



High *In Vitro* Susceptibility to the First-in-Class Spiropyrimidinetrione Zoliflodacin among Consecutive Clinical *Neisseria gonorrhoeae* Isolates from Thailand and South Africa

Susanne Jacobsson,^a Ranmini Kularatne,^b Rossaphorn Kittiyaowamarn,^c Venessa Maseko,^b Porntip Paopang,^c Pongsathorn Sangprasert,^c Pachara Sirivongrangson,^d Laura Piddock,^e Teodora Wi,^f Emilie Alirol,^e Magnus Unemo^a

^aWHO Collaborating Centre for Gonorrhoea and Other STIs, National Reference Laboratory for Sexually Transmitted Infections, Department of Laboratory Medicine, Faculty of Medicine and Health, Örebro University, Örebro, Sweden

^bCentre for HIV and STIs, National Institute for Communicable Diseases, National Health Laboratory Service, Johannesburg, South Africa

^cBangrak STI Center, Bureau of AIDS, TB, and STIs, Department of Disease Control, Ministry of Public Health, Bangkok, Thailand

^dDepartment of Disease Control, Ministry of Public Health, Bangkok, Thailand

^eGlobal Antibiotic Research & Development Partnership, Geneva, Switzerland

^fDepartment of Reproductive Health, World Health Organization, Geneva, Switzerland

ABSTRACT We evaluated the *in vitro* susceptibility to the first-in-class spiropyrimidinetrione zoliflodacin among recent consecutive clinical *Neisseria gonorrhoeae* isolates cultured in Thailand ($n = 99$) (in 2018) and South Africa ($n = 100$) (in 2015 to 2017). Zoliflodacin was highly active *in vitro* against all tested isolates (MIC range, 0.004 to 0.25 $\mu\text{g/ml}$; MIC₅₀, 0.064 $\mu\text{g/ml}$; MIC₉₀, 0.125 $\mu\text{g/ml}$), with no cross-resistance to any of the seven comparator antimicrobials. Our data support the initiation of the global phase 3 randomized controlled clinical trial of zoliflodacin for uncomplicated gonorrhea.

KEYWORDS South Africa, Thailand, gonorrhea, resistance, spiropyrimidinetrione, susceptibility, treatment, zoliflodacin

The sexually transmitted infection (STI) gonorrhea, including its severe complications and sequelae, is a significant public health concern globally. The World Health Organization (WHO) estimated 87 million new cases of gonorrhea globally among adults in 2016 (1). Antimicrobial resistance in *Neisseria gonorrhoeae* has been a major concern for many decades, which has severely limited current treatment options internationally. The emergence of resistance to the extended-spectrum cephalosporins threatens the effectiveness of ceftriaxone, the last remaining option for empirical first-line monotherapy of gonorrhea globally (2–17). Novel effective antimicrobials for the treatment of urogenital and extragenital gonorrhea are essential (2, 3, 18, 19).

Zoliflodacin is the first-in-class spiropyrimidinetrione and the drug in most advanced clinical development for the treatment of uncomplicated gonorrhea (19, 20). Zoliflodacin is a topoisomerase II inhibitor, like the fluoroquinolones, but with a novel target in GyrB and a distinct mechanism of action (20). A phase 2 randomized controlled clinical trial (RCT) showed that zoliflodacin (3 g, orally), in the per-protocol analyses, achieved high cure rates among patients with uncomplicated urogenital gonorrhea (100% [47/47 patients]) and rectal gonorrhea (100% [6/6 patients]) but a reduced cure rate for the small number of pharyngeal gonorrhea cases (78% [7/9 patients]). Zoliflodacin was well tolerated, with transient gastrointestinal upset being the most frequently reported adverse event (21). A multicontinent phase 3 RCT to evaluate the clinical efficacy, tolerability, and safety in patients with uncomplicated urogenital and extragenital gonorrhea is planned in 2019 (22). This phase 3 RCT will include gonorrhea patients in the United States, the Netherlands, Thailand, and South Africa. *In vitro*

Citation Jacobsson S, Kularatne R, Kittiyaowamarn R, Maseko V, Paopang P, Sangprasert P, Sirivongrangson P, Piddock L, Wi T, Alirol E, Unemo M. 2019. High *in vitro* susceptibility to the first-in-class spiropyrimidinetrione zoliflodacin among consecutive clinical *Neisseria gonorrhoeae* isolates from Thailand and South Africa. *Antimicrob Agents Chemother* 63:e01479-19. <https://doi.org/10.1128/AAC.01479-19>.

Copyright © 2019 American Society for Microbiology. All Rights Reserved.

Address correspondence to Magnus Unemo, magnus.unemo@regionorebrolan.se.

Received 22 July 2019

Returned for modification 22 August 2019

Accepted 15 September 2019

Accepted manuscript posted online 23 September 2019

Published 21 November 2019

TABLE 1 MIC values for zoliflodacin and therapeutic antimicrobials currently or previously recommended for gonorrhea treatment in *Neisseria gonorrhoeae* isolates obtained in Thailand (in 2018) and South Africa (in 2015 to 2017)

Antimicrobial and isolate group	MIC ($\mu\text{g/ml}$) ^a				Susceptibility status (%) ^b		
	Range	Modal MIC	MIC ₅₀	MIC ₉₀	S	I	R
Zoliflodacin							
All isolates (<i>n</i> = 199)	0.004 to 0.25	0.064	0.064	0.125	ND		
South African isolates (<i>n</i> = 100)	0.004 to 0.25	0.064	0.064	0.125	ND		
Thai isolates (<i>n</i> = 99)	0.004 to 0.125	0.032	0.032	0.064	ND		
Ciprofloxacin-resistant isolates ^b (<i>n</i> = 177)	0.004 to 0.125	0.064	0.064	0.125	ND		
Ciprofloxacin-susceptible isolates ^b (<i>n</i> = 22)	0.032 to 0.25	0.064	0.064	0.125	ND		
Ceftriaxone, all isolates (<i>n</i> = 199)	<0.002 to 0.064	0.004	0.004	0.008	100	ND	0
Azithromycin, all isolates (<i>n</i> = 199)	0.032 to 4	0.125	0.125	0.25	99.0	ND	1.0
Ciprofloxacin, all isolates (<i>n</i> = 199)	<0.002 to >32	2	2	4	11.1	ND	88.9
Penicillin G, all isolates (<i>n</i> = 199)	0.125 to >32	>32	4	>32	0	31.7	68.3
Spectinomycin, all isolates (<i>n</i> = 199)	8 to 32	16	16	16	100	ND	0
Tetracycline, all isolates (<i>n</i> = 199)	0.25 to 128	32	32	32	5.5	6.5	87.9
Gentamicin, all isolates (<i>n</i> = 199)	2 to 8	8	8	8	ND		

^aMICs were determined using the agar dilution technique for zoliflodacin and Etest for the other antimicrobials.

^bS, susceptible; I, intermediate susceptible; R, resistant; ND, not determined (due to the lack of interpretative criteria). The EUCAST clinical breakpoints were applied for all antimicrobials with the exception of azithromycin, for which no resistance breakpoint exists; instead, the ECOFF value (1 $\mu\text{g/ml}$) was used to distinguish isolates with azithromycin resistance determinants (31).

susceptibility to zoliflodacin was proven to be high in the United States (23) and the Netherlands (24). However, the susceptibility to zoliflodacin among *N. gonorrhoeae* strains circulating in Thailand and South Africa remains unknown. In Thailand, gonorrhea is highly prevalent, particularly in certain high-risk groups, e.g., men who have sex with men (MSM) (25), and it is a major concern that extensively drug-resistant gonococcal isolates might be spreading in Thailand and other countries in Southeast Asia (13, 14). In South Africa, the prevalence of gonorrhea is very high in both women and men (26). Furthermore, the *N. gonorrhoeae* resistance to traditional therapeutic antimicrobials for gonorrhea is also high (27), and treatment failures with the extended-spectrum cephalosporin cefixime have been verified (28).

We assessed the *in vitro* susceptibility to zoliflodacin among consecutive clinical *N. gonorrhoeae* isolates cultured in Thailand (*n* = 99) (in 2018) and South Africa (*n* = 100) (in 2015 to 2017), to provide data that are crucial to obtain prior to initiating the multicountry phase 3 RCT of zoliflodacin. The *N. gonorrhoeae* isolates were cultured from male patients with urethritis presenting to primary health care centers in Kwa Zulu Natal (*n* = 40) (in 2015) and Gauteng (*n* = 60) (in 2017) Provinces, South Africa, and from male patients (*n* = 37) and female patients (*n* = 62) attending the Bangrak STI center in Bangkok, Thailand, in 2018. Antimicrobial susceptibility profiles, including MICs, were obtained for all *N. gonorrhoeae* isolates by the agar dilution technique, according to CLSI guidelines (29), for zoliflodacin as described previously (30) and by Etest, according to the manufacturer's instructions (bioMérieux, Marcy-l'Étoile, France), for seven other antimicrobials (ceftriaxone, azithromycin, ciprofloxacin, penicillin G, spectinomycin, tetracycline, and gentamicin). The agar dilution technique and Etests were performed in parallel, using the same inoculum of the *N. gonorrhoeae* isolates. The EUCAST clinical breakpoints were applied for all antimicrobials, with the exception of azithromycin, for which no resistance breakpoint exists; instead, the epidemiological cutoff (ECOFF) value was used to distinguish isolates with azithromycin resistance determinants (31). For quality control, the WHO reference strains A, F, and P (32, 33) were included in each antimicrobial susceptibility testing run.

The results of all antimicrobial susceptibility tests are summarized in Table 1, in which the zoliflodacin MIC data are categorized as the zoliflodacin susceptibility in all isolates (*n* = 199), in South African isolates (*n* = 100), in Thai isolates (*n* = 99), in ciprofloxacin-resistant isolates (*n* = 177), and in ciprofloxacin-susceptible isolates (*n* = 22). The MIC data for all other tested antimicrobials are also summarized. Briefly, zoliflodacin was highly active *in vitro* against all *N. gonorrhoeae* isolates (*n* = 199), with

TABLE 2 Zoliflodacin MIC distributions

Country	No. (cumulative %) with zoliflodacin MIC of:						Zoliflodacin MIC ₅₀ (μg/ml)	Zoliflodacin MIC ₉₀ (μg/ml)
	0.004 μg/ml	0.016 μg/ml	0.032 μg/ml	0.064 μg/ml	0.125 μg/ml	0.25 μg/ml		
Thailand (<i>n</i> = 99)	10 (10)		49 (60)	37 (97)	3 (100)		0.032	0.064
South Africa (<i>n</i> = 100)	2 (2)	1 (3)	21 (24)	48 (72)	27 (99)	1 (100)	0.064	0.12

a MIC range of 0.004 to 0.25 μg/ml, a modal MIC of 0.064 μg/ml, a MIC₅₀ of 0.064 μg/ml, and a MIC₉₀ of 0.125 μg/ml. The modal MIC, MIC₅₀, and MIC₉₀ were 1 MIC log₂ dilution higher in the South African isolates than in the Thai isolates (Table 1). The majority of isolates (93% [*n* = 185]) displayed MICs ranging from 0.032 μg/ml to 0.125 μg/ml, 6.5% (*n* = 13) of the isolates displayed lower MICs (0.004 or 0.016 μg/ml), and only 0.5% (*n* = 1) had a zoliflodacin MIC of 0.25 μg/ml (Table 2).

The levels of resistance to conventional gonorrhea therapeutic antimicrobials in South African isolates and Thai isolates were 78% and 100%, respectively, for ciprofloxacin, 85% and 91% for tetracycline, 57% and 80% for penicillin G, 0% and 2% for azithromycin, 0% and 0% for spectinomycin, and 0% and 0% for ceftriaxone (Table 1). With the exception of ceftriaxone, the modal MIC, MIC₅₀, and MIC₉₀ of all antimicrobials were substantially higher than those observed for zoliflodacin. The zoliflodacin MIC₉₀ (0.125 μg/ml) was 16-fold higher than that of ceftriaxone (0.008 μg/ml), 2-fold lower than that of azithromycin (0.25 μg/ml), 32-fold lower than that of ciprofloxacin (4 μg/ml), and >32-fold lower than those of gentamicin, spectinomycin, tetracycline, and penicillin G (8 μg/ml, 16 μg/ml, 32 μg/ml, and >32 μg/ml, respectively). No cross-resistance between zoliflodacin and the tested conventional gonorrhea therapeutic antimicrobials was found. The modal MIC, MIC₅₀, and MIC₉₀ values were identical for ciprofloxacin-susceptible and ciprofloxacin-resistant isolates; additionally, the Spearman's correlation coefficient for the zoliflodacin MICs versus the ciprofloxacin MICs was -0.064, suggesting a lack of correlation between the MICs of the two antimicrobials.

In conclusion, zoliflodacin was highly active *in vitro* against contemporary consecutive clinical *N. gonorrhoeae* isolates from Thailand and South Africa, and no cross-resistance with any of the tested conventional gonorrhea therapeutic antimicrobials was found. The levels of susceptibility to the internationally recommended therapeutic antimicrobials ceftriaxone, cefixime, azithromycin, and spectinomycin were also high. Zoliflodacin previously demonstrated potent *in vitro* activity against temporally and genetically diverse clinical *N. gonorrhoeae* isolates and international reference strains from 21 European Union/European Economic Area countries (MIC₅₀, 0.064 μg/ml; MIC₉₀, 0.125 μg/ml; MIC range, 0.002 to 0.25 μg/ml), the United States (MIC₅₀, 0.06 μg/ml; MIC₉₀, 0.125 μg/ml; MIC range, 0.008 to 0.25 μg/ml), and China (MIC₅₀, 0.03 μg/ml; MIC₉₀, 0.06 μg/ml; MIC range, ≤0.002 to 0.125 μg/ml) (23, 24, 30, 34). Zoliflodacin has additionally shown *in vitro* activity against other STI pathogens, i.e., *Chlamydia trachomatis* and *Mycoplasma genitalium* (35, 36). The results of our study support the idea that zoliflodacin is a promising novel oral therapeutic option for gonorrhea, and they were critical for the timely initiation of the multicontinent phase 3 RCT of zoliflodacin for uncomplicated gonorrhea.

ACKNOWLEDGMENTS

We are grateful to Entasis Therapeutics (Waltham, MA, USA) for providing zoliflodacin and to Alita Miller and John Mueller for critical review of the manuscript.

The present study was supported by the Global Antibiotic Research and Development Partnership (Geneva, Switzerland), with funding from the United Kingdom Department for International Development, the Dutch Ministry of Health, Welfare, and Sport, the South African Medical Research Council, and the German Federal Ministry of Education and Research. The work was performed at the WHO Collaborating Centre for Gonorrhoea and Other STIs, National Reference Laboratory for Sexually Transmitted

Infections, Department of Laboratory Medicine, Clinical Microbiology, Örebro University Hospital, Örebro, Sweden.

REFERENCES

- Rowley J, Vander Hoorn S, Korenromp E, Low N, Unemo M, Abu-Raddad LJ, Chico RM, Smolak A, Newman L, Gottlieb S, Thwin SS, Broutet N, Taylor MM. 2019. Chlamydia, gonorrhoea, trichomoniasis and syphilis: global prevalence and incidence estimates, 2016. *Bull World Health Organ* 97:548–562. <https://doi.org/10.2471/BLT.18.228486>.
- Wi T, Lahra MM, Ndowa F, Bala M, Dillon JR, Ramon-Pardo P, Eremin SR, Bolan G, Unemo M. 2017. Antimicrobial resistance in *Neisseria gonorrhoeae*: global surveillance and a call for international collaborative action. *PLoS Med* 14:e1002344. <https://doi.org/10.1371/journal.pmed.1002344>.
- Unemo M, Shafer WM. 2014. Antimicrobial resistance in *Neisseria gonorrhoeae* in the 21st century: past, evolution, and future. *Clin Microbiol Rev* 27:587–613. <https://doi.org/10.1128/CMR.00010-14>.
- Ohnishi M, Golparian D, Shimuta K, Saika T, Hoshina S, Iwasaku K, Nakayama S, Kitawaki K, Unemo M. 2011. Is *Neisseria gonorrhoeae* initiating a future era of untreatable gonorrhoea? Detailed characterization of the first strain with high-level resistance to ceftriaxone. *Antimicrob Agents Chemother* 55:3538–3545. <https://doi.org/10.1128/AAC.00325-11>.
- Unemo M, Golparian D, Nicholas R, Ohnishi M, Galloway A, Sednaoui P. 2012. High-level cefixime- and ceftriaxone-resistant *Neisseria gonorrhoeae* in France: novel *penA* mosaic allele in a successful international clone causes treatment failure. *Antimicrob Agents Chemother* 56:1273–1280. <https://doi.org/10.1128/AAC.05760-11>.
- Cámara J, Serra J, Ayats J, Bastida T, Carnicer-Pont D, Andreu A, Ardanuy C. 2012. Molecular characterization of two high-level ceftriaxone-resistant *Neisseria gonorrhoeae* isolates detected in Catalonia, Spain. *J Antimicrob Chemother* 67:1858–1860. <https://doi.org/10.1093/jac/dks162>.
- Lahra MM, Ryder N, Whitley DM. 2014. A new multidrug-resistant strain of *Neisseria gonorrhoeae* in Australia. *N Engl J Med* 371:1850–1851. <https://doi.org/10.1056/NEJMc1408109>.
- Deguchi T, Yasuda M, Hatazaki K, Kameyama K, Horie K, Kato T, Mizutani K, Seike K, Tsuchiya T, Yokoi S, Nakano M, Yoh M. 2016. New clinical strain of *Neisseria gonorrhoeae* with decreased susceptibility to ceftriaxone, Japan. *Emerg Infect Dis* 22:142–144. <https://doi.org/10.3201/eid2201.150868>.
- Nakayama S, Shimuta K, Furubayashi K, Kawahata T, Unemo M, Ohnishi M. 2016. New ceftriaxone- and multidrug-resistant *Neisseria gonorrhoeae* strain with a novel mosaic *penA* gene isolated in Japan. *Antimicrob Agents Chemother* 60:4339–4341. <https://doi.org/10.1128/AAC.00504-16>.
- Lefebvre B, Martin I, Demczuk W, Deshaies L, Michaud S, Labbé AC, Beaudoin MC, Longtin J. 2018. Ceftriaxone-resistant *Neisseria gonorrhoeae*, Canada, 2017. *Emerg Infect Dis* 24:381–383. <https://doi.org/10.3201/eid2402.171756>.
- Terkelsen D, Tolstrup J, Johnsen CH, Lund O, Larsen HK, Worning P, Unemo M, Westh H. 2017. Multidrug-resistant *Neisseria gonorrhoeae* infection with ceftriaxone resistance and intermediate resistance to azithromycin, Denmark, 2017. *Euro Surveill* 22:17-00659. <https://doi.org/10.2807/1560-7917.ES.2017.22.42.17-00659>.
- Poncin T, Fouere S, Braille A, Camelena F, Agsoug M, Bebear C, Kumanski S, Lot F, Mercier-Delarue S, Ngangro NN, Salmona M, Schnepf N, Timsit J, Unemo M, Bercot B. 2018. Multidrug-resistant *Neisseria gonorrhoeae* failing treatment with ceftriaxone and doxycycline in France, November 2017. *Euro Surveill* 23:1800264. <https://doi.org/10.2807/1560-7917.ES.2018.23.21.1800264>.
- Eyre DW, Sanderson ND, Lord E, Regisford-Reimmer N, Chau K, Barker L, Morgan M, Newnham R, Golparian D, Unemo M, Crook DW, Peto TE, Hughes G, Cole MJ, Fifer H, Edwards A, Andersson MI. 2018. Gonorrhoea treatment failure caused by a *Neisseria gonorrhoeae* strain with combined ceftriaxone and high-level azithromycin resistance, England, February 2018. *Euro Surveill* 23:1800323. <https://doi.org/10.2807/1560-7917.ES.2018.23.27.1800323>.
- Whitley DM, Jennison A, Pearson J, Lahra MM. 2018. Genetic characterization of *Neisseria gonorrhoeae* resistant to both ceftriaxone and azithromycin. *Lancet Infect Dis* 18:717–718. [https://doi.org/10.1016/S1473-3099\(18\)30340-2](https://doi.org/10.1016/S1473-3099(18)30340-2).
- Eyre DW, Town K, Street T, Barker L, Sanderson N, Cole MJ, Mohammed H, Pitt R, Gobin M, Irish C, Gardiner D, Sedgwick J, Beck C, Saunders J, Turbitt D, Cook C, Phin N, Nathan B, Horner P, Fifer H. 2019. Detection in the United Kingdom of the *Neisseria gonorrhoeae* FC428 clone, with ceftriaxone resistance and intermediate resistance to azithromycin, October to December 2018. *Euro Surveill* 24:1900147. <https://doi.org/10.2807/1560-7917.ES.2019.24.10.1900147>.
- Golparian D, Rose L, Lynam A, Mohamed A, Bercot B, Ohnishi M, Crowley B, Unemo M. 2018. Multidrug-resistant *Neisseria gonorrhoeae* isolate, belonging to the internationally spreading Japanese FC428 clone, with ceftriaxone resistance and intermediate resistance to azithromycin in Ireland, August 2018. *Euro Surveill* 23:1800617. <https://doi.org/10.2807/1560-7917.ES.2018.23.47.1800617>.
- Ko KKK, Chio MTW, Goh SS, Tan AL, Koh TH, Rahman N. 2019. First case of ceftriaxone-resistant multidrug-resistant *Neisseria gonorrhoeae* in Singapore. *Antimicrob Agents Chemother* 63:e02624-18. <https://doi.org/10.1128/AAC.02624-18>.
- Alirol E, Wi TE, Bala M, Bazzo ML, Chen XS, Deal C, Dillon JR, Kularatne R, Heim J, Hooft van Huijsduijnen R, Hook EW, Lahra MM, Lewis DA, Ndowa F, Shafer WM, Tayler L, Workowski K, Unemo M, Balasegaram M. 2017. Multidrug-resistant gonorrhoea: a research and development roadmap to discover new medicines. *PLoS Med* 14:e1002366. <https://doi.org/10.1371/journal.pmed.1002366>.
- Unemo M, Bradshaw CS, Hocking JS, de Vries HJC, Francis SC, Mabey D, Marrazzo JM, Sonder GJB, Schwebke JR, Hoornenborg E, Peeling RW, Philip SS, Low N, Fairley CK. 2017. Sexually transmitted infections: challenges ahead. *Lancet Infect Dis* 17:e235–e279. [https://doi.org/10.1016/S1473-3099\(17\)30310-9](https://doi.org/10.1016/S1473-3099(17)30310-9).
- Basarab GS, Kern GH, McNulty J, Mueller JP, Lawrence K, Vishwanathan K, Alm RA, Barvian K, Doig P, Galullo V, Gardner H, Gowravaram M, Huband M, Kimzey A, Morningstar M, Kutschke A, Lahiri SD, Perros M, Singh R, Schuck VJ, Tommasi R, Walkup G, Newman JV. 2015. Responding to the challenge of untreatable gonorrhoea: ETX0914, a first-in-class agent with a distinct mechanism-of-action against bacterial type II topoisomerases. *Sci Rep* 5:11827. <https://doi.org/10.1038/srep11827>.
- Taylor SN, Marrazzo J, Batteiger BE, Hook EW, III, Seña AC, Long J, Wierzbicki MR, Kwak H, Johnson SM, Lawrence K, Mueller J. 2018. Single-dose zoliflodacin (ETX0914) for treatment of urogenital gonorrhoea. *N Engl J Med* 379:1835–1845. <https://doi.org/10.1056/NEJMoa1706988>.
- Global Antibiotic Research and Development Partnership. 7 November 2018. Press release: novel antibiotic shows promise in treatment of uncomplicated gonorrhoea. <https://www.gardp.org/2018/news-resources/press-releases/novel-antibiotic-treatment-uncomplicated-gonorrhoea>.
- Papp JR, Lawrence K, Sharpe S, Mueller J, Kirkcaldy RD. 2016. *In vitro* growth of multidrug-resistant *Neisseria gonorrhoeae* isolates is inhibited by ETX0914, a novel spiroprimidinetriene. *Int J Antimicrob Agents* 48:328–330. <https://doi.org/10.1016/j.ijantimicag.2016.05.018>.
- Unemo M, Ringlander J, Wiggins C, Fredlund H, Jacobsson S, Cole M. 2015. High *in vitro* susceptibility to the novel spiroprimidinetriene ETX0914 (AZD0914) among 873 contemporary clinical *Neisseria gonorrhoeae* isolates from 21 European countries from 2012 to 2014. *Antimicrob Agents Chemother* 59:5220–5225. <https://doi.org/10.1128/AAC.00786-15>.
- Tongtoyai J, Todd CS, Chonwattana W, Pattanasin S, Chaikummao S, Varangrat A, Lokpichart S, Holtz TH, van Griensven F, Curlin ME. 2015. Prevalence and correlates of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* by anatomic site among urban Thai men who have sex with men. *Sex Transm Dis* 42:440–449. <https://doi.org/10.1097/OLQ.0000000000000311>.
- Kularatne RS, Niit R, Rowley J, Kufa-Chakezha T, Peters RPH, Taylor MM, Johnson LF, Korenromp EL. 2018. Adult gonorrhoea, chlamydia and syphilis prevalence, incidence, treatment and syndromic case reporting in South Africa: estimates using the Spectrum-STI model, 1990–2017. *PLoS One* 13:e0205863. <https://doi.org/10.1371/journal.pone.0205863>.
- Kularatne R, Maseko V, Gumede L, Kufa T. 2018. Trends in *Neisseria gonor-*

- rhoeae* antimicrobial resistance over a ten-year surveillance period, Johannesburg, South Africa, 2008–2017. *Antibiotics* (Basel) 7:E58. <https://doi.org/10.3390/antibiotics7030058>.
28. Lewis DA, Sriruttan C, Müller EE, Golparian D, Gumedde L, Fick D, de Wet J, Maseko V, Coetzee J, Unemo M. 2013. Phenotypic and genetic characterization of the first two cases of extended-spectrum-cephalosporin-resistant *Neisseria gonorrhoeae* infection in South Africa and association with cefixime treatment failure. *J Antimicrob Chemother* 68:1267–1270. <https://doi.org/10.1093/jac/dkt034>.
 29. Clinical and Laboratory Standards Institute. 2015. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard—10th ed. M07-A10. Clinical and Laboratory Standards Institute, Wayne, PA.
 30. Jacobsson S, Golparian D, Alm RA, Huband M, Mueller J, Jensen JS, Ohnishi M, Unemo M. 2014. High *in vitro* activity of the novel spiropyrimidinetrione AZD0914, a DNA gyrase inhibitor, against multidrug-resistant *Neisseria gonorrhoeae* isolates suggests a new effective option for oral treatment of gonorrhea. *Antimicrob Agents Chemother* 58:5585–5588. <https://doi.org/10.1128/AAC.03090-14>.
 31. European Committee on Antimicrobial Susceptibility Testing. 2019. Breakpoint tables for interpretation of MICs and zone diameters, version 9.0. eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_9.0_Breakpoint_Tables.pdf.
 32. Unemo M, Fasth O, Fredlund H, Limnios A, Tapsall J. 2009. Phenotypic and genetic characterization of the 2008 WHO *Neisseria gonorrhoeae* reference strain panel intended for global quality assurance and quality control of gonococcal antimicrobial resistance surveillance for public health purposes. *J Antimicrob Chemother* 63:1142–1151. <https://doi.org/10.1093/jac/dkp098>.
 33. Unemo M, Golparian D, Sánchez-Busó L, Grad Y, Jacobsson S, Ohnishi M, Lahra MM, Limnios A, Sikora AE, Wi T, Harris SR. 2016. The novel 2016 WHO *Neisseria gonorrhoeae* reference strains for global quality assurance of laboratory investigations: phenotypic, genetic and reference genome characterization. *J Antimicrob Chemother* 71:3096–3108. <https://doi.org/10.1093/jac/dkw288>.
 34. Su XH, Wang BX, Le WJ, Liu YR, Wan C, Li S, Alm RA, Mueller JP, Rice PA. 2016. Multidrug-resistant *Neisseria gonorrhoeae* isolates from Nanjing, China, are sensitive to killing by a novel DNA gyrase inhibitor, ETX0914 (AZD0914). *Antimicrob Agents Chemother* 60:621–623. <https://doi.org/10.1128/AAC.01211-15>.
 35. Kohlhoff SA, Huband MD, Hammerschlag MR. 2014. In vitro activity of AZD0914, a novel DNA gyrase inhibitor, against *Chlamydia trachomatis* and *Chlamydia pneumoniae*. *Antimicrob Agents Chemother* 58:7595–7596. <https://doi.org/10.1128/AAC.03920-14>.
 36. Damião Gouveia AC, Unemo M, Jensen JS. 2018. In vitro activity of zoliflodacin (ETX0914) against macrolide-resistant, fluoroquinolone-resistant and antimicrobial-susceptible *Mycoplasma genitalium* strains. *J Antimicrob Chemother* 73:1291–1294. <https://doi.org/10.1093/jac/dky022>.