



Complete Genome Sequence of *Citrobacter freundii* Myophage Maroon

James R. McDermott,^a Qiuyan Shao,^a Chandler O'Leary,^a  Rohit Kongari,^a Mei Liu^a

^aCenter for Phage Technology, Texas A&M University, College Station, Texas, USA

ABSTRACT *Citrobacter freundii* is a nosocomial opportunistic pathogen that can cause urinary and bloodstream infections. Phage therapies against *C. freundii* may prove useful in treating infections caused by this ubiquitous bacterium. Here, we report the complete genome of a T4-like myophage, Maroon, that infects *C. freundii*.

Citrobacter freundii is a facultative anaerobic Gram-negative bacterium that can cause opportunistic nosocomial infections within the bloodstream, respiratory tract, and urinary tract in immunocompromised patients (1, 2). With the increase of multidrug-resistant *Citrobacter* infections (3), phage therapy is being investigated as an alternative solution (4, 5). Here, we present the complete genome of the *C. freundii* myophage Maroon.

Phage Maroon was isolated from a municipal wastewater sample collected from Brazos County, Texas, in 2015 using a *C. freundii* strain as the host. LB broth or agar (Difco) was used to culture the host bacterium and phage enrichment at 37°C with aeration. Phage isolation and propagation were conducted using the soft-agar overlay method (6). It was identified as a siphophage using negative-stain transmission electron microscopy performed at the Texas A&M University Microscopy and Imaging Center as described previously (7). Phage genomic DNA was prepared using a modified Promega Wizard DNA cleanup kit protocol (7). Pooled indexed DNA libraries were prepared using the Illumina TruSeq Nano LT kit, and the sequence was obtained from the Illumina MiSeq platform using the MiSeq V2 500-cycle reagent kit following the manufacturer's instructions, producing 616,387 paired-end reads for the index containing the Maroon genome. FastQC 0.11.5 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) was used to quality control the reads. The reads were trimmed with FastX Toolkit 0.0.14 (http://hannonlab.cshl.edu/fastx_toolkit/download.html) before being assembled using SPAdes 3.5.0 (8). Contig completion was confirmed by PCR using primers (5'-CCTGGG ATATCCGTAATTGG-3' and 5'-TATCGAAGCCATTTTGACCA-3') facing off the ends of the assembled contig and Sanger sequencing of the resulting product, with the contig sequence manually corrected to match the resulting Sanger sequencing read. GLIMMER 3.0 (9) and MetaGeneAnnotator 1.0 (10) were used to predict protein-coding genes with manual verification, and tRNA genes were predicted with ARAGORN 2.36 (11). Rho-independent termination sites were identified using Transtern (<http://transtern.cbc.umd.edu/>). Sequence similarity searches were done using BLASTp 2.2.28 (12) against the NCBI nonredundant (nr), UniProt Swiss-Prot (13), and TrEMBL databases with a 0.001 maximum expectation value cutoff. InterProScan 5.15-54.0 (14), Lipop (15), and TMHMM 2.0 (16) were used to predict protein function. All analyses were conducted at default settings via the CPT Galaxy (17) and Web Apollo (18) interfaces (<https://cpt.tamu.edu/galaxy-pub>).

Maroon was assembled at 32.2-fold coverage into a 178,830-bp genome. The GC content of the genome is 44.9%. A total of 277 coding sequences were annotated, resulting in a protein-coding density of 95%. Maroon shares nucleotide-level similarity

Citation McDermott JR, Shao Q, O'Leary C, Kongari R, Liu M. 2019. Complete genome sequence of *Citrobacter freundii* myophage Maroon. *Microbiol Resour Announc* 8:e01145-19. <https://doi.org/10.1128/MRA.01145-19>.

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2019 McDermott et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Mei Liu, meiliu@tamu.edu.

Received 13 September 2019

Accepted 5 October 2019

Published 24 October 2019

across the genome with many T4-like phages, such as *Citrobacter* phage Margaery (GenBank accession no. [KT381880](#)) and *Cronobacter* phage vB_CsaM_leB (GenBank accession no. [KX431559](#)) (sharing 99% and 94% DNA similarity, respectively). While 168 of the identified genes do not have a predicted function, proteins with annotated functions have homologs (via BLASTp against the NCBI nr database at an E value of <0.001) in T4 (GenBank accession no. [NC_000866](#)) or T4-like phages. Genes involved in DNA replication and nucleotide biosynthesis (DNA helicase, *nrdA*, *nrdB*, polynucleotide kinase, etc.) and virion morphogenesis (capsid proteins, tail fibers, tail sheath stabilizer, tail baseplate, etc.) were all identified. Lysis genes coding for a class III holin, a soluble L-alanyl-D-glutamate peptidase, and a separated spanin pair were also found.

Data availability. The genome sequence of phage Maroon was deposited under GenBank accession no. [MH823906](#). The associated BioProject, SRA, and BioSample accession numbers are [PRJNA222858](#), [SRR8770019](#), and [SAMN11233125](#), respectively.

ACKNOWLEDGMENTS

This work was supported by funding from the National Science Foundation (awards EF-0949351 and DBI-1565146). Additional support came from the Center for Phage Technology (CPT), an Initial University Multidisciplinary Research Initiative supported by Texas A&M University and Texas AgriLife, and from the Department of Biochemistry and Biophysics at Texas A&M University.

We are grateful for the advice and support of the CPT staff and the Texas A&M University Microscopy and Imaging Center. This announcement was prepared in partial fulfillment of the requirements for BICH464 Phage Genomics, an undergraduate course at Texas A&M University.

REFERENCES

- Ranjan KP, Ranjan N. 2013. *Citrobacter*: an emerging health care associated urinary pathogen. *Urol Ann* 5:313–314.
- Huang SH, Stins MF, Kim KS. 2000. Bacterial penetration across the blood-brain barrier during the development of neonatal meningitis. *Microbes Infect* 2:1237–1244. [https://doi.org/10.1016/S1286-4579\(00\)01277-6](https://doi.org/10.1016/S1286-4579(00)01277-6).
- Liu LH, Wang NY, Wu AY, Lin CC, Lee CM, Liu CP. 2018. *Citrobacter freundii* bacteremia: risk factors of mortality and prevalence of resistance genes. *J Microbiol Immunol Infect* 51:565–572. <https://doi.org/10.1016/j.jmii.2016.08.016>.
- Chaudhry W, Haq IU, Andleeb S, Qadri I. 2014. Characterization of a virulent bacteriophage LK1 specific for *Citrobacter freundii* isolated from sewage water. *J Basic Microbiol* 54:531. <https://doi.org/10.1002/jobm.201200710>.
- Hamdi S, Rousseau GM, Labrie SJ, Kourda RS, Tremblay DM, Moineau S, Slama KB. 2016. Characterization of five podoviridae phages infecting *Citrobacter freundii*. *Front Microbiol* 7:1023–1023. <https://doi.org/10.3389/fmicb.2016.01023>.
- Adams MK. 1959. *Bacteriophages*. Interscience Publishers, Inc., New York.
- Gill JJ, Berry JD, Russell WK, Lessor L, Escobar-Garcia DA, Hernandez D, Kane A, Keene J, Maddox M, Martin R, Mohan S, Thorn AM, Russell DH, Young R. 2012. The *Caulobacter crescentus* phage phiCbK: genomics of a canonical phage. *BMC Genomics* 13:542. <https://doi.org/10.1186/1471-2164-13-542>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledgebase. *Nucleic Acids Res* 46:2699. <https://doi.org/10.1093/nar/gky092>.
- Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
- Juncker AS, Willenbrock H, Von Heijne G, Brunak S, Nielsen H, Krogh A. 2003. Prediction of lipoprotein signal peptides in Gram-negative bacteria. *Protein Sci* 12:1652–1662. <https://doi.org/10.1110/ps.0303703>.
- Krogh A, Larsson B, von Heijne G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- Cock PJ, Gruning BA, Paszkiewicz K, Pritchard L. 2013. Galaxy tools and workflows for sequence analysis with applications in molecular plant pathology. *PeerJ* 1:e167. <https://doi.org/10.7717/peerj.167>.
- Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elsik CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. *Genome Biol* 14:R93. <https://doi.org/10.1186/gb-2013-14-8-r93>.