



Complete Genome Sequence of *Pseudomonas* sp. Strain KUIN-1, a Model Strain for Studies on the Production of Cell-Free Ice Nucleation Proteins

Taisei Yamamoto,^a Yoshie Hasegawa,^a Hidehisa Kawahara,^a  Hiroaki Iwaki^a

^aDepartment of Life Science and Biotechnology, Kansai University, Suita, Osaka, Japan

ABSTRACT *Pseudomonas* sp. (formerly *Pseudomonas fluorescens*) strain KUIN-1 is an ice-nucleating bacterium that was isolated from the leaves of field beans (*Phaseolus vulgaris* L.). This microorganism can release cell-free ice nucleation proteins and shows cold shock-induced freezing tolerance. Here, we report the 6,028,589-bp complete genome sequence of *Pseudomonas* sp. KUIN-1.

Pseudomonas sp. (formerly *Pseudomonas fluorescens*) strain KUIN-1 is an ice-nucleating bacterium that was isolated from the leaves of field beans (*Phaseolus vulgaris* L.), grown on Ellerslie Farm at the University of Alberta, Canada, by Obata et al. (1). Bacterial ice nuclei (or ice nucleation proteins) are outer membrane-associated lipoglycoproteins that are widely used in artificial snow production and also have the potential to be utilized as cloud-seeding agents and in the processed and frozen food industries (2). However, their commercial applications have been hampered because they are normally produced by plant-pathogenic species, such as those from the genera *Pseudomonas* and *Pantoea* (synonym *Erwinia*) (2). *Pseudomonas* sp. strain KUIN-1 releases cell-free ice nucleation proteins (3), and the cell-free character of the proteins makes it unnecessary to consider its plant-pathogenic character when utilizing them. Additionally, strain KUIN-1 shows cold shock-induced freezing tolerance (4). Here, we report the complete genome sequence of *Pseudomonas* sp. KUIN-1 in order to facilitate its further characterization and taxonomic identification.

Pseudomonas sp. strain KUIN-1 was provided to us by Hitoshi Obata (Kansai University, Osaka, Japan), and the genomic DNA was isolated from cells after culture for 18 h in 50 ml of Miller's LB medium (Merck Millipore) at 30°C, using Wilson's procedure with some modifications (5). Cells were then washed twice with Tris-EDTA buffer and then resuspended in 15 ml of the same buffer supplemented with 1 mg/ml lysozyme; the amounts of the subsequent reagents were scaled up in relation to the volume of the cell suspension. A 20-kb SMRTbell template library was prepared from approximately 8 μ g of input genomic DNA, using the SMRTbell template prep kit 1.0 (Pacific Biosciences). The SMRTbell library was sequenced using single-molecule real-time (SMRT) cell 8Pac version 3 and P6-C4 chemistry, and 240-min movies were captured for each SMRT cell using the PacBio RS II instrument (Pacific Biosciences). The default parameters were used for all software, unless otherwise noted. Subreads were filtered using PreAssembler Filter version 1 (minimum subread and polymerase read lengths, 500 and 100 bp, respectively; minimum polymerase read quality, 0.80) in SMRT Analysis version 2.3.0 (Pacific Biosciences), and 59,461 reads, composed of 609,693,648 bp, with an N_{50} value of 16,730 bp, were obtained. The reads were *de novo* assembled with the Hierarchical Genome Assembly Process (HGAP) protocol version 3 in SMRT Analysis to produce one circular contig with 85 \times coverage depth (6). The complete genome sequence of strain KUIN-1 consisted of a 6,028,589-bp circular chromosome that had a G+C content of 59.23%, and no plasmid was found.

Citation Yamamoto T, Hasegawa Y, Kawahara H, Iwaki H. 2019. Complete genome sequence of *Pseudomonas* sp. strain KUIN-1, a model strain for studies on the production of cell-free ice nucleation proteins. Microbiol Resour Announc 8:e01204-19. <https://doi.org/10.1128/MRA.01204-19>.

Editor Frank J. Stewart, Georgia Institute of Technology

Copyright © 2019 Yamamoto et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Hiroaki Iwaki, iwaki@kansai-u.ac.jp.

Received 8 October 2019

Accepted 17 October 2019

Published 7 November 2019

TABLE 1 16S rRNA gene sequence identity and ANI values for *Pseudomonas* sp. strain KUIN-1 compared to closely related species

<i>Pseudomonas</i> species	16S rRNA gene % identity (GenBank accession no.)	dDDH ^a (%)	ANI ^b (%)	Genome sequence GenBank accession no.
<i>P. congelans</i>	99.87 (NR_028985)	55.8	93.61	FNJH00000000
<i>P. cerasi</i>	99.86 (NR_146827)	57.4	93.74	LT222319
<i>P. syringae</i>	99.73 (NR_043716)	60.6	94.65	JALK00000000
<i>P. savastanoi</i>	99.72 (NR_117822)	37.1	88.26	LJRJ00000000
<i>P. cannabina</i>	99.60 (NR_025550)	30.5	84.84	FNKU00000000
<i>P. fluorescens</i>	97.96 (NR_115715)	22.6	76.33	LT907842

^a dDDH, digital DNA-DNA hybridization.

^b ANI, average nucleotide identity.

Strain KUIN-1 was reclassified using the 16S rRNA gene sequence, digital DNA-DNA hybridization (dDDH), and average nucleotide identity (ANI) (Table 1). The 16S rRNA gene sequence identities were calculated using the BLASTN program (<https://www.ncbi.nlm.nih.gov/BLAST/>) from the National Center for Biotechnology Information against the 16S ribosomal RNA sequences (*Bacteria* and *Archaea*) database. The ANI values were calculated with JSpeciesWS (<http://jspecies.ribohost.com/jspeciesws/>) using BLAST (7, 8). The dDDH values were calculated with the Genome-to-Genome Distance Calculator 2.1, available on the DSMZ website (<https://ggdc.dsmz.de>), using formula 2 (9, 10). The 16S rRNA gene sequence analysis revealed that strain KUIN-1 is affiliated with the *Pseudomonas syringae* group (11) rather than with *Pseudomonas fluorescens* (Table 1). The ANI and dDDH values were determined by using the five species that were most closely related to strain KUIN-1 based on the 16S rRNA gene sequence analysis. The ANI and dDDH values showed that strain KUIN-1 was most closely related to *P. syringae* (Table 1), and that these values were slightly below the accepted threshold for prokaryotic species boundaries, which are 95 to 96% for ANI and 70% for dDDH. This indicates that although strain KUIN-1 is closely related to *P. syringae*, the strain should be classified as a novel species in the genus *Pseudomonas*. Therefore, we tentatively identified strain KUIN-1 as a *Pseudomonas* sp.

The genome sequence was annotated with the DNA Data Bank of Japan (DDBJ) Fast Annotation and Submission Tool (DFAST; <https://dfast.nig.ac.jp>) (12). In total, 5,230 protein-coding sequences, 63 tRNA genes, and 16 rRNA genes were detected. As expected, because this species produces ice nuclei (3), one gene encoding the ice nucleation protein (1,352 amino acids) was predicted in the genome of strain KUIN-1. The genome also contains five CspA cold shock protein-like genes (13), which are considered to function as RNA chaperones and to prevent the formation of the secondary structures of mRNAs that enable efficient translation of mRNAs at low temperatures (14). The complete genome sequence will help clarify the mechanisms involved in the release of the ice nucleation proteins and the cold shock-induced freezing tolerance of strain KUIN-1.

Data availability. The genome sequence reported here was deposited in DDBJ under accession number AP020337. The associated BioProject, BioSample, and DDBJ Sequence Read Archive (DRA) accession numbers are PRJDB8624, SAMD00182269, and DRA009026, respectively.

ACKNOWLEDGMENT

This work was supported by a grant-in-aid from Kansai University for progress of research in a graduate course, 2018.

REFERENCES

- Obata H, Saeki Y, Tanishita J, Tokuyama T, Hori H, Higashi Y. 1987. Identification of an ice-nucleating bacterium KUIN-1 as *Pseudomonas fluorescens* and its ice nucleation properties. *Agric Biol Chem* 51: 1761–1766. <https://doi.org/10.1080/00021369.1987.10868291>.
- Margaritis A, Bassi AS. 1991. Principles and biotechnological applications of bacterial ice nucleation. *Crit Rev Biotechnol* 11:277–295. <https://doi.org/10.3109/07388559109069185>.
- Obata H, Tanaka T, Kawahara H, Tokuyama T. 1993. Properties of cell-free ice nuclei from ice nucleation-active *Pseudomonas fluorescens* KUIN-1. *J Ferment Bioeng* 76:19–24. [https://doi.org/10.1016/0922-338X\(93\)90046-B](https://doi.org/10.1016/0922-338X(93)90046-B).
- Obata H, Ishigaki H, Kawahara H, Yamade K. 1998. Purification and characterization of a novel cold-regulated protein from an ice-nucleating bacterium, *Pseudomonas fluorescens* KUIN-1. *Biosci Biotechnol Biochem* 62:2091–2097. <https://doi.org/10.1271/bbb.62.2091>.

5. Chachaty E, Saulnier P. 2000. Isolating chromosomal DNA from bacteria, p 29–32. In Rapley R (ed), *The nucleic acid protocols handbook*, vol 1. Humana Press, Totowa, NJ.
6. Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
7. Richter M, Rosselló-Móra R, Glöckner FO, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
8. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.
9. Auch AF, von Jan M, Klenk H-P, Göker M. 2010. Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison. *Stand Genomic Sci* 2:117–134. <https://doi.org/10.4056/sigs.531120>.
10. Meier-Kolthoff JP, Auch AF, Klenk H-P, Göker M. 2013. Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinformatics* 14:60. <https://doi.org/10.1186/1471-2105-14-60>.
11. Gomila M, Peña A, Mulet M, Lalucat J, García-Valdés E. 2015. Phylogenomics and systematics in *Pseudomonas*. *Front Microbiol* 6:214. <https://doi.org/10.3389/fmicb.2015.00214>.
12. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. *Bioinformatics* 34:1037–1039. <https://doi.org/10.1093/bioinformatics/btx713>.
13. Yamanaka K, Fang L, Inouye M. 1998. The CspA family in *Escherichia coli*: multiple gene duplication for stress adaptation. *Mol Microbiol* 27:247–255. <https://doi.org/10.1046/j.1365-2958.1998.00683.x>.
14. Jiang W, Hou Y, Inouye M. 1997. CspA, the major cold-shock protein of *Escherichia coli*, is an RNA chaperone. *J Biol Chem* 272:196–202. <https://doi.org/10.1074/jbc.272.1.196>.