



Complete Genome Sequence of *Klebsiella pneumoniae* Siphophage Seifer

Adam J. Salazar,^a Lauren Lessor,^a Chandler O'Leary,^a  Jason Gill,^a Mei Liu^a

^aCenter for Phage Technology, Texas A&M University, College Station, Texas, USA

ABSTRACT Carbapenemase-producing *Klebsiella pneumoniae* poses a significant public health threat due to its resistance to antibiotics. Siphophage Seifer was isolated and characterized as part of an effort to develop phage therapeutics to control this pathogen. This report describes the complete genome sequence of phage Seifer, which is a distant member of the χ -like siphovirus phage cluster.

Klebsiella pneumoniae is an opportunistic human pathogen often linked to hospital-acquired infections (1). *K. pneumoniae* strains that carry the plasmid-borne and highly mobile *K. pneumoniae* carbapenemases (*bla*_{KPC}) are of special concern due to their ability to degrade carbapenem antibiotics (1–3). Phages targeting KPC-positive *K. pneumoniae* could be used as alternatives to antibiotic treatments.

Phage Seifer was isolated from a wastewater sample collected from Brazos County, TX, in 2015 using a KPC-positive *K. pneumoniae* clinical isolate of sequence type 258 as the host. Host bacteria were cultured on tryptic soy broth or agar (Difco) at 37°C with aeration. Phages were cultured and propagated using the soft-agar overlay method (4). The phage was identified as a siphophage using negative-stain transmission electron microscopy performed at the Texas A&M University Microscopy and Imaging Center as described previously (5). Phage genomic DNA was prepared using a modified Promega Wizard DNA cleanup kit protocol as described previously (5). Pooled indexed DNA libraries were prepared using the Illumina TruSeq Nano LT kit, and the sequence was obtained from the Illumina MiSeq platform using the MiSeq V2 500-cycle reagent kit following the manufacturer's instructions, producing 697,877 paired-end 250-bp reads for the index containing the phage Seifer genome. FastQC 0.11.5 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) was used to quality control the reads. The reads were trimmed with FastX Toolkit 0.0.14 (http://hannonlab.cshl.edu/fastx_toolkit/download.html) before being assembled using SPAdes 3.5.0 (6). Contig completion was confirmed with PCR using primers (5'-TGTATCCTACGCTCGTCCCC-3', 5'-CCGATTATGACCGCTATG-3') facing off the ends of the assembled contig and Sanger sequencing of the resulting product, with the contig sequence manually corrected to match the resulting Sanger sequencing read. GLIMMER 3.0 (7) and MetaGeneAnnotator 1.0 (8) were used to predict protein coding genes with manual verification, and tRNA genes were predicted with ARAGORN 2.36 (9). Rho-independent terminators were identified via TransTerm (<http://transterm.cbcb.umd.edu/>). Sequence similarity searches were performed using BLASTp 2.2.28 (10) with a maximum expectation cutoff of 0.001 against the NCBI nonredundant (nr), UniProt Swiss-Prot (11), and TrEMBL databases. InterProScan 5.15-54.0 (12), LipoP (13), and TMHMM 2.0 (14) were used to predict protein function. All analyses were conducted at default settings via the CPT Galaxy (15) and WebApollo (16) interfaces (<https://cpt.tamu.edu/galaxy-pub>).

Phage Seifer has a complete genome of 58,197 bp assembled at 212-fold coverage. It has a GC content of 56% and a coding density of 95%. Seifer is a distant member of the χ -like phage cluster (17), with similarity to phage χ in the left arm of the genome.

Citation Salazar AJ, Lessor L, O'Leary C, Gill J, Liu M. 2019. Complete genome sequence of *Klebsiella pneumoniae* siphophage Seifer. *Microbiol Resour Announc* 8:e01289-19. <https://doi.org/10.1128/MRA.01289-19>.

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2019 Salazar et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Mei Liu, meiliu@tamu.edu.

Received 11 October 2019

Accepted 24 October 2019

Published 14 November 2019

Out of 82 total predicted proteins in phage Seifer, 43 and 45 proteins show similarity to proteins encoded by *Salmonella* phage χ (GenBank accession no. [NC_025442](#)) (18, 19) and *Escherichia* phage Utah (GenBank accession no. [KY014601](#)) (20), respectively, based on BLASTp with an E value of <0.001 . The 12-bp 5' extended overhang sequences (5'-GGTGCAGAGC-3') in phages χ and Utah were identified in phage Seifer at the beginning of the genome. As determined by BLASTn against the NCBI nucleotide (nt) database, Seifer is closely related to *Klebsiella* phage Soft (GenBank accession no. [MN106244](#)) at the nucleotide level by sharing 85% overall identity (E value, 0).

Data availability. The genome sequence of phage Seifer was submitted to GenBank under the accession no. [MH817999](#). The associated BioProject, SRA, and BioSample numbers are [PRJNA222858](#), [SRR8556780](#), and [SAMN10904482](#), respectively.

ACKNOWLEDGMENTS

This work was supported by funding from the National Science Foundation (awards EF-0949351 and DBI-1565146) and by the National Institutes of Health (NIAID award AI121689). Additional support came from the Center for Phage Technology (CPT), an Initial University Multidisciplinary Research Initiative supported by Texas A&M University and Texas AgriLife, and from the Department of Biochemistry and Biophysics at Texas A&M University.

We thank Karen Frank at the NIH Clinical Center for providing bacterial isolates. We are grateful for the advice and support of the CPT staff and the Texas A&M University Microscopy and Imaging Center. This announcement was prepared in partial fulfillment of the requirements for BICH464 Phage Genomics, an undergraduate course at Texas A&M University.

REFERENCES

- Satlin MJ, Chen L, Patel G, Gomez-Simmonds A, Weston G, Kim AC, Seo SK, Rosenthal ME, Sperber SJ, Jenkins SG, Hamula CL, Uhlemann AC, Levi MH, Fries BC, Tang YW, Juretschko S, Rojzman AD, Hong T, Mathema B, Jacobs MR, Walsh TJ, Bonomo RA, Kreiswirth BN. 2017. Multicenter clinical and molecular epidemiological analysis of bacteremia due to carbapenem-resistant Enterobacteriaceae (CRE) in the CRE epicenter of the United States. *Antimicrob Agents Chemother* 61:e02349-16. <https://doi.org/10.1128/AAC.02349-16>.
- Sanchez GV, Master RN, Clark RB, Fyyaz M, Duvvuri P, Ekta G, Bordon J. 2013. *Klebsiella pneumoniae* antimicrobial drug resistance, United States, 1998–2010. *Emerg Infect Dis* 19:133–136. <https://doi.org/10.3201/eid1901.120310>.
- Tumbarello M, Viale P, Viscoli C, Trecarichi EM, Tumietto F, Marchese A, Spanu T, Ambretti S, Ginocchio F, Cristini F, Losito AR, Tedeschi S, Cauda R, Bassetti M. 2012. Predictors of mortality in bloodstream infections caused by *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*: importance of combination therapy. *Clin Infect Dis* 55: 943–950. <https://doi.org/10.1093/cid/cis588>.
- Adams MK. 1959. *Bacteriophages*. Interscience Publishers, Inc., New York, NY.
- Gill JJ, Berry JD, Russell WK, Lessor L, Escobar-Garcia DA, Hernandez D, Kane A, Keene J, Maddox M, Martin R, Mohan S, Thorn AM, Russell DH, Young R. 2012. The *Caulobacter crescentus* phage phiCbK: genomics of a canonical phage. *BMC Genomics* 13:542. <https://doi.org/10.1186/1471-2164-13-542>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27: 4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledgebase. *Nucleic Acids Res* 46:2699. <https://doi.org/10.1093/nar/gky092>.
- Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30: 1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
- Juncker AS, Willenbrock H, Von Heijne G, Brunak S, Nielsen H, Krogh A. 2003. Prediction of lipoprotein signal peptides in Gram-negative bacteria. *Protein Sci* 12:1652–1662. <https://doi.org/10.1110/ps.0303703>.
- Krogh A, Larsson B, von Heijne G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- Cock PJ, Gruning BA, Paszkiewicz K, Pritchard L. 2013. Galaxy tools and workflows for sequence analysis with applications in molecular plant pathology. *PeerJ* 1:e167. <https://doi.org/10.7717/peerj.167>.
- Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elsik CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. *Genome Biol* 14:R93. <https://doi.org/10.1186/gb-2013-14-8-r93>.
- Grose JH, Casjens SR. 2014. Understanding the enormous diversity of bacteriophages: the tailed phages that infect the bacterial family Enterobacteriaceae. *Virology* 468–470:421–443. <https://doi.org/10.1016/j.virol.2014.08.024>.
- Lee JH, Shin H, Choi Y, Ryu S. 2013. Complete genome sequence analysis of bacterial-flagellum-targeting bacteriophage chi. *Arch Virol* 158: 2179–2183. <https://doi.org/10.1007/s00705-013-1700-0>.
- Hendrix RW, Ko CC, Jacobs-Sera D, Hatfull GF, Erhardt M, Hughes KT, Casjens SR. 2015. Genome sequence of *Salmonella* phage chi. *Genome Announc* 3:e01229-14. <https://doi.org/10.1128/genomeA.01229-14>.
- Leavitt JC, Heitkamp AJ, Bhattacharjee AS, Gilcrease EB, Casjens SR. 2017. Genome sequence of *Escherichia coli* tailed phage Utah. *Genome Announc* 5:e01494-16. <https://doi.org/10.1128/genomeA.01494-16>.