Microglial activation in regions related to cognitive function predicts disease onset in Huntington's Disease: A multimodal imaging study

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Abstract: Huntington's disease (HD) is an inherited neurodegenerative disorder associated with motor, cognitive and psychiatric deficits. This study, using a multimodal imaging approach, aims to assess in vivo the functional and structural integrity of regions and regional networks linked with motor, cognitive and psychiatric function. Predicting disease onset in at risk individuals is problematic and thus we sought to investigate this by computing the 5-year probability of HD onset (p5 HD) and relating it to imaging parameters. Using MRI, ¹¹C-PK11195 and ¹¹C-raclopride PET, we have investigated volumes, levels of microglial activation and D2/D3 receptor binding in CAG repeat-matched groups of premanifest and symptomatic HD gene carriers. Findings were correlated with disease-burden and UHDRS scores. Atrophy was detected in sensorimotor striatum (SMST), substantia nigra, orbitofrontal and anterior prefrontal cortex in the premanifest HD. D2/D3 receptor binding was reduced and microglial activation increased in SMST and associative striatum (AST), bed nucleus of the stria terminalis, the amygdala and the hypothalamus. In symptomatic HD cases this extended to involve atrophy in globus pallidus, limbic striatum, the red nuclei, anterior cingulate cortex, and insula. D2/D3 receptor binding was additionally reduced in substantia nigra, globus pallidus, limbic striatum, anterior cingulate cortex and insula, and microglial activation increased in globus pallidus, limbic striatum and anterior prefrontal cortex. In premanifest HD, increased levels of microglial activation in the AST and in the regional network associated with cognitive function correlated with p5 HD onset. These data suggest that pathologically activated microglia in AST and other areas related to cognitive function, maybe better predictors of clinical onset and stresses the importance of early cognitive assessment in HD. Hum Brain Mapp 32:258–270, 2011. © 2010 Wiley-Liss, Inc.

Keywords: PET; MRI; microglia; dopamine; biomarkers; associative striatum; PK; RAC; ROI; VOI

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Published online 28 May 2010 in Wiley Online Library (wileyonlinelibrary.com).

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INTRODUCTION

Huntington's disease (HD) is a progressive neurodegenerative disorder with typical onset between 35 and 50 years of age. HD is inherited in an autosomal dominant pattern and the responsible IT15 gene mutation consists of an unstable expansion of CAG repeats within its coding region that is located in the distal portion of the short arm of chromosome 4 (4p16.3) and encodes the production of the protein huntingtin [The Huntington's Disease Collaborative Research Group, 1993]. The toxic gain of function of the mutant huntingtin and its associated proteins contribute to the disruption of multiple intracellular pathways leading to the formation of aggregates that progressively result in the dysfunction and then degeneration of important neuronal pathways and cell loss in regions of the HD brain (for review see Roze et al. [2008]).

Twenty-five years ago, Vonsattel et al. [1985] developed a grading system to assess the severity of HD striatal degeneration based on a large post-mortem study. From that time, although progressive atrophy of the striatum remains one of the main pathological hallmarks of HD, advances in neuroimaging techniques have greatly contributed to a better understanding of HD pathology as more widespread [Tabrizi et al., 2009], complex and challenging for the development of potential treatments.

Structural and functional neuroimaging studies have now established important correlations between morphological and functional brain changes and the development of clinical deficits during the course of the disease. A combined neuroimaging approach incorporating both structural and functional techniques could potentially serve as a biomarker for use in future clinical disease modifying drug trials in HD.

This study has been designed taking into account several issues in HD: (1) The size of CAG repeats expansion inversely correlates with the age at onset [Kieburtz et al., 1994] and positively with the rate of HD progression [Penney et al., 1997]. (2) The extent and localization of atrophy in HD brain remains controversial (for review see Bohanna et al. [2008]). (3) Altered dopaminergic neurotransmission is evident from the premanifest stage of the disease in striatal [Andrews et al., 1999; van Oostrom et al., 2009] and extrastriatal areas [Pavese et al., 2003; Politis et al., 2008] and has been implicated as potentially pathogenic, contributing to the neuronal cell death [Jakel and Maragos, 2000]. (4) There is detectable ongoing neuroinflammation and pathogenic immune activation very early in the course of the disease [Björkqvist et al., 2008] and activated microglia have been proved as sensitive reactors to pathological changes in HD brain such as the presence of mutant huntingtin [Sapp et al., 2001; Shin et al., 2005].

11C-raclopride (RAC) with positron emission tomography (PET) is a marker of dopamine D2 and D3 receptor availability and an indirect marker of synaptic dopamine fluxes in the living human brain. In Huntington's disease, this imaging technique has been largely employed for the

assessment of striatal and extrastriatal functional integrity [Andrews et al., 1999; Pavese et al., 2003; Politis et al., 2008; van Oostrom et al., 2009]. Furthermore, in healthy brain, microglia exist in a resting state characterized by ramified morphology, and monitor the brain milieu. In response to a neuronal insult (such as mutant huntingtin), however, microglia become activated and are transformed from their resting ramified state into an amoeboid morphology releasing cytokines and growth factors and having phagic properties [Davalos et al., 2005; Nimmerjahn et al., 2005]. 11C-PK11195 (PK) is a specific radioligand of the translocator protein (TSPO) previously known as the peripheral benzodiazepine receptor. The TSPO is expressed by the mitochondria of activated microglia and PK PET has been used as a marker for this transition of microglia from a resting to an activated state [Cagnin et al., 2001].

We have measured volumetric and functional (D2/D3 receptor binding and activation of microglia) parameters in HD to investigate the structure and function of individual brain regions and estimated regional networks with known connection to psychiatric, cognitive and motor symptoms. Furthermore, we have looked for associations between imaging findings and clinical scales related to daily living, disease severity, motor dysfunction and in the case of premanifest HD gene carriers, the probability of HD onset. For this imaging paradigm, we have created averaged imaging profiles derived from three cohorts of subjects. Regional volume changes were studied with structural magnetic resonance imaging (MRI) volumetric analysis, whereas for the investigation of the integrity of post-synaptic dopamine D2/D3 receptors and the presence of altered levels of activated microglia, RAC and PK PET have been employed, respectively.

MATERIALS AND METHODS

Subjects

Thirty-two subjects participated in this study. Comparisons between the functional and structural imaging profiles of eight premanifest HD brains (4M: 4F; 45 \pm 3 CAG repeats, 41 ± 10 years of age, 361 ± 65 disease-burden score, p5 HD 0.4 \pm 0.1; mean \pm SD) that all had genetically proven disease, eight premanifest HD brains (4M: 4F; 45 \pm 3 CAG repeats, 48 \pm 5 years of age, 428 \pm 126 disease-burden score, 5 \pm 3 years of disease, 16 \pm 11 Unified Huntington's Disease Rating Scale (UHDRS) motor score, Total Functional Capacity (TFC) score 6.5 \pm 2, IS score 75% \pm 5; mean \pm SD) that all had clinically manifest disease and 16 normal brains (MRI: n = 16, 14M, 2F, 46 \pm 6 years \pm SD; RAC: n = 8, 8M, 44 \pm 4 years \pm SD; PK: n =8, 6M, 2F, 48 \pm 7 years \pm SD) all in good health with no history of neurological or psychiatric disorder, were performed (Table 1). None of these subjects were on any medication.

Averaged profile	Premanifest HD brain	Symptomatic HD brain	MRI normal brain	RAC normal brain	PK normal brain
No. of subjects	8	8	16	8	8
Sex	4M/4F	4M/4F	14M/2F	8M/0F	6M/2F
Age (years \pm SD)	40.8 ± 9.5	47.7 ± 5.0	45.9 ± 6.1	43.7 ± 4.3	48.4 ± 7.2
CAG repeats (mean \pm SD)	44.9 ± 2.6	44.7 ± 3.1	-	-	_
Disease-burden score ^a (mean \pm SD)	361.2 ± 64.9	427.4 ± 126.1	_	-	-
Disease duration ^b (years \pm SD)	-	5.2 ± 2.7	_	-	-
UHDRS motor score ^c (mean \pm SD)	0.0 ± 0.0	15.9 ± 11.1	_	-	-
p5 HD ^d	0.4 ± 0.1	-	_	-	-
TFC ^e	13.0 ± 0.0	6.5 ± 2.3	-	_	_
IS^{f}	$100.0\%\pm0.0$	$74.3\%\pm5.3$	-	-	_

TABLE I. Clinical profile and characteristics of the subjects formulate the averaged premanifest HD, symptomatic HD, and MRI, RAC and PK normal brains

^aDisease-burden score = CAG index = age \times (CAG repeats length – 35.5) [Penney et al., 1997].

^bDisease duration measured from the time where the subject could remember his/her first symptoms.

 $^{\circ}$ UHDRS = Unified Huntington's Disease Rating Scale [Huntington Study Group, 1996]. UHDRS motor scores: Range (Best = 0, Worst = 124).

^dProbability of developing Huntington's disease symptoms in the following 5 years. Taken from the age-CAG repeats correlation tables [Langbehn et al., 2004].

 e TFC = Total Functional Capacity scores (Best = 13, Worst = 0).

 $^{f}IS = Independence Scale scores (Best = 100\%, Worst = 0\%).$

Clinical Evaluation

The symptomatic HD and premanifest HD gene carriers were assessed clinically in three major domains: (1) daily living using the TFC and Independence Scale (IS) scores; (2) disease severity using CAG repeat length and the disease-burden score (CAG index = age \times [CAG = 35.5], Penney et al., 1997) and; (3) motor dysfunction using the motor component of the UHDRS. Additionally, the 5-year probability of symptomatic HD onset (probability of diagnosis calculated based on CAG repeat length and current age, see Langbehn et al. [2004]) was calculated in the premanifest HD gene carriers (Table 1).

Imaging Techniques

MRI, RAC and PK PET scanning were performed no more than 2 months apart.

MRI scanning

Subjects had a volumetric T1 MRI with a 1.5 Tesla scanner using a RF-fast pulse sequence, TR = 30 ms and TE = 3 ms acquisition time and a flip angle of 30° (Philips Medical Systems; Picker Eclipse, Picker International Inc., Highland Heights, OH, USA). This scanner allows the acquisition of high-resolution RF spoiled T1-weighted volume datasets. Subjects were positioned supine with their transaxial planes parallel to the line intersecting anterior and posterior commissure (AC–PC line). They were made comfortable and their head position was maintained still with the help of individualized foam holders.

PET scanning

All symptomatic and premanifest HD gene carriers had both RAC and PK PET scan, whereas normal controls had either RAC or PK PET (half-half). PET was performed using an ECAT EXACT HR++ (CTI/Siemens 966; Siemens, Knoxville, TN) camera with 23.4 cm total axial field of view. The camera has a reconstructed (image) transaxial spatial resolution of 5.1 ± 0.6 mm and an axial resolution of 5.9 ± 0.6 mm over a 10-cm radius FOV from the centre [Spinks et al., 2000]. This high-resolution, high-sensitivity tomograph permits interrogation of the function of small volume structures.

The subjects were positioned supine with their transaxial planes parallel to the line intersecting the AC–PC line. They were made comfortable and their head position maintained with the help of individualised foam holders. In addition, their position relative to the camera's laser light was monitored throughout and was repositioned if movement was detected. Subjects were in a resting state with low light and no noise in the room. Smoking and consumption of alcohol, coffee and other caffeinated beverages was not allowed for 12 h before scanning. Eating and drinking was not allowed for 8 h before PET.

A 5-min transmission scan was performed before injection of each radiotracer to correct for tissue attenuation of 511 keV gamma radiation. The mean tracer dose for RAC administered was 188 MBq (range: 181–196 MBq) and for PK was 297 MBq (ranging from 293 to 304 MBq). The radiotracers were infused intravenously over 30 s. Scanning began at the start of the tracer injection generating 20 time frames over 65 min for RAC and 18 time frames over 60 min for PK. RAC and PK tracers were supplied by

Hammersmith Imanet plc, London, UK and permission to administer RAC and PK was obtained from the Administration of Radioactive Substances Advisory Committee (ARSAC) of the Department of Health, UK.

Data Analysis

MRI

The MRI images after transformation to DICOM format were reoriented parallel to the AC–PC line and then transformed to ANALYZE 7.5 format. Volume-of-interest (VOI) analysis was performed on individual MRIs using ANA-LYZE software (version 8.1, BRU, Mayo Foundation, Rochester, MN). This method allows a direct volume comparison expressed in cubic millimetres between brain regions under investigation after calculating the sum of sampled slices on each MRI.

RAC PET

Region-of-interest (ROI) image analysis was performed using ANALYZE software to detect functional changes in dopaminergic D2 and D3 receptor binding. Parametric images of RAC binding potentials (BP_{ND}) were generated from the dynamic RAC scan series, using BASIS function implementation of the simplified reference region compartmental model with the cerebellum as the reference tissue for non-specific tracer binding [Gunn et al., 1997]. Each parametric RAC BP_{ND} image was anatomically coregistered and resliced to the corresponding volumetric T1weighted MRI. This was done using the Mutual Information Registration algorithm in the SPM2 software package (Wellcome Department of Cognitive Neuroscience, Institute of Neurology, London, UK) implemented in Matlab6 (Version 6.5.1, The Mathworks Inc., Natick, MA). BP_{ND} values for the ROIs were obtained by defining ROIs on the individual MR images that were subsequently used to sample the parametric images.

PK PET

Parametric BP_{ND} images of PK were generated from the dynamic PK scan series, using cluster analysis to identify a reference cluster of voxels representing non-specific PK uptake and thus to provide a tissue input function as previously described [Banati, 2002]. In brief, the cluster analysis divides the brain voxels into 10 clusters based on the shape of their time-activity curves. The cluster with the time-activity curve which most resembles that of the mean for a normal population of cortical voxels, tested for similarity using a χ^2 test (P < 0.05), is selected as the reference cluster. Parametric PK BP_{ND} images were anatomically coregistered, resliced and sampled as for RAC.

Regions targeted

The regions selected were based on the study's aims, the sufficiency of PET camera resolution and the post-mortem [Guverich and Joyce, 1999; Hurd et al., 2001] and imaging [Ito et al., 2008] studies regarding the density of D2 and D3 receptors in human brain.

Regions were defined manually on each subject's volumetric MRI guided by a three-dimensional sectional atlas [Duvernoy, 1999]. Sensorimotor striatum (SMST), associative striatum (AST), limbic striatum (LST), anterior cingulate cortex (ACC), posterior cingulate cortex (PCC), anterior prefrontal cortex (aPFC), orbitofrontal cortex (OFC), thalamus, hypothalamus, insula, hippocampus, globus pallidus, substantia nigra, red nucleus, superior and inferior colliculus, bed nucleus of the stria terminalis (BNST) and Amygdala were interrogated.

The striatal subregions were examined according to functional criteria previously described [Martinez et al., 2003]. SMST included the post-commissural putamen; AST included the pre-commissural dorsal putamen and caudate and the post-commissural caudate; LST included the Ventral striatum (VST) consisting of nucleus accumbens, and ventral caudate and putamen rostral to the AC.

Volumes and RAC and PK BP_{ND} were calculated after PET coregistration to MRI, providing equal structural VOI and functional ROI object maps for each structure. RAC BP_{ND} , PK BP_{ND} and volume values for left and right hemisphere regions were averaged.

Statistics

Statistical analyses of MRI, PET and clinical data were performed using SPSS software package (Version 16, SPSS Inc, Chicago, IL, USA). The Kruskal-Wallis test with post-test (Dunn's) was employed for between-group comparisons and Spearman's correlation with multiple comparisons for interrogation of correlations between MRI, PET data and clinical parameters. The significance level was set at P < 0.05.

The MRI and PET data were investigated in two ways: first by examining individual regional volumes, RAC BP_{ND} and PK BP_{ND} values and second by combining the values of regions that are known to be associated with motor, psychiatric and cognitive disorders [Mega and Cummings, 1994].

Brain regions related to motor functions (Regions-Motor Group) include SMST, globus pallidus, substantia nigra and red nucleus. Regions related to cognitive functions (Regions-Cognitive Group) include ACC, PCC, AST, aPFC, OFC, insula and hippocampus. Regions related to psychiatric disorders (Regions-Psychiatric Group) include ACC, LST, OFC, hypothalamus, insula, hippocampus, BNST and amygdala. Volumes and RAC and PK BP_{ND} for the regional group investigations were calculated using formula (1) and (2), respectively. Group = Group of regions related to motor or cognitive or psychiatric function; Tracer = PK or RAC; R= Region.

$$\text{Group}_{mm^3} = R1_{mm^3} + R2_{mm^3} + \dots RV_{mm^3} \qquad (1)$$

(

$$(\text{Group})_{\text{PET}(\text{tracer})\text{BP}_{\text{ND}}} = \frac{\left(R1_{\text{PET}(\text{tracer})\text{BP}_{\text{ND}}} \times R1_{\text{mm}^3}\right) + \left(R2_{\text{PET}(\text{tracer})\text{BP}_{\text{ND}}} \times R2_{\text{mm}^3}\right) + \dots + \left(RV_{\text{PET}(\text{tracer})\text{BP}_{\text{ND}}} \times RV_{\text{mm}^3}\right)}{(R1 + R2 + \dots + RV)_{\text{mm}^3}}$$
(2)

RESULTS

Regional Individual Analyses

MRI-VOI

Significant decreases in volume were detected in SMST, aPFC, OFC and substantia nigra in premanifest HD compared with normal controls. Additional significant decreases in volume were detected in ACC, AST, LST, insula, globus pallidus and red nucleus in symptomatic HD. A direct comparison of symptomatic and premanifest HD found relative volume decreases in ACC, SMST, insula, globus pallidus and substantia nigra in the former (Table 2; Figures 1–3).

PET-RAC ROI

Significant decreases in RAC BP_{ND} were detected in SMST, AST, hypothalamus, BNST and amygdala in the premanifest HD with additional decreases in ACC, LST, insula, globus pallidus and substantia nigra in symptomatic HD compared with normal controls. When the symptomatic and premanifest HD groups were directly compared there were relative RAC BP_{ND} decreases in ACC, SMST and AST in the former (Table 2; Figures 1–3).

PET-PK ROI

Significant increases in PK BP_{ND} were detected in SMST, AST, hypothalamus, BNST and amygdala of the premanifest HD cases with additional increases in LST, aPFC and globus pallidus of symptomatic HD compared with normal controls. A direct comparison of symptomatic and premanifest HD groups revealed relative PK BP_{ND} increases in aPFC in the former (Table 2; Figures 1–3).

Regional Group Analyses

Motor group

When the motor group of regions were assessed together, we found significant decreases in volume and RAC BP_{ND} and increases in PK BP_{ND} in both the premanifest and symptomatic HD groups compared with normal controls. Volume and RAC BP_{ND} were relatively decreased in the symptomatic HD compared with the premanifest HD cases (Table 2).

Cognitive group

When the cognitive group of regions were assessed together, we found significant decreases in volume and in RAC BP_{ND} and significant PK BP_{ND} increases in the symptomatic HD cases compared with normal controls (Table 2).

Psychiatric Group

When the psychiatric group of regions were assessed together, we found significant decreases in volume and in RAC BP_{ND} and significant PK BP_{ND} increases in the symptomatic HD cases compared with normal controls (Table 2).

Correlations

MRI/PET correlations

No significant correlations are reported.

RAC PET/PK PET correlations

RAC BP_{ND} values were negatively correlated with PK BP_{ND} values for both HD groups in SMST (premanifest HD: r = -0.6571, P < 0.05; Symptomatic HD: r = -0.68, P < 0.05), hypothalamus (premanifest HD: r = -0.7055, P < 0.05; Symptomatic HD: r = -0.6672, P < 0.01) and BNST (premanifest HD: r = -0.7107, P < 0.05; Symptomatic HD: r = -0.7505, P < 0.05).

Imaging/clinical correlations

For the symptomatic HD, TFC scores negatively correlated with the cognitive group of regions PK BP_{ND} values (r = -0.6980, P < 0.05). Disease-burden scores positively correlated with SMST PK BP_{ND} values (r = 0.8176, P < 0.01). UHDRS motor scores correlated positively with the SMST PK BP_{ND} values (r = 0.7092, P < 0.05) and negatively with SMST RAC BP_{ND} values (r = -0.7465, P < 0.05). The 5-year probability of developing HD onset positively correlated with PK BP_{ND} for both AST (r = 0.8095, P < 0.05) and the cognitive group of regions (r = 0.8810, P < 0.01) in premanifest HD cases (Figures 4 and 5).

DISCUSSION

In this study, we have created averaged brain-imaging profiles for CAG repeat-matched premanifest and symptomatic HD groups in order to investigate changes in volume, post-synaptic dopamine D2/D3 receptor binding and microglial activation in brain regions known to be involved in the motor, cognitive and psychiatric features of HD. The imaging data were interrogated by targeting individual regions as well as combining them into groups according to their broad involvement in motor, cognitive

me RAC BP _{ND} PK BP _{ND} Volume RAC BP _{ND} Increases Increasese Increases Incr		Premanif	Premanifest HD vs normal brain	nal brain	Symptom	Symptomatic HD vs normal brain	nal brain	Premanifest	Premanifest HD vs symptomatic HD	omatic HD
20.9* 46.5* 14.3 11.8 28.1* 27.4* 28.14 36.0** 14.1 24.2 6.6 11.8 10.8 9.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 22.3 17.7 nnc 26.6* 17.3 18.5 17.7 nnc 22.3 nnc 23.5 nnc 22.3 nnc 22.3 nnc 23.5 nnc 23		Volume loss (mm ³) (%)	RAC BP _{ND} decreases (%)	PK BP _{ND} increases (%)	Volume loss (mm ³) (%)	RAC BP _{ND} decreases (%)	PK BP _{ND} increases (%)	Volume loss (mm ³) (%)	RAC BP _{ND} decreases (%)	PK BP _{ND} increases (%)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Autonica since (ACC)	L L	6 0	UN	**0 70	17 7 7 *	UN	*0 OC	16 E*	UN
28.1* 27.4* 28.1* 27.4* 21.4 36.0* 14.1 24.2 6.6 11.8 10.8 9.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 23.2 11.2 24.6 26.8* 23.2 11.2 25.8 13.1 12.2 7.1 NC NC NC NC 22.3 NC NC 22.3 17.3 18.5 17.7 nc 20.6* 17.3 18.5 17.7 nc 20.6* 17.3 18.5 17.7 nc 20.6* 17.3 18.5 17.7 nc 20.6* 17.3 18.5 17.7 nc 20.6* 17.3 17.3 nc 20.6* 17.3 17.3 nc 20.6* 17.3 17.3 nc 20.6* 17.3 17.4 nc 20.6* 17.3 17.5 nc 20.5* 17.7 17.5 nc 20.6* 17.5 17.5 nc 20.5* 17.7 17.5 nc 20.5* 17.5 17.5 nc 20.5* 17.5 nc 20.5*		1.1	t: 0		20.7 7 1	1.10		20.2	0.04	
28.1* 27.4* 21.4 24.2 6.6 11.8 10.8 9.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 22.3 17.3 18.5 17.7 ten rostral to the AC].	Posterior cingulate cortex (PCC)	NC	NC	NC	15.4	14.0	17.7	14.3	11.8	15.2
21.4 36.0** 14.1 24.2 6.6 11.8 10.8 9.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 23.2 11.2 5.8 13.1 12.2 7.1 NC NC NC NC 25.8 13.1 12.2 7.1 NC NC 25.8 13.1 12.2 7.1 NC NC 25.8 17.3 17.3 17.3 17.3 17.3 17.3 17.3 17.3 17.3 17.3 17.5 17.5 17.7 NC	Sensorimotor striatum (SMST) ^a	26.7^{*}	35.4**	54.2***	47.0***	58.2***	57.0***	28.1^{*}	27.4*	NC
14.1 24.2 6.6 11.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 22.3 NC 22.3 NC 22.3 17.3 17.3 18.5 17.7 ren rostral to the AC].	Associative striatum (AST) ^b	14.5	28.9*	46.7**	32.7**	54.5***	60.0***	21.4	36.0**	21.0
6.6 11.8 10.8 9.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 22.3 NC 22.3 NC 22.3 17.3 18.5 17.7 17.3 18.5 17.7 ten rostral to the AC].	Limbic striatum (LST) ^c	19.2	17.0	26.5	30.5*	40.5^{**}	38.9*	14.1	24.2	14.5
10.8 9.8 NC 7.7% 5.5 NC 7.7% 34.2* 23.2 11.2 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 22.3 NC 22.3 17.3 17.3 18.5 17.7 and 17.3 18.5 17.7 ten rostral to the AC].	Anterior pre-frontal cortex (aPFC)	18.2^{*}	6.0	8.2	23.5***	15.9	28.4*	9.9	11.8	23.1*
NC 7.7% 5.5 NC 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC 22.3 NC 22.3 NC 22.3 17.3 17.8 17.3 17.3 18.5 17.7 are rostral to the AC].	Orbitofrontal cortex (OFC)	23.0*	NC	NC	31.4^{**}	13.9	9.6	10.8	9.8	14.3
 5.5 NC 34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 26.6* 23.5* 17.3 18.5 17.7 nen rostral to the ACJ. 	Thalamus	6.4	9.9	22.0	8.5	13.8	23.0	NC	7.7%	NC
34.2* 23.2 11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC NC NC 22.3 NC 22.3 17.8 17.3 18.5 17.7 17.8 17.3 18.5 17.7 are no the AC].	Hypothalamus	9.5	22.5*	36.6**	16.0	28.5**	40.6^{***}	5.5	NC	NC
11.2 5.1 31.1* 24.6 26.8* 25.8 13.1 12.2 NC NC NC NC NC 22.3 NC 22.3 17.3 17.3 18.5 17.7 17.3 18.5 17.7 17.3 18.5 17.7	Insula	6.6	7.4	7.9	38.5**	30.9*	16.7	34.2*	23.2	9.6
31.1* 24.6 26.8* 25.8 13.1 12.2 7.1 NC NC NC NC 22.3 NC 22.3 17.8 17.3 18.5 17.7 18.5 17.7 18.5 17.7	Hippocampus	6.0	13.4	12.8	16.5	17.8	19.9	11.2	5.1	8.1
26.8* 25.8 13.1 12.2 7.1 NC NC NC NC 22.3 NC 22.3 17.8 17.3 18.5 17.7 18.5 17.7 18.5 17.7 hen rostral to the AC].	Globus pallidus	18.5	21.4	28.9	44.2**	44.4***	51.2*	31.1^{*}	24.6	24.7
13.1 12.2 7.1 NC NC NC NC 22.3 NC 22.3 NC 23.5* 17.3 17.3 18.5 17.7 17.7 18.5 17.7 NC	Substantia nigra	22.1*	14.7	NC	41.4^{**}	33.9*	19.6	26.8*	25.8	16.3
7.1 NC NC NC NC NC 22.3 NC 26.6* 23.5* 17.3 17.3 18.5 17.7 hen rostral to the AC].	Red nucleus	15.4	9.9	7.7	26.5*	18.0	15.0	13.1	12.2	7.9
NC NC NC NC NC 22.3 NC 22.3 NC 23.5* 17.8 17.3 18.5 17.7 18.5 17.7 nen rostral to the AC].	Superior colliculus	NC	20.9	NC	10.4	21.8	12.7	7.1	NC	8.6
NC 22.3 NC NC 22.3 16.6* 23.5* 17.3 18.5 17.3 18.5 17.7 nen rostral to the AC].	Inferior colliculus	NC	19.9	13.4	NC	21.1	24.3	NC	NC	17.2
NC NC 26.6* 23.5* 17.8 17.3 18.5 17.3 18.5 17.7 nen rostral to the AC].	Bed nucleus of the stria terminalis (BNST)	NC	34.4^{*}	31.2*	7.5	54.3**	36.3*	NC	22.3	6.5
26.6* 23.5* 17.8 17.3 18.5 17.3 18.5 17.7 nen rostral to the AC].	Amygdala	10.4	28.8*	43.6^{*}	13.2	29.9*	45.2*	NC	NC	NC
17.8 17.3 18.5 17.3 18.5 17.7 nen rostral to the AC].	Motor group ^d	22.6*	25.0*	37.6**	42.7**	45.7***	43.8**	26.6^{*}	23.5*	11.1
18.5 nen rostral to the AC]	Cognitive group ^e	10.8	9.2	10.2	26.7**	24.9*	21.7^{*}	17.8	17.3	13.3
^a Sensorimotor striatum (SMST) = post-commissural putamen. ^b Associative striatum (AST) = pre-commissural dorsal putamen + pre-commissural dorsal caudate + post-commissural caudate. ^c Limbic striatum (LST) = ventral striatum (VST) [VST = nucleus accumbens + ventral caudate rostral to the AC + ventral putamen rostral to the AC]. ^d Motor group = SMST + globus pallidus + substantia nigra + red nucleus. ^e Coentive group = ACC + PCC + AST + aPFC + OFC + insula + hippocampus.	Psychiatric group ^f	10.6	12.0	14.4	27.4^{*}	29.1*	21.5*	18.5	17.7	9.4
	^a Sensorimotor striatum (SMST) = post-commissural ^b Associative striatum (AST) = pre-commissural dor ^c Limbic striatum (LST) = ventral striatum (VST) [Vi ^d Motor group = SMST + globus pallidus + substar ^e Cognitive group = ACC + PCC + AST + aPFC +	uissural putam ral dorsal put ST) [VST = nu substantia nigi PFC + OFC +	ten. amen + pre-co accumbi ra + red nuclei insula + hipp	mmissural do ens + ventral us. ocampus.	rsal caudate + caudate rostral	post-commissu: to the AC $+ v\epsilon$	ral caudate. entral putamer	n rostral to the	ACJ.	

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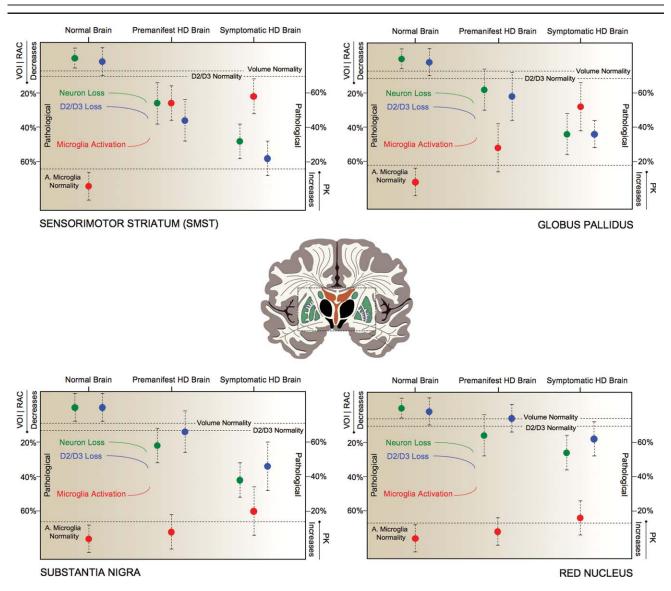
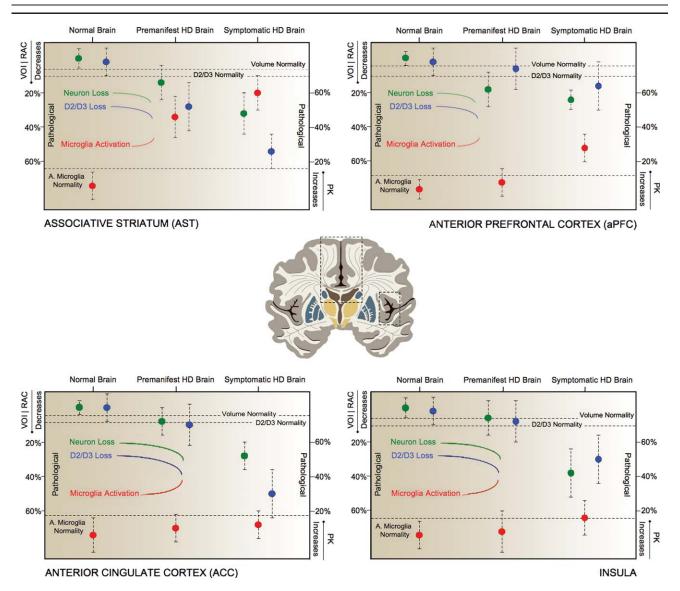


Figure 1.

Brain regions involved in motor symptoms of Huntington's disease. Decreases in volume (green), loss of D2/D3 receptors (blue) and increases in activated microglia (red) in the averaged 45-CAG repeat premanifest HD, the averaged 45-CAG repeat Symptomatic HD and Normal Brain. Decreases in RAC BP_{ND} , mm³ and increases in PK BP_{ND} in the premanifest and sympto-

and psychiatric functions. The regional group analyses is an estimation and in some cases overlapped as some, if not all, of the regions share more than one functional component. Where applicable, we have accounted the values for a region in more than one group. For example dysfunctions of the ACC has been connected with both cognitive and psychiatric dysfunction even though the cognitive component seems to be more pronounced [Bush et al., 2000]. With regards to the striatum, in this study, we have matic Huntington's disease brain are expressed as percentage changes from the normal brain values. Dotted lines represent normality range defined as mean RAC $BP_{ND} - SD$, mean $mm^3 - SD$, mean PK $BP_{ND} + SD$ for D2/D3 receptors, volume and microglia activation, respectively. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

applied and sampled a striatal object map that considers the anatomical sections with respect to functionality as described by Martinez et al. [2003]. This is a significant improvement in comparison to the traditional way of sampling the striatum by using a strict anatomical definition as it allows us to calculate separately the values for the motor (SMST), cognitive (AST) and psychiatric (LST) components of this brain region. Limitations of the study include the small sample sizes of the groups under





Regions involved in cognitive symptoms of Huntington's disease. Decreases in volume (green), loss of D2/D3 receptors (blue) and increases in activated microglia (red) in the averaged 45-CAG repeat premanifest HD, the averaged 45-CAG repeat Symptomatic HD and Normal Brain. Decreases in RAC BP_{ND} , mm³ and increases in PK BP_{ND} in the premanifest and sympto-

investigation and the resolution of PET camera for detecting changes in very small regions.

The Premanifest HD Striatum

We found that in premanifest HD cases, higher levels of activated microglia in the AST correlated with a higher probability of HD onset over the next 5 years. We did not matic Huntington's disease brain are expressed as percentage changes from the normal brain values. Dotted lines represent normality range defined as mean RAC $BP_{ND} - SD$, mean mm³ – SD, mean PK $BP_{ND} + SD$ for D2/D3 receptors, volume and microglia activation, respectively. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

observe any similar correlation in SMST or LST or in any other individual region under investigation and, therefore, it is reasonable to assume that increased levels of activated microglia in the cognitive component of the striatum may be a good predictor of clinical disease onset. Moreover, the functional and structural modalities under investigation were not similarly affected in the striatal functional subdivisions in the premanifest HD cases. SMST and AST seem to be affected whereas none of the changes under investigation in

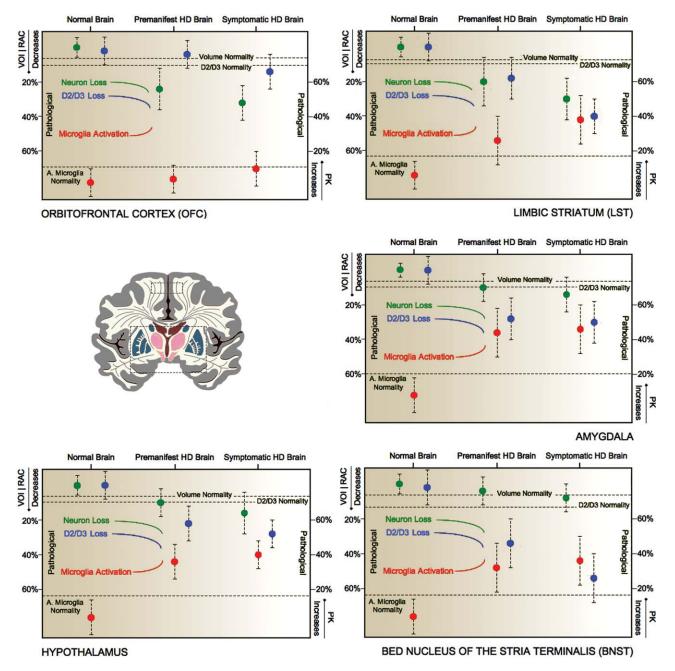


Figure 3.

Regions involved in psychiatric symptoms of Huntington's disease. Decreases in volume (green), loss of D2/D3 receptors (blue) and increases in activated microglia (red) in the averaged 45-CAG repeat premanifest HD, the averaged 45-CAG repeat Symptomatic HD and Normal Brain. Decreases in RAC BP_{ND} , mm³ and increases in PK BP_{ND} in the premanifest and sympto-

matic Huntington's disease brain are expressed as percentage changes from the normal brain values. Dotted lines represent normality range defined as mean RAC BP_{ND} – SD, mean mm³ – SD, mean PK BP_{ND} + SD for D2/D3 receptors, volume and microglia activation, respectively. [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

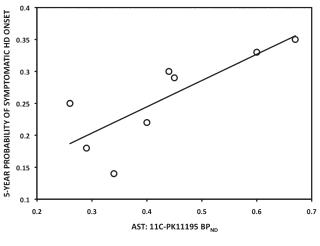


Figure 4.

Correlation between AST 11C-PK11195 BP_{ND} and the predicted 5-year probability of symptomatic HD onset in the averaged 45-CAG repeat premanifest HD brain (r = 0.8095, P = 0.0218).

LST reached statistical significance. Taken together these findings suggest, that the more dorsal part of the striatum (related to motor and cognitive function) is affected earliest in HD, whereas the ventral part of the striatum (related to psychiatric function) is affected later, possibly after the onset of motor symptoms.

The Motor Regions

From the regions comprising the motor group, SMST and substantia nigra showed significant changes in the premanifest HD, whereas globus pallidus and red nucleus did not show any changes. In symptomatic HD, we detected volume loss in all four regions, and changes from PET in SMST, globus pallidus and substantia nigra and globus pallidus. Furthermore, in SMST, increased levels of activated microglia correlated with D2/D3 binding loss in both premanifest and symptomatic HD cases and higher disease-burden scores correlated with higher levels of activated microglia in the symptomatic HD group. The worst UHDRS motor scores correlated with increased levels of activated microglia and D2/D3 receptor binding loss in the symptomatic HD group.

These findings support the primary involvement of SMST dysfunction in the motor component of the disease which appears early and the biological substrate responsible could be related to the close relationship between D2/D3 receptor loss and increased levels of activated microglia. Also, the levels of microglia activation appear to be related to the severity of the clinical disease once the motor symptoms have been established.

The Cognitive Regions

When the cognitive group of regions was assessed together we found significant changes in symptomatic HD for all the imaging modalities under investigation. In the premanifest HD, AST, aPFC and OFC showed significant changes, whereas ACC, PCC, insula and hippocampus did not show any statistical significant differences. The symptomatic HD cases showed additional changes in ACC, insula, in the AST and aPFC.

These findings provide evidence for the central early role of AST pathology in HD, which may explain some of the cognitive dysfunction seen early in the disease [Ho et al., 2003]. Furthermore, the loss of D2/D3 receptors in the ACC may also underlie in part the cognitive dysfunction seen in the disease as in this area D2/D3 receptors modulate executive functioning [Lumme et al., 2007]. Insula, OFC and aPFC are regions involved in many 'high-level' cognitive processes [Peinemann et al., 2005; Ramnani and Owen, 2004; Rosas et al., 2005] and their dysfunction reported in this study could also contribute to the cognitive dysfunction seen in HD.

As discussed earlier, the increased levels of activated microglia in AST, but not in SMST or elsewhere, proved to be a strong regional predictor of developing symptomatic onset over the next 5 years. However, when we investigated for a similar correlation averaging the PK BP_{ND} per unit of volume across all the sampled regions with possible connection to cognitive disease, this positive correlation became even stronger. This intriguing finding provides evidence that increased levels of activated microglia in the regional network with possible connection to cognitive disease could serve as a good predictor of clinical onset, highlighting the importance of a regular cognitive assessment from an early premanifest stage.

The importance of the combined assessment of the cognitive group of regions is highlighted even further in the symptomatic HD cases as in these, increased levels of

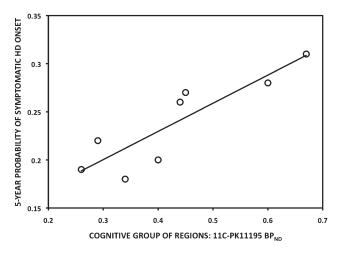


Figure 5.

Correlation between Cognitive Group of Regions 11C-PK11195 BP_{ND} and the predicted 5-year probability of symptomatic HD onset in the averaged 45-CAG repeat premanifest HD brain (r = 0.8810, P = 0.0072).

activated microglia significantly correlated with worse TFC scores and consequently with the capacity to maintain a job, with the need for assistance in dealing with finance or domestic chores and with the level of care needed for the patients. However, these findings from functional imaging need to be confirmed with neuropsychological testing in a further study.

The Psychiatric Regions

Assessing the psychiatric regions together, volume and D2/D3 binding were significantly reduced and levels of activated microglia significantly increased only in symptomatic HD cases. The function of ACC, OFC, insula and hippocampus along with their role in cognitive function, share a psychiatric component and OFC is known to be involved in emotion and reward and insula in anger, fear, disgust, happiness and sadness. Also, D2/D3 receptor loss in ACC has been correlated with the positive symptoms in Schizophrenia [Suhara et al., 2002]. Similar symptoms are observed in HD and the D2/D3 receptor loss that we found in the ACC could play a role in the development of some of the psychiatric symptoms observed in these patients.

We also found significant loss of D2/D3 binding and increased levels of activated microglia in the hypothalamus, BNST and amygdala of the premanifest HD cases.

While the hypothalamic involvement in HD has been extensively discussed in a previous report [Politis et al., 2008], BSTN and amygdala are limbic structures well connected to each other and to hypothalamus and brainstem and they mediate many autonomic and behavioural responses to aversive or threatening stimuli [Walker et al., 2003]. Their dysfunction may partly explain the prevalent and early emergence of psychiatric problems in HD.

Interpretation of Atrophy

One of the main issues with PET studies that this combined neuroimaging approach wanted to address, is the interpretation of the functional PET data with regards to volume loss. In other words, are the D2/D3 binding loss and increases in activated microglia a unique pathogenic event or are they reflecting structural changes and the secondary effect of atrophy? To address this important question, grey matter volumes were extracted for each region under investigation. These volumes were the same as used for all MRI, RAC BP_{ND} and PK BP_{ND} sampling. Hence, we investigated whether MR volume changes associate with PET BP_{ND} values changes between normality and disease. Examining each of the regions individually or as a combined group of regions none of these associations reached statistical significance and in many cases the nonparametric correlation coefficient was in the opposite direction providing supporting evidence that the PET BP_{ND} values are possibly independent of the reductions in volume.

D2 Receptors

Raised dopaminergic neurotransmission has been implicated in HD as underlying hyperkinetic symptoms and the psychotic features. The cellular localization of dopamine D2 receptors has been demonstrated on both pyramidal neurons and nonpyramidal interneurons, which use GABA as an inhibitory transmitter [Le Moine and Gaspar, 1998]. Unfortunately, RAC cannot discriminate D2 receptors on between different types of neurons. However, loss of inhibitory GABAergic neurons may cause dopaminergic activation, while loss of dopaminergic receptors may lead to compensatory hyperactivity in the remaining neurons and this process, as it is shown in this study, begins in the premanifest stage and intensifies with the progression of the illness affecting regional networks responsible for the motor, cognitive and psychiatric component of the disease.

Activated Microglia

In this study we have also demonstrated a close association between the levels of microglial activation and clinical scales of disease severity and motor dysfunction, measures of daily living and most importantly the 5-year prediction of HD onset. Pathologically activated microglia could result in neuronal death through the release of a variety of cytokines and other neurotoxic factors [Nakanishi, 2003] and could potentially trigger a cell-autonomous immune reaction. Also, monocytes are the peripheral counterparts of microglia and it was recently shown that monocytes from HD patients express mutant huntingtin and are dysfunctional even in the premanifest disease stage in both HD patients and mice [Björkqvist et al., 2008]. Our findings that increased levels of activated microglia in premanifest disease and in areas related to motor, cognitive and psychiatric functions along with the aforementioned clinical correlations, enhance the possible pathological importance of activated microglia in the development of clinical signs and symptoms and disease pathogenesis.

CONCLUSIONS

The identification of new biomarkers is a constant need for HD and we believe that a combined neuroimaging approach has the potential to serve this role. This study provides an example of how combined neuroimaging could be used in this way, as we were able first to relate PK PET to MRI findings and show that the increased levels of activated microglia represent a potential pathogenic effect that is not simply secondary to structural changes. Then we were able to correlate the microglial activation effect with the 5-year probability of premanifest HD gene carriers developing the disease [Langbehn et al., 2004]. However, for such a biomarker to be useful, larger longitudinal studies are needed in order for normality values to be derived and the range of variance to be ascertained in establishing the normal biological range. These biomarkers identify potential pathogenic processes and will be useful for studying the pharmacological responses to novel therapies.

In conclusion, this study provides evidence for the involvement of multiple brain regions in the premanifest and symptomatic HD, although further studies are needed to better understand the relation of these changes to clinical parameters. The use of multimodal imaging has a potential as a biomarker for HD and in this study we showed that imaging activated microglia in the AST and related regions associated with cognitive function provides a prognostic tool for the disease clinical onset. Further studies are needed to investigate the amount of time ahead of disease that activated microglia in these regions can predict clinical disease as well as their prognostic role in the disease progression.

ACKNOWLEDGMENTS

We would specially like to thank the patients and their families. Nicola Pavese and David J. Brooks received consultancy fees from GE Healthcare.

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