

Antiretroviral Treatment of Acute HIV Infection Normalizes Levels of Cerebrospinal Fluid Markers of Central Nervous System (CNS) Inflammation: A Consequence of a Reduced CNS Reservoir?

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(See the Major Article by Hellmuth et al, on pages 1885–91.)

Antiretroviral treatment (ART) has transformed human immunodeficiency virus (HIV) infection from a generally lethal infection into a chronic disease, enabling persons living with HIV (PLWH) to have a near-normal life. Still, HIV-infected individuals continue to have a shorter life expectancy than the general population, which may in part be explained by differences in lifestyle factors such as smoking, alcohol consumption, and drug use among PLWH [1]. Persistent immune activation and inflammation may also contribute to increased age-related morbidity and mortality in PLWH. Levels of immune activation markers commonly continue to be increased despite suppressive ART. This is probably driven by HIV release from latently infected resting T cells that are randomly activated, as well as other cells. Other sources may be microbial translocation, lifestyle factors, and coinfections, such as that due to cytomegalovirus [1, 2]. The clinical consequences of this low-grade chronic

immune activation are not fully known, but it is clear that increased levels of inflammatory biomarkers are predictive of subsequent morbidity and mortality [3, 4].

It is beneficial to initiate ART in patients with high CD4⁺ T-cell counts, rather than postponing treatment [5]. The reduction of systemic inflammation by ART may help explain many of the benefits [4]. Nevertheless, not all comorbidities are reduced by early initiation of ART in persons with high CD4⁺ T-cell counts. For example, cardiovascular complications (or surrogate markers of cardiovascular disease), pulmonary disease markers, and neurocognitive dysfunction did not appear to improve significantly with early ART initiation in the START trial, despite major declines in infections and infection-related cancers [6–8]. While the relatively young age of START participants, in addition to measurement issues, may have contributed to the lack of any clear beneficial impact of immediate ART initiation on these morbidities, it is also possible, that irreversible HIV-associated organ injury in the relevant tissues (ie, the vasculature, lung, and brain) may not yet have been established at such early disease stages [9].

HIV is a neurotropic virus and can be found in the central nervous system (CNS) early after infection [10]. While infected CD4⁺ T cells often traffic through the cerebrospinal fluid (CSF) during untreated

disease, an active, productive infection is also established within the CNS itself, mainly in microglia and brain macrophages, leading to chronic CNS immune activation that may contribute to neuronal damage if left untreated [11]. Similar to its effect outside the CNS, ART substantially decreases intrathecal immune activation, although not to normal levels [12]. Notably, CD4⁺ T-cell trafficking through the CSF is often reduced to near normal levels with ART [13]. Yet, CSF levels of neopterin, a pteridine marker of primarily macrophage activation [14], have been found to be stably increased in the majority of PLWH who begin treatment during the chronic phase of HIV infection and also after long-term receipt (ie, for >10 years) of suppressive ART [15, 16].

By contrast, Hellmuth et al [17] report in this issue of *The Journal of Infectious Diseases* that, in the CSF, levels of markers of residual CNS inflammation are essentially normalized when ART is initiated early, during acute HIV infection (AHI), while plasma levels of markers remain abnormal [17]. This important finding has several potential implications. The most plausible explanation for the difference is that a stable, permanent infection of cells in the CNS is established later than in systemic viral reservoirs, which are highly concentrated in memory T-cell compartments within the first days of systemic HIV infection [9]. This hypothesis is supported by a recent study

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examining anti-HIV antibody formation as a surrogate marker for antigen load and the size of the viral reservoir [18] in patients followed longitudinally during early HIV infection [19]. While serum anti-HIV antibodies emerged in blood by day 30 in untreated early infection, CSF antibodies reached similar levels about 2 weeks later. In addition, high antibody levels, comparable to those observed in chronically infected subjects, were reached several months later in CSF, compared with blood. Furthermore, while treatment of chronic infection resulted in only small reductions in levels of anti-HIV antibodies in both CSF and serum, treatment during early infection substantially reduced CSF levels of antibodies, sometimes to levels close to those in HIV-negative controls, whereas antibody levels in serum were less affected [19]. To the extent that differential CSF and blood antibodies indicate HIV persistence, these data strongly suggest a relative delay in the establishment of the CNS reservoir, compared with the systemic HIV reservoir. If true, this provides an opportunity for early treatment to have a greater impact on the magnitude of long-term CNS infection, supported by the normalization of CSF levels of immune activation biomarkers by early ART initiation, as reported by Hellmuth et al [17].

CNS infection is generally well controlled by systemic suppressive ART, although sometimes the HIV RNA load is increased in the CSF despite suppression of the plasma viral load, a phenomenon referred to as “CSF viral escape” [20]. Occasionally, CSF viral escape is accompanied by neurological and neurocognitive signs and symptoms (symptomatic CSF escape) [21]. However, asymptomatic escape, found when lumbar puncture is performed in study protocols, is more common [22, 23]. CSF escape is associated with increased intrathecal immune activation and may be related to the size of the CNS HIV reservoir, although this has not been definitely proven. Correspondingly, the residual CSF viral load below the limit of quantification of standard assays also correlates with the degree of CSF immune activation in PLWH receiving suppressive ART [24,

25], reinforcing the view that intrathecal immune activation is driven by persistent virus in the CNS. Nevertheless, similar to findings during systemic infection [26], treatment intensification does not seem to decrease the residual CSF viral load or inflammation [27, 28], suggesting there is no ongoing HIV replication during effective treatment. Altogether, these data suggest a close association between the CNS reservoir and CSF immune activation.

Macrophages and microglia are the main target cells for HIV replication in the CNS, which is different from systemic infection, where lymphocytic infection dominates [29]. This, together with CNS being an immune-privileged anatomic compartment protected by the blood-brain barrier, provides a foundation for compartmentalized HIV-1 replication within the CNS reservoir [30]. The CNS reservoir has not yet been adequately characterized, either in morphological terms or with regard to the mechanisms leading to its establishment and maintenance. A better understanding of the CNS reservoir formation prior to treatment and of its preservation during ART is important when exploring HIV eradication strategies, but also if one is to avoid severe CNS adverse events during the course of HIV cure studies [31]. It would be of great benefit for cure strategies if the HIV CNS reservoir were substantially reduced by early ART, as indicated in the study by Hellmuth et al [17].

The clinical implications of normalized intrathecal immune activation in PLWH treated during acute HIV infection are unclear. Will the long-term risk for cerebrovascular events, HIV-associated neurocognitive disease, and non-HIV-associated dementia, such as Alzheimer disease and vascular dementia, decline? Although there are some indications of an association between immune activation and these conditions [32, 33], there is yet no evidence that early initiation of ART prevents neurological or neurocognitive diseases.

Notes

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References

1. De Francesco D, Wit FW, Bürkle A, et al. Do people living with HIV experience greater age advancement than their HIV-negative counterparts? *AIDS* **2019**; 33:259–68.
2. Hunt PW, Martin JN, Sinclair E, et al. Valganciclovir reduces T cell activation in HIV-infected individuals with incomplete CD4+ T cell recovery on antiretroviral therapy. *J Infect Dis* **2011**; 203:1474–83.
3. Borges ÁH, Silverberg MJ, Wentworth D, et al.; INSIGHT SMART; ESPRIT; SILCAAT Study Groups. Predicting risk of cancer during HIV infection: the role of inflammatory and coagulation biomarkers. *AIDS* **2013**; 27:1433–41.
4. Baker JV, Sharma S, Grund B, et al. Systemic inflammation, coagulation, and clinical risk in the START trial. *Open forum Infect Dis* **2017**; 4:ofx262.
5. Lundgren JD, Babiker AG, Gordin F, et al.; INSIGHT START Study Group. Initiation of antiretroviral therapy in early asymptomatic HIV infection. *N Engl J Med* **2015**; 373:795–807.
6. Wright EJ, Grund B, Robertson KR, et al. No neurocognitive advantage for immediate antiretroviral treatment in adults with greater than 500 CD4+ T Cell Counts. *AIDS* **2018**; 32:1.

7. Kunisaki KM, Niewoehner DE, Collins G, et al; INSIGHT START Pulmonary Substudy Group. Pulmonary effects of immediate versus deferred antiretroviral therapy in HIV-positive individuals: a nested substudy within the multicentre, international, randomised, controlled Strategic Timing of Antiretroviral Treatment (START) trial. *Lancet Respir Med* **2016**; 4:980–9.
8. Baker JV, Sharma S, Achhra AC, et al. Changes in cardiovascular disease risk factors with immediate versus deferred antiretroviral therapy initiation among HIV-positive participants in the START (Strategic Timing of Antiretroviral Treatment) Trial. *J Am Heart Assoc* **2017**; 6 (5). PMID: 28533305.
9. Hunt PW, Lee SA, Siedner MJ. Immunologic Biomarkers, Morbidity, and Mortality in Treated HIV Infection. *J Infect Dis* **2016**; 214(Suppl 2):S44–50.
10. Spudich S, Gisslen M, Hagberg L, et al. Central nervous system immune activation characterizes primary human immunodeficiency virus 1 infection even in participants with minimal cerebrospinal fluid viral burden. *J Infect Dis* **2011**; 204:753–60.
11. Hagberg L, Fuchs D, Rosengren L, Gisslén M. Intrathecal immune activation is associated with cerebrospinal fluid markers of neuronal destruction in AIDS patients. *J Neuroimmunol* **2000**; 102:51–5.
12. Price RW, Spudich S. Antiretroviral therapy and central nervous system HIV type 1 infection. *J Infect Dis* **2008**; 197(Suppl 3):S294–306.
13. Sinclair E, Ronquillo R, Lollo N, et al. Antiretroviral treatment effect on immune activation reduces cerebrospinal fluid HIV-1 infection. *J Acquir Immune Defic Syndr* **2008**; 47:544–52.
14. Hagberg L, Cinque P, Gisslen M, et al. Cerebrospinal fluid neopterin: an informative biomarker of central nervous system immune activation in HIV-1 infection. *AIDS Res Ther* **2010**; 7:15.
15. Ulfhammer G, Edén A, Mellgren Å, et al. Persistent central nervous system immune activation following more than 10 years of effective HIV antiretroviral treatment. *AIDS* **2018**; 32:2171–8.
16. Yilmaz A, Yiannoutsos CT, Fuchs D, et al. Cerebrospinal fluid neopterin decay characteristics after initiation of antiretroviral therapy. *J Neuroinflammation* **2013**; 10:62.
17. Hellmuth J, Slike BM, Sacdalan C, et al. Very Early Initiation of Antiretroviral Therapy During Acute HIV Infection Is Associated With Normalized Levels of Immune Activation Markers in Cerebrospinal Fluid but Not in Plasma. *J Infect Dis* **2019**; 220:1885–91.
18. Lee SA, Bacchetti P, Chomont N, et al. Anti-HIV antibody responses and the hiv reservoir size during antiretroviral therapy. *PLoS One* **2016**; 11:e0160192.
19. Burbelo PD, Price RW, Hagberg L, et al. Anti-human immunodeficiency virus antibodies in the cerebrospinal fluid: evidence of early treatment impact on central nervous system reservoir? *J Infect Dis* **2018**; 217:1024–32.
20. Ferretti F, Gisslen M, Cinque P, Price RW. Cerebrospinal fluid HIV escape from antiretroviral therapy. *Curr HIV/AIDS Rep* **2015**; 12:280–8.
21. Canestri A, Lescure FX, Jaureguiberry S, et al. Discordance between cerebral spinal fluid and plasma HIV replication in patients with neurological symptoms who are receiving suppressive antiretroviral therapy. *Clin Infect Dis* **2010**; 50:773–8.
22. Edén A, Nilsson S, Hagberg L, et al. Asymptomatic cerebrospinal fluid HIV-1 viral blips and viral escape during antiretroviral therapy: a longitudinal study. *J Infect Dis* **2016**; 214:1822–5.
23. Edén A, Fuchs D, Hagberg L, et al. HIV-1 viral escape in cerebrospinal fluid of subjects on suppressive antiretroviral treatment. *J Infect Dis* **2010**; 202:1819–25.
24. Dahl V, Peterson J, Fuchs D, Gisslen M, Palmer S, Price RW. Low levels of HIV-1 RNA detected in the cerebrospinal fluid after up to 10 years of suppressive therapy are associated with local immune activation. *AIDS* **2014**; 28:2251–8.
25. Yilmaz A, Svennerholm B, Hagberg L, Gisslén M. Cerebrospinal fluid viral loads reach less than 2 copies/ml in HIV-1-infected patients with effective antiretroviral therapy. *Antivir Ther* **2006**; 11:833–7.
26. Dinoso JB, Kim SY, Wiegand AM, et al. Treatment intensification does not reduce residual HIV-1 viremia in patients on highly active antiretroviral therapy. *Proc Natl Acad Sci U S A* **2009**; 106:9403–8.
27. Dahl V, Lee E, Peterson J, et al. Raltegravir treatment intensification does not alter cerebrospinal fluid HIV-1 infection or immunoactivation in subjects on suppressive therapy. *J Infect Dis* **2011**; 204:1936–45.
28. Yilmaz A, Verhofstede C, D'Avolio A, et al. Treatment intensification has no effect on the HIV-1 central nervous system infection in patients on suppressive antiretroviral therapy. *J Acquir Immune Defic Syndr* **2010**; 55:590–6.
29. Joseph SB, Arrildt KT, Sturdevant CB, Swanstrom R. HIV-1 target cells in the CNS. *J Neurovirol* **2015**; 21:276–89.
30. Bednar MM, Sturdevant CB, Tompkins LA, et al. Compartmentalization, viral evolution, and viral latency of HIV in the CNS. *Curr HIV/AIDS Rep* **2015**; 12:262–71.
31. Winston A, Julie F, Fidler S. HIV cure strategies: response to ignore the central nervous system at your patients' peril. *AIDS* **2017**; 31(7):1051–2. doi:10.1097/QAD.0000000000001430. PMID: 28350582
32. Edén A, Marcotte TD, Heaton RK, et al. Increased intrathecal immune activation in virally suppressed HIV-1 infected patients with neurocognitive impairment. *PLoS One* **2016**; 11:e0157160.
33. Chitnis T, Weiner HL. CNS inflammation and neurodegeneration. *J Clin Invest* **2017**; 127:3577–87.