



Review

Interplay Between MicroRNAs and Oxidative Stress in Ovarian Conditions with a Focus on Ovarian Cancer and Endometriosis

Josep Mari-Alexandre ^{1,*}, Antonio Pellín Carcelén ^{2,†}, Cristina Agababyan ^{1,3},
Andrea Moreno-Manuel ^{4,5}, Javier García-Oms ^{1,3}, Silvia Calabuig-Fariñas ^{4,5,6,7} and
Juan Gilabert-Estellés ^{1,3,8}

¹ Research Laboratory in Biomarkers in Reproduction, Gynaecology and Obstetrics, Fundació Hospital General Universitario de València, 46014 València, Spain; dra.kristina.agababyan@gmail.com (C.A.); javiergoms@yahoo.es (J.G.-O.); juangilaeste@yahoo.es (J.G.-E.)

² Department of Physiology, Universitat de València, 46010 València, Spain; apellincarcelen@gmail.com

³ Comprehensive Multidisciplinary Endometriosis Unit, Consorcio Hospital General Universitario de València, 46014 València, Spain

⁴ Molecular Oncology Laboratory, Fundació para la Investigació del Hospital General Universitario de València, 46014, València, Spain; andrea.morenomanuel@gmail.com (A.M.-M.); calabuix_sil@gva.es (S.C.-F.)

⁵ TRIAL Mixed Unit, Centro de Investigación Príncipe Felipe-Fundación para la Investigación del Hospital General Universitario de València, 46014 València, Spain

⁶ Department of Pathology, Universitat de València, 46010 València, Spain

⁷ Centro de Investigación Biomédica en Red en Cáncer (CIBERONC), 46014 València, Spain

⁸ Department of Paediatrics, Obstetrics and Gynaecology, University of València, 46010 València, Spain

* Correspondence: mari_josale@gva.es; Tel.: +34-96-313-1893 (ext. 437211)

† These authors contributed equally to this work.

Received: 23 September 2019; Accepted: 23 October 2019; Published: 25 October 2019



Abstract: Ovarian cancer and endometriosis are two distinct gynaecological conditions that share many biological aspects including proliferation, invasion of surrounding tissue, inflammation, inhibition of apoptosis, deregulation of angiogenesis and the ability to spread at a distance. miRNAs are small non-coding RNAs (19–22 nt) that act as post-transcriptional modulators of gene expression and are involved in several of the aforementioned processes. In addition, a growing body of evidence supports the contribution of oxidative stress (OS) to these gynaecological diseases: increased peritoneal OS due to the decomposition of retrograde menstruation blood facilitates both endometriotic lesion development and fallopian tube malignant transformation leading to high-grade serous ovarian cancer (HGSOC). Furthermore, as HGSOC develops, increased OS levels are associated with chemoresistance. Finally, continued bleeding within ovarian endometrioma raises OS levels and contributes to the development of endometriosis-associated ovarian cancer (EAOC). Therefore, this review aims to address the need for a better understanding of the dialogue between miRNAs and oxidative stress in the pathophysiology of ovarian conditions: endometriosis, EAOC and HGSOC.

Keywords: oxidative stress; miRNAs; endometriosis; high-grade serous ovarian cancer; endometriosis-associated ovarian cancer; epithelial-to-mesenchymal transition; chemoresistance

1. Introduction

The Great Oxidative Event occurred between 2.4 and 2.1 billion years ago when high O₂ concentrations appeared in the Earth atmosphere as a metabolic product of cyanobacteria oxygenic photosynthesis [1]. This phenomenon is considered a breakthrough for life on Earth, as living organisms had to develop an arsenal of antioxidant strategies to adapt to this powerful compound. However,

in some circumstances the balance between oxidants and antioxidants might be shifted in favour of the former, giving rise to oxidative stress (OS). Remarkably, high partial oxygen pressures might also lead to OS through the formation of highly reactive oxygen species (ROS), including superoxide anion radical ($O_2^{\bullet-}$), hydroxyl radical (OH^{\bullet}), hydrogen peroxide (H_2O_2), and peroxy radical (ROO^{\bullet}) (Figure 1).

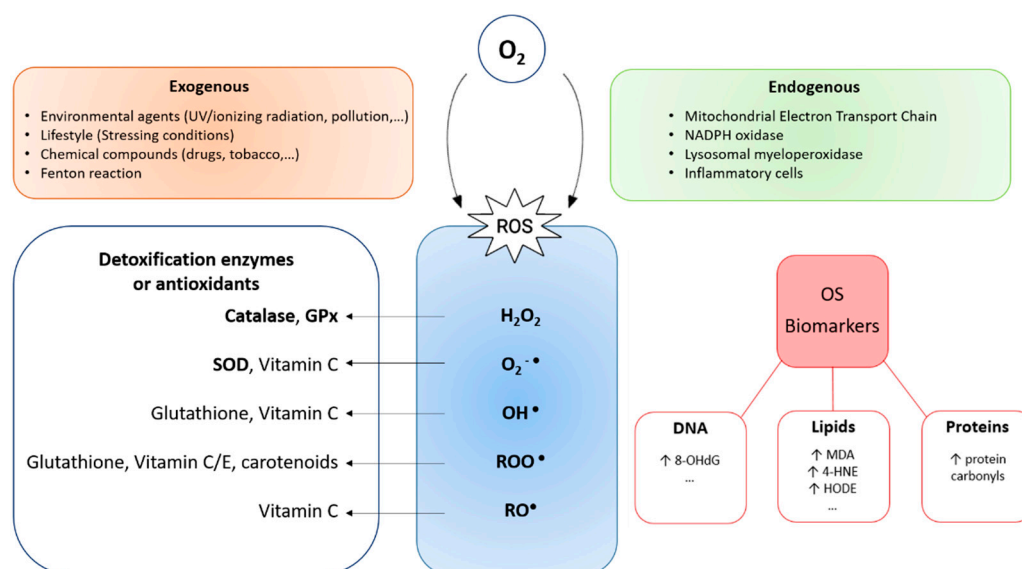


Figure 1. Diagram representing reactive oxygen species (ROS) production, their detoxification mechanisms and the oxidative stress alterations produced by their action, which can serve as oxidative stress (OS) biomarkers. Abbreviations: NADPH, nicotinamide adenine dinucleotide phosphate; GPx, glutathione peroxidase; SOD, superoxide dismutase; H_2O_2 , hydrogen peroxide; $O_2^{\bullet-}$, superoxide anion radical; OH^{\bullet} , hydroxyl radical; ROO^{\bullet} , peroxy radical; RO^{\bullet} , alkoxy radical; 8-OHdG, 8-hydroxy-2'-deoxyguanosine; MDA, malondialdehyde; 4-HNE, 4-hydroxynonenal; HODE, hydroxyoctadecadienoic acid.

Certain levels of ROS can be produced endogenously as a sub-product of countless chemical reactions essential for cell life (including those mediated by the mitochondrial electron transport chain reactions and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase), playing an important role in the regulation of cellular signalling processes [2]. However, an excess of ROS provokes the disruption of redox signalling and control and/or molecular damage [3] (Figure 1). On the other hand, ROS can also be generated exogenously, as a result of the exposure of biological systems to environmental agents (i.e., ultraviolet or ionizing radiation) or by the action of free iron (via the Fenton reaction) [4,5] (Figure 1).

The antioxidant mechanisms in charge of avoiding oxidative damage to cells include: a) enzymes (such as superoxide dismutase, glutathione peroxidase, catalase and metal binding proteins), b) non-enzymatic protectors (such as glutathione, vitamin E, vitamin C, uric acid, bilirubin and albumin) and c) repairers of damaged molecules (such as DNA repair enzymes, methionine, sulfoxide reductase) [6] (Figure 1). Contrary to the popular belief, growing evidence suggests a predominant role of antioxidant enzymes over dietary antioxidants in protection against OS [7]. When these antioxidant defences are overwhelmed (due to an excess of prooxidant substances, a deficiency of antioxidant agents or both [8]) OS produces damage to biomolecules essential for life, as lipids (malondialdehyde) [6,9], proteins (protein carbonyls) [3,9] and DNA (8-hydroxy-2'-deoxyguanosine (8-OHdG) [6,7,10] (Figure 1). This contributes to the pathophysiology of many pathological conditions such as Alzheimer's disease [11], frailty [9], ovarian cancer [4], and endometriosis [1].

On the other hand, epigenetics refers to the heritable changes in gene function that cannot be explained by changes in the DNA sequence [12]. These changes are produced through four

epigenetic mechanisms that are dynamic and reversible, and include: DNA methylation, histone modifications, chromatin remodelling, and the expression of non-coding RNAs, including miRNAs [13]. A growing body of evidence suggests that epigenetics could be involved in the pathophysiology of endometriosis [14] and that carcinogenesis cannot be explained only by DNA mutations, but also that epigenetic alterations need to be included in the equation [15]. In this respect, distinct epigenetic mechanisms could contribute to carcinogenesis either by repressing the expression of tumour suppressor genes (TSG) (i.e., DNA hypermethylation at gene promoters, over-expression of miRNA targeting TSG, histone modifications, and heterochromatin conformation at TSG coding regions) or allowing the activation of oncogenes (OG) (i.e., global DNA hypomethylation, down-regulation of miRNA targeting OG, histone modifications, and euchromatin conformation at OG coding regions).

Importantly, great research endeavours have been conducted to decipher the role of miRNAs in these pathologies. miRNAs are small (19–22 nt) non-coding RNAs that can act as post-transcriptional regulators of gene expression, reducing the expression of their target mRNAs either by inhibiting its translation or by promoting its degradation. Thus, the levels of their target mRNAs are opposed to those of their targeting miRNAs (Figure 2B). It is worth mentioning that several miRNAs can target a given mRNA and a single miRNA can target several mRNAs, increasing the complexity of the regulatory mechanism mediated by these molecules [16–19]. miRNAs are involved in pivotal biological processes including development, differentiation, apoptosis, and proliferation. Remarkably, miRNAs themselves can also act as OG or TSG, depending on their targets [20]. Extensive literature supports the role of miRNAs in the development of endometriosis (reviewed in [21–24]) as well as in endometriosis-associated ovarian cancer (EAOC) [25,26] and in high-grade serous ovarian cancer (HGSOC) [27–29], as hereafter described.

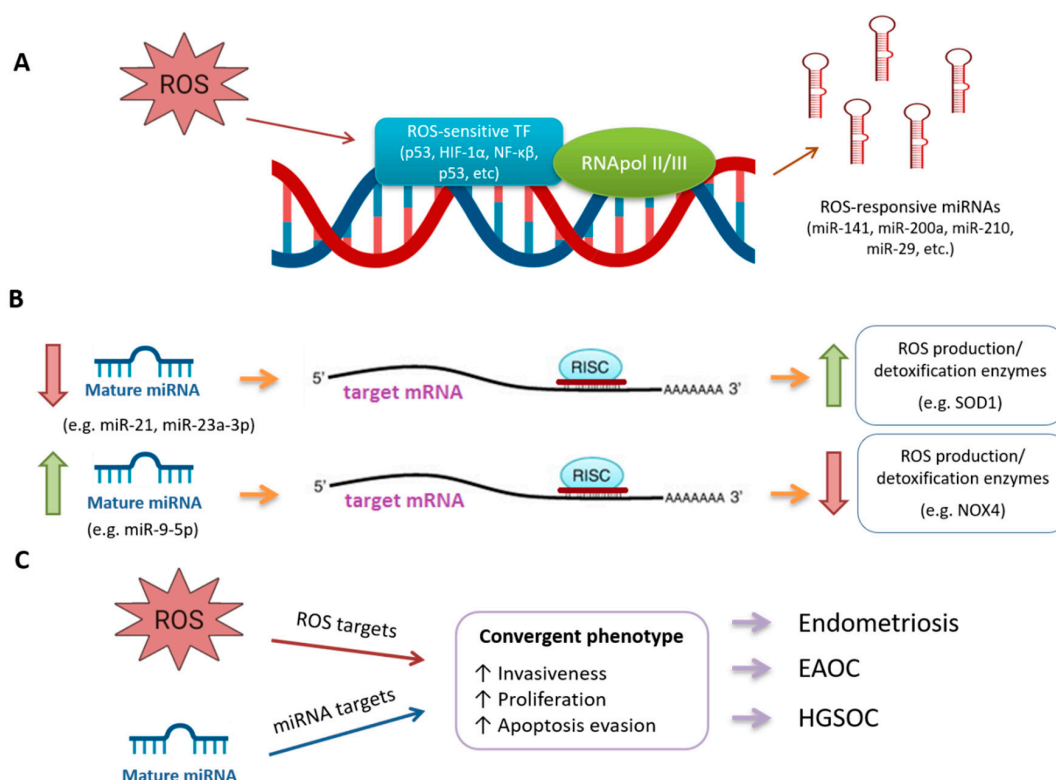


Figure 2. Schematic representation of the possible mechanisms of interplay between miRNAs and oxidative stress. (A) ROS can activate ROS-sensitive transcription factors to induce the transcription of specific primary miRNA (pri-miRNA) that will conduce to mature miRNAs; (B) The levels of a given miRNA inversely correlate with those of their target mRNAs, that could belong to ROS production/detoxification enzymes; (C) ROS and miRNAs can produce separate effects that converge

in a common phenotype, leading to endometriosis, EAOC and HGOSC. Abbreviations: ROS, reactive oxygen species; TF, transcription factors; RNAPol II/III, RNA polymerase II or III; HIF-1 α , hypoxia-inducible factor 1 α ; NF- κ B, nuclear factor κ B; RISC, RNA-induced silencing complex; SOD1, superoxide dismutase 1; NOX4, NADPH oxidase 4; EAOC, endometriosis-associated ovarian cancer; HGOSC, high-grade serous ovarian cancer.

As for the goal of this review, the complex interplay between OS and miRNAs represents an active research area in different pathologies that develops on the basis of several premises: (1) OS triggers the expression of responsive miRNAs through ROS-sensitive transcription factors (reviewed in [30,31]) (Figure 2A); (2) miRNAs can regulate the expression of enzymes involved in ROS production or detoxification (reviewed in [31,32]) (Figure 2B); (3) miRNAs and OS independently act to reach convergent phenotypes (Figure 2C).

Therefore, the aim of the present review is to compile existing evidence on how miRNAs and OS interact through the aforementioned mechanisms into the pathophysiology of three important gynaecological diseases: endometriosis, EAOC, and HGOSC.

2. Endometriosis

Endometriosis is an oestrogen-dependent inflammatory disorder defined by the presence of endometrial-like tissue in ectopic locations, which limits the quality of life of affected women [33–35]. This pathology affects 10% of reproductive-aged women from all ethnic and social groups, although the prevalence in those patients experiencing pain, infertility, or both is as high as 35%–50% [36], being the estimated prevalence of this condition around 176 million worldwide. The most frequent involvement is in the peritoneum (superficial and deep endometriotic implants) and ovaries (ovarian endometrioma (OMA) or endometriotic cysts), although cases of pulmonary [37] and cerebral endometriosis [38] have also been documented.

While a unifying theory regarding the exact aetiopathogenic mechanism of endometriosis is still lacking [13], nowadays the most widely accepted theory is Sampson's proposal of retrograde menstruation and implantation [39]. This theory postulates that desquamated endometrial cells reach the peritoneal cavity by retrograde flow through the fallopian tubes, where they are able to implant and survive (Figure 3). Since retrograde menstruation occurs in 90% of healthy women of reproductive age with patent fallopian tubes [40], the fact that only a small percentage develops the disease suggests that there must be additional mechanisms that allow the migrated tissue to implant and survive [41].

miRNAs and Oxidative Stress in Endometriosis

Sampson's proposal not only postulates an origin for the endometrial-like ectopic tissue, but also provides a mechanism for the action of OS in the pathogenesis of endometriosis. Remarkably, compelling evidence demonstrates an increase in OS markers in several fluids (serum, peritoneal and follicular fluid) and tissues in women with endometriosis (reviewed in [42,43]).

From the point of view of the desquamated cells, several studies including ours have observed that eutopic endometria expresses higher levels of vascular endothelial growth factor A (VEGF-A) (the main pro-angiogenic factor) [44,45], urokinase-type plasminogen activator (uPA), and matrix metalloproteinase 3 (MMP-3) (proteolytic factors) [46] in comparison to control endometria, a process that could be mediated by miRNAs [47]. Upon menstruation, endometrial cells lose their blood supply, entering into a hypoxic state by the time they reach the peritoneal cavity. This upregulates the expression of hypoxia-inducible factor 1-alpha (HIF-1 α), which further stimulates the transcription of several hypoxia-inducible miRNAs or hypoxamiRs [30,48], as the prototypical hypoxamiR, miR-210, which is overexpressed in OMA tissues, promoting cell survival [49] (Figure 2A).

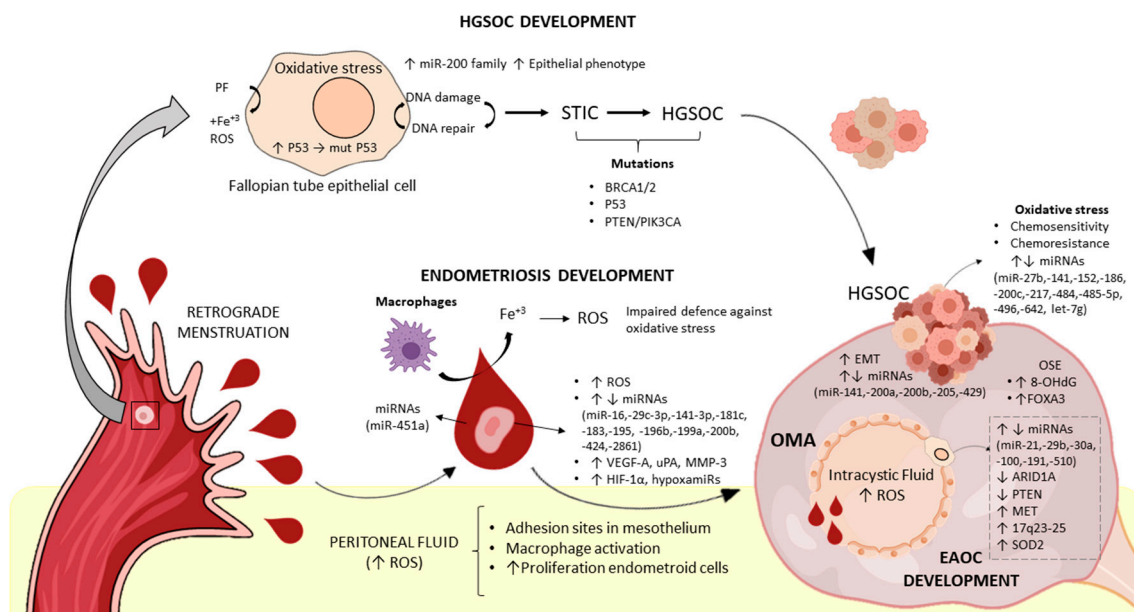


Figure 3. Global vision of aetiopathogenic mechanisms leading to endometriosis, EAO and HGSOC development. Regarding HGSOC development, oxidative stress contributes to fallopian tube epithelial cells alterations, through the action of ROS. Repeated cycles of DNA damage and repair produce mutations in driver genes *BRCA1/2*, *P53*, *PTEN* and *PIK3CA*. Additionally, miRNA deregulation contributes to tumour progression. Once the malignant lesion is established in the ovary, oxidative stress is initially involved in first-line chemotherapy mechanism of action, although excessive oxidative stress is linked to tumour chemoresistance. Regarding endometriosis development, refluxed endometrial cells from patients show some features predisposing them to the development of this condition (i.e., increased angiogenesis and proteolysis, disbalanced miRNAs profile, etc). Upon menstruation, endometrial cells lose their blood supply and activate hypoxia-responsive miRNAs (hypoxamiRs) that together with erythrocyte-derived miRNAs contribute to the development of the condition. Blood decomposition by pelvic macrophages contribute to ROS production, which alters the peritoneal microenvironment to enhance endometrial cells attachment and proliferation. Finally, the intra-cystic fluid of OMAs presents with higher levels of ROS, triggering subsequent events such as miRNAs disbalance, decreased expression of ARID1A and PTEN, amplification of *MET* and 17q24–25, and increased generation of SOD2, all of which enhance the development of EAO. Abbreviations: EAO, endometriosis-associated ovarian cancer; HGSOC, High-Grade Serous Ovarian Cancer; STIC, Serous Tubal Intraepithelial Carcinoma; OSE, Ovarian Surface Epithelium; OMA, Ovarian endometrioma; ROS, Reactive Oxygen Species; PF, Peritoneal Fluid; BRCA, Breast Cancer gene; PTEN, Phosphatase and tensin homolog; PIK3CA, Phosphatidylinositol-4,5 Biphosphonate 3-Kinase Catalytic Subunit Alpha; VEGF-A, Vascular Endothelial Growth Factor A; uPA, Urokinase-type plasminogen Activator; MMP-3; Matrix Metalloproteinase 3; HIF-1 α ; Hypoxia-inducible factor 1-alpha; ARID1A, AT-Rich Interaction Domain A; MET, mesenchymal-to-epithelial transcription factor; EMT, Epithelial-Mesenchymal Transition; SOD2, Superoxide Dismutase 2; FOXA3, Hepatocyte Nuclear Factor 3-gamma; 8-OHdG, 8-Oxo-2'-deoxyguanosine.

Once in the peritoneal cavity, endometrial cells floating in a mixture of blood and peritoneal fluid (PF) need to undergo a continuum of important events if they are to implant and survive. These events include the attachment to ectopic sites, extracellular matrix degradation, invasion, and angiogenesis (a complex and sequential process devoted to the formation of new blood vessels from pre-existing ones, to assure oxygen and nutrient supply for proliferation and survival [50]). Notably, *in vitro* studies revealed that several miRNAs are involved in distinct processes leading to the establishment and survival of the endometriotic lesions, including invasiveness (miR-200b [51], miR-183 [52], miR-199a [53]), proliferation (miR-210 [54], miR-200b [51], miR-2861 [55], miR-195 [56], miR-196b [57]), apoptosis

evasion (miR-181c [58], miR-141-3p [59], miR-2861 [55], miR-195 [56], miR-196b [57], miR-210 [54], and increased angiogenesis (miR-16, miR-29c-3p, miR-424 [60]) (Table 1). A myriad of studies reveals the putative role of miRNAs in endometriosis, recently reviewed by Panir and collaborators [61]. Among these miRNAs are miR-21, miR-23a-3p, and miR-9-5p, which have been linked to the regulation or redox enzymes [30] (Figure 2B).

Table 1. Deregulated miRNAs in selected in vitro studies in endometriosis.

References	Main Biological Function Promoted	Experimental Design	Main Deregulated miRNAs in Patients	
[49]	Cell survival	OMA cell line under hypoxia	↑ miR-210	
[51]	Invasiveness	Immortalized endometriotic cell line 12Z, the stromal cell line ST-T1b and primary endometriotic stromal cells	↓ miR-200b	
[52]		Primary eutopic and control stromal cells	↓ miR-183	
[53]		Primary ectopic, eutopic and control stromal cells	↓ miR-199a	
[54]	Proliferation	Primary ectopic and control stromal cells	↑ miR-210	
[51]		Immortalized endometriotic cell line 12Z, the stromal cell line ST-T1b and primary endometriotic stromal cells	↓ miR-200b	
[55]		Ectopic endometrial cells	↓ miR-2861	
[56]		Primary ectopic, eutopic and control stromal cells	↓ miR-195	
[57]		Primary ectopic and control stromal cells	↓ mi-196b	
[58]		Endometrial cell lines	↑ miR-181c	
[59]		Apoptosis evasion	Ectopic endometrial stromal cells	↓ miR-143-3p
[55]			Ectopic endometrial cells	↓ miR-2861
[56]	Primary ectopic, eutopic and control stromal cells		↓ miR-195	
[57]	Primary ectopic and control stromal cells		↓ mi-196b	
[49]	Angiogenesis	OMA cell line under hypoxia	↑ miR-210	
[54]		Primary ectopic and control stromal cells	↓ miR-16, ↓ miR-29c-3p, ↓ miR-424	

↑, up-regulated levels; ↓, down-regulated levels; OMA: ovarian endometrioma.

From the point of view of the milieu into which these cells arrive, PF from patients might favour endometriosis development through several mechanisms: (a) Firstly, increased OS in PF might create adhesion sites for the migrated cells by damaging the mesothelial wall [1]; (b) acting on endometrial cells, PF from patients is involved in the over-expression of the proteolytic factors uPA and MMP-3 [62], and not yet resolved components in this biofluid increase angiogenesis by down-regulation of angiogenesis-related miRNAs (miR-16-5p, miR-29c-3p, and miR-424-5p), mainly in eutopic cells from patients, favouring their survival [60]. The putative role of OS in mediating these effects is reinforced by recent results from Wright and co-workers [63], who observed that global down-regulation in miRNAs could be recapitulated by stimulating endometrial cells with oxidized low-density lipoprotein (LDL) (an OS marker present in patient's PF and associated with pain). These in vitro findings have also been observed in tissues, since several authors, including us, have reported an increase in VEGF-A in endometriotic tissues, which might also be regulated by miRNAs such as miR-16, miR-29c-3p, and miR-424 [45,60,64]; and (c) accompanying erythrocytes might be a source not only of OS but also of miRNAs [65], as observed by the presence of miR-451, the most abundant miRNA in erythrocytes, in PF [47]. Surprisingly, in vitro and in vivo evidence suggests that miR-451 is uptaken by endometriotic tissues, correlating its expression with survival status of the lesions [66] (Table 2, Figure 3).

Table 2. Deregulated miRNAs in selected studies considering distinct biofluids from patients with endometriosis, EAOC or HGSOC compared to control women.

miRNAs in Biofluids			
Reference	Gynaecological Condition	Biofluid Specimen	Main Deregulated miRNAs in Patients
[47]	Endometriosis	Peritoneal fluid	↑ miR-106b-3p, miR-451a and miR-486-5p
[67]		Serum	↓ let-7b and miR-135
[68]		Serum	↓ miR-9 *, miR-141 *, miR-145 * and miR-542-3p ↑ miR-122 and miR-199a
[69]		Serum	↑ miR-122 and miR-199a
[70]		Serum	↓ miR-30c-5p, miR-127-3p, miR-99b-5p, miRNA-15b-5p and miRNA-20a-5p ↑ miR-424-3p and miR-185-5p
[71]		Plasma	↓ miR-17-5p, miR-20a and miR-22
[72]		Plasma	↓ miR-200a-3p, miR-200b-3p and miR-414-3p
[73]		Plasma	↑ miR-154-5p
[74]	EAOC	Plasma	Three distinct miRNA signatures, including ↑ miR-15b, miR-16, miR-21, and miR-195
[75]	HGSOC	Serum	↑ miR-1290
[76]		Serum	↓ miR-375 + CA-125 levels
[77]		Serum	↑ miR-1246
[78]		Serum	↑ miR-200b, miR-200c

↑, up-regulated levels; ↓ down-regulated levels; CA-125, cancer antigen 125.

Provided that several authors do not distinguish the type of ectopic lesions into their analyses, the precise effect of OS in different endometriotic lesions is difficult to evaluate. In spite of this, some conclusions can be drawn for the OMA landscape which will be commented on from the extra- to the intra-cystic space. Firstly, several authors observed an increase in 8-OHdG [79,80], forkhead box A3 (FOXA3) and advanced glycation end products [80] in the normal ovarian cortex surrounding OMAs in comparison to the normal ovarian cortex surrounding benign ovarian cysts, which might postulate OS as a specific mechanism in endometriosis. Secondly, Ngô and co-workers [81] observed that the pro-oxidant/antioxidant balance is shifted towards enhanced OS in both epithelial and stromal cells within the cyst wall, which increased cell proliferation through ERK1/2 pathway activation. Additionally, Chen and co-workers [82] observed an increase in generation of both ROS and of superoxide dismutase 2 (SOD2) by mitochondria in stromal cells, which may support the development of the disease by allowing a high metabolic rate within these lesions. Finally, the increased ROS production in endometriotic cells might also be a consequence of the pro-oxidative inner cyst fluid stimulation [83]. Interestingly, stimulation of immortalized ovarian surface epithelium (OSE) and endometrial glandular cells with endometriotic cyst content produced more ROS than treatment with non-endometriotic cyst content [84]. Altogether, the increased OS might have consequences in gene expression, regarding the down-regulation of the tumour suppressor gene AT-rich interaction domain A (*ARID1A*) via promoter hypermethylation [85], which is considered an early event in endometriosis malignant transformation (Figure 3).

Opposite to this evidence, Santulli and collaborators [86] did not find any significant differences in protein OS markers in the PF of women with ovarian or peritoneal endometriosis when compared with control PF, in contrast to women with deep infiltrating endometriosis. This counterintuitive finding might find a rationale either when considering OMA as an encapsulated lesion within the ovary unlikely to influence PF composition, or a possible bias in these observations due to the inclusion of patients with benign pathologies in the control population.

3. Endometriosis-Associated Ovarian Cancer

Endometriosis malignant transformation occurs in OMA at a higher rate than in other endometriotic lesions [87], producing the so-called EAOC. This clinical entity might be the result of a sequential process of malignant transformation from endometriotic lesions through atypical endometriosis [88] and, finally, to ovarian cancer, especially to the endometrioid (EOC) and clear cell (OCCC) histological subtypes [89]. Importantly, the estimated risk of malignancy of ovarian endometriosis may be close to 1% [90,91]. This enhanced rate of transformation of OMA, together with the specific histotypes to which it leads, points to unique carcinogenic processes within these endometriotic lesions, with a plausible involvement of OS and miRNAs.

miRNAs and Oxidative Stress in Endometriosis-Associated Ovarian Cancer

The intra-cystic fluid within OMA represents a unique milieu that may underlay the initiation of the malignant transformation process [92], probably triggered by the release of free iron as a result of monthly bleeding and the subsequent raise in OS [84] (Figure 3). At this respect, endometriotic cysts fluid present with higher levels of free iron, lactate dehydrogenase, lipid peroxide, 8-OHdG and potential antioxidant in comparison to non-endometriotic cysts and OCCC. Also, higher levels of 8-OHdG are reportedly associated with OCCC, being almost negative in EOC [93]. In agreement with these results, Fujimoto and co-workers [94] found increased levels of 8-OHdG and heme-oxygenase 1 in cyst fluid from OMA when compared to EAOC (mainly OCCC). Additionally, the expression of 8-OHdG is decreased in EAOC tissues (OCCC and EOC) when compared to paired adjacent endometriotic tissue and OMA [95]. Therefore, one might conclude that the levels of OS are higher in OMA and decrease in OCCC, and further in EOC, which suggests their involvement mainly in the initiation process of EAOC.

Regarding antioxidants, decreased MnSOD expression and increased malondialdehyde (MDA) expression has been observed in OMA and EOAC in comparison to non-EAOC and control endometria, corroborating the increased OS levels commonly found in these tissues [96]. In vitro studies denoted that the over-expression of the antioxidant lipocalin2 increases intracellular iron concentrations in OCCC cell lines but reduces the levels of ROS and DNA damage, probably through increasing glutathione, xCT (a cystine transporter protein) and CD44v (a stem cell marker), resulting in reduced apoptosis and prolonged cell survival of OCCC [97]. Altogether, it seems plausible to acknowledge that OS is involved in early steps of the malignant transformation of endometriosis.

In this context, some of the specific alterations produced by this pro-oxidant milieu have been unravelled. Opposed to HGSOE, EAOC is characterized by mutations in several genes, including the TSG *ARID1A*, *PTEN*, and the OG phosphatidylinositol-4,5 biphosphonate 3-kinase catalytic subunit alpha (*PIK3CA*) [98]. Winarto and collaborators [96] investigated tissue samples from patients with endometriosis, EAOC, or non-EAOC to observe that *ARID1A* expression decreases with increased OS a finding also corroborated in vitro. Interestingly, an epigenetic modification (*ARID1A* promoter hypermethylation in OMA) might underlay this observation [85]. In addition, chromosomal aberrations are more frequently found in EOC over OMA and less frequently in extragonadal endometriosis, which might reflect a clonal expansion of aberrant OMA cells produced as a result of the harmful intra-cystic milieu [87]. In addition, copy number variation in OCCC has been identified by several authors: for instance, mesenchymal-to-epithelial transcription factor (*MET*) gene amplification is a frequent event in OCCC and this genomic amplification can be recapitulated in an ROS-induced rat carcinoma model [99]. Furthermore, amplification at loci 17q23-25 occurs in approximately 40% of OCCC, which might over-express the encoded miR-21, decreasing the expression of their target *PTEN*. Therefore, both OS and miRNAs converge in some cases with genetic alterations to provoke the loss of function of important tumour suppressor genes in EAOC carcinogenesis (Figure 3).

Apart from its role in tumour initiation, a growing body of evidence has shed light into the role of deregulated miRNAs in other processes linked to EAOC carcinogenesis, as cell proliferation [100–102], migration [100,101], invasion [100,102], and epithelial-to-mesenchymal transition [100,103–106].

However, differences in study design and methodological approaches render low overlapping between reported deregulated miRNAs (Table 3), that is limited to miR-21 [106,107], miR-510 [104,105], miR-29b [108,109], miR-191 [102,110] and miR-30a [111,112].

Table 3. Deregulated miRNAs in selected studies in EAOC.

Reference	Effect	Experimental Design	Main Deregulated miRNAs in Patients
[100]	Promoted proliferation, migration, invasion	OCCC and adjacent non-tumor tissues	↓ miR-424
[101]	Increased cell motility, growth and colony formation	OCCC and EOC cell lines and OMA primary stromal cells	↓ miR-381
[102]	Increased cell proliferation and invasion	EOC, OCCC, OMA and control endometria tissues	↑ miR-191
[103]	Increased MET phenotype and good prognosis	HGSOC, EOC, OCCC and mucinous ovarian cancer tissues	↓ miR-506
[104]	Increased EMT phenotype	HGSOC and OSE tissues	↑ miR-205-5p
	EMT (miR-200s), poor PFS and OS (miR-200c -3p)	HGSOC, OCCC and OSE tissues	↑ miR-200s, miR-182-5p ↓ miR-383
	Hystology differentiators	OCCC and HGSOC tissues	↑ miR-509-3-5p, miR-509-3p, miR-509-5p, miR-510
[105]	Poor overall survival	OCCC and HGSOC tissues	↓ miR-510, miR-129-3p
[106]	Increased EMT phenotype	OCCC and HGSOC tissues	↑ miR-9
[107]	Down-regulation of the TSG PTEN	OCCC tissues	↑ miR-21
[108]	Increased paclitaxel chemosensitivity	OCCC cell lines	↑ miR-29b
[109]	Poor prognosis	OCCC, HGSOC, mucinous ovarian cancer and control tissues	↓ miR-29b
[110]	Increased apoptosis evasion	EOC, OMA and control tissues	↑ miR-191
[111]	Hystology differentiators	OCCC, EOC, HGSOC and mucinous ovarian cancer	↑ miR-30a and miR-30a *
[112]	Poor overall survival in ovarian papillary serous carcinoma tissues	OCCC and ovarian papillary serous carcinoma tissues	↓ miR-30a, miR-30e and miR-505
[113]	Enhanced sensitivity to everolimus	OCCC and OSE cell lines	↓ miR-100

↑, up-regulated levels; ↓, down-regulated levels; EOC, endometrioid ovarian cancer; OCCC, ovarian clear cell carcinoma; OSE, ovarian surface epithelium; HGSOC, high-grade serous ovarian cancer; EMT, epithelial-to-mesenchymal transition; MET, mesenchymal-to-epithelial transition. PTEN, phosphatase and tensin homologue; TSG, tumour suppressor gene; PFS, progression-free survival.

Regarding treatment, first-line chemotherapy agents in ovarian cancer (EAOC and HGSOC) are represented by taxanes and carboplatin, which exert their antitumour effect partly mediated by an increase in OS (Figure 3). A very recent study from Amano and co-workers [114] observed an association between higher mitochondrial SOD2 levels in EAOC and poor prognosis, without any difference regarding histology (OCCC vs. EOC), which could reflect an improved scavenging of platinum-mediated ROS production during EAOC treatment. On the other hand, the acquisition of chemoresistance by both EAOC and HGSOC is directly linked to the poor overall results in these patients. Specifically, the clear cell subtype is known to be more chemoresistant and associated to worse prognosis than the endometrioid subtype.

In this context, Sugio and co-workers [108] observed that increased miR-29b levels correlate with progression-free survival in OCCC patients. Of note, the down-regulation of the protein Bcl2-associated athanogene 3 (BAG3) seemed to induce the miR-29b expression, which finally sensitized cells to paclitaxel. In an attempt to overwhelm chemoresistance in OCCC, new therapeutic approaches have been developed, including the use of the rapamycin analogue everolimus, for which resistances have also been documented [115]. However, Nagaraja and collaborators [113] found that induced over-expression of miR-100 increased the sensitivity to everolimus in OCCC cell lines, an effect

mediated by the inhibition of mammalian target of rapamycin (mTOR) signalling (Table 3). All in all, these results might pave the way for a miRNA-based therapy to overcome chemoresistance in EAO.

Therefore, there exists compelling evidence that miRNAs and OS cooperate in the distinct steps of EAO carcinogenesis to accomplish the malignant transformation of endometriosis and that both of them influence therapeutic outcomes in these conditions.

4. High-Grade Serous Ovarian Cancer

Ovarian cancer is the fifth cause of cancer death in women and the most lethal of gynaecological malignancies [116]. Although this term designates a group of multiple malignant diseases sharing the same anatomical location and with different histological types [117], the poor prognosis data is mainly related to the most frequent (70%) histological subtype, HGSOC. Actually, HGSOC causes 70%–80% of gynaecological cancer-related deaths [118]. Regarding prognosis, more than 80% of HGSOC patients are diagnosed in advanced stages and the 5-year survival rates are below 50% [119,120]. This poor prognosis in HGSOC may be attributable to the asymptomatic nature, the lack of diagnostic methods in initial stages, and the presence of metastases at the time of diagnosis, although major determinants of HGSOC-related deaths are the frequent recurrences and the acquisition of chemoresistance by the tumour [121]. To this respect, slight changes have been produced in the medical treatment algorithm for HGSOC in the last decades, which relies on taxanes and cisplatin derivatives.

Despite its clinical importance, the exact aetiopathogenic mechanism of HGSOC remains elusive, since an unsolved debate on whether HGSOC arises from the ovary itself or from the fimbriae of the fallopian tubes still exists. Initially, the OSE was proposed as the focus for epithelial ovarian cancers, postulating the repeated ovulation and inclusion of cysts during regeneration as an initiation mechanism. The later theory of the fallopian tube epithelium (FTE) as a source of this tumour has gained growing attention [122], considering the premalignant lesion serous tubal intraepithelial carcinoma (STIC) as an early event. Accordingly, the tumour formed in the fallopian tube spreads to the ovary helped by the retrograde menstruation, where it is more capable of metastasizing [123] (Figure 3).

miRNAs and Oxidative Stress in High-Grade Serous Ovarian Cancer

Multiple studies based on both OS and miRNAs have reinforced an FTE origin of HGSOC. Vercellini and co-workers [124] proposed the “incessant menstruation” hypothesis, which states that pelvic macrophages decompose menstrual blood in the peritoneal cavity, and the released free-iron can damage epithelial fimbriae via ROS production. Specifically, the action of ROS on the FTE provokes a continued process of DNA damage and repair, which permits the sequential acquisition of mutations, and genomic instability. This genomic instability is represented by very frequent structural and numerical aberrations in chromosomes 3, 8, 11, 17, and 21 [125] and the mutations in HGSOC-driver genes, breast cancer gene 1 and 2 (*BRCA1/2*), *TP53*, or *PTEN/PIK3CA* [126] (Figure 3). Additionally, the loss of function of these genes has been involved with an impaired defence against OS in these tumours [127], what determines a positive feed-back loop. Nevertheless, we cannot withstand that 17% of HGSOC patients carry germline mutations in *BRCA1/2* [128].

From an epigenetic standpoint, DNA methylation [129] and miRNA [130] studies endorse the fallopian tube origin of HGSOC. For the former, Klinkebiel and co-workers [129] examined a small cohort of paired HGSOC, FTE, and OSE found that DNA methylomes are more highly conserved between HGSOC and FTE than between HGSOC and OSE. For the latter, Yang and co-workers [130] examined the expression of the miR-200 family (i.e., miR-200a, -200b, -141, and -429) and miR-205, their target genes and downstream effectors in a panel of HGSOC, STIC, FTE, and OSE tissues. As a result, the authors observed an over-expression of miR-200 family in HGSOC, STIC, and FTE and an increase of the epithelial phenotype through down-regulation of the target genes *ZEB1*, *ZEB2*, *TGFβ1*, and *TGFβ2*. These effects were not observed in OSE. Interestingly, pre-miR-200 transfection in FTE cells increased the levels of CA-125, recapitulating the high expression of this mucin in HGSOC.

Compelling literature shows evidence that epithelial-mesenchymal transition (EMT), a process by which epithelial cells lose their characteristic organization and acquire the motility of mesenchymal cells, plays a central role in HGSOc tumour progression and chemoresistance acquisition [131,132]. Notably, several miRNAs are among their master regulators. Boac and collaborators [133] serially treated four ovarian cancer cell lines (A2780CP, A2780S, IGROV1, and OVCAR5) with six cycles of cisplatin, and assessed the miRNA patterns in each of the treatment-recovery cycles. They identified five known miRNAs positively (namely miR-496, miR-485-5p, let-7g, and miR-152) or negatively (miR-27b) correlated with cisplatin chemoresistance, being the modulated pathways mainly involved in EMT regulation. Zhu and collaborators [134] demonstrated that decreased expression of miR-186 is associated with increased cisplatin resistance in HGSOc patients. Remarkably, decreased miR-186 levels up-regulate those of their target Twist1, an EMT driver, promoting the mesenchymal phenotype. Hellman and collaborators [135] analysed nine studies involving gene data sets to discover pathways associated with platinum resistance in ovarian cancer. Interestingly, despite the low degree of gene overlapping due to study design heterogeneity and technology employed, pathways related to OS (“oxidative stress”, “oxidative stress response mediated by nuclear factor (NF)-E2-related factor 2”) and to EMT (“TGFbeta signalling”, “cell migration”, “cellular movement”, and “cell-to-cell signalling”) were among the most over-represented in the studied datasets. In addition, components of the miR-17-92 cluster, which down-regulates two key TGFβ signalling molecules, and let-7 family members were also associated with platinum resistance in these analyses. Finally, Brozovic and collaborators [136] found that decreased miR-200s (miR-200a, miR200b, miR-200c, miR-429, and miR-141) expression is associated with a partial EMT phenotype in the ovarian cancer paclitaxel resistant cell lines OVCAR-3/TP and MES-OV/TP. Consistently, miR-200c and miR-141 inhibition increased the mesenchymal phenotype and the resistance to paclitaxel in non-resistant OVCAR-3 cell lines. As expected, miR-200c and miR-141 over-expression sensitized MES-OV/TP cells to paclitaxel through a mesenchymal-to-epithelial transition, and increased the levels of a set of redox enzymes, mainly reductases. It is important to mention that oxidative stress induces the expression of both miR-141 and miR-200c [137] (Table 4, Figure 2A). As can be observed, the vast majority of studies regarding the miRNA regulation of EMT have been developed in cell cultures. Although they represent a valuable source of information, established cell lines do not completely mirror the biological complexity of a tumour tissue sample. At the light of the importance of the EMT phenomenon in HGSOc patients’ prognosis, it becomes clear that there is a need for major number of studies in HGSOc tissue specimens that would increase the knowledge about the miRNA regulation of the EMT.

Table 4. Deregulated miRNAs in selected studies in HGSOc.

Reference	Effect	Experimental Design	Main Deregulated miRNAs in Patients
[130]	Susceptibility to oncogenic mutations and histologic differentiation. FTE cells increase CA-125 upon pre-miR-200 transfection.	HGSOc, STIC and FTE vs. OSE	↑ miR-200a, miR-200b, miR-141 and miR-429 and miR-205
[133]	Five miRNAs associated with cisplatin resistance EMT phenotype associated with higher chemoresistance Two pathways associated with overall patient survival (TGF/WNT and Regulation of EMT)	Four ovarian cancer cell lines, public ovarian cancer dataset	Positively correlated namely miR-496, miR-485-5p, let-7g and miR-152 Negatively correlated miR-27b
[134]	EMT phenotype, cisplatin resistance and worse prognosis	HGSOc tissue and ovarian cancer cell lines (chemosensitive and chemoresistant)	↓ miR-186, ↑ miR-200 family (significantly miR-141 and miR-200a)
[135]	Platinum resistance, related to EMT and stemness	Exploratory study based on nine published gene sets associated with platinum resistance in ovarian cancer.	↓ miR-17-92 cluster, let-7 family members

Table 4. Cont.

Reference	Effect	Experimental Design	Main Deregulated miRNAs in Patients
[136]	Stronger EMT phenotype and paclitaxel resistance	Two ovarian cancer cell lines (sensitive and resistant to paclitaxel and carboplatin)	↓ miR-200s (miR-200a, miR200b, miR-200c, miR-429 and miR-141)
[138]	Increased chemoresistance by regulation of the VEGFB and VEGFR2 pathway	198 serous epithelial ovarian carcinomas, six epithelial ovarian carcinoma cell lines	↓ miR-484 (tumour angiogenesis), miR-642, miR-217
[139]	Poor prognosis, increased paclitaxel resistance	HGSOC tissues relative to normal control tissues. Paclitaxel resistant ovarian cell lines.	↓ miR-136
[140]	Decreased cisplatin resistance by PARP1 regulation	Cisplatin-resistant and cisplatin-sensitive ovarian cancer cell lines	↓ miR-216b
[141]	Longer progression-free survival (PFS), increased platinum sensitivity to cisplatin and PARP inhibitors by directly targeting BRCA1	Serous ovarian cancer patients and tumour xenografts	↑ miR-9

↑, up-regulated levels; ↓, down-regulated levels; BRCA1, Breast Cancer type 1 susceptibility protein; EMT, epithelial-to-mesenchymal transition; VEGFB, vascular endothelial growth factor B; VEGFR2, vascular endothelial growth factor receptor 2; FTE, Fallopian Tube Epithelial; HGSOC, high-grade serous ovarian cancer; OSE, ovarian surface epithelium; PARP1, Poly [ADP-ribose] polymerase 1; STIC, Serous Tubal Intraepithelial Carcinoma, TGF, Transforming Growth Factor; Wnt, Wingless-related integration site.

Apart from EMT, Vecchione and collaborators [138] analysed the miRNA expression profiles in 198 HGSOC patients and validated a signature of three miRNAs (miR-484, miR-642 and miR-217) involved in chemoresistance, of which, miR-484 was associated with angiogenesis regulation. In addition, Jeong and co-workers [139] observed that miR-136 behaves as a TSG and that miR-136 down-regulation is associated with poor overall results in HGSOC patients. Specifically, miR-136 targets Notch3, and miR-136 over-expression re-sensitized paclitaxel-resistant ovarian cancer cells and significantly reduced cell viability, proliferation, cancer stem cell spheroid formation, and angiogenesis, as well as increased apoptosis when compared with the effects of isolated paclitaxel treatment.

As aforementioned, first-line chemotherapy schemes (cis-platin derivatives and taxanes) in HGSOC exert their anticancer effect partially mediated by increased oxidative stress. Accordingly, Ayyagari and co-workers [142] showed a synergistical effect on reduced cell viability and increased apoptosis when ovarian cancer cell lines were simultaneously treated with the anti-parasitic drug bithionol and paclitaxel, and this effect was attributable to an increase in intracellular ROS production. These findings are in agreement with previous studies from the same research group considering the combination of bithionol and cis-platin [143].

On the other hand, several PARP inhibitors (namely olaparib, niraparib, and rucaparib) have been approved for the treatment of HGSOC. These drugs act by preventing the poly [ADP-ribose] polymerase (PARP)-mediated repair of DNA damage and are especially effective in *BRCA1/2* mutation carriers. In this respect, Hou and collaborators [144] observed that the anti-tumour effect of PARP is mediated by increased ROS production and that antioxidant treatment with *N*-acetylcysteine rescued the effect. Notably, several miRNAs have been associated with PARP inhibitors effectiveness. In vitro studies have linked miR-622 and miR-493-5p over-expression with platinum and PARP inhibitor resistance in *BRCA1* and *BRCA2* mutated ovarian cancer cell lines, respectively [145,146].

A growing body of evidence suggests that OS is involved in the acquisition of chemoresistance in HGSOC as the tumour develops (Figure 3). Belotte and co-workers [147] reported that chemoresistant MDAH-2774 and SKOV-3 ovarian cancer cell lines display a pro-oxidant state, with reduced expression of the antioxidant enzyme glutathione reductase and increased expression of reactive nitrogen species nitrate/nitrite and their synthesizing enzyme iNOs. One step further, Fletcher and co-workers [148] observed that first-line chemotherapy agents induce point mutations in key redox enzymes, allowing a pro-oxidant state in ovarian cancer cells and favouring chemoresistance. Specifically, the authors observed decreased levels of SOD2, cytochrome b-245 alpha chain (CYBA, a NADPH oxidase subunit)

and glutathione reductase and an increase in iNOS activity, nitrate/nitrite levels and glutathione peroxidase in chemoresistant cells. As expected, the combination of SOD with chemotherapy (cisplatin or taxanes) significantly increased the sensitivity to chemotherapy. In addition, miR-216b increases cisplatin sensitivity by directly targeting PARP1 [140] whilst miR-9 increases sensitivity to cisplatin and PARP inhibitor by directly targeting BRCA1 [141].

Regarding the antioxidant response, Pei and co-workers [149] observed a significant inhibition of cell adhesion, migration, invasion, metastasis, and oxidative stress levels in SKOV-3 cells when treated with the antioxidant bisdemethoxycurcumin. In an interesting approach, Pons and collaborators [150] wondered whether the initial activation state of the antioxidant response could influence patients' outcomes. The authors observed a significant reduction of the antioxidant enzymes glutathione reductase and catalase, as well as of the uncoupling proteins (UCP) UCP2 and UCP5 in HGSOc patients resistant to carboplatin/paclitaxel. Nevertheless, the small cohort assessed and the few markers of OS preclude further conclusions on this study.

Altogether, it seems clear that whichever the origin of HGSOc, both OS and miRNAs play a crucial role in its initiation, promotion, and progression, including chemotherapy outcomes.

5. Potential Role of miRNAs and Oxidative Stress in Diagnosis and Treatment of Endometriosis, EAOc and HGSOc

As expected, the opportunity that alterations in both miRNAs and OS markers confer as potential biomarkers of disease has not been overlooked by researchers. However, the small number of patients included in the majority of studies and the high variability in the assessed analytes preclude any molecule being proposed as a reliable biomarker at this point. A common finding in studies evaluating OS as biomarkers for endometriosis is an increase in either plasma or serum oxidative stress biomarkers [151,152] and reduced levels of thiols [153,154] in patients in comparison to control women. In addition, other authors evaluated urine as a source of biomarkers, observing higher concentration of metabolites related to inflammation and oxidative stress (namely *N*(1)-methyl-4-pyridone-5-carboxamide, guanidinosuccinate, creatinine, taurine, valine, and 2-hydroxyisovalerate) in patients with endometriosis [155]. In EAOc, two studies demonstrated the utility of examining OS-related molecules as prognosis biomarkers. Amano and collaborators [114] observed that increased levels of SOD2 in EAOc specimens associated with worse prognosis (overall survival and progression-free survival). Additionally, protease-activated receptor-2 (PAR-2) expression, which is up-regulated by OS, correlated with shorter survival in OCCc specimens [93]. Further immunohistochemical analyses also revealed that higher 8-OHdG levels is associated with poor differentiation, higher stage, and non-optimal surgical outcomes in epithelial ovarian cancer, including HGSOc, EOC, and OCCc specimens [156]. Importantly, these observations might have a counterpart in peripheral markers, since higher serum 8-OHdG levels were associated with poor prognosis and platinum resistance in epithelial ovarian cancers, especially in EOC patients [157]. Similar results were also obtained in stage I-II epithelial ovarian cancer studies [158].

Interestingly, miRNAs can be found as circulating miRNAs in a number of biofluids, including blood, urine, and peritoneal fluid [159]. Although far from the scope of this review, several mechanisms explain the higher stability of circulating miRNAs in these biofluids [160], which make them attractive biomarkers in a myriad of pathologies, including gynaecological conditions. A limited number of studies have explored the potential of circulating miRNAs as non-invasive biomarkers for endometriosis [67–73]. As a matter of fact, slight reproducibility has been found among studies and neither a single miRNA nor a combination of them has demonstrated a higher performance when compared to current diagnostic techniques. These differences in results might find a rationale in the type of blood sample analyzed (either serum or plasma), the distinct stages of endometriosis considered and the heterogeneous cohort considered as control. Cho and co-workers [67] found decreased levels of let-7b and miR-135 in serum samples from patients with endometriosis. Wang and co-workers [68] observed decreased levels of miR-9 *, miR-141 *, miR-145 * and miR-542-3p * and increased levels of miR-122 and miR-199a in sera

from patients in comparison to control women. Interestingly, these results considering miR-122 and miR-199a have been recently corroborated by other authors [69]. Wang and co-workers [70] performed a deep sequencing approach and qRT-PCR validation to determine that down-regulated miR-30c-5p, miR127-3p, miR-99b-5p, miRNA-15b-5p, and miRNA-20a-5p and up-regulated miR-424-3p and miR-185-5p could be putative biomarkers of the disease. Regarding plasma samples, decreased levels of miR-17-5p, miR-20a, and miR-22 [71], decreased levels of miR-200a-3p, miR-200b-3p, miR-414-3p [72] and increased levels of miR-154-5p [73] have been proposed as biomarkers of endometriosis. Perhaps due to the scarcity of cases, only one study reports circulating miRNA profiles in EAO patients [74]. Suryawanshi and collaborators identified three distinct miRNA signatures in plasma capable of discriminating among patients with EAO, endometriosis and healthy individuals. Interestingly, four miRNAs distinguishing EAO patients from healthy women (namely miR-15b, -16, -21, and -195) also discriminated cancer and control mice in a pre-clinical murine model. Finally, several recent studies have evidenced the utility of miRNAs as biomarkers in HGSO. Kobayashi and co-workers [75] observed that miR-1290 is elevated in sera from HGSO patients in comparison to control women, but not in ovarian cancers of other histological types, and that its expression correlates with tumour burden. Shah and co-workers [76] observed that the combination of sera miR-375 and CA-125 is a diagnostic biomarker of HGSO. Todeschini and collaborators [77] employed two independent patient cohorts to identify miR-1246 as the best diagnostic marker in HGSO. Finally, Kan and collaborators [78] built a predictive model including sera levels of miR-200b and miR-200c, with an area under the curve of 0.784, distinguishing HGSO patients from control individuals (Table 2).

With respect to treatment, a number of studies have considered the antioxidant treatment in endometriosis, based on supplementation with Vitamin C and E [161], or preparations with different antioxidants [162,163]. As a result, peripheral oxidative stress markers diminished [161], and the pain symptoms were improved [162,163] albeit without any benefit on pregnancy rates [161]. In the last 10 years, a few clinical trials have evaluated the utility of oxidative-stress based treatments for HGSO, either aiming to increase oxidative stress or employing antioxidants. Monk and co-workers report a very recent two-stage phase II clinical trial, which failed to provide any benefit in objective tumour responses in a panel of platinum-resistant recurrent ovarian, tubal and peritoneal cancer patients treated with i.v. elesclomol, a ROS inducer, plus weekly paclitaxel in comparison to paclitaxel alone [164]. This observation might be in agreement with the involvement of ROS in the acquisition of chemoresistance by the tumour. In agreement with this rationale, a recent phase II trial evaluated the combination of the vitamin E analogue, delta tocotrienol, with bevacizumab in 23 patients with refractory ovarian cancer. The combination produced an improvement in progression-free survival (median 6.9 months) and overall survival (median 10.9 months) with regards to the data in current literature [165]. Trudel and co-workers [166] performed a two-stage, single-arm, phase II study (NCT00721890) of the tea drink enriched with the polyphenol epigallocatechin gallate (EGCG) for maintenance treatment in advanced ovarian cancer. Sixteen participants (13 HGSO and three EOC) were included in the study since they were in complete remission after completion of their first line treatment and were followed for 18 months thereafter. Unfortunately, daily nutritional intervention with 500mL of the drink failed to prove any benefit regarding recurrence improvement at 18 months.

Regarding miRNAs, the vast literature involving the outstanding role of miRNAs in the three considered gynaecological conditions pave the way for a miRNA-based therapy in order to restore deregulated miRNA levels. Indeed, pathologically down- or over-expressed levels of a given miRNA could be in vitro restored by the employment of miRNA mimics or antimiRs, respectively. Although a limited number of miRNAs has been tested in clinical trials (i.e., anti-miR-122 for Hepatitis C, antimir-103/107 for type 2 diabetes and non-alcoholic fatty liver diseases, antimir-155 for cutaneous T cell lymphoma and mycosis fungoides, miR-29 mimic for scleroderma, miR-16 mimic for mesothelioma and non-small cell lung cancer, and miR-34 mimics for multiple solid tumors (reviewed in [167]), none of them have reached phase III clinical trials and there is no single ongoing trial considering gynecological diseases, to the best of our knowledge. Several limitations might provide a rationale

for the lack of translation of miRNA-based therapeutics, as the high probability of off-targets cellular and systemic effects (provided the multiple mRNAs targeted by a single miRNA and the difficulty of delivering miRNA therapies to a specific organ or even to a specific cellular type, respectively). However, a major limitation might lay in the lack of knowledge of the dynamic expression of miRNA patterns and the overall effect produced due to the interaction among them, since researchers usually only have access to a still picture of disease specimens.

6. Conclusions

In conclusion, extensive literature shows that there exists an interplay between miRNAs and oxidative stress in several gynaecological conditions, highlighting endometriosis, EAO and HGSO. In endometriosis, oxidative stress and miRNAs contribute to the establishment and development of endometriotic lesions. Regarding OMA, repeated ROS stimulation and miRNA deregulation on the cells of the cystic wall seem to play an important role in the malignant transformation of endometriosis. Besides, a mechanistic model for oxidative stress and miRNAs in OMA malignant transformation may be established: Within endometrial cysts, oxidative stress might be involved in the carcinogenesis of EAO. Initially, repeated intra-cystic bleeding generates ROS and OS that act on the cystic wall cells. To cope with this adverse event, OMA epithelial cells increase their antioxidant response, but eventually are overwhelmed by repeated ROS, producing genetic and epigenetic alterations in crucial tumour suppressor genes. Additionally, miRNAs can contribute not only to the loss of function of these tumour suppressor genes but also to important carcinogenic events. Yet, once EAO is established, this increased antioxidant response diminishes the platinum-mediated ROS injury and the efficiency of the chemotherapy treatment, which is also modulated by specific miRNAs. Finally, oxidative stress and miRNA deregulation is involved in the carcinogenesis of HGSO and crucially influences the response to first-line chemotherapeutics, both regarding initial treatment outcomes and acquisition of chemoresistance.

With respect to diagnosis, increased circulating levels of OS markers have been involved with endometriosis diagnosis, and increased circulating levels of SOD and OS markers have been associated with poor prognosis in EAO and HGSO. Additionally, several studies have proven the putative role of miRNAs as biomarkers of these three gynaecological conditions. Regarding therapeutics, antioxidant treatment in endometriosis seems to improve the associated pain and the combination of antioxidants with bevacizumab is a promising approach in refractory ovarian cancer patients. On the other hand, miRNA treatment is still in its infancy, with few ongoing clinical trials and none of them in gynaecological diseases. The difficulty in developing miRNA-based therapies could be related to the likely off-target effects and the lack of knowledge of the precise dynamics and interactions of miRNAs throughout the disease development.

Altogether, future research endeavours are guaranteed to enlarge the knowledge on the action of miRNAs and oxidative stress in the pathophysiology of these important gynaecological pathologies and to propose targeted therapeutic strategies to deal with their pernicious effect.

Author Contributions: J.M.-A. conceived and designed the manuscript, revised the literature and wrote the manuscript. A.P.C. revised the literature and drafted the manuscript. C.A. and J.G.-O. contributed to literature revision and revised the manuscript. A.M.-M. and S.C.-F. prepared the figures and tables and revised the manuscript. J.G.-E. has written the manuscript.

Funding: This work was supported by grants from ISCIII and FEDER (PI17/01945). J. M.-A. is supported by a post-doctoral grant (APOSTD/2019/087) and A.M.-M. is supported by PhD scholarship from Asociación Española Contra el Cáncer (AECC).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Donnez, J.; Binda, M.M.; Donnez, O.; Dolmans, M.M. Oxidative stress in the pelvic cavity and its role in the pathogenesis of endometriosis. *Fertil. Steril.* **2016**, *106*, 1011–1017. [[CrossRef](#)] [[PubMed](#)]

2. Dröge, W. Free radicals in the physiological control of cell function. *Physiol. Rev.* **2002**, *82*, 47–95. [[CrossRef](#)] [[PubMed](#)]
3. Sies, H. Oxidative stress: A concept in redox biology and medicine. *Redox Biol.* **2015**, *4*, 180–183. [[CrossRef](#)] [[PubMed](#)]
4. Rockfield, S.; Raffel, J.; Mehta, R.; Rehman, N.; Nanjundan, M. Iron overload and altered iron metabolism in ovarian cancer. *Biol. Chem.* **2017**, *398*, 995–1007. [[CrossRef](#)]
5. Dixon, S.J.; Stockwell, B.R. The role of iron and reactive oxygen species in cell death. *Nat. Chem. Biol.* **2014**, *10*, 9–17. [[CrossRef](#)] [[PubMed](#)]
6. Sies, H.; Berndt, C.; Jones, D.P. Oxidative Stress. *Annu. Rev. Biochem.* **2017**, *86*, 715–748. [[CrossRef](#)] [[PubMed](#)]
7. Bokov, A.; Chaudhuri, A.; Richardson, A. The role of oxidative damage and stress in aging. *Mech. Ageing Dev.* **2004**, *125*, 811–826. [[CrossRef](#)] [[PubMed](#)]
8. Sies, H. *Oxidative Stress: Introductory Remarks*; Academic Press: London, UK, 1985; pp. 1–8.
9. Inglés, M.; Gambini, J.; Carnicero, J.A.; García-García, F.J.; Rodríguez-Mañas, L.; Olaso-González, G.; Dromant, M.; Borrás, C.; Viña, J. Oxidative stress is related to frailty, not to age or sex, in a geriatric population: Lipid and protein oxidation as biomarkers of frailty. *J. Am. Geriatr. Soc.* **2014**, *62*, 1324–1328. [[CrossRef](#)]
10. Sies, H.; Jones, D. Oxidative stress. In *Encyclopedia of Stress*, 2nd ed.; Fink, G., Ed.; Elsevier: Amsterdam, The Netherlands, 2007; pp. 45–48.
11. Smith, M.A.; Perry, G.; Richey, P.L.; Sayre, L.M.; Anderson, V.E.; Beal, M.F.; Kowall, N. Oxidative damage in Alzheimer's. *Nature* **1996**, *382*, 120–121. [[CrossRef](#)]
12. Fincham, J. *Epigenetic Mechanisms of Gene Regulation*; Russo, V.E.A., Martienssen, R.A., Riggs, A.D., Eds.; Cold Spring Harbor Laboratory: Cold Spring Harbor, NY, USA, 1997; pp. 159–162.
13. Portela, A.; Esteller, M. Epigenetic modifications and human disease. *Nat. Biotechnol.* **2010**, *28*, 1057–1068. [[CrossRef](#)]
14. Guo, S.W. Epigenetics of endometriosis. *Mol. Hum. Reprod.* **2009**, *15*, 587–607. [[CrossRef](#)] [[PubMed](#)]
15. Esteller, M. Epigenetics in cancer. *N. Engl. J. Med.* **2008**, *358*, 1148–1159. [[CrossRef](#)] [[PubMed](#)]
16. Lee, R.C.; Feinbaum, R.L.; Ambros, V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* **1993**, *75*, 843–854. [[CrossRef](#)]
17. Lee, Y.; Ahn, C.; Han, J.; Choi, H.; Kim, J.; Yim, J.; Lee, J.; Provost, P.; Rådmark, O.; Kim, S.; et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* **2003**, *425*, 415–419. [[CrossRef](#)]
18. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* **2004**, *116*, 281–297. [[CrossRef](#)]
19. Caporali, A.; Emanuelli, C. MicroRNA regulation in angiogenesis. *Vascul. Pharmacol.* **2011**, *55*, 79–86. [[CrossRef](#)]
20. Gailhouste, L.; Ochiya, T. Cancer-related microRNAs and their role as tumor suppressors and oncogenes in hepatocellular carcinoma. *Histol. Histopathol.* **2013**, *28*, 437–451.
21. Santamaria, X.; Taylor, H. MicroRNA and gynecological reproductive diseases. *Fertil. Steril.* **2014**, *101*, 1545–1551. [[CrossRef](#)]
22. Marí-Alexandre, J.; Sánchez-Izquierdo, D.; Gilabert-Estellés, J.; Barceló-Molina, M.; Braza-Boils, A.; Sandoval, J. miRNAs Regulation and Its Role as Biomarkers in Endometriosis. *Int. J. Mol. Sci.* **2016**, *17*, E93. [[CrossRef](#)]
23. Borghese, B.; Zondervan, K.T.; Abrao, M.S.; Chapron, C.; Vaiman, D. Recent insights on the genetics and epigenetics of endometriosis. *Clin. Genet.* **2017**, *91*, 254–264. [[CrossRef](#)]
24. Saare, M.; Rekker, K.; Laisk-Podar, T.; Rahmioglu, N.; Zondervan, K.; Salumets, A.; Götte, M.; Peters, M. Challenges in endometriosis miRNA studies-From tissue heterogeneity to disease specific miRNAs. *Biochim. Biophys. Acta* **2017**, *1863*, 2282–2292. [[CrossRef](#)] [[PubMed](#)]
25. He, J.; Chang, W.; Feng, C.; Cui, M.; Xu, T. Endometriosis Malignant Transformation: Epigenetics as a Probable Mechanism in Ovarian Tumorigenesis. *Int. J. Genom.* **2018**, *2018*, 1465348. [[CrossRef](#)] [[PubMed](#)]
26. Siufi Neto, J.; Kho, R.M.; Siufi, D.F.; Baracat, E.C.; Anderson, K.S.; Abrão, M.S. Cellular, histologic, and molecular changes associated with endometriosis and ovarian cancer. *J. Minim. Invasive Gynecol.* **2014**, *21*, 55–63. [[CrossRef](#)] [[PubMed](#)]
27. Sun, Y.; Guo, F.; Bagnoli, M.; Xue, F.X.; Sun, B.C.; Shmulevich, I.; Mezzanzanica, D.; Chen, K.X.; Sood, A.K.; Yang, D.; et al. Key nodes of a microRNA network associated with the integrated mesenchymal subtype of high-grade serous ovarian cancer. *Chin. J. Cancer.* **2015**, *34*, 28–40. [[CrossRef](#)]

28. Mittempergher, L. Genomic Characterization of High-Grade Serous Ovarian Cancer: Dissecting Its Molecular Heterogeneity as a Road Towards Effective Therapeutic Strategies. *Curr. Oncol. Rep.* **2016**, *18*, 44. [[CrossRef](#)]
29. Samuel, P.; Pink, R.C.; Brooks, S.A.; Carter, D.R. miRNAs and ovarian cancer: A miRiad of mechanisms to induce cisplatin drug resistance. *Expert Rev. Anticancer Ther.* **2016**, *16*, 57–70. [[CrossRef](#)]
30. Lan, J.; Huang, Z.; Han, J.; Shao, J.; Huang, C. Redox regulation of microRNAs in cancer. *Cancer Lett.* **2018**, *418*, 250–259. [[CrossRef](#)]
31. He, J.; Jiang, B.H. Interplay between Reactive oxygen Species and MicroRNAs in Cancer. *Curr. Pharmacol. Rep.* **2016**, *2*, 82–90. [[CrossRef](#)]
32. Cheng, X.; Ku, C.H.; Siow, R.C. Regulation of the Nrf2 antioxidant pathway by microRNAs: New players in micromanaging redox homeostasis. *Free Radic. Biol. Med.* **2013**, *64*, 4–11. [[CrossRef](#)]
33. Giudice, L.C.; Kao, L.C. Endometriosis. *Lancet* **2004**, *364*, 1789–1799. [[CrossRef](#)]
34. Giudice, L.C. Endometriosis. *N. Engl. J. Med.* **2010**, *362*, 2389–2398. [[CrossRef](#)] [[PubMed](#)]
35. Tamaresis, J.S.; Irwin, J.C.; Goldfien, G.A.; Rabban, J.T.; Burney, R.O.; Nezhat, C.; DePaolo, L.V.; Giudice, L.C. Molecular classification of endometriosis and disease stage using high-dimensional genomic data. *Endocrinology* **2014**, *155*, 4986–4999. [[CrossRef](#)] [[PubMed](#)]
36. Burney, R.O.; Giudice, L.C. Pathogenesis and pathophysiology of endometriosis. *Fertil. Steril.* **2012**, *98*, 511–519. [[CrossRef](#)] [[PubMed](#)]
37. Huang, H.; Li, C.; Zarogoulidis, P.; Darwiche, K.; Machairiotis, N.; Yang, L.; Simoff, M.; Celis, E.; Zhao, T.; Zarogoulidis, K.; et al. Endometriosis of the lung: Report of a case and literature review. *Eur. J. Med. Res.* **2013**, *18*, 13. [[CrossRef](#)] [[PubMed](#)]
38. Thibodeau, L.L.; Prioleau, G.R.; Manuelidis, E.E.; Merino, M.J.; Heafner, M.D. Cerebral endometriosis. Case report. *J. Neurosurg.* **1987**, *66*, 609–610. [[CrossRef](#)] [[PubMed](#)]
39. Sampson, J.A. Metastatic or Embolic Endometriosis, due to the Menstrual Dissemination of Endometrial Tissue into the Venous Circulation. *Am. J. Pathol.* **1927**, *3*, 93–110.
40. Halme, J.; Hammond, M.G.; Hulka, J.F.; Raj, S.G.; Talbert, L.M. Retrograde menstruation in healthy women and in patients with endometriosis. *Obstet. Gynecol.* **1984**, *64*, 151–154.
41. Koninckx, P.R.; Kennedy, S.H.; Barlow, D.H. Pathogenesis of endometriosis: The role of peritoneal fluid. *Gynecol. Obstet. Investig.* **1999**, *47*, 23–33. [[CrossRef](#)]
42. Scutiero, G.; Iannone, P.; Bernardi, G.; Bonaccorsi, G.; Spadaro, S.; Volta, C.A.; Greco, P.; Nappi, L. Oxidative Stress and Endometriosis: A Systematic Review of the Literature. *Oxid. Med. Cell. Longev.* **2017**, *2017*, 7265238. [[CrossRef](#)]
43. Carvalho, L.F.; Samadder, A.N.; Agarwal, A.; Fernandes, L.F.; Abrão, M.S. Oxidative stress biomarkers in patients with endometriosis: Systematic review. *Arch. Gynecol. Obstet.* **2012**, *286*, 1033–1040. [[CrossRef](#)]
44. Ramón, L.A.; Braza-Boils, A.; Gilabert-Estellés, J.; Gilabert, J.; España, F.; Chirivella, M.; Estellés, A. microRNAs expression in endometriosis and their relation to angiogenic factors. *Hum. Reprod.* **2011**, *26*, 1082–1090. [[CrossRef](#)] [[PubMed](#)]
45. Braza-Boils, A.; Mari-Alexandre, J.; Gilabert, J.; Sánchez-Izquierdo, D.; España, F.; Estellés, A.; Gilabert-Estellés, J. MicroRNA expression profile in endometriosis: Its relation to angiogenesis and fibrinolytic factors. *Hum. Reprod.* **2014**, *29*, 978–988. [[CrossRef](#)] [[PubMed](#)]
46. Gilabert-Estellés, J.; Ramón, L.A.; España, F.; Gilabert, J.; Vila, V.; Réganon, E.; Castelló, R.; Chirivella, M.; Estellés, A. Expression of angiogenic factors in endometriosis: Its relation to fibrinolytic and metalloproteinase (MMP) systems. *Hum. Reprod.* **2007**, *22*, 2120–2127. [[CrossRef](#)] [[PubMed](#)]
47. Mari-Alexandre, J.; Barceló-Molina, M.; Belmonte-López, E.; García-Oms, J.; Estellés, A.; Braza-Boils, A.; Gilabert-Estellés, J. Micro-RNA profile and proteins in peritoneal fluid from women with endometriosis: Their relationship with sterility. *Fertil. Steril.* **2018**, *109*, 675–684. [[CrossRef](#)]
48. Gee, H.E.; Ivan, C.; Calin, G.A.; Ivan, M. HypoxamiRs and cancer: From biology to targeted therapy. *Antioxid. Redox Signal.* **2014**, *21*, 1220–1238. [[CrossRef](#)]
49. Xu, T.X.; Zhao, S.Z.; Dong, M.; Yu, X.R. Hypoxia responsive miR-210 promotes cell survival and autophagy of endometriotic cells in hypoxia. *Eur. Rev. Med. Pharmacol. Sci.* **2016**, *20*, 399–406.
50. Augustin, H.G. Vascular morphogenesis in the ovary. *Baillieres Best Pract. Res. Clin. Obstet. Gynaecol.* **2000**, *14*, 867–882. [[CrossRef](#)]

51. Eggers, J.C.; Martino, V.; Reinbold, R.; Schäfer, S.D.; Kiesel, L.; Starzinski-Powitz, A.; Schüring, A.N.; Kemper, B.; Greve, B.; Götte, M. microRNA miR-200b affects proliferation, invasiveness and stemness of endometriotic cells by targeting ZEB1, ZEB2 and KLF4. *Reprod. Biomed. Online* **2016**, *32*, 434–445. [[CrossRef](#)]
52. Chen, J.; Gu, L.; Ni, J.; Hu, P.; Hu, K.; Shi, Y.L. MiR-183 Regulates ITGB1P Expression and Promotes Invasion of Endometrial Stromal Cells. *Biomed. Res. Int.* **2015**, *2015*, 340218. [[CrossRef](#)]
53. Dai, L.; Gu, L.; Di, W. MiR-199a attenuates endometrial stromal cell invasiveness through suppression of the IKK β /NF- κ B pathway and reduced interleukin-8 expression. *Mol. Hum. Reprod.* **2012**, *18*, 136–145. [[CrossRef](#)]
54. Okamoto, M.; Nasu, K.; Abe, W.; Aoyagi, Y.; Kawano, Y.; Kai, K.; Moriyama, M.; Narahara, H. Enhanced miR-210 expression promotes the pathogenesis of endometriosis through activation of signal transducer and activator of transcription 3. *Hum. Reprod.* **2015**, *30*, 632–641. [[CrossRef](#)]
55. Yu, H.; Zhong, Q.; Xia, Y.; Li, E.; Wang, S.; Ren, R. MicroRNA-2861 targets STAT3 and MMP2 to regulate the proliferation and apoptosis of ectopic endometrial cells in endometriosis. *Pharmazie* **2019**, *74*, 243–249. [[PubMed](#)]
56. Wang, Y.; Chen, H.; Fu, Y.; Ai, A.; Xue, S.; Lyu, Q.; Kuang, Y. MiR-195 inhibits proliferation and growth and induces apoptosis of endometrial stromal cells by targeting FKN. *Int. J. Clin. Exp. Pathol.* **2013**, *15*, 2824–2834.
57. Abe, W.; Nasu, K.; Nakada, C.; Kawano, Y.; Moriyama, M.; Narahara, H. miR-196b targets c-myc and Bcl-2 expression, inhibits proliferation and induces apoptosis in endometriotic stromal cells. *Hum. Reprod.* **2013**, *28*, 750–761. [[CrossRef](#)] [[PubMed](#)]
58. Zhao, Q.; Ye, M.; Yang, W.; Wang, M.; Li, M.; Gu, C.; Zhao, L.; Zhang, Z.; Han, W.; Fan, W.; et al. Effect of Mst1 on Endometriosis Apoptosis and Migration: Role of Drp1-Related Mitochondrial Fission and Parkin-Required Mitophagy. *Cell Physiol. Biochem.* **2018**, *45*, 1172–1190. [[CrossRef](#)] [[PubMed](#)]
59. Zhang, Y.; Yan, J.; Pan, X. miR-141-3p affects apoptosis and migration of endometrial stromal cells by targeting KLF-12. *Pflugers Arch.* **2019**, *471*, 1055–1063. [[CrossRef](#)]
60. Braza-Boils, A.; Salloum-Asfar, S.; Marí-Alexandre, J.; Arroyo, A.B.; González-Conejero, R.; Barceló-Molina, M.; García-Oms, J.; Vicente, V.; Estellés, A.; Gilibert-Estellés, J.; et al. Peritoneal fluid modifies the microRNA expression profile in endometrial and endometriotic cells from women with endometriosis. *Hum. Reprod.* **2015**, *30*, 2292–2302. [[CrossRef](#)]
61. Panir, K.; Schjenken, J.E.; Robertson, S.A.; Hull, M.L. Non-coding RNAs in endometriosis: A narrative review. *Hum. Reprod. Update* **2018**, *24*, 497–515. [[CrossRef](#)]
62. Cosín, R.; Gilibert-Estellés, J.; Ramón, L.A.; Gómez-Lechón, M.J.; Gilibert, J.; Chirivella, M.; Braza-Boils, A.; España, F.; Estellés, A. Influence of peritoneal fluid on the expression of angiogenic and proteolytic factors in cultures of endometrial cells from women with endometriosis. *Hum. Reprod.* **2010**, *25*, 398–405. [[CrossRef](#)]
63. Wright, K.R.; Mitchell, B.; Santanam, N. Redox regulation of microRNAs in endometriosis-associated pain. *Redox Biol.* **2017**, *12*, 956–966. [[CrossRef](#)]
64. Yang, R.Q.; Teng, H.; Xu, X.H.; Liu, S.Y.; Wang, Y.H.; Guo, F.J.; Liu, X.J. Microarray analysis of microRNA deregulation and angiogenesis-related proteins in endometriosis. *Genet. Mol. Res.* **2016**, *15*, 1–8. [[CrossRef](#)] [[PubMed](#)]
65. Teruel-Montoya, R.; Kong, X.; Abraham, S.; Ma, L.; Kunapuli, S.P.; Holinstat, M.; Shaw, C.A.; McKenzie, S.E.; Edelstein, L.C.; Bray, P.F. MicroRNA expression differences in human hematopoietic cell lineages enable regulated transgene expression. *PLoS ONE* **2014**, *9*, e102259. [[CrossRef](#)] [[PubMed](#)]
66. Nothnick, W.B.; Swan, K.; Flyckt, R.; Falcone, T.; Graham, A. Human endometriotic lesion expression of the miR-144-3p/miR-451a cluster, its correlation with markers of cell survival and origin of lesion content. *Sci. Rep.* **2019**, *9*, 8823. [[CrossRef](#)] [[PubMed](#)]
67. Cho, S.; Mutlu, L.; Grechukhina, O.; Taylor, H.S. Circulating microRNAs as potential biomarkers for endometriosis. *Fertil Steril.* **2015**, *103*, 1252–1260. [[CrossRef](#)] [[PubMed](#)]
68. Wang, W.T.; Zhao, Y.N.; Han, B.W.; Hong, S.J.; Chen, Y.Q. Circulating microRNAs identified in a genome-wide serum microRNA expression analysis as non-invasive biomarkers for endometriosis. *J. Clin. Endocrinol. Metab.* **2013**, *98*, 281–289. [[CrossRef](#)]
69. Maged, A.M.; Deeb, W.S.; El Amir, A.; Zaki, S.S.; El Sawah, H.; Al Mohamady, M.; Metwally, A.A.; Katta, M.A. Diagnostic accuracy of serum miR-122 and miR-199a in women with endometriosis. *Int. J. Gynaecol. Obstet.* **2018**, *141*, 14–19. [[CrossRef](#)]

70. Wang, L.; Huang, W.; Ren, C.; Zhao, M.; Jiang, X.; Fang, X.; Xia, X. Analysis of Serum microRNA Profile by Solexa Sequencing in Women with Endometriosis. *Reprod. Sci.* **2016**, *23*, 1359–1370. [[CrossRef](#)]
71. Jia, S.Z.; Yang, Y.; Lang, J.; Sun, P.; Leng, J. Plasma miR-17-5p, miR20a and miR-22 are down-regulated in women with endometriosis. *Hum. Reprod.* **2013**, *28*, 322–330. [[CrossRef](#)]
72. Rekker, K.; Saare, M.; Roost, A.M.; Kaart, T.; Sõritsa, D.; Karro, H.; Sõritsa, A.; Simón, C.; Salumets, A.; Peters, M. Circulating miR-200-family microRNAs have altered plasma levels in patients with endometriosis and vary with blood collection time. *Fertil Steril.* **2015**, *104*, 938–946. [[CrossRef](#)]
73. Pateisky, P.; Pils, D.; Szabo, L.; Kuessel, L.; Husslein, H.; Schmitz, A.; Wenzl, R.; Yotova, I. hsa-miRNA-154-5p expression in plasma of endometriosis patients is a potential diagnostic marker for the disease. *Reprod. Biomed. Online* **2018**, *37*, 449–466. [[CrossRef](#)]
74. Suryawanshi, S.; Vlad, A.M.; Lin, H.M.; Mantia-Smaldone, G.; Laskey, R.; Lee, M.; Lin, Y.; Donnellan, N.; Klein-Patel, M. Plasma microRNAs as novel biomarkers for endometriosis and endometriosis-associated ovarian cancer. *Clin. Cancer Res.* **2013**, *19*, 1213–1224. [[CrossRef](#)] [[PubMed](#)]
75. Kobayashi, M.; Sawada, K.; Nakamura, K.; Yoshimura, A.; Miyamoto, M.; Shimizu, A.; Ishida, K.; Nakatsuka, E.; Kodama, M.; Hashimoto, K.; et al. Exosomal miR-1290 is a potential biomarker of high-grade serous ovarian carcinoma and can discriminate patients from those with malignancies of other histological types. *J. Ovarian Res.* **2018**, *11*, 81. [[CrossRef](#)] [[PubMed](#)]
76. Shah, J.S.; Gard, G.B.; Yang, J.; Maidens, J.; Valmadre, S.; Soon, P.S.; Marsh, D.J. Combining serum microRNA and CA-125 as prognostic indicators of preoperative surgical outcome in women with high-grade serous ovarian cancer. *Gynecol. Oncol.* **2018**, *148*, 181–188. [[CrossRef](#)] [[PubMed](#)]
77. Todeschini, P.; Salviato, E.; Paracchini, L.; Ferracin, M.; Petrillo, M.; Zanotti, L.; Tognon, G.; Gambino, A.; Calura, E.; Caratti, G.; et al. Circulating miRNA landscape identifies miR-1246 as promising diagnostic biomarker in high-grade serous ovarian carcinoma: A validation across two independent cohorts. *Cancer Lett.* **2017**, *388*, 320–327. [[CrossRef](#)]
78. Kan, C.W.; Hahn, M.A.; Gard, G.B.; Maidens, J.; Huh, J.Y.; Marsh, D.J.; Howell, V.M. Elevated levels of circulating microRNA-200 family members correlate with serous epithelial ovarian cancer. *BMC Cancer* **2012**, *12*, 627. [[CrossRef](#)]
79. Matsuzaki, S.; Schubert, B. Oxidative stress status in normal ovarian cortex surrounding ovarian endometriosis. *Fertil. Steril.* **2010**, *93*, 2431–2432. [[CrossRef](#)]
80. Di Emidio, G.; D'Alfonso, A.; Leocata, P.; Parisse, V.; Di Fonso, A.; Artini, P.G.; Patacchiola, F.; Tatone, C.; Carta, G. Increased levels of oxidative and carbonyl stress markers in normal ovarian cortex surrounding endometriotic cysts. *Gynecol. Endocrinol.* **2014**, *30*, 808–812. [[CrossRef](#)]
81. Ngô, C.; Chéreau, C.; Nicco, C.; Weill, B.; Chapron, C.; Batteux, F. Reactive oxygen species controls endometriosis progression. *Am. J. Pathol.* **2009**, *175*, 225–234. [[CrossRef](#)]
82. Chen, C.; Zhou, Y.; Hu, C.; Wang, Y.; Yan, Z.; Li, Z.; Wu, R. Mitochondria and oxidative stress in ovarian endometriosis. *Free Radic. Biol. Med.* **2019**, *136*, 22–34. [[CrossRef](#)]
83. Kobayashi, Y.; Osanai, K.; Tanaka, K.; Nishigaya, Y.; Matsumoto, H.; Momomura, M.; Hashiba, M.; Mita, S.; Kyo, S.; Iwashita, M. Endometriotic cyst fluid induces reactive oxygen species (ROS) in human immortalized epithelial cells derived from ovarian endometrioma. *Redox Rep.* **2017**, *22*, 361–366. [[CrossRef](#)]
84. Yamaguchi, K.; Mandai, M.; Toyokuni, S.; Hamanishi, J.; Higuchi, T.; Takakura, K.; Fujii, S. Contents of endometriotic cysts, especially the high concentration of free iron, are a possible cause of carcinogenesis in the cysts through the iron-induced persistent oxidative stress. *Clin. Cancer Res.* **2008**, *14*, 32–40. [[CrossRef](#)] [[PubMed](#)]
85. Xie, H.; Chen, P.; Huang, H.W.; Liu, L.P.; Zhao, F. Reactive oxygen species downregulate ARID1A expression via its promoter methylation during the pathogenesis of endometriosis. *Eur. Rev. Med. Pharmacol. Sci.* **2017**, *21*, 4509–4515. [[PubMed](#)]
86. Santulli, P.; Chouzenoux, S.; Fiorese, M.; Marcellin, L.; Lemarechal, H.; Millischer, A.E.; Batteux, F.; Borderie, D.; Chapron, C. Protein oxidative stress markers in peritoneal fluids of women with deep infiltrating endometriosis are increased. *Hum Reprod.* **2015**, *30*, 49–60. [[CrossRef](#)] [[PubMed](#)]
87. Körner, M.; Burckhardt, E.; Mazzucchelli, L. Higher frequency of chromosomal aberrations in ovarian endometriosis compared to extragonadal endometriosis: A possible link to endometrioid adenocarcinoma. *Mod. Pathol.* **2006**, *19*, 1615–1623. [[CrossRef](#)] [[PubMed](#)]

88. Níguez-Sevilla, I.; Machado-Linde, F.; Marín-Sánchez, M.D.P.; Areñse, J.J.; Torroba, A.; Nieto-Díaz, A.; Sánchez Ferrer, M.L. Prognostic importance of atypical endometriosis with architectural hyperplasia versus cytologic atypia in endometriosis-associated ovarian cancer. *J. Gynecol. Oncol.* **2019**, *30*, e63. [[CrossRef](#)]
89. Pearce, C.L.; Templeman, C.; Rossing, M.A.; Lee, A.; Near, A.M.; Webb, P.M.; Nagle, C.M.; Doherty, J.A.; Cushing-Haugen, K.L.; Wicklund, K.G.; et al. Association between endometriosis and risk of histological subtypes of ovarian cancer: A pooled analysis of case-control studies. *Lancet Oncol.* **2012**, *13*, 385–394. [[CrossRef](#)]
90. Guo, S.W. Endometriosis and ovarian cancer: Potential benefits and harms of screening and risk-reducing surgery. *Fertil Steril.* **2015**, *104*, 813–830. [[CrossRef](#)]
91. Poole, E.M.; Lin, W.T.; Kvaskoff, M.; De Vivo, I.; Terry, K.L.; Missmer, S.A. Endometriosis and risk of ovarian and endometrial cancers in a large prospective cohort of U.S. nurses. *Cancer Causes Control.* **2017**, *28*, 437–445. [[CrossRef](#)]
92. Mandai, M.; Matsumura, N.; Baba, T.; Yamaguchi, K.; Hamanishi, J.; Konishi, I. Ovarian clear cell carcinoma as a stress-responsive cancer: Influence of the microenvironment on the carcinogenesis and cancer phenotype. *Cancer Lett.* **2011**, *310*, 129–133. [[CrossRef](#)]
93. Aman, M.; Ohishi, Y.; Imamura, H.; Shinozaki, T.; Yasutake, N.; Kato, K.; Oda, Y. Expression of protease-activated receptor-2 (PAR-2) is related to advanced clinical stage and adverse prognosis in ovarian clear cell carcinoma. *Hum Pathol.* **2017**, *64*, 156–163. [[CrossRef](#)]
94. Fujimoto, Y.; Imanaka, S.; Yamada, Y.; Ogawa, K.; Ito, F.; Kawahara, N.; Yoshimoto, C.; Kobayashi, H. Comparison of redox parameters in ovarian endometrioma and its malignant transformation. *Oncol. Lett.* **2018**, *16*, 5257–5264. [[CrossRef](#)] [[PubMed](#)]
95. Sova, H.; Kangas, J.; Puistola, U.; Santala, M.; Liakka, A.; Karihtala, P. Down-regulation of 8-hydroxydeoxyguanosine and peroxiredoxin II in the pathogenesis of endometriosis-associated ovarian cancer. *Anticancer Res.* **2012**, *32*, 3037–3044. [[PubMed](#)]
96. Winarto, H.; Tan, M.I.; Sadikin, M.; Wanandi, S.I. ARID1A Expression is Down-Regulated by Oxidative Stress in Endometriosis and Endometriosis-Associated Ovarian Cancer. *Transl. Oncogenom.* **2017**, *24*, 9.
97. Yamada, Y.; Miyamoto, T.; Kashima, H.; Kobara, H.; Asaka, R.; Ando, H.; Higuchi, S.; Ida, K.; Shiozawa, T. Lipocalin 2 attenuates iron-related oxidative stress and prolongs the survival of ovarian clear cell carcinoma cells by up-regulating the CD44 variant. *Free Radic Res.* **2016**, *50*, 414–425. [[CrossRef](#)] [[PubMed](#)]
98. Mabuchi, S.; Sugiyama, T.; Kimura, T. Clear cell carcinoma of the ovary: Molecular insights and future therapeutic perspectives. *J. Gynecol. Oncol.* **2016**, *27*, e31. [[CrossRef](#)] [[PubMed](#)]
99. Yamashita, Y.; Akatsuka, S.; Shinjo, K.; Yatabe, Y.; Kobayashi, H.; Seko, H.; Kajiyama, H.; Kikkawa, F.; Takahashi, T.; Toyokuni, S. Met is the most frequently amplified gene in endometriosis-associated ovarian clear cell adenocarcinoma and correlates with worsened prognosis. *PLoS ONE* **2013**, *8*, e57724. [[CrossRef](#)]
100. Wu, X.; Ruan, Y.; Jiang, H.; Xu, C. MicroRNA-424 inhibits cell migration, invasion, and epithelial mesenchymal transition by downregulating doublecortin-like kinase 1 in ovarian clear cell carcinoma. *Int. J. Biochem. Cell Biol.* **2017**, *85*, 66–74. [[CrossRef](#)]
101. Hsu, C.Y.; Hsieh, T.H.; Er, T.K.; Chen, H.S.; Tsai, C.C.; Tsai, E.M. MiR-381 regulates cell motility, growth and colony formation through PIK3CA in endometriosis-associated clear cell and endometrioid ovarian cancer. *Oncol. Rep.* **2018**, *40*, 3734–3742. [[CrossRef](#)]
102. Dong, M.; Yang, P.; Hua, F. MiR-191 modulates malignant transformation of endometriosis through regulating TIMP3. *Med. Sci. Monit.* **2015**, *21*, 915–920.
103. Sun, Y.; Hu, L.; Zheng, H.; Bagnoli, M.; Guo, Y.; Rupaimoole, R.; Rodriguez-Aguayo, C.; Lopez-Berestein, G.; Ji, P.; Chen, K.; et al. MiR-506 inhibits multiple targets in the epithelial-to-mesenchymal transition network and is associated with good prognosis in epithelial ovarian cancer. *J. Pathol.* **2015**, *235*, 25–36. [[CrossRef](#)]
104. Vilming-Elgaen, B.; Olstad, O.K.; Haug, K.B.; Brusletto, B.; Sandvik, L.; Staff, A.C.; Gautvik, K.M.; Davidson, B. Global miRNA expression analysis of serous and clear cell ovarian carcinomas identifies differentially expressed miRNAs including miR-200c-3p as a prognostic marker. *BMC Cancer.* **2014**, *14*, 80. [[CrossRef](#)] [[PubMed](#)]
105. Zhang, X.; Guo, G.; Wang, G.; Zhao, J.; Wang, B.; Yu, X.; Ding, Y. Profile of differentially expressed miRNAs in high-grade serous carcinoma and clear cell ovarian carcinoma, and the expression of miR-510 in ovarian carcinoma. *Mol. Med. Rep.* **2015**, *12*, 8021–8031. [[CrossRef](#)] [[PubMed](#)]

106. Yanaihara, N.; Noguchi, Y.; Saito, M.; Takenaka, M.; Takakura, S.; Yamada, K.; Okamoto, A. MicroRNA Gene Expression Signature Driven by miR-9 Overexpression in Ovarian Clear Cell Carcinoma. *PLoS ONE* **2016**, *11*, e0162584. [[CrossRef](#)] [[PubMed](#)]
107. Hirata, Y.; Murai, N.; Yanaihara, N.; Saito, M.; Saito, M.; Urashima, M.; Murakami, Y.; Matsufuji, S.; Okamoto, A. MicroRNA-21 is a candidate driver gene for 17q23-25 amplification in ovarian clear cell carcinoma. *BMC Cancer* **2014**, *14*, 799. [[CrossRef](#)]
108. Sugio, A.; Iwasaki, M.; Habata, S.; Mariya, T.; Suzuki, M.; Osogami, H.; Tamate, M.; Tanaka, R.; Saito, T. BAG3 upregulates Mcl-1 through downregulation of miR-29b to induce anticancer drug resistance in ovarian cancer. *Gynecol. Oncol.* **2014**, *134*, 615–623. [[CrossRef](#)]
109. Dai, F.; Zhang, Y.; Chen, Y. Involvement of miR-29b signaling in the sensitivity to chemotherapy in patients with ovarian carcinoma. *Hum. Pathol.* **2014**, *45*, 1285–1293. [[CrossRef](#)]
110. Tian, X.; Xu, L.; Wang, P. MiR-191 inhibits TNF- α induced apoptosis of ovarian endometriosis and endometrioid carcinoma cells by targeting DAPK1. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 4933–4942.
111. Calura, E.; Fruscio, R.; Paracchini, L.; Bignotti, E.; Ravaggi, A.; Martini, P.; Sales, G.; Beltrame, L.; Clivio, L.; Ceppi, L.; et al. MiRNA landscape in stage I epithelial ovarian cancer defines the histotype specificities. *Clin. Cancer Res.* **2013**, *19*, 4114–4123. [[CrossRef](#)]
112. Zhao, H.; Ding, Y.; Tie, B.; Sun, Z.F.; Jiang, J.Y.; Zhao, J.; Lin, X.; Cui, S. miRNA expression pattern associated with prognosis in elderly patients with advanced OPSC and OCC. *Int. J. Oncol.* **2013**, *43*, 839–849. [[CrossRef](#)]
113. Nagaraja, A.K.; Creighton, C.J.; Yu, Z.; Zhu, H.; Gunaratne, P.H.; Reid, J.G.; Olokpa, E.; Itamochi, H.; Ueno, N.T.; Hawkins, S.M.; et al. A link between mir-100 and FRAP1/mTOR in clear cell ovarian cancer. *Mol. Endocrinol.* **2010**, *24*, 447–463. [[CrossRef](#)]
114. Amano, T.; Chano, T.; Isono, T.; Kimura, F.; Kushima, R.; Murakami, T. Abundance of mitochondrial superoxide dismutase is a negative predictive biomarker for endometriosis-associated ovarian cancers. *World J. Surg. Oncol.* **2019**, *17*, 24. [[CrossRef](#)] [[PubMed](#)]
115. Ho, C.M.; Lee, F.K.; Huang, S.H.; Cheng, W.F. Everolimus following 5-aza-2-deoxycytidine is a promising therapy in paclitaxel-resistant clear cell carcinoma of the ovary. *Am. J. Cancer Res.* **2018**, *8*, 56–69. [[PubMed](#)]
116. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer Statistics, 2019. *CA Cancer J. Clin.* **2019**, *69*, 7–34. [[CrossRef](#)] [[PubMed](#)]
117. Vaughan, S.; Coward, J.; Bast, R.C., Jr.; Berchuck, A.; Berek, J.S.; Brenton, J.D.; Coukos, G.; Crum, C.C.; Drapkin, R.; Etemadmoghadam, D. Rethinking ovarian cancer: Recommendations for improving outcomes. *Nat. Rev. Cancer* **2011**, *11*, 719–725. [[CrossRef](#)] [[PubMed](#)]
118. Lisio, M.A.; Fu, L.; Goyeneche, A.; Gao, Z.H.; Telleria, C. High-Grade Serous Ovarian Cancer: Basic Sciences, Clinical and Therapeutic Standpoints. *Int. J. Mol. Sci.* **2019**, *20*, E952. [[CrossRef](#)] [[PubMed](#)]
119. Gilabert-Estelles, J.; Braza-Boils, A.; Ramon, L.A.; Zorio, E.; Medina, P.; España, F.; Estelles, A. Role of microRNAs in gynecological pathology. *Curr. Med. Chem.* **2012**, *19*, 2406–2413. [[CrossRef](#)]
120. Lluca, A.; Escrig, J. Prognostic value of peritoneal cancer index in primary advanced ovarian cancer. *Eur. J. Surg. Oncol.* **2018**, *44*, 163–169. [[CrossRef](#)]
121. Arend, R.C.; Londoño-Joshi, A.I.; Straughn, J.M.; Buchsbaum, D.J. The Wnt/ β -catenin pathway in ovarian cancer: A review. *Gynecol. Oncol.* **2013**, *131*, 772–779. [[CrossRef](#)]
122. Kim, J.; Park, E.Y.; Kim, O.; Schilder, J.M.; Coffey, D.M.; Cho, C.H.; Bast, R.C., Jr. Cell Origins of High-Grade Serous Ovarian Cancer. *Cancers* **2018**, *10*, E433. [[CrossRef](#)]
123. Labidi-Galy, S.I.; Papp, E.; Hallberg, D.; Niknafs, N.; Adleff, V.; Noe, M.; Bhattacharya, R.; Novak, M.; Jones, S.; Phallen, J.; et al. High grade serous ovarian carcinomas originate in the fallopian tube. *Nat. Commun.* **2017**, *8*, 1093. [[CrossRef](#)]
124. Vercellini, P.; Crosignani, P.; Somigliana, E.; Viganò, P.; Buggio, L.; Bolis, G.; Fedele, L. The ‘incessant menstruation’ hypothesis: A mechanistic ovarian cancer model with implications for prevention. *Hum. Reprod.* **2011**, *26*, 2262–2273. [[CrossRef](#)] [[PubMed](#)]
125. Bayani, J.; Brenton, J.D.; Macgregor, P.F.; Beheshti, B.; Albert, M.; Nallainathan, D.; Karaskova, J.; Rosen, B.; Murphy, J.; Laframboise, S.; et al. Parallel Analysis of Sporadic Primary Ovarian Carcinomas by Spectral Karyotyping, Comparative Genomic Hybridization, and Expression Microarrays. *Cancer Res.* **2002**, *62*, 3466–3476.

126. Kobayashi, H.; Ogawa, K.; Kawahara, N.; Iwai, K.; Niuro, E.; Morioka, S.; Yamada, Y. Sequential molecular changes and dynamic oxidative stress in high-grade serous ovarian carcinogenesis. *Free Radic Res.* **2017**, *51*, 755–764. [[CrossRef](#)]
127. Martinez-Outschoorn, U.E.; Balliet, R.M.; Lin, Z.; Whitaker-Menezes, D.; Howell, A.M.; Sotgia, F.; Lisanti, M.P. Hereditary ovarian cancer and two-compartment tumor metabolism: Epithelial loss of BRCA1 induces hydrogen peroxide production, driving oxidative stress and NF κ B activation in the tumor stroma. *Cell Cycle* **2012**, *11*, 4152–4166. [[CrossRef](#)]
128. Alsop, K.; Fereday, S.; Meldrum, C.; DeFazio, A.; Emmanuel, C.; George, J.; Dobrovic, A.; Birrer, M.J.; Webb, P.M.; Stewart, C.; et al. BRCA mutation frequency and patterns of treatment response in BRCA mutation-positive women with ovarian cancer: A report from the Australian Ovarian Cancer Study Group. *J. Clin. Oncol.* **2012**, *30*, 2654–2663. [[CrossRef](#)] [[PubMed](#)]
129. Klinkebiel, D.; Zhang, W.; Akers, S.N.; Odunsi, K.; Karpf, A.R. DNA Methylome Analyses Implicate Fallopian Tube Epithelia as the Origin for High-Grade Serous Ovarian Cancer. *Mol. Cancer Res.* **2016**, *14*, 787–794. [[CrossRef](#)] [[PubMed](#)]
130. Yang, J.; Zhou, Y.; Ng, S.K.; Huang, K.C.; Ni, X.; Choi, P.W.; Hasselblatt, K.; Muto, M.G.; Welch, W.R.; Berkowitz, R.S.; et al. Characterization of MicroRNA-200 pathway in ovarian cancer and serous intraepithelial carcinoma of fallopian tube. *BMC Cancer* **2017**, *17*, 422. [[CrossRef](#)]
131. Katsuno, Y.; Lamouille, S.; Derynck, R. TGF- β signaling and epithelial-mesenchymal transition in cancer progression. *Curr. Opin. Oncol.* **2013**, *25*, 76–84. [[CrossRef](#)]
132. Prieto-García, E.; Díaz-García, C.V.; García-Ruiz, I.; Agulló-Ortuño, M.T. Epithelial-to-mesenchymal transition in tumor progression. *Med. Oncol.* **2017**, *34*, 122. [[CrossRef](#)]
133. Boac, B.M.; Xiong, Y.; Marchion, D.C.; Abbasi, F.; Bush, S.H.; Ramirez, I.J.; Khulpateea, B.R.; Clair-McClung, E.; Berry, A.L.; Bou-Zgheib, N.; et al. Micro-RNAs associated with the evolution of ovarian cancer cisplatin resistance. *Gynecol. Oncol.* **2016**, *140*, 259–263. [[CrossRef](#)]
134. Zhu, X.; Shen, H.; Yin, X.; Long, L.; Xie, C.; Liu, Y.; Hui, L.; Lin, X.; Fang, Y.; Cao, Y.; et al. miR-186 regulation of Twist1 and ovarian cancer sensitivity to cisplatin. *Oncogene* **2016**, *35*, 323–332. [[CrossRef](#)] [[PubMed](#)]
135. Helleman, J.; Smid, M.; Jansen, M.P.; van der Burg, M.E.; Berns, E.M. Pathway analysis of gene lists associated with platinum-based chemotherapy resistance in ovarian cancer: The big picture. *Gynecol Oncol.* **2010**, *17*, 170–176. [[CrossRef](#)] [[PubMed](#)]
136. Brozovic, A.; Duran, G.E.; Wang, Y.C.; Francisco, E.B.; Sikic, B.I. The miR-200 family differentially regulates sensitivity to paclitaxel and carboplatin in human ovarian carcinoma OVCAR-3 and MES-OV cells. *Mol Oncol.* **2015**, *9*, 1678–1693. [[CrossRef](#)] [[PubMed](#)]
137. Mateescu, B.; Batista, L.; Cardon, M.; Gruosso, T.; de Feraudy, Y.; Mariani, O.; Nicolas, A.; Meyniel, J.P.; Cottu, P.; Sastre-Garau, X.; et al. miR-141 and miR-200a act on ovarian tumorigenesis by controlling oxidative stress response. *Nat Med.* **2011**, *17*, 1627–1635. [[CrossRef](#)] [[PubMed](#)]
138. Vecchione, A.; Belletti, B.; Lovat, F.; Volinia, S.; Chiappetta, G.; Giglio, S.; Sonogo, M.; Cirombella, R.; Onesti, E.C.; Pellegrini, P.; et al. A microRNA signature defines chemoresistance in ovarian cancer through modulation of angiogenesis. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 9845–9850. [[CrossRef](#)]
139. Jeong, J.Y.; Kang, H.; Kim, T.H.; Kim, G.; Heo, J.H.; Kwon, A.Y.; Kim, S.; Jung, S.G.; An, H.J. MicroRNA-136 inhibits cancer stem cell activity and enhances the anti-tumor effect of paclitaxel against chemoresistant ovarian cancer cells by targeting Notch3. *Cancer Lett.* **2017**, *386*, 168–178. [[CrossRef](#)]
140. Liu, Y.; Niu, Z.; Lin, X.; Tian, Y. MiR-216b increases cisplatin sensitivity in ovarian cancer cells by targeting PARP1. *Cancer Gene Ther.* **2017**, *24*, 208–214. [[CrossRef](#)]
141. Sun, C.; Li, N.; Yang, Z.; Zhou, B.; He, Y.; Weng, D.; Fang, Y.; Wu, P.; Chen, P.; Yang, X.; et al. miR-9 regulation of BRCA1 and ovarian cancer sensitivity to cisplatin and PARP inhibition. *J. Natl. Cancer Inst.* **2013**, *105*, 1750–1758. [[CrossRef](#)]
142. Ayyagari, V.N.; Diaz-Sylvester, P.L.; Hsieh, T.J.; Brard, L. Evaluation of the cytotoxicity of the Bithionol-paclitaxel combination in a panel of human ovarian cancer cell lines. *PLoS ONE* **2017**, *12*, e0185111. [[CrossRef](#)]
143. Ayyagari, V.N.; Brard, L. Bithionol inhibits ovarian cancer cell growth in vitro—Studies on mechanism(s) of action. *BMC Cancer* **2014**, *14*, 61. [[CrossRef](#)]

144. Hou, D.; Liu, Z.; Xu, X.; Liu, Q.; Zhang, X.; Kong, B.; Wei, J.J.; Gong, Y.; Shao, C. Increased oxidative stress mediates the antitumor effect of PARP inhibition in ovarian cancer. *Redox Biol.* **2018**, *17*, 99–111. [[CrossRef](#)] [[PubMed](#)]
145. Choi, Y.E.; Meghani, K.; Brault, M.E.; Leclerc, L.; He, Y.J.; Day, T.A.; Elias, K.M.; Drapkin, R.; Weinstock, D.M.; Dao, F.; et al. Platinum and PARP Inhibitor Resistance Due to Overexpression of MicroRNA-622 in BRCA1-Mutant Ovarian Cancer. *Cell Rep.* **2016**, *14*, 429–439. [[CrossRef](#)] [[PubMed](#)]
146. Meghani, K.; Fuchs, W.; Detappe, A.; Drané, P.; Gogola, E.; Rottenberg, S.; Jonkers, J.; Matulonis, U.; Swisher, E.M.; Konstantinopoulos, P.A.; et al. Multifaceted Impact of MicroRNA 493-5p on Genome-Stabilizing Pathways Induces Platinum and PARP Inhibitor Resistance in BRCA2-Mutated Carcinomas. *Cell Rep.* **2018**, *23*, 100–111. [[CrossRef](#)] [[PubMed](#)]
147. Belotte, J.; Fletcher, N.M.; Awonuga, A.O.; Alexis, M.; Abu-Soud, H.M.; Saed, M.G.; Diamond, M.P.; Saed, G.M. The role of oxidative stress in the development of cisplatin resistance in epithelial ovarian cancer. *Reprod. Sci.* **2014**, *21*, 503–508. [[CrossRef](#)]
148. Fletcher, N.M.; Belotte, J.; Saed, M.G.; Memaj, I.; Diamond, M.P.; Morris, R.T.; Saed, G.M. Specific point mutations in key redox enzymes are associated with chemoresistance in epithelial ovarian cancer. *Free Radic Biol. Med.* **2017**, *102*, 122–132. [[CrossRef](#)]
149. Pei, H.; Yang, Y.; Cui, L.; Yang, J.; Li, X.; Yang, Y.; Duan, H. Bisdemethoxycurcumin inhibits ovarian cancer via reducing oxidative stress mediated MMPs expressions. *Sci. Rep.* **2016**, *6*, 28773. [[CrossRef](#)]
150. Pons, D.G.; Sastre-Serra, J.; Nadal-Serrano, M.; Oliver, A.; García-Bonafé, M.; Bover, I.; Roca, P.; Oliver, J. Initial activation status of the antioxidant response determines sensitivity to carboplatin/paclitaxel treatment of ovarian cancer. *Anticancer Res.* **2012**, *32*, 4723–4728.
151. Montoya-Estrada, A.; Coria-García, C.F.; Cruz-Orozco, O.P.; Aguayo-González, P.; Torres-Ramos, Y.D.; Flores-Herrera, H.; Hicks, J.J.; Medina-Navarro, R.; Guzmán-Grenfell, A.M. Increased systemic and peritoneal oxidative stress biomarkers in endometriosis are not related to retrograde menstruation. *Redox Rep.* **2019**, *24*, 51–55. [[CrossRef](#)]
152. Lambrinoudaki, I.V.; Augoulea, A.; Christodoulakos, G.E.; Economou, E.V.; Kaparos, G.; Kontoravdis, A.; Papadias, C.; Creasas, G. Measurable serum markers of oxidative stress response in women with endometriosis. *Fertil Steril.* **2009**, *91*, 46–50. [[CrossRef](#)]
153. Turkyilmaz, E.; Yildirim, M.; Cendek, B.D.; Baran, P.; Alisik, M.; Dalgaci, F.; Yavuz, A.F. Evaluation of oxidative stress markers and intra-extracellular antioxidant activities in patients with endometriosis. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2016**, *199*, 164–168. [[CrossRef](#)]
154. Rosa e Silva, J.C.; do Amara, V.F.; Mendonça, J.L.; Rosa e Silva, A.C.; Nakao, L.S.; Poli Neto, O.B.; Ferriani, R.A. Serum markers of oxidative stress and endometriosis. *Clin. Exp. Obstet. Gynecol.* **2014**, *41*, 371–374. [[PubMed](#)]
155. Vicente-Muñoz, S.; Morcillo, I.; Puchades-Carrasco, L.; Payá, V.; Pellicer, A.; Pineda-Lucena, A. Nuclear magnetic resonance metabolomic profiling of urine provides a noninvasive alternative to the identification of biomarkers associated with endometriosis. *Fertil Steril.* **2015**, *104*, 1202–1209. [[CrossRef](#)] [[PubMed](#)]
156. Karihtala, P.; Soini, Y.; Vaskivuo, L.; Bloigu, R.; Puistola, U. DNA adduct 8-hydroxydeoxyguanosine, a novel putative marker of prognostic significance in ovarian carcinoma. *Int. J. Gynecol. Cancer.* **2009**, *19*, 1047–1051. [[CrossRef](#)] [[PubMed](#)]
157. Pylväs-Eerola, M.; Karihtala, P.; Puistola, U. Preoperative serum 8-hydroxydeoxyguanosine is associated with chemoresistance and is a powerful prognostic factor in endometrioid-type epithelial ovarian cancer. *BMC Cancer* **2015**, *15*, 493. [[CrossRef](#)]
158. Pylväs, M.; Puistola, U.; Laatio, L.; Kauppila, S.; Karihtala, P. Elevated serum 8-OHdG is associated with poor prognosis in epithelial ovarian cancer. *Anticancer Res.* **2011**, *31*, 1411–1415.
159. Weber, J.A.; Baxter, D.H.; Zhang, S.; Huang, D.Y.; Huang, K.H.; Lee, M.J.; Galas, D.J.; Wang, K. The microRNA spectrum in 12 body fluids. *Clin. Chem.* **2010**, *56*, 1733–1741. [[CrossRef](#)]
160. Shah, M.Y.; Calin, G.A. The mix of two worlds: Non-coding RNAs and hormones. *Nucl. Acid Ther.* **2013**, *23*, 2–8. [[CrossRef](#)]
161. Mier-Cabrera, J.; Genera-García, M.; De la Jara-Díaz, J.; Perichart-Perera, O.; Vadillo-Ortega, F.; Hernández-Guerrero, C. Effect of vitamins C and E supplementation on peripheral oxidative stress markers and pregnancy rate in women with endometriosis. *Int. J. Gynaecol. Obstet.* **2008**, *100*, 252–256. [[CrossRef](#)]

162. Lete, I.; Mendoza, N.; de la Viuda, E.; Carmona, F. Effectiveness of an antioxidant preparation with N-acetyl cysteine, alpha lipoic acid and bromelain in the treatment of endometriosis-associated pelvic pain: LEAP study. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2018**, *228*, 221–224. [[CrossRef](#)]
163. Ray, K.; Fahrman, J.; Mitchell, B.; Paul, D.; King, H.; Crain, C.; Cook, C.; Golovko, M.; Brose, S.; Golovko, S.; et al. Oxidation-sensitive nociception involved in endometriosis-associated pain. *Pain* **2015**, *156*, 528–539. [[CrossRef](#)]
164. Monk, B.J.; Kauderer, J.T.; Moxley, K.M.; Bonebrake, A.J.; Dewdney, S.B.; Secord, A.A.; Ueland, F.R.; Johnston, C.M.; Aghajanian, C. A phase II evaluation of elesclomol sodium and weekly paclitaxel in the treatment of recurrent or persistent platinum-resistant ovarian, fallopian tube or primary peritoneal cancer: An NRG oncology/gynecologic oncology group study. *Gynecol. Oncol.* **2018**, *151*, 422–427. [[CrossRef](#)] [[PubMed](#)]
165. Thomsen, C.B.; Andersen, R.F.; Steffensen, K.D.; Adimi, P.; Jakobsen, A. Delta tocotrienol in recurrent ovarian cancer. A phase II trial. *Pharmacol. Res.* **2019**, *141*, 392–396. [[CrossRef](#)] [[PubMed](#)]
166. Trudel, D.; Labbé, D.P.; Araya-Farias, M.; Doyen, A.; Bazinet, L.; Duchesne, T.; Plante, M.; Grégoire, J.; Renaud, M.C.; Bachvarov, D.; et al. A two-stage, single-arm, phase II study of EGCG-enriched green tea drink as a maintenance therapy in women with advanced stage ovarian cancer. *Gynecol. Oncol.* **2013**, *131*, 357–361. [[CrossRef](#)] [[PubMed](#)]
167. Rupaimoole, R.; Slack, F.J. MicroRNA therapeutics: Towards a new era for the management of cancer and other diseases. *Nat. Rev. Drug Discov.* **2017**, *16*, 203–222. [[CrossRef](#)] [[PubMed](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).