




The anti-inflammatory potency of biologics targeting tumour necrosis factor- α , interleukin (IL)-17A, IL-12/23 and CD20 in hidradenitis suppurativa: an *ex vivo* study

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Summary

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Background Biologics targeting inflammatory mediators can achieve clinical improvements in hidradenitis suppurativa (HS). However, their clinical efficacy shows great interpatient variability in daily practice.

Objectives To investigate the anti-inflammatory potency of a selection of currently available biologics and prednisolone for the treatment of HS in an *ex vivo* skin culture system using lesional HS biopsies.

Methods Lesional skin samples from 10 patients with HS and skin samples from five healthy controls were cultured *ex vivo* and exposed to prednisolone or biologics targeting tumour necrosis factor (TNF)- α , interleukin (IL)-17A, IL-12/23p40 or CD20 (adalimumab, infliximab, secukinumab, ustekinumab and rituximab, respectively). Real-time quantitative polymerase chain reaction and cytokine bead arrays were used to measure the inhibitory effect of the biologics on cytokines and antimicrobial peptides (AMPs).

Results The relative mRNA expression of all tested cytokines and AMPs was significantly downregulated by all anti-inflammatory agents ($P < 0.001$). The protein production of the proinflammatory cytokines TNF- α , interferon γ , IL-1 β , IL-6 and IL-17A was significantly inhibited by adalimumab, infliximab, ustekinumab, prednisolone (all $P < 0.001$) and rituximab ($P = 0.0071$), but not by secukinumab ($P = 0.0663$). On both mRNA and protein levels, adalimumab, infliximab and prednisolone reduced the levels of a broader mix of individual cytokines than secukinumab, ustekinumab and rituximab. Moreover, a significant inhibitory effect on mRNA expression levels of inflammatory markers in healthy control skin was observed only for TNF- α inhibitors ($P < 0.001$) and prednisolone ($P = 0.0015$).

Conclusions This *ex vivo* study suggests that TNF- α inhibitors and prednisolone are the most powerful inhibitors of proinflammatory cytokines and AMPs in HS lesional skin, which concurs with our clinical experience in patients with HS.

What's already known about this topic?

- A key element of hidradenitis suppurativa (HS) is an aberrant immune response characterized by the overexpression of several proinflammatory cytokines and antimicrobial peptides in lesional skin.
- Biologics targeting inflammatory cytokines have the potential to improve HS disease activity.
- There is still need for efficacious drugs in the treatment of HS.

What does this study add?

- We sought to quantify the anti-inflammatory effects of currently available biologics in an *ex vivo* disease model.

- Adalimumab, infliximab, secukinumab, ustekinumab and rituximab in addition to prednisolone significantly inhibited a selected panel of proinflammatory cytokines and antimicrobial peptides in *ex vivo* HS lesional skin.
- Adalimumab, infliximab and prednisolone reduced the levels of a broader mix of individual cytokines than secukinumab, ustekinumab and rituximab.

What is the translational message?

- The significant inhibition of important proinflammatory cytokines by tumour necrosis factor- α inhibitors in HS correlates with their clinical efficacy.
- Our *ex vivo* skin culture system represents an adequate model for studies in search of novel candidate drugs for the treatment of HS and to personalize the treatment in specific patients.

Hidradenitis suppurativa (HS) is a chronic skin disease characterized by painful, deep-seated, inflamed nodules, abscesses and, in later stages, sinus tracts.¹ Occlusion of the hair follicle with subsequent rupture, followed by a fierce local inflammatory response, are considered primary pathogenic events. The aberrant innate immune response involves the upregulation of various proinflammatory cytokines and antimicrobial peptides (AMPs).²

Several studies have found elevated mRNA and protein levels of tumour necrosis factor (TNF)- α in lesional and perilesional HS skin.^{3,4} Targeting TNF- α with adalimumab and infliximab has been shown to be clinically efficacious in HS in randomized placebo-controlled trials.^{5–7} In addition, the upregulation of interleukin (IL)-1 β , IL-17A and IL-23 in HS lesions points to the importance of the IL-17 pathway in the pathophysiology of HS.^{8–10} Targeting IL-1 in HS yielded ambiguous clinical outcomes. In a randomized controlled trial anakinra proved to be efficacious in moderate-to-severe HS, yet cases of failure for patients on anakinra therapy have been reported.^{11–13}

Most recently, IL-17 has been targeted in two clinical trials in HS (NCT02421172, NCT03248531), but the results have not yet been released. Treatment with secukinumab or ustekinumab induced amelioration of HS in a few cases and case series.^{14–16} In addition, enhanced levels of AMPs in lesional HS skin have been described for S100A7 (psoriasin), S100A8, S100A9, human β -defensin (hBD)-2, hBD-3 and LL-37.^{17–19} Upregulation of these AMPs in HS lesional skin is mainly driven by IL-17.¹⁸

Chronic HS lesions are characterized by a marked increase in the number of CD20+, CD79A+ B cells and CD138+ plasma cells.²⁰ One case report described a clear improvement in HS after treatment with rituximab for concomitant idiopathic carpotarsal osteolysis syndrome.²¹ This highlights the contribution of these B cells and plasma cells to the inflammatory process in HS.

Although biologics targeting inflammatory mediators are now widely used for the treatment of HS, their clinical

efficacy shows great interpatient variability. Our *ex vivo* skin culture system is a fast and simple method to investigate and simultaneously compare the effect of biologics in fresh, human, lesional HS skin samples.²² It approaches the *in vivo* situation by maintaining the patients' skin architecture and allows close monitoring of the events following response to immunostimulators or suppressors in the same experiment.²³ Therefore, this study sought to investigate the anti-inflammatory potency of a selection of currently available biologics and prednisolone for the treatment of HS in an *ex vivo* skin culture system using lesional HS biopsies.

Materials and methods

Ethics and informed consent

HS lesional skin was collected from excised skin after HS surgery at the Department of Dermatology of the Erasmus University Medical Center in Rotterdam, the Netherlands, from October 2017 to February 2018. According to the opt-out principle used in the Erasmus University Medical Center no informed consent is required for the use of excised tissue for research purposes as this is considered waste material. Control skin samples were obtained from healthy individuals in the Sint Franciscus Hospital in Rotterdam, the Netherlands. All healthy volunteers provided written informed consent for the use of their excised skin in this study.

Patients with hidradenitis suppurativa

A total of 10 patients with HS (five men and five women) who had chronic, active disease (seven Hurley stage II, three Hurley stage III) requiring surgical excision under general or sedative anaesthesia were included. HS lesional skin samples were obtained from the inguinogenital area in five of these 10 patients, from the axillae in four patients and from the gluteal area in one patient. The mean age was 43.7 ± 7.4 years (SD), the mean body mass index

(BMI) was $30.3 \pm 6.5 \text{ kg m}^{-2}$, and seven of 10 patients were current smokers. Eight patients received a stable (≥ 28 days) dosage of systemic antibiotics for their HS at the time of surgery (four patients used azithromycin 500 mg three times per week; two patients moxifloxacin 400 mg once daily, rifampicin 600 mg once daily, and metronidazole 500 mg once daily; one patient clindamycin 300 mg twice daily, rifampicin 600 mg once daily and metronidazole 500 mg once daily; one patient doxycycline 100 mg once daily). None of the patients had been using immunosuppressive or immunomodulatory therapies including biologics for at least 2 months prior to surgery.

Healthy control skin

Healthy skin samples were obtained from the submammary and abdominal waste material of five women who underwent breast or abdominal reduction surgery. We argue that skin from these regions is suitable as a control sample because the inframammary and abdominal folds are predilection sites for HS. The controls (all female, mean age 41.0 ± 10.7 years, mean BMI $29.8 \pm 4.2 \text{ kg m}^{-2}$, no current smokers) were otherwise healthy, had no family history of HS, and were not using any immunosuppressive or immunomodulatory treatment at the time of surgery.

Biopsy procedure

A total of seven fresh 4-mm punch biopsies were taken per participant. HS skin biopsies were obtained from the same palpable actively inflamed lesion/infiltrate, and at least 1 cm away from the excision border. Abscesses were not biopsied in order to avoid sampling only the roof of the abscess. Each biopsy was weighed prior to culture as described in an earlier publication.⁴ HS biopsies had a mean weight of $2.4 \pm 6.6 \text{ mg}$, which was significantly heavier than biopsies from healthy skin with a mean weight of $16.1 \pm 3.0 \text{ mg}$ ($P < 0.001$). This potential confounder was addressed by normalizing all protein levels for the mg tissue weight.

Ex vivo skin culture

The 4-mm biopsies were immediately cultured in a transwell system (Netwell; Costar, Cambridge, MA, U.S.A.) as described previously.^{22,23} In brief, samples were placed in punched-out holes in the transwell membrane of a 12-well plate with the epidermis exposed to the air and the dermis immersed in 1 mL Iscove's modified Dulbecco's medium (Gibco, Paisley, U.K.) containing 0.5% human AB serum, penicillin (100 U mL^{-1}) and streptomycin (100 U mL^{-1}) with or without an anti-inflammatory agent. Biologics and prednisolone were separately added to the culture media resulting in the following seven conditions: (i) culture media as negative control; (ii) prednisolone $100 \mu\text{g mL}^{-1}$ as positive control; (iii) adalimumab $30 \mu\text{g mL}^{-1}$; (iv) infliximab

$20 \mu\text{g mL}^{-1}$; (v) secukinumab $30 \mu\text{g mL}^{-1}$; (vi) ustekinumab $10 \mu\text{g mL}^{-1}$ and (vii) rituximab $200 \mu\text{g mL}^{-1}$. The concentrations of the monoclonal antibodies were derived from the reported trough levels – at least twofold – in patients with plaque psoriasis (adalimumab,^{24,25} infliximab,²⁵ ustekinumab,²⁶ secukinumab),²⁷ and CD20+ B-cell malignancies (rituximab).²⁸ Our previous experiments using the *ex vivo* model showed that the cytokine concentrations measured in HS samples were usually in the picogram range.^{4,7} The biologic concentrations used in our experiments were at least 1000-fold higher and should suffice for neutralization purposes. Skin biopsies were incubated for 24 h at 37°C in an atmosphere of 5% CO_2 and 98% humidity. Subsequently, the biopsies were placed in 250 μL lysis buffer containing 1% β -mercaptoethanol. Both the supernatants and the biopsies were transferred to a polypropylene tube and stored at -20°C until further analysis.

Messenger RNA expression analysis

Total mRNA was extracted using the GenElute Mammalian Total RNA Miniprep Kit (Sigma-Aldrich, St. Louis, MA, U.S.A.). RNA was treated with $0.1 \text{ U } \mu\text{L}^{-1}$ DNase (Invitrogen, Carlsbad, CA, U.S.A.) and cDNA was subsequently synthesized using 1 μg total RNA template, with SuperScript II reverse transcriptase, random hexamer primers (Invitrogen) and oligo (dT)15 (Promega Co., Madison, WI, U.S.A.). Primers and probes were designed and chosen using ProbeFinder Software and the Universal Probe library (Roche Applied Science, Indianapolis, IN, U.S.A.). ABL1 was chosen as a reference house-keeping gene.²⁹ Real-time quantitative polymerase chain reaction (qPCR) was performed for the following 12 genes using the ViiA7 sequence-detection system (Applied Biosystems, Waltham, MA, U.S.A.): TNF- α , IL-1 β , IL-6, CXCL-8 (IL-8), IL-12p19, IL-17A, interferon (IFN) α /MxA, S100A7, S100A8, S100A9, LL37 and HBD-2. Messenger RNA expression was analysed with QuantStudio real-time qPCR software version 1.3 (Applied Biosystems).

Protein quantification

The following 18 inflammation-related cytokines were simultaneously measured in the supernatant using a customized bead-based multi-analyte profiling assay (Luminex, R&D systems, Minneapolis, MN, U.S.A.): CCL-20, TNF- α , IFN γ , IL-1 β , IL-1R1, IL-5, IL-6, IL-10, IL-12/23p40, IL-17A, IL-17E, IL-18, IL-19, IL-22, IL-27, IL-31, IL-33 and IL-36 β . Assays were used according to the manufacturers' protocol. A dilution factor of two was used for all supernatants.

Statistical analyses

The relative mRNA expression and protein production per sample compared with culture media (negative control) was calculated for every condition. Protein levels were normalized according to input weight and expressed in pg mL^{-1} per mg

Table 1 Relative mRNA expression and modulation by biologics in hidradenitis suppurativa (HS) lesional skin

	HS lesional skin (n = 10)																	
	Prednisolone			Adalimumab			Infliximab			Secukinumab ^a			Ustekinumab			Rituximab		
	Median (IQR)	Unadj. P-values	Unadj. P-values	Median (IQR)	Unadj. P-values	Unadj. P-values	Median (IQR)	Unadj. P-values	Unadj. P-values	Median (IQR)	Unadj. P-values	Unadj. P-values	Median (IQR)	Unadj. P-values	Unadj. P-values	Median (IQR)	Unadj. P-values	
IL-1β	0.27 (0.15–0.48)	< 0.0001 ^b	0.0073 ^b	0.53 (0.23–0.74)	0.0073 ^b	0.60 (0.50–0.73)	0.0306	0.67 (0.53–0.87)	0.2377	0.76 (0.39–1.06)	0.6194	0.49 (0.35–0.72)	0.0597					
IL-6	0.42 (0.36–0.51)	< 0.0001 ^b	0.0956	0.62 (0.52–0.89)	0.0956	0.65 (0.41–0.71)	0.0047 ^b	0.70 (0.44–0.90)	0.1063	0.55 (0.37–0.90)	0.0122 ^b	0.75 (0.55–0.97)	0.4396					
CXCL-8 (IL-8)	0.44 (0.34–0.60)	0.1675	0.0042 ^b	0.30 (0.22–0.49)	0.0042 ^b	0.29 (0.26–0.38)	0.0002 ^b	0.28 (0.16–0.47)	0.0015 ^b	0.39 (0.32–0.48)	0.0296	0.69 (0.41–0.86)	0.9779					
IL-17A	0.13 (0.07–0.37)	0.0049 ^b	1.0000	0.93 (0.59–1.37)	1.0000	0.55 (0.43–0.84)	1.0000	0.95 (0.62–1.42)	1.0000	0.96 (0.56–1.29)	1.0000	0.99 (0.43–1.61)	1.0000					
IL-23p19	0.92 (0.33–1.20)	1.0000	0.3934	0.48 (0.34–0.66)	0.3934	0.49 (0.31–0.88)	0.4901	0.48 (0.14–1.01)	0.3836	0.72 (0.46–1.79)	1.0000	0.97 (0.51–1.99)	1.0000					
TNF-α	1.09 (0.64–1.58)	1.0000	1.0000	0.92 (0.52–1.02)	1.0000	0.74 (0.37–1.03)	0.7250	0.53 (0.28–1.15)	0.5127	0.83 (0.53–2.56)	1.0000	0.71 (0.60–1.11)	1.0000					
IFN-MXA	0.75 (0.60–1.23)	1.0000	0.7914	0.56 (0.35–1.08)	0.7914	0.84 (0.59–1.19)	1.0000	0.67 (0.43–0.81)	0.3345	0.83 (0.55–1.17)	1.0000	0.91 (0.57–1.17)	1.0000					
S100A7	0.80 (0.51–1.02)	0.3934	0.0266	0.56 (0.50–0.70)	0.0266	0.56 (0.45–0.72)	0.0116 ^b	0.50 (0.34–0.88)	0.0105 ^b	0.64 (0.54–0.68)	0.0223	0.56 (0.46–0.74)	0.0180					
S100A8	0.74 (0.55–1.11)	1.0000	0.0060 ^b	0.50 (0.39–0.54)	0.0060 ^b	0.52 (0.44–0.60)	0.0248	0.67 (0.52–0.73)	0.2826	0.79 (0.59–1.12)	1.0000	0.69 (0.56–0.93)	0.7091					
S100A9	0.80 (0.51–1.07)	0.8442	0.0005 ^b	0.35 (0.30–0.53)	0.0005 ^b	0.29 (0.27–0.42)	< 0.0001 ^b	0.36 (0.25–0.68)	0.001 ^b	0.52 (0.42–0.78)	0.0296	0.66 (0.52–0.93)	0.9779					
IL-37	0.67 (0.14–1.28)	0.8997	0.5915	0.40 (0.37–0.83)	0.5915	0.64 (0.38–1.10)	1.0000	0.51 (0.32–1.84)	1.0000	0.77 (0.48–1.37)	1.0000	0.89 (0.67–1.22)	1.0000					
HBD-2	0.48 (0.19–0.87)	0.0416	0.0306	0.43 (0.29–0.65)	0.0306	0.32 (0.20–0.91)	0.0445	0.69 (0.13–1.07)	0.1763	0.49 (0.43–0.76)	0.1538	0.76 (0.68–1.13)	1.0000					
All (n = 12)	0.57 (0.34–1.03)	< 0.0001	< 0.0001	0.51 (0.33–0.92)	< 0.0001	0.53 (0.32–0.78)	< 0.0001	0.56 (0.29–0.99)	< 0.0001	0.64 (0.44–1.00)	< 0.0001	0.73 (0.50–1.19)	< 0.0001					
Cytokines ^c (n = 7)	0.41 (0.20–0.77)	< 0.0001	< 0.0001	0.52 (0.32–0.95)	< 0.0001	0.54 (0.34–0.75)	< 0.0001	0.60 (0.37–0.98)	< 0.0001	0.61 (0.39–1.08)	< 0.0002	0.76 (0.44–1.21)	0.4780					
AMPs ^d (n = 5)	0.75 (0.44–1.11)	< 0.0001	< 0.0001	0.48 (0.33–0.68)	< 0.0001	0.47 (0.29–0.73)	< 0.0001	0.52 (0.30–0.96)	< 0.0001	0.64 (0.45–0.89)	< 0.0001	0.73 (0.53–0.98)	0.0001					

AMPs, antimicrobial peptides; IQR, interquartile range; IL, interleukin; IFN, interferon; TNF, tumour necrosis factor; Unadj., unadjusted. P-values marked in bold indicate analyses that are significant at the 0.05 level. ^aOne HS sample for secukinumab was excluded for real-time quantitative polymerase chain reaction analysis as a result of a human error during the process of cDNA synthesis (n = 9). ^bSignificant after correction with the Benjamini–Hochberg test (P < 0.0156). ^cCytokines: pooled effect of IL-1β, IL-6, IL-8, IL-17A, IL-23p19, TNF-α and IFN-MXA. ^dAMPs: pooled effect of S100A7, S100A8, S100A9, IL-37 and HBD-2.

tissue weight. The lower limit of quantification was imputed when protein concentrations in the supernatant (pg mL^{-1}) were below the limit of detection. The Kruskal–Wallis test, i.e. a one-way ANOVA on ranks, was used to compare the variance in mRNA and protein levels per inflammatory marker. If one condition stochastically dominated another condition, the Dunnnett's post-test was subsequently used to test conditions vs. culture media (pairwise comparisons) separately. This approach was chosen in order to increase power. A two-sided *P*-value < 0.05 was considered significant. The level of significance for the relative mRNA expression and protein production was separately adjusted by the Benjamini–Hochberg procedure for multiple comparisons. A correlation between relative mRNA and protein levels per cytokine was calculated using the Spearman's rho test for non-normally distributed continuous variables. GraphPad Prism version 6 (GraphPad Software, La Jolla, CA, U.S.A.) was used for all statistical analyses.

Results

Sample flow for analysis

In total, samples from 10 patients with HS and five healthy controls were analysed by real-time qPCR. Supernatants from one patient with HS were excluded from multiplex assay because of erroneous sample processing, resulting in samples from nine patients with HS with protein data. For control skin, supernatants of three healthy volunteers were analysed by Luminex assay. However, protein levels were below the level of detection or in the very low range of the calibration line in these control samples (data not shown). Therefore, samples of the other two healthy volunteers were not analysed.

Significant downregulation of mRNA expression of cytokines and antimicrobial peptides by prednisolone and different biologics

The relative changes in mRNA expression of the 12 genes in the HS samples, including significance levels, are shown in Table 1. The overall median inhibitory effect on the mix of cytokines and AMPs per condition was as follows: prednisolone 0.57 [interquartile range (IQR) 0.34–1.03], adalimumab 0.51 (0.33–0.92), infliximab 0.53 (0.32–0.78), secukinumab 0.56 (0.29–0.99), ustekinumab 0.64 (0.44–1.00) and rituximab 0.73 (0.50–1.19). The inhibitory impact on AMPs was stronger for the biologics than for prednisolone (Table 1, Fig. 1). Prednisolone, adalimumab and infliximab significantly inhibited IL-1 β mRNA expression. Expression of CXCL-8 (IL-8) was significantly downregulated by adalimumab, infliximab, secukinumab and ustekinumab. Messenger RNA levels of two other important proinflammatory cytokines, TNF- α and IL-23p19, were not significantly downregulated by any of the biologics or prednisolone. Messenger RNA expression of members of the S100 family and HBD-2 was significantly reduced by adalimumab, infliximab, secukinumab and ustekinumab (Table 1). In healthy control skin, prednisolone, adalimumab and infliximab significantly downregulated mRNA expression of all tested cytokines and AMPs (Table S1, Fig. S1; see Supporting Information).

Significant *ex vivo* reduction in tumour necrosis factor- α , interferon γ , interleukin-1 β , interleukin-6, and interleukin-17A protein levels in hidradenitis suppurativa lesional skin

In total, 18 cytokines were measured in the culture media. Six cytokines were not detected in the majority of HS samples throughout all conditions, namely IL-1RI, IL-5, IL-12/23p40,

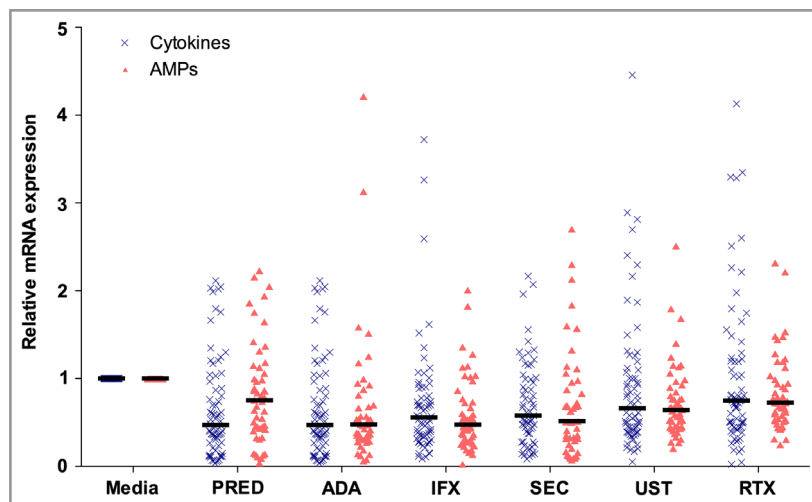


Fig 1. Anti-inflammatory impact on the relative mRNA expression of the mix of cytokines and antimicrobial peptides (AMPs) in hidradenitis suppurativa (HS) lesional skin ($n = 10$). Horizontal bars display the median. Media, culture media; PRED, prednisolone $100 \mu\text{g mL}^{-1}$; ADA, adalimumab $30 \mu\text{g mL}^{-1}$; IFX, infliximab $20 \mu\text{g mL}^{-1}$; SEC, secukinumab $30 \mu\text{g mL}^{-1}$; UST, ustekinumab $10 \mu\text{g mL}^{-1}$; RTX, rituximab $200 \mu\text{g mL}^{-1}$.

IL-22, IL-31 and IL-33. Relative changes in protein production including significance levels for the other 12 cytokines are shown in Table 2. The overall median effect on cytokines per condition was as follows: prednisolone 0.51 (0.28–0.64); adalimumab 0.78 (0.54–0.91); infliximab 0.70 (0.49–0.94); secukinumab 0.85 (0.69–0.95); ustekinumab 0.71 (0.57–0.79) and rituximab 0.81 (0.70–0.94). Specifically the release of TNF- α , IFN γ , IL-1 β , IL-6 and IL-17A, was significantly inhibited by all tested drugs with the exception of secukinumab (Fig. 2). In addition, prednisolone significantly inhibited the release of TNF- α , IFN γ , IL-1 β , IL-6, IL-17A, IL-10 and IL-25/17E. As expected, adalimumab and infliximab almost completely neutralized TNF- α levels in the supernatants (Fig. 3). Of note, for the two important cytokines TNF- α and IL-17A we observed both a strong intrapatient and interpatient variability among all biologic conditions (Fig. 3).

Great variation in correlation between mRNA expression and protein production levels in hidradenitis suppurativa samples

The cytokines IL-1 β , IL-6, IL-17A and TNF- α , were measured by both real-time qPCR and Luminex. There was great variation in the correlation between mRNA and protein levels of these individual cytokines (Fig. 4). A high correlation between mRNA and protein levels was found for IL-17A ($r = 0.86$, $P < 0.001$), while the correlation for TNF- α was almost zero ($r = 0.04$, $P = 0.7480$).

Discussion

In this study, we show that the commercially available biologics, used in daily practice for the treatment of HS (adalimumab, infliximab, ustekinumab and rituximab), significantly inhibited mRNA and protein expression of various cytokines and AMPs in lesional HS skin, cultured *ex vivo* for 24 h. Secukinumab demonstrated significant downregulation of inflammatory markers in the lesional skin only at the mRNA level. The anti-inflammatory effect of prednisolone and all biologics, except secukinumab, was the strongest at the protein levels of TNF- α , IFN γ , IL-1 β , IL-6 and IL-17A, which are important cytokines in the pathogenesis of HS. Prednisolone and TNF- α inhibitors seemed to be the most effective in reducing the release of a broader range of proinflammatory cytokines and AMPs in lesional HS skin. Only prednisolone, adalimumab and infliximab had an inhibitory effect on the mRNA expression levels of inflammatory markers in healthy control skin (see supporting information).

In patients with moderate-to-severe HS, the strongest evidence has been documented for the anti-TNF agents adalimumab and infliximab.^{5,30–32} Our translational findings correspond with the observed efficacy of TNF- α inhibitors in HS in daily practice. In addition, we confirmed the previously reported decrease of *ex vivo* IL-1 β protein expression after adalimumab treatment on the mRNA level.⁷ As TNF- α is a multifunctional cytokine with numerous actions, a simultaneous

downregulation of other cytokines, such as IL-6 and cytokines of the IL-1 family, could also explain the results of the anti-TNF agents in our study.^{9,33}

Remarkably, secukinumab did not reduce the IL-17 protein levels in the same way that adalimumab and infliximab reduced TNF- α protein levels. Unfortunately, IL-12p40 protein (target of ustekinumab), an important indicator of the IL-17 pathway, fell below the level of detection in the Luminex assay. It could be that other cytokines, such as TNF- α and IL-1 β , not being blocked by secukinumab in this culture system, were still able to induce production of IL-17. Moreover, levels of IL-17A were in the lower range of detection in most patients. Another explanation may be that the anti-IL-17A antibodies used in the Luminex assay detect a different epitope of IL-17A than that recognized by secukinumab. The lower mRNA expression of S100A7, S100A9, IL-6 and CXCL-8 could be considered to be the result of blocking of IL-17A bioactivity by secukinumab. Similarly, at the mRNA level, ustekinumab exerted its action predominantly via IL-6, CXCL-8, S100A7 and S100A9, although the last three markers were nonsignificant after Benjamini–Hochberg correction.

The pan-cytokine inhibitory characteristics of prednisolone were demonstrated by inhibition of multiple cytokines on both the mRNA and protein level, which supports the efficacy of systemic and intralesional corticosteroids for acute HS flares in clinical practice.³⁴ However, prolonged high-dose systemic corticosteroids are not recommended as HS rapidly flares after tapering, especially after a long course.³⁵ Nonetheless, low-dose systemic prednisolone could be a valuable adjunct therapy for recalcitrant HS.³⁶

Rituximab was the only biologic without a significant inhibitory effect on individual inflammatory mRNA and protein levels. This is not surprising as B-cell blockade in inflammatory diseases acts via inhibition of antibody production, antigen presentation and indirectly via cytokine reduction.³⁷ Therefore, the presence of the complete immune system is required for B-cell blockade to be effective. Moreover, HS is considered primarily a disease of a deficient innate immunity. On the other hand, chronic HS lesions are full of B cells and plasma cells, indicating that adaptive humoral immunity is also activated in longstanding HS.^{20,38}

The impact of biologics on AMP expression has never been investigated in HS lesional skin. Our findings indicate a potential supporting role for AMPs in HS pathophysiology as it is known that AMPs are capable of activating keratinocytes and attracting innate immune cells to amplify the local immune response.³⁹ Cytokines produced by innate and adaptive immune cells, such as TNF- α , IL-17 and IL-12, drive AMP production in the keratinocytes.^{40–42} Moreover, AMPs can be activated by damage- and pathogen-associated molecular patterns after follicle rupture with the release of keratin fibres and skin commensals in the dermis.

Although it is assumed that levels of mRNA and protein have a one-to-one correlation, the absence of correlation between TNF- α mRNA and its protein levels has been

Table 2 Relative protein production and modulation by biologics in hidradenitis suppurativa (HS) lesional skin

	HS lesional skin (n = 9)											
	Prednisolone		Adalimumab		Infliximab		Secukinumab		Ustekinumab		Rituximab	
	Median (IQR)	Unadj. P-values	Median (IQR)	Unadj. P-values	Median (IQR)	Unadj. P-values	Median (IQR)	Unadj. P-values	Median (IQR)	Unadj. P-values	Median (IQR)	Unadj. P-values
TNF- α	0.20 (0.15–0.29)	0.0022^a	0.09 (0.07–0.20)	< 0.0001^a	0.03 (0.02–0.05)	< 0.0001^a	0.89 (0.53–0.93)	1.0000	0.54 (0.23–0.76)	0.3305	0.70 (0.65–1.10)	1.0000
IFN γ	0.37 (0.11–0.53)	0.0068^a	0.43 (0.30–1.17)	0.4169	0.46 (0.13–0.91)	0.0673	0.56 (0.51–0.66)	0.6285	0.39 (0.33–0.61)	0.0673	0.49 (0.41–0.60)	0.2596
CCL-20	0.50 (0.20–0.81)	0.2845	0.58 (0.28–1.80)	1.0000	0.71 (0.53–1.13)	1.0000	0.94 (0.53–1.19)	1.0000	0.73 (0.62–1.28)	1.0000	0.95 (0.70–1.38)	1.0000
IL-1 β	0.23 (0.17–0.45)	0.0049^a	0.41 (0.29–0.73)	0.3114	0.44 (0.31–0.75)	0.3305	1.08 (0.46–1.43)	1.0000	0.59 (0.45–2.06)	1.0000	0.70 (0.52–1.03)	1.0000
IL-6	0.59 (0.46–0.73)	0.0006^a	0.81 (0.68–0.85)	0.0724	0.76 (0.64–0.91)	0.1035	0.99 (0.76–1.08)	1.0000	0.76 (0.69–0.99)	0.1556	0.87 (0.71–0.92)	0.1035
IL-17A	0.09 (0.07–0.19)	< 0.0001^a	0.83 (0.68–1.38)	1.0000	0.50 (0.32–0.52)	0.2085	0.70 (0.60–1.60)	1.0000	0.54 (0.25–0.81)	0.3022	0.96 (0.34–1.24)	1.0000
IL-18	0.86 (0.59–1.03)	1.0000	0.75 (0.66–3.48)	1.0000	1.05 (0.74–1.11)	1.0000	0.90 (0.71–1.04)	1.0000	0.82 (0.78–0.90)	0.7341	0.93 (0.78–2.75)	1.0000
IL-36 β	0.94 (0.48–1.45)	1.0000	0.98 (0.81–2.04)	1.0000	1.04 (0.82–1.47)	1.0000	1.88 (0.94–2.34)	1.0000	1.05 (0.78–1.37)	1.0000	1.21 (0.85–1.65)	1.0000
IL-10 ^b	0.29 (0.20–0.41)	0.0062^a	1.12 (0.22–1.94)	1.0000	0.93 (0.37–1.05)	1.0000	0.81 (0.62–2.02)	1.0000	0.91 (0.55–1.42)	1.0000	0.62 (0.43–0.91)	0.6975
IL-25/17E ^b	0.52 (0.33–0.64)	0.0043^a	0.59 (0.48–0.95)	0.2517	0.50 (0.44–1.01)	0.0931	0.68 (0.55–0.88)	0.3208	0.67 (0.55–0.90)	0.2932	0.74 (0.52–0.90)	0.2440
IL-19 ^b	0.68 (0.64–0.99)	0.9862	0.91 (0.51–1.67)	1.0000	0.69 (0.56–1.31)	1.0000	0.79 (0.48–0.92)	1.0000	0.77 (0.65–1.12)	1.0000	0.92 (0.51–1.52)	1.0000
IL-27 ^b	0.63 (0.26–0.85)	0.0751	0.92 (0.72–1.01)	1.0000	0.95 (0.61–1.12)	1.0000	0.66 (0.64–1.02)	0.963	0.69 (0.62–0.83)	0.3609	0.72 (0.63–0.79)	0.2365
All ^c (n = 12)	0.51 (0.28–0.64)	< 0.0001	0.78 (0.54–0.91)	0.0006	0.70 (0.49–0.94)	< 0.0001	0.85 (0.69–0.95)	0.0684	0.71 (0.57–0.79)	0.0010	0.81 (0.70–0.94)	0.0121
Proinflammatory (n = 8)	0.45 (0.19–0.82)	NT	0.69 (0.29–1.18)	NT	0.64 (0.30–1.04)	NT	0.91 (0.53–1.43)	NT	0.71 (0.42–0.99)	NT	0.80 (0.58–1.22)	NT
Anti-inflammatory (n = 4)	0.54 (0.29–0.70)	NT	0.87 (0.51–1.42)	NT	0.70 (0.46–1.11)	NT	0.69 (0.54–1.13)	NT	0.75 (0.61–1.05)	NT	0.74 (0.50–0.98)	NT
Important ^d (n = 5)	0.23 (0.20–0.37)	< 0.0001	0.43 (0.41–0.81)	< 0.0001	0.46 (0.44–0.50)	< 0.0001	0.89 (0.70–0.99)	0.0663	0.54 (0.54–0.59)	< 0.0001	0.70 (0.70–0.87)	0.0071

AMPs, antimicrobial peptides; IQR, interquartile range; Unadj, unadjusted; NT, not tested; IL, interleukin; IFN, interferon; TNF, tumour necrosis factor. P-values marked in bold indicate analytes that are significant at the 0.05 level. ^aSignificant after correction with the Benjamini–Hochberg test ($P < 0.0156$). ^bCytokines with an anti-inflammatory function, note that IL-17E and IL-27 also have an immunoregulatory/proinflammatory function. ^cAll: pooled effect of 12 cytokines. ^dImportant: five key proinflammatory cytokines in the pathophysiology of HS; TNF- α , IFN γ , IL-1 β , IL-6 and IL-17A.

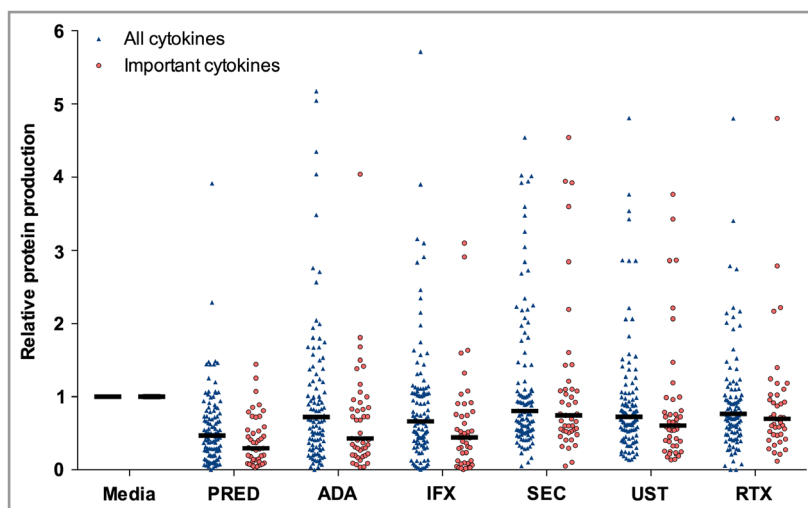


Fig 2. Anti-inflammatory impact on protein production of all cytokines and five important cytokines as measured by Luminex assay ($n = 9$). The important inflammatory cytokines are tumour necrosis factor- α , interferon γ , interleukin (IL)-1 β , IL-6, IL-17A. A total of 12 data points are outside the y-axis range. Media, culture media; PRED, prednisolone 100 $\mu\text{g mL}^{-1}$; ADA, adalimumab 30 $\mu\text{g mL}^{-1}$; IFX, infliximab 20 $\mu\text{g mL}^{-1}$; SEC, secukinumab 30 $\mu\text{g mL}^{-1}$; UST, ustekinumab 10 $\mu\text{g mL}^{-1}$; RTX, rituximab 200 $\mu\text{g mL}^{-1}$.

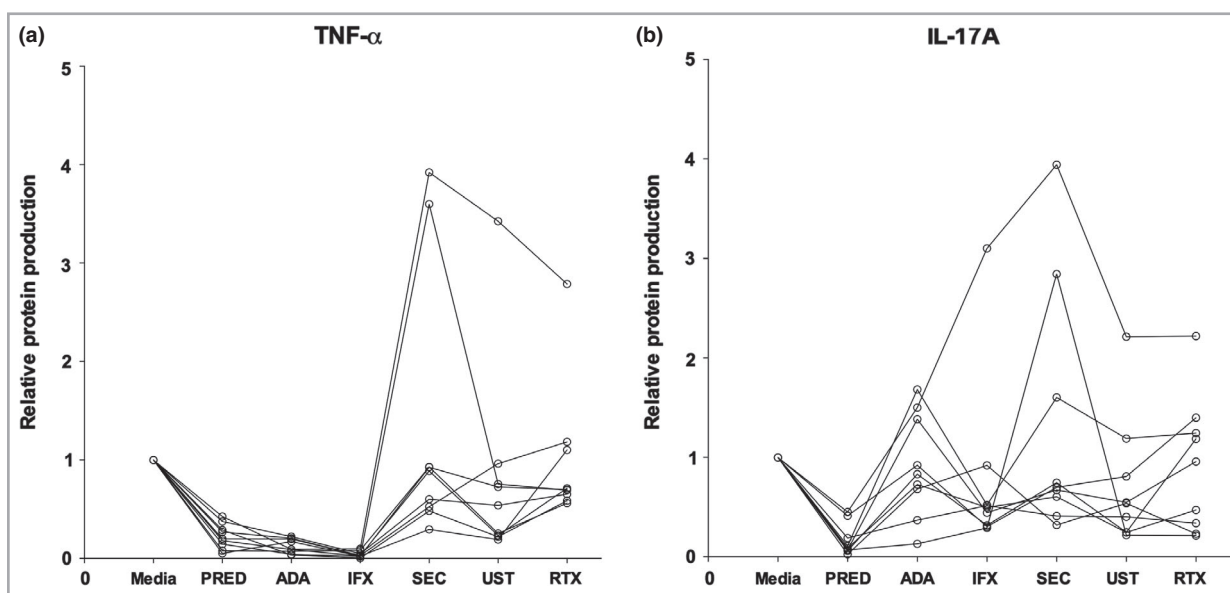


Fig 3. Interpatient and inpatient variability demonstrated by fluctuating protein levels of tumour necrosis factor (TNF)- α and interleukin (IL)-17A ($n = 9$). (a) TNF- α . Connected dots represent one patient. High inpatient variability as demonstrated by the fluctuating per-patient protein levels between conditions. Low interpatient variability was observed for conditions PRED, ADA and IFX. (b) IL-17A. Connected dots represent one patient. Very high (more than TNF- α) inpatient variability. Low interpatient variability for was observed for the condition PRED. PRED, prednisolone 100 $\mu\text{g mL}^{-1}$; ADA, adalimumab 30 $\mu\text{g mL}^{-1}$; IFX, infliximab 20 $\mu\text{g mL}^{-1}$. SEC, secukinumab 30 $\mu\text{g mL}^{-1}$; UST, ustekinumab 10 $\mu\text{g mL}^{-1}$; RTX, rituximab 200 $\mu\text{g mL}^{-1}$.

previously reported.^{43–45} In addition, other factors such as the half-life of proteins and the degradation and stability of mRNA may vary widely.^{43,46,47}

Major strengths of our study are the use of a standardized *ex vivo* transwell culture system and weighing each biopsy in order to normalize all protein levels for mg tissue weight. Cytokines were evaluated on both mRNA and protein level,

including four cytokines that were assessed for validation purposes. Furthermore, the use of healthy control skin from regions that are suitable to function as control samples further increases the validity of our study. Possible limitations include the relatively small sample size, lack of dose–response relationships, and AMPs that have only been evaluated on the mRNA level. Moreover, further analysis using staining techniques at

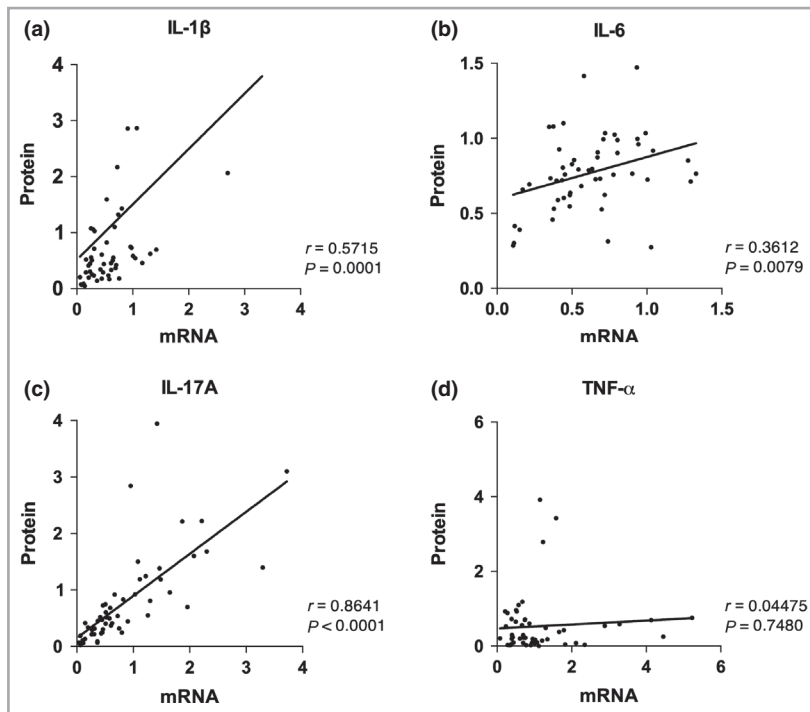


Fig 4. Correlation between the relative protein production (y-axis) and mRNA (x-axis) expression of hidradenitis suppurativa (HS) lesional skin ($n = 9$). (a) Interleukin (IL)-1 β . (b) IL-6. (c) IL-17A. (d) Tumour necrosis factor (TNF)- α . Values on x-axis and y-axis display the relative production and expression values of a cytokine. P-values were calculated according to Spearman's rho test.

the cellular level could potentially indicate the effects of the biologics on the mRNA expression and protein production of inflammatory markers in different cellular subsets.

In conclusion, prednisolone, adalimumab, infliximab, secukinumab, ustekinumab and rituximab significantly inhibited the expression of inflammatory cytokines and AMPs in *ex vivo* actively inflamed lesional skin of patients with HS. The *ex vivo* model will enable studies with combinations of biologics and the targeting of novel important cytokines, alone or in combination with low-dose prednisolone.

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Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's website:

Fig S1. Relative mRNA expression of all cytokines and all antimicrobial peptides (AMPs) in healthy control skin, (n = 5).

Table S1. Relative expression of mRNA and modulation by biologics in healthy control skin.