



Draft Genome Sequences of Type Strains of Adlercreutzia muris and Ellagibacter urolithinifaciens, Belonging to the Family Eggerthellaceae

Nicolas Danylec,^a Dominic A. Stoll,^a Melanie Huch^a

^aMax Rubner-Institut, Federal Research Institute of Nutrition and Food, Department of Safety and Quality of Fruit and Vegetables, Karlsruhe, Germany

ABSTRACT Here, we report the annotated draft genome sequences of two type strains belonging to the family *Eggerthellaceae* within the class *Coriobacteriia* (phylum *Actinobacteria*), *Adlercreutzia muris* WCA-131-CoC-2 (= DSM 29508 = KCTC 15543) and *Ellagibacter urolithinifaciens* CEBAS 4A (= CCUG 70284 = DSM 104140).

S pecies of the family *Eggerthellaceae* are typical members of the mammalian gut and have been isolated from, e.g., humans (1, 2), mice (3–5), and sheep (6). Several *Eggerthellaceae* species have been reported to metabolize secondary plant compounds, especially polyphenols like ellagic acid (2), daidzein (3), and resveratrol (7). *Ellagibacter urolithinifaciens* CEBAS 4A^T was isolated from the feces of a 41-year-old healthy male during the search for a bacterial strain that is able to metabolize ellagic acid. This strain is capable of metabolizing ellagic acid into isourolithin A (2). The type strain of *Adlercreutzia muris* was isolated as *Enterorhabdus muris* from the colonic content of a wild-type C57BL/6 mouse (5, 8). The genus *Enterorhabdus* showed only a low genetic divergence with the genus *Adlercreutzia* (9, 10).

In this study, we announce the annotated draft genome sequences of the type strains of *Adlercreutzia muris* and *Ellagibacter urolithinifaciens*. With this announcement, we provide the remaining annotated draft genome sequences of type strains of the family *Eggerthellaceae* with standing names in nomenclature (11).

The following type strains were obtained from the German Collection of Microorganisms and Cell Cultures (DSMZ): *A. muris* DSM 29508 and *E. urolithinifaciens* DSM 104140. Strains were cultured, and genomic DNA was isolated as previously described (1, 12, 13). *Adlercreutzia muris* DSM 29508^T was cultured (37°C) under anaerobic conditions of N₂-CO₂ (80:20) in flushed brain heart infusion medium (Merck) supplemented with 0.5% yeast extract, 0.05% L-cysteine monohydrochloride (Roth), 1 mg ml⁻¹ resazurin sodium salt, 2.5 mg liter⁻¹ heme solution, and 2 μ g ml⁻¹ vitamin K₁ solution (Sigma-Aldrich). *Ellagibacter urolithinifaciens* DSM 104140^T was cultured (37°C) under anaerobic conditions of N₂-CO₂ (80:20) in flushed anaerobe basal broth (Oxoid). DNA extraction was done using a Qiagen blood and tissue kit. DNA was quantified with a double-stranded DNA (dsDNA) high-sensitivity (HS) assay on a Qubit version 2.0 fluorometer (Thermo Fisher Scientific), according to the manufacturer's instructions, and adjusted to a concentration of 0.2 ng μ l⁻¹.

The sequencing library was constructed using a Nextera XT DNA library prep kit and a Nextera XT index kit (Illumina). Whole-genome shotgun sequencing was performed on an Illumina MiSeq benchtop sequencer using a 500-cycle v2 kit (read length, 2×250 bp). In total, 1,469,853 reads were generated for *A. muris* DSM 29508^T, while 1,067,866 reads were generated for *E. urolithinifaciens* DSM 104140^T. Data processing was done as previously described (12, 13). Sequence reads were quality trimmed using Trimmomatic version 0.39 (14) and assembled using SPAdes version 3.13.1 with default Citation Danylec N, Stoll DA, Huch M. 2019. Draft genome sequences of type strains of Adlercreutzia muris and Ellagibacter urolithinifaciens, belonging to the family Eggerthellaceae. Microbiol Resour Announc 8:e01306-19. https://doi.org/10.1128/MRA .01306-19.

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Address correspondence to Dominic A. Stoll, Dominic.Stoll@mri.bund.de.

N.D. and D.A.S. contributed equally to this work.

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	Genome	No. of contigs		Total no.	G+C	GenBank
Bacterial species (strain)	size (bp)	(avg coverage [×])	N ₅₀ (bp)	of genes	content (%)	accession no.
Adlercreutzia muris (DSM 29508)	2,765,182	94 (134.19)	79,375	2,303	65.1	WAJS0000000
Ellagibacter urolithinifaciens (DSM 104140)	2,462,117	81 (127.67)	100,872	2,161	60.4	WAJR0000000

TABLE 1 Accession numbers, assembly metrics, and annotated features of the type strains of *Adlercreutzia muris* and *Ellagibacter urolithinifaciens*

settings in the "careful" mode (15, 16). The estimated insert sizes of A. muris DSM 29508^T and *E. urolithinifaciens* DSM 104140^T were 255.724 bp and 297.597 bp, respectively. Adequate trimming was verified by mapping the adapter sequences to the assembled contigs using Bowtie 2 version 2.3.3.1 (17). To eliminate sequence contamination, the contigs were aligned to the genome of coliphage phi-X174 (GenBank accession number NC_001422) using a BLASTn search (18). All contigs of <500 bp were manually excluded, and renaming of contigs was done using awk (19). To calculate genome coverage of each strain, trimmed reads were mapped against remaining contigs by Bowtie 2. In the case of A. muris DSM 29508[⊤], 99.15% of 1,463,425 trimmed reads could be mapped against the respective contigs, whereas in the case of E. urolithinifaciens DSM 104140^T, 99.18% of 1,064,956 trimmed reads could be mapped against the contigs. Draft genome sequences were annotated using the automated NCBI Prokaryotic Genome Annotation Pipeline (20). The assembly metrics and annotated features of both type strains are given in Table 1. ResFinder version 3.2 (21) was used to evaluate the draft genome sequences for antimicrobial resistance (AMR) genes. For both strains, no AMR gene was identified.

Data availability. This whole-genome shotgun project, including raw reads of *A. muris* DSM 29508^T and *E. urolithinifaciens* DSM 104140^T, has been deposited at DDBJ/ ENA/GenBank under BioProject number PRJNA574580. The versions described in this publication are the first versions and are listed in Table 1. The raw reads of *A. muris* DSM 29508^T have been deposited under SRA accession number SRX6974936, while the raw reads of *E. urolithinifaciens* DSM 104140^T have been deposited under accession number SRX6974937.

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We declare that there are no conflicts of interest.

REFERENCES

- Danylec N, Gobl A, Stoll DA, Hetzer B, Kulling SE, Huch M. 2018. *Rubneribacter badeniensis* gen. nov., sp. nov. and *Enteroscipio rubneri* gen. nov., sp. nov., new members of the *Eggerthellaceae* isolated from human faeces. Int J Syst Evol Microbiol 68:1533–1540. https://doi.org/10.1099/ ijsem.0.002705.
- Beltrán D, Romo-Vaquero M, Espín JC, Tomás-Barberán FA, Selma MV. 2018. *Ellagibacter isourolithinifaciens* gen. nov., sp. nov., a new member of the family *Eggerthellaceae*, isolated from human gut. Int J Syst Evol Microbiol 68:1707–1712. https://doi.org/10.1099/ijsem.0.002735.
- Clavel T, Charrier C, Braune A, Wenning M, Blaut M, Haller D. 2009. Isolation of bacteria from the ileal mucosa of TNF^{ΔARE} mice and description of *Enterorhabdus mucosicola* gen. nov., sp. nov. Int J Syst Evol Microbiol 59:1805–1812. https://doi.org/10.1099/ijs.0.003087-0.
- Clavel T, Duck W, Charrier C, Wenning M, Elson C, Haller D. 2010. Enterorhabdus caecimuris sp. nov., a member of the family Coriobacteriaceae isolated from a mouse model of spontaneous colitis, and emended description of the genus Enterorhabdus Clavel et al. 2009. Int J Syst Evol Microbiol 60: 1527–1531. https://doi.org/10.1099/ijs.0.015016-0.
- Lagkouvardos I, Pukall R, Abt B, Foesel BU, Meier-Kolthoff JP, Kumar N, Bresciani A, Martinez I, Just S, Ziegler C, Brugiroux S, Garzetti D, Wenning M, Bui TP, Wang J, Hugenholtz F, Plugge CM, Peterson DA, Hornef MW, Baines JF, Smidt H, Walter J, Kristiansen K, Nielsen HB, Haller D, Overmann J, Stecher B, Clavel T. 2016. The Mouse Intestinal Bacterial Collec-

tion (miBC) provides host-specific insight into cultured diversity and functional potential of the gut microbiota. Nat Microbiol 1:16131. https://doi.org/10.1038/nmicrobiol.2016.131.

- Lanigan GW. 1976. Peptococcus heliotrinreducans, sp. nov., a cytochromeproducing anaerobe which metabolizes pyrrolizidine alkaloids. J Gen Microbiol 94:1–10. https://doi.org/10.1099/00221287-94-1-1.
- Bode LM, Bunzel D, Huch M, Cho GS, Ruhland D, Bunzel M, Bub A, Franz CM, Kulling SE. 2013. *In vivo* and *in vitro* metabolism of *trans*-resveratrol by human gut microbiota. Am J Clin Nutr 97:295–309. https://doi.org/ 10.3945/ajcn.112.049379.
- Oren A, Garrity GM. 2016. List of new names and new combinations previously effectively, but not validly, published. Int J Syst Evol Microbiol 66:4299–4305.
- Nouioui I, Carro L, García-López M, Meier-Kolthoff JP, Woyke T, Kyrpides NC, Pukall R, Klenk H-P, Goodfellow M, Göker M. 2018. Genome-based taxonomic classification of the phylum *Actinobacteria*. Front Microbiol 9:2007. https://doi.org/10.3389/fmicb.2018.02007.
- Oren A, Garrity GM. 2019. Notification of changes in taxonomic opinion previously published outside the IJSEM. Int J Syst Evol Microbiol 69: 13–32. https://doi.org/10.1099/ijsem.0.003171.
- Parte AC. 2018. LPSN—List of Prokaryotic names with Standing in Nomenclature (bacterio.net), 20 years on. Int J Syst Evol Microbiol 68: 1825–1829. https://doi.org/10.1099/ijsem.0.002786.

- Danylec N, Stoll DA, Dotsch A, Huch M. 2019. Draft genome sequences of type strains of Gordonibacter faecihominis, Paraeggerthella hongkongensis, Parvibacter caecicola, Slackia equolifaciens, Slackia faecicanis, and Slackia isoflavoniconvertens. Microbiol Resour Announc 8:e01532-18. https://doi.org/10.1128/MRA.01532-18.
- Stoll DA, Danylec N, Dotsch A, Becker B, Huch M. 2018. Draft genome sequence of Salmonella enterica subsp. enterica serovar Enteritidis MS 501, a potential human pathogen isolated from red lettuce (Lactuca sativa var. capitata) in Karlsruhe, Germany. Microbiol Resour Announc 7:e00938-18. https://doi.org/10.1128/MRA.00938-18.
- 14. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. J Comput Biol 20:714–737. https://doi.org/10.1089/cmb .2013.0084.
- 16. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS,

Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.

- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. Nat Methods 9:357–359. https://doi.org/10.1038/nmeth.1923.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic Local Alignment Search Tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- Aho AV, Kernighan BW, Weinberger PJ. 1979. Awk—a pattern scanning and processing language. Softw Pract Exp 9:267–279. https://doi.org/10 .1002/spe.4380090403.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi .org/10.1093/nar/gkw569.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi.org/ 10.1093/jac/dks261.