EDITORIAL

doi: 10.1093/jnci/djz052 First published online April 16, 2019 Editorial

TP53 Status and Estrogen Receptor-Beta in Triple-Negative Breast Cancer: Company Matters

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Estrogen receptor β (ESR2) shares a structural homology at the DNA and ligand binding domains (96% and 58%, respectively) with estrogen receptor α (ESR1), the major type of estrogen receptor in breast cancer ([1,2](#page-1-0)). Similarities notwithstanding, ESR2 has functions and expression patterns distinct from ESR1 and is widely expressed in both basal and luminal epithelial cells [\(3–6](#page-1-0)). The exact role of ESR2 in breast cancer is not clear, with both antiproliferative and proliferative roles being described [\(7](#page-1-0), [8](#page-1-0)). The mechanisms for these opposing actions of ESR2 in breast tumorigenesis have not been fully elucidated; this, in part, is due to different isoforms and binding partners.

In this issue of the Journal, Mukhopadhyay et al. [\(9](#page-1-0)) provide a mechanistic explanation for the plastic nature of ESR2 function in triple-negative breast cancer (TNBC) related to its interactions with TP53 status (wildtype or mutant). In wild-type TP53-expressing cells, silencing of ESR2 augmented apoptosis, whereas its over expression resulted in increased proliferation. Opposite effects were observed following silencing or overexpression of ESR2 in mutant TP53 cells, suggesting the important role of TP53 status in determining ESR2's function. Mechanistically, ESR2-mutant TP53 interaction mediates sequestration of mutant TP53, leading to the TP73 activation and antiproliferative effects. Treatment with tamoxifen (4-hydroxy tamoxifen) also increases ESR2 expression and reactivates TP73 in mutant TP53 cells, providing an explanation for its beneficiary effects. Analysis of the Molecular Taxonomy of Breast Cancer International Consortium TNBC subgroup of basal-like tumors ($n = 259$), based on ESR2 levels and TP53 mutation status, confirmed the impact of these interactions on survival, that is, mutant TP53-expressing tumors with high ESR2 levels have better survival.

The strengths of this study include provision of a mechanistic understanding for the dual role of ESR2 in breast cancer based on TP53 mutational status with further validation of the hypothesis in clinical cohorts. Considering that basal-like TNBC cases are enriched in TP53 mutations ([10\)](#page-1-0), Mukhopadhyay et al. [\(9](#page-1-0)) suggest

that the company of ESR2 with mutant TP53 can prognosticate TNBC patients and more importantly help select a population for tamoxifen therapy. The beneficial effects of endocrine therapy in unselected ESR1-negative breast cancer and TNBC cohorts have been previously described [\(11–14](#page-1-0)). The ability to selectively administer endocrine therapy should, in principle, lead to greater response rates. It is unclear what the impact of ESR2-TP53 interactions have in ER-positive breast cancer, particularly because all patients are offered endocrine therapy.

Many tumor-related genes have been documented to have a dualistic nature being associated with progression in some, but not all, cancers. The opposing effects exist for many biomarkers even within the same cancer as in the case of ESR2 in breast cancer. Understanding the molecular basis of this phenomenon, although not always possible, is a laudable goal. A number of different mechanisms have been described to explain the duality of protein function. The first and foremost is the tissue type. The cellular milieu of different organs is distinct, and the role that individual pathways play in maintaining cellular phenotype can be dramatically different. This is, at least in part, the explanation offered for the tissue-specific impact of mutations in BRCA1, a gene involved in DNA repair. Mutations can also lead to altered splicing pattern or posttranslational modifications resulting in mislocalization of proteins and acquisition of novel functionality. Abnormal nuclear localization of EGFR, and MUC1 and cytoplasmic localization of BRCA1, and TP53 have been described in breast cancer and represent good examples for this concept ([15\)](#page-1-0); these may be because of mutations in the gene itself or its binding partners. Duality of function can also be induced by splicing factors inducing alternative transcripts of the gene as illustrated by progesterone A and B isoforms in breast cancer. Mutations can lead to constitutive activation or suppression of function. Mutations leading to stabilized mutant TP53 proteins may simultaneously gain novel functions,

Received: March 21, 2019; Accepted: April 5, 2019

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primarily through protein–protein interactions with other transcription factors within the cellular neighborhood (16). Proteins that partner with mutant TP53 may transactivate or disrupt target gene activation with consequent changes in cellular function, suggesting the importance of the neighborhood actors. Epithelial splicing regulatory protein (ESRP1), a splicing factor, exhibits a dual role based on the tissue and cancer type (17). Low ESRP1 expression has been associated with the development of epithelial to mesenchymal transformation (EMT) by alternative splicing in ER-negative breast cancer models (MDA-MB-231 cells) (18, 19). In contrast, knockdown of ESRP1 in ERpositive models did not result in development of mesenchymal phenotype (16). This may be because of the lack of key EMT transcription factors in ER-positive breast cancer, suggesting that "company matters."

Beyond the obvious, the current study has broader implications. It documents the important principle of company matters in understanding the impact of markers and mutations in cancers, including breast cancer. The intracellular environment is a complex milieu wherein changes in one player can have a dramatic impact on DNA, RNA, and protein interactions. The players in the neighborhood could further affect cellular phenotype. Acknowledging these processes also provides a reality check for those of us involved in precision medicine, wherein treatments are being prescribed based on the presence of single gene mutations (20). The cooperativity and interactions of cellular networks may, to a large extent, determine the prognostic and predictive utility of mutations in patients. The study by Mukhopadhyay et al. (9) is a good step in this direction and provides compelling reasons to understand the combinatorial impact to determine clinically actionable strategies and solutions.

Notes

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The authors have no conflicts of interest to disclose directly related to this editorial.

References

1. Kuiper GG, Enmark E, Pelto-Huikko M, Nilsson S, Gustafsson JA. Cloning of a novel receptor expressed in rat prostate and ovary. Proc Natl Acad Sci USA. 1996;93(12):5925–5930.

- 2. Mosselman S, Polman J, Dijkema R. ER beta: Identification and characterization of a novel human estrogen receptor. FEBS Lett. 1996;392(1):
- 49–53. 3. Speirs V, Skliris GP, Burdall SE, Carder PJ. Distinct expression patterns of ER alpha and ER beta in normal human mammary gland. J Clin Pathol. 2002;55(5): 371–374.
- 4. Skliris GP, Leygue E, Watson PH, Murphy LC. Estrogen receptor alpha negative breast cancer patients: estrogen receptor beta as a therapeutic target. J Steroid Biochem Mol Biol. 2008;109(1–2):1–10.
- 5. Marotti JD, Collins LC, Hu R, Tamimi RM. Estrogen receptor-beta expression in invasive breast cancer in relation to molecular phenotype: results from the Nurses' Health Study. Mod Pathol. 2010;23(2):197–204.
- 6. Leung YK, Lee MT, Lam HM, Tarapore P, Ho SM. Estrogen receptor-beta and breast cancer: translating biology into clinical practice. Steroids. 2012;77(7): 727–737.
- 7. Palmieri C, Cheng GJ, Saji S, et al. Estrogen receptor beta in breast cancer. Endocr Relat Cancer. 2002;9(1):1–13.
- 8. Leygue E, Murphy LC. A bi-faceted role of estrogen receptor beta in breast cancer. Endocr Relat Cancer. 2013;20(3):R127–R139.
- 9. Mukhopadhyay UK, Oturkar CC, Adams C, et al. TP53 status as a determinant of pro- versus anti-tumorigenic effects of estrogen receptor-beta in breast cancer. J Natl Cancer Inst. 2019;11(11):djz051.
- 10. Garrido-Castro AC, Lin NU, Polyak K. Insights into molecular classifications of triple-negative breast cancer: improving patient selection for treatment. Cancer Discov. 2019;9(2):176–198.
- 11. Gruvberger-Saal SK, Bendahl PO, Saal LH, et al. Estrogen receptor beta expression is associated with tamoxifen response in ERalpha-negative breast carcinoma. Clin Cancer Res. 2007;13(7):1987–1994.
- 12. Honma N, Horii R, Iwase T, et al. Clinical importance of estrogen receptorbeta evaluation in breast cancer patients treated with adjuvant tamoxifen therapy. J Clin Oncol. 2008;26(22):3727–3734.
- 13. Yan Y, Li X, Blanchard A, et al. Expression of both estrogen receptor-beta 1 (ER-beta1) and its co-regulator steroid receptor RNA activator protein (SRAP) are predictive for benefit from tamoxifen therapy in patients with estrogen receptor-alpha (ER-alpha)-negative early breast cancer (EBC). Ann Oncol. 2013; 24(8):1986–1993.
- 14. Mishra AK, Abrahamsson A, Dabrosin C. Fulvestrant inhibits growth of triple negative breast cancer and synergizes with tamoxifen in ERalpha positive breast cancer by up-regulation of ERbeta. Oncotarget. 2016;7(35): 56876–56888.
- 15. Wang X, Li S. Protein mislocalization: mechanisms, functions and clinical applications in cancer. Biochim Biophys Acta. 2014;1846(1):13–25.
- 16. Kim MP, Lozano G. Mutant p53 partners in crime. Cell Death Differ. 2018;25(1): 161–168.
- 17. Gokmen-Polar Y, Neelamraju Y, Goswami CP, et al. Splicing factor ESRP1 controls ER-positive breast cancer by altering metabolic pathways. EMBO Rep. 2019;20(2):e46078. [http://embor.embopress.org/content/20/2/e46078.](http://embor.embopress.org/content/20/2/e46078)
- 18. Warzecha CC, Jiang P, Amirikian K, et al. An ESRP-regulated splicing programme is abrogated during the epithelial-mesenchymal transition. EMBO J. 2010;29(19):3286–3300.
- 19. Warzecha CC, Shen S, Xing Y, Carstens RP. The epithelial splicing factors ESRP1 and ESRP2 positively and negatively regulate diverse types of alternative splicing events. RNA Biol. 2009;6(5):546–562.
- 20. Abramovitz A, Williams C, De PK, et al. Personalized cancer treatment and patient stratification using massive parallel sequencing (MPS) and other OMICs data. In: Badve SS, Kumar GL, eds. Predictive Biomarkers in Oncology: Applications in Precision Medicine. Switzerland: Springer; 2019:131–147.