

Figure 1 Sucrase-isomaltase (SI) 15Phe-driven IBS risk effects are stronger in low-starch consumers. The prevalence of IBS (%) across quartiles of starch intake (g/day) is reported, together with respective counts and number of individuals in each quartile group (Q1–Q4).

correlate with more frequent stools. These observations were not assessed in relation to key dietary factors including carbohydrate (ie, SI substrates) consumption.¹

drate (ie, SI substrates) consumption.¹ 1 Here, we studied two large German si population-based cross-sectional cohorts, (i namely PopGen (n=639; average age 61.4; 44.8% female) and FoCus (n=759; p average age 53.0; 58.5% female), with a available genotype (genome-wide arrays), c dietary (12-month food frequency questionnaire, FFQ), faecal microbiota (16S a sequencing) and IBS status (self-reported v from questionnaire) data, as previously In described in detail.^{2–4}

In a combined age/sex/body mass index (BMI)-adjusted logistic regression analysis of the two data sets, carriers of the 15Phe variant (52.86%) reported IBS significantly more often than non-carriers (3.69% vs 1.84%, respectively; P=0.044, OR=2.04), thus replicating and extending previous findings.¹ When taking into account the consumption of SI substrate carbohydrates (polysaccharides and disaccharides; g/day) estimated from FFQ, this association appeared strongest for individuals with lowest intake (not shown). In particular, as illustrated in figure 1, starch was the individual carbohydrate



Figure 2 15Phe genotype influences *Blautia* faecal abundance. (Left) Genotype-stratified correlation between starch intake and *Blautia* faecal microbiota abundance (each circle represents an individual). A trend was identified when comparing the two sucrase-isomaltase (*SI*) 15Phe genotype groups for their starch-bacteria correlations (age/sex/body mass index (BMI)/total energy (TE)-adjusted generalised linear model (GLM) with negative-binomial distribution, and interaction term for genotype and starch intake), in that increasing starch intake corresponds to higher *Blautia* abundance in 15Phe carriers compared with non-carriers (uncorrected P=0.054). (Right) *Blautia* faecal microbiota abundance in the two *SI* genotype groups stratified according to IBS status was significantly increased in IBS cases carrying the 15Phe variant (P=0.00041, beta=0.80), while there was no significant association in non-carriers (P=0.31, beta=0.33). Association analysis was performed using GLM age/sex/BMI/TE adjusted (glm.nb in stats/R). Plots were made using ggplot in ggplot2/R with stat_smooth and method=Im (left panel), and square root transformation of *Blautia* relative abundance (right panel).

Sucrase-isomaltase 15Phe IBS risk variant in relation to dietary carbohydrates and faecal microbiota composition

Recently in *Gut*, a coding sucrase-isomaltase (*SI*) variant (15Phe at single nucleotide polymorphism rs9290264) with 35% reduced disaccharidase activity was reported to increase IBS risk and to component where the largest difference in IBS prevalence was observed between 15Phe carriers and non-carriers (7.8% vs 1.9%, respectively; P=0.029, OR=4.17). This suggests that 15Phe-driven genetic IBS risk effects may be better detectable in low-carbohydrate consumers (possibly driven by starch intake), where relative differences in SI enzymatic activity might have more pronounced consequences on the presence of symptom-generating undigested carbohydrates in the large bowel (compared with other intake groups, where colonic accumulation of undigested carbohydrates may result from higher intake irrespective of genotype).

We then studied PopGen and FoCus faecal microbiota profiles in relation to carbohydrate consumption and SI 15Phe genotype. Expectedly, intake of polysaccharides (P=0.008), disaccharides (P=0.008), their sum (P=0.01) and starch (P=0.007) correlated with microbiota composition in an age/sex/BMI/ total energy (TE)-adjusted multivariate analysis of variance model (mvabund/R using default settings, after excluding rare taxa with >95% zeros).⁵ Of note, similar effects were also observed when comparing 15Phe carriers with non-carriers (mvabund/R as above with genotype as covariate, P=0.016) irrespective of carbohydrate intake, thus suggesting SI genotype may be relevant to faecal microbiota composition. In order to gain further insight into the SI genotype-carbohydrate-microbiota interaction, we focused on 26 genera known to use intestinally available polysaccharides and disaccharides for their growth, namely 'carb-digesters' as defined and characterised previously by others.⁶ Although multiple testing correction returned no significant results, nominal trends for genotype-dependent starch-microbiota correlations were observed for Blautia, Oscillibacter, Ruminococcus and unclassified Enterobacteriaceae (typifying results for Blautia shown in figure 2). This is noteworthy, since similar changes in the relative abundance of most of these genera have been previously detected in patients with IBS.7-9 Of note, while we observed increased Blautia abundance in faecal samples from IBS cases also in our data set (generalised linear model age/sex/BMI/TE adjusted, P=0.00035, beta=0.66vs controls), this was strongly affected by SI genotype and only significant in 15Phe carriers (P=0.00041, beta=0.80 vs P = 0.31. beta=0.33 for non-carriers) (figure 2).

In conclusion, we report here preliminary evidence linking the IBS-associated *SI* 15Phe variant to detectable diet-mediated effects on faecal microbiota composition and IBS risk. This adds to previous findings, and warrants further studies of the complex *SI* genotype-dietary carbohydrate-microbiota interactions in order to infer causality in relation to overall risk of IBS.

Louise Thingholm, ¹ Malte Rühlemann, ¹ Jun Wang, ² Matthias Hübenthal, ¹ Wolfgang Lieb, ³ Matthias Laudes, ⁴ Andre Franke, ¹ Mauro D'Amato^{5,6,7}

¹Institute of Clinical Molecular Biology, Christian-Albrechts-University of Kiel, Kiel, Germany ²CAS Key Laboratory of Pathogenic Microbiology and Immunology, Institute of Microbiology, Chinese Academy of Sciences, Beijing, China ³Institute of Epidemiology and Biobank PopGen, Christian-Albrechts-University of Kiel, Kiel, Germany ⁴Department of Internal Medicine I, University Hospital S-H (UKSH, Campus Kiel), Kiel, Germany ⁵Gastrointestinal Genetics Unit, Biodonostia Health Research Institute, San Sebastian, Spain ⁶Ikerbasque, Basque Foundation for Science, Bilbao, Spain

⁷Clinical Epidemiology Unit and Center for Molecular Medicine, Department of Medicine Solna, Karolinska Institutet, Stockholm, Sweden

Correspondence to Professor Mauro D'Amato, Unit of Clinical Epidemiology, Department of Medicine Solna, Karolinska Institutet, T2, Karolinska University Hospital, SE-17176 Stockholm, Sweden; mauro. damato@ki.se

Acknowledgements We thank Ms Ilona Urbach, Ines Wulf and Tonio Hauptmann of the IKMB microbiome laboratory for excellent technical support.

Contributors LT and MDA: study concept and design. WL and ML: data acquisition. LT, MR, JW and MH: statistical analyses. LT and MDA: data analysis and interpretation. AF and MDA: obtained funding, administrative and technical support, and study supervision. MDA drafted the manuscript, with input and critical revision from all other authors. All authors approved the final draft of the manuscript.

Funding This study was supported by the German Ministry of Education and Research (BMBF) programme e:Med sysINFLAME (http://www.gesundheitsforschungbmbf.de/de/5111.php, no: 01ZX1306A), the Swedish Research Council (VR 2013-3862) to MDA, and received infrastructure support from the Deutsche Forschungsgemeinschaft (DFG) Cluster of Excellence 306 'Inflammation at Interfaces' (http://www. inflammation-at-interfaces.de, no: EXC306 and EXC306/2) and from the PopGen Biobank (Kiel, Germany).

Competing interests MDA has received unrestricted research grants from QOL Medical.

Ethics approval Institutional ethical review committee for the PopGen Biobank.

Provenance and peer review Not commissioned; internally peer reviewed.



Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http:// creativecommons.org/licenses/by-nc/4.0/

© Article author(s) (or their employer(s) unless otherwise stated in the text of the article) 2019. All rights reserved. No commercial use is permitted unless otherwise expressly granted.

AF and MD'A contributed equally.



To cite Thingholm L, Rühlemann M, Wang J, *et al. Gut* 2019;**68**:177–178.

Received 12 December 2017 Revised 28 December 2017 Accepted 29 December 2017 Published Online First 13 January 2018



▶ http://dx.doi.org/10.1136/gutjnl-2016-312456

Gut 2019;**68**:177–178. doi:10.1136/ gutjnl-2017-315841

REFERENCES

- Henström M, Diekmann L, Bonfiglio F, et al. Functional variants in the sucrase-isomaltase gene associate with increased risk of irritable bowel syndrome. Gut 2018;67:263–70.
- 2 Krawczak M, Nikolaus S, von Eberstein H, et al. PopGen: population-based recruitment of patients and controls for the analysis of complex genotype-phenotype relationships. Community Genet 2006;9:55–61.
- 3 Müller N, Schulte DM, Türk K, et al. IL-6 blockade by monoclonal antibodies inhibits apolipoprotein (a) expression and lipoprotein (a) synthesis in humans. J Lipid Res 2015;56:1034–42.
- 4 Wang J, Thingholm LB, Skiecevičienė J, et al. Genomewide association analysis identifies variation in vitamin D receptor and other host factors influencing the gut microbiota. *Nat Genet* 2016;48:1396–406.
- 5 Yi Wang UN, Wright S, Warton DI. mvabund: an R package for model-based analysis of multivariate data. *Methods Ecol Evol* 2012:471–4.
- 6 Vieira-Silva S, Falony G, Darzi Y, et al. Species-function relationships shape ecological properties of the human gut microbiome. Nat Microbiol 2016;1:16088.
- 7 Labus JS, Hollister EB, Jacobs J, *et al.* Differences in gut microbial composition correlate with regional brain volumes in irritable bowel syndrome. *Microbiome* 2017;5:49.
- 8 Rajilić-Stojanović M, Biagi E, Heilig HG, et al. Global and deep molecular analysis of microbiota signatures in fecal samples from patients with irritable bowel syndrome. Gastroenterology 2011;141:1792–801.
- 9 Rajilić-Stojanović M, Jonkers DM, Salonen A, et al. Intestinal microbiota and diet in IBS: causes, consequences, or epiphenomena? *Am J Gastroenterol* 2015;110:278–87.