





Complete Genome Sequence of Staphylococcus arlettae Strain P2, Isolated from a Laboratory Environment

Hiroki Yu,^{a,b} Makoto Taniguchi,^c Kazuma Uesaka,^d Apirak Wiseschart,^{e,f} Kusol Pootanakit,^e Yudai Nishitani,^{a,b*} Yota Murakami, a,b,g Koichiro Ishimori, a,b,g (Kentaro Miyazaki, f,h Kei Kitaharab,f,g

^aGraduate School of Chemical Sciences and Engineering, Hokkaido University, Sapporo, Hokkaido, Japan

ABSTRACT Staphylococcus arlettae is one coagulase-negative species in the bacterial genus Staphylococcus. Here, we describe the closed complete genome sequence of S. arlettae strain P2, which was obtained using a hybrid approach combining Oxford Nanopore long-read and Illumina MiSeq short-read sequencing data.

taphylococcus arlettae is a species of the genus Staphylococcus and was reportedly ifirst isolated from skin or nares of poultry or goats (1). S. arlettae has since been additionally isolated from veterinary (2-6) and clinical (7-10) samples. Other S. arlettae isolation sites include soil (11), cell phone surfaces (12), and a disused biological safety cabinet (13).

The S. arlettae strain selected for whole-genome sequencing reported here was isolated from a biological laboratory in a university in Sapporo, Japan. Microbes on the laboratory floor surface were swabbed using a sterilized moistened swab. The swab was used to streak an LB plate, and one colony, which appeared on the plate after incubation overnight at 37°C, was purified by colony streaking onto a fresh LB plate. The procedure was performed thrice. Analysis of the 16S rRNA gene of the resulting isolate (named P2) revealed that it shared 99.9% identity with the 16S rRNA sequence of S. arlettae strain CVD059 (9). Although draft genome sequences of S. arlettae strains, including CVD059, have already been reported by some groups (2, 14), none have been shown as closed sequences, motivating us to determine the first closed genome of an S. arlettae strain.

Before DNA extraction, strain P2 was inoculated in LB broth, and cells were cultured at 37°C until early stationary phase (the doubling time of P2 was approximately 38 min). High-molecular-weight genomic DNA was prepared from a harvested bacterial pellet using the MagAttract high-molecular-weight (HMW) DNA kit (Qiagen) according to the manufacturer's instructions. The obtained genomic DNA was subjected to long-read and short-read sequencing at the Oral Microbiome Center at Taniguchi Dental Clinic in Japan. Default parameters were used for all software, unless otherwise specified.

Long-read sequencing was performed using a GridION X5 system (Oxford Nanopore Technologies [ONT]); 1.0- μ g unfragmented genomic DNA was used for library construction using a ligation sequencing kit (ONT). The prepared library was applied to a FLO-MIN106 R9.41 flow cell (ONT). The long-read sequences, which were base called using Guppy v.3.0.3 (ONT), generated 59,502 reads (756 Mb) with an average length of Citation Yu H, Taniguchi M, Uesaka K, Wiseschart A, Pootanakit K, Nishitani Y, Murakami Y, Ishimori K, Miyazaki K, Kitahara K. 2019. Complete genome sequence of Staphylococcus arlettae strain P2, isolated from a laboratory environment. Microbiol Resour Announc 8:e00696-19. https://doi.org/10.1128/ MRA.00696-19.

Editor Steven R. Gill, University of Rochester School of Medicine and Dentistry

Copyright © 2019 Yu et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kentaro Miyazaki, miyazaki-kentaro@aist.go.jp, or Kei Kitahara, keikitahara@sci.hokudai.ac.jp.

* Present address: Yudai Nishitani, University of Hyogo, Akashi, Hyogo, Japan.

Received 10 June 2019 Accepted 30 September 2019 Published 7 November 2019

bAmbitious Leader's Program Fostering Future Leaders to Open New Frontiers in Materials Science (ALP), Hokkaido University, Sapporo, Hokkaido, Japan

^cOral Microbiome Center, Taniguchi Dental Clinic, Takamatsu, Kagawa, Japan

^dGraduate School of Bioagricultural Sciences, Nagoya University, Nagoya, Aichi, Japan

eInstitute of Molecular Biosciences, Mahidol University, Salaya, Nakhon Pathom, Thailand

Department of Life Science and Biotechnology, Bioproduction Research Institute, National Institute of Advanced Industrial Science and Technology, Tsukuba, Ibaraki, Japan

⁹Department of Chemistry, Faculty of Science, Hokkaido University, Sapporo, Hokkaido, Japan

Department of Computational Biology and Medical Sciences, Graduate School of Frontier Sciences, The University of Tokyo, Kashiwa, Chiba, Japan

Yu et al.

♣ Microbiologia

12,711 bp during a 10-h runtime (numbers are those for reads after quality filtering, with an average Phred quality value of >8.0 using NanoFilt v.2.3.0 [15]; the raw data contained 104,000 reads, with an average length of 8,957 bp).

For short-read sequencing, the paired-end (2×156 -bp) Nextera DNA library (prepared using Nextera DNA Flex library prep kit [Illumina]) was sequenced on a MiSeq instrument. Raw sequencing data were processed using the FASTQ preprocessing program fastp v.0.19.5 (16) for the purpose of trimming adapters and low-quality data, yielding 1.05 million short reads with an average length of 152.8 bp.

For complete *de novo* genome assembly, both long-read and short-read data were processed using Unicycler v.0.4.4 (17), followed by a final polishing step using Pilon v.1.23 (18), generating a single circular contig for the chromosome with a length of 2,629,900 bp (G+C content of 33.7%) and another circular contig for a plasmid with a length of 22,364 bp (G+C content of 30.1%). To confirm that both circular contigs have no structural misassembly, we used the software program SV-Quest (K. Uesaka, unpublished data), which maps the short-read sequences back to the two contigs, detecting no signals for structural gaps and other inconsistencies. Automatic annotation was then performed using the annotation pipeline DFAST v.1.1.0 (19), provided by DDBJ, which predicted 2,550 coding sequences as well as 22 rRNA genes and 60 tRNA genes. Compared with CVD059, P2 had a chromosome that was 45 kbp shorter, which showed a symmetrical identity of 93.8% and gapped identity of 99.2%. CVD059 is reported to have 2,439 coding sequences (9). There has been no report that CVD059 contains a plasmid.

To our knowledge, this represents the first closed genome sequence report for an *S. arlettae* strain registered to a public database, providing an essential basis for detailed comparative analysis of *S. arlettae* genomes in the future.

Data availability. The closed complete chromosomal and plasmid sequences were deposited at DDBJ/EMBL/GenBank under accession numbers AP019698 and AP019699, respectively. The versions described in the manuscript are the first versions, AP019698.1 and AP019699.1, respectively. Raw sequencing data were deposited in the DDBJ SRA database under the accession numbers DRX167894 and DRX167895.

ACKNOWLEDGMENTS

This work was supported, in part, by the Japan Society for the Promotion of Science (JSPS) Grant-in-Aid for Challenging Research (Pioneering) number 17H06254 (to K.K.) and JSPS Grant-in-Aid for Young Scientists (B) number 25830132 (to K.K.). A.W. is a recipient of a Thailand Research Fund (PHD/0029/2557) through the Royal Golden Jubilee Ph.D. Program. The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

REFERENCES

- Schleifer KH, Kilpper-Bälz R, Devriese LA. 1984. Staphylococcus arlettae sp. nov., S. equorum sp. nov. and S. kloosii sp. nov.: three new coagulasenegative, novobiocin-resistant species from animals. Syst Appl Microbiol 5:501–509. https://doi.org/10.1016/S0723-2020(84)80007-7.
- Adkins PRF, Dufour S, Spain JN, Calcutt MJ, Reilly TJ, Stewart GC, Middleton JR. 2018. Cross-sectional study to identify staphylococcal species isolated from teat and inguinal skin of different-aged dairy heifers. J Dairy Sci 101: 3213–3225. https://doi.org/10.3168/jds.2017-13974.
- Liu B-H, Lei C-W, Zhang A-Y, Pan Y, Kong L-H, Xiang R, Wang Y-X, Yang Y-X, Wang H-N. 2017. Colocation of the multiresistance gene cfr and the fosfomycin resistance gene fosd on a novel plasmid in *Staphylococcus arlettae* from a chicken farm. Antimicrob Agents Chemother 61:e01388-17. https:// doi.org/10.1128/AAC.01388-17.
- Nobrega DB, Naushad S, Naqvi SA, Condas LAZ, Saini V, Kastelic JP, Luby C, De Buck J, Barkema HW. 2018. Prevalence and genetic basis of antimicrobial resistance in non-aureus staphylococci isolated from canadian dairy herds. Front Microbiol 9:256. https://doi.org/10.3389/fmicb .2018.00256.
- Sanz S, Olarte C, Alonso CA, Hidalgo-Sanz R, Gomez P, Ruiz-Ripa L, Torres C. 2018. Identification of enterococci, staphylococci, and *Enterobacteri*-

- aceae from slurries and air in and around two pork farms. J Food Prot 81:1776–1782. https://doi.org/10.4315/0362-028X.JFP-18-098.
- Suwannarach N, Kaewyana C, Yodmeeklin A, Kumla J, Matsui K, Lumyong S. 2017. Evaluation of *Muscodor cinnamomi* as an egg biofumigant for the reduction of microorganisms on eggshell surfaces and its effect on egg quality. Int J Food Microbiol 244:52–61. https://doi.org/ 10.1016/j.ijfoodmicro.2016.12.021.
- Dziri R, Klibi N, Lozano C, Ben Said L, Bellaaj R, Tenorio C, Boudabous A, Ben Slama K, Torres C. 2016. High prevalence of *Staphylococcus haemolyticus* and *Staphylococcus saprophyticus* in environmental samples of a Tunisian hospital. Diagn Microbiol Infect Dis 85:136–140. https://doi.org/ 10.1016/j.diagmicrobio.2016.03.006.
- Andreis SN, Perreten V, Schwendener S. 2017. Novel beta-lactamase bla_{ARL} in Staphylococcus arlettae. mSphere 2:e00117-17. https://doi.org/ 10.1128/mSphere.00117-17.
- Dinakaran V, Shankar M, Jayashree S, Rathinavel A, Gunasekaran P, Rajendhran J. 2012. Genome sequence of *Staphylococcus arlettae* strain CVD059, isolated from the blood of a cardiovascular disease patient. J Bacteriol 194:6615–6616. https://doi.org/10.1128/JB.01732-12.
- 10. Teeraputon S, Santanirand P, Wongchai T, Songjang W, Lapsomthob N,

Volume 8 Issue 45 e00696-19 mra.asm.org **2**



- Jaikrasun D, Toonkaew S, Tophon P. 2017. Prevalence of methicillin resistance and macrolide-lincosamide-streptogramin B resistance in *Staphylococcus haemolyticus* among clinical strains at a tertiary-care hospital in Thailand. New Microbes New Infect 19:28–33. https://doi.org/10.1016/j.nmni.2017.05.007.
- Nanjani SG, Soni HP. 2014. Characterization of an extremely halotolerant Staphylococcus arlettae HPSSN35C isolated from Dwarka Beach, India. J Basic Microbiol 54:843–850. https://doi.org/10.1002/jobm .201200690.
- Kurli R, Chaudhari D, Pansare AN, Khairnar M, Shouche YS, Rahi P. 2018. Cultivable microbial diversity associated with cellular phones. Front Microbiol 9:1229. https://doi.org/10.3389/fmicb.2018.01229.
- Lavecchia A, Chiara M, Manzari C, Trotta M, Marzano M, Horner D, Pesole G, Placido A. 2018. Draft genome sequences of three novel Staphylococcus arlettae strains isolated from a disused biological safety cabinet. Microbiol Resour Announc 7:e01012-18. https://doi.org/10.1128/MRA .01012-18.
- Naushad S, Barkema HW, Luby C, Condas LA, Nobrega DB, Carson DA, De Buck J. 2016. Comprehensive phylogenetic analysis of bovine non-

- aureus staphylococci species based on whole-genome sequencing. Front Microbiol 7:1990. https://doi.org/10.3389/fmicb.2016.01990.
- De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. Bioinformatics 34:2666–2669. https://doi.org/10.1093/bioinformatics/bty149.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10 .1093/bioinformatics/bty560.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal .pone.0112963.
- Tanizawa Y, Fujisawa T, Kaminuma E, Nakamura Y, Arita M. 2016. DFAST and DAGA: Web-based integrated genome annotation tools and resources. Biosci Microbiota Food Health 35:173–184. https://doi.org/10 .12938/bmfh.16-003.

Volume 8 Issue 45 e00696-19 mra.asm.org **3**