzers when used for dipstick analysis. The results obtained using these machines were also comparable to those obtained with quantitative assays. The three URiSCAN devices were comparable to or better than were the Roche analyzers at detecting erythrocytes and leukocytes. This URiSCAN series will likely be valuable for routine urinalysis in clinical laboratories.

CONFLICT OF INTEREST

There are no conflicts of interest regarding the publication of this article.

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