



Published in final edited form as:

Toxicol Pathol. 2019 December ; 47(8): 1004–1011. doi:10.1177/0192623319878398.

Lupus, Silica, and Dietary Omega-3 Fatty Acid Interventions

Kathryn A. Wierenga^{1,2}, Jack R. Harkema^{2,3}, James J. Pestka^{2,4,5}

¹Department of Biochemistry and Molecular Biology, Michigan State University, East Lansing, Michigan

²Institute for Integrative Toxicology, Michigan State University, East Lansing, Michigan

³Department of Pathobiology and Diagnostic Investigation, Michigan State University, East Lansing, Michigan

⁴Department of Food Science and Human Nutrition, Michigan State University, East Lansing, Michigan

⁵Department of Microbiology and Molecular Genetics, Michigan State University, East Lansing, Michigan

Abstract

Two environmental factors, crystalline silica (cSiO₂), a toxic airborne particle encountered occupationally, and docosahexaenoic acid (DHA), a dietary omega-3 highly unsaturated fatty acid (HUFA), have the potential to influence the development of systemic lupus erythematosus (lupus). Using the NZBWF1 mouse, which spontaneously develops lupus, we found that intranasal exposure to cSiO₂ significantly decreases latency and promotes rapid progression of the disease. Specifically, cSiO₂ induces the development of ectopic lymphoid structures (ELS) containing germinal centers in the lungs that yield vigorous and diverse autoantibody responses locally and systemically. Transcriptomic analysis revealed that cSiO₂ promotes a robust type I interferon gene signature that likely precipitates ELS neogenesis. Intriguingly, dietary supplementation with human-relevant doses of DHA impedes cSiO₂-induced gene expression, ELS neogenesis, autoantibody elevation, and glomerulonephritis in this lupus-prone mouse model. Together our findings point to the feasibility of enhancing tissue omega-3 HUFAs as a personalized nutritional intervention to impede onset and progression of environment-triggered autoimmune disease.

Keywords

Autoimmunity; systemic lupus erythematosus; ectopic lymphoid structure; tertiary lymphoid organ; omega-3 fatty acid; silica; lung

Corresponding Author: Kathryn Wierenga, Department of Biochemistry and Molecular Biology, Michigan State University, 469 Wilson Road, East Lansing, MI 48824, royerkat@msu.edu.

Conflicting Interests Statement

The authors declare no real, perceived or potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Lupus results from gene-environment interactions

Systemic lupus erythematosus (lupus) is a prototypical autoimmune disease characterized by inflammation and a loss of self-tolerance. There are an estimated 250,000 patients in the US with lupus,¹ and this is likely an underestimate, as lupus is frequently misdiagnosed.² This devastating disease disproportionately impacts women of child-bearing age and of non-Caucasian descent. Lupus begins with benign but detectable elevations in plasma autoantibodies and inflammatory biomarkers, but eventually develops into full-scale systemic autoimmune disease. Rampant autoantibodies form immune complexes with their cognate autoantigens and can deposit in the kidney, triggering glomerulonephritis and ultimately renal failure if left untreated. While the genome is the primary predisposing factor for autoimmunity, environmental factors such as toxicants³ and diet⁴ modulate underlying hereditary susceptibilities, playing major roles in the development and/or exacerbation of symptoms. To illustrate the interactive actions of such factors, we review here investigations in a widely used preclinical mouse model of lupus demonstrating that 1) early autoimmune disease onset can be triggered by the respiratory toxicant crystalline silica (cSiO₂) and 2) this triggering can be ameliorated by consumption of the dietary omega-3 fatty acid docosahexaenoic acid (DHA).

cSiO₂ is an autoimmune trigger

SiO₂ is the most abundant mineral on earth⁵ and an estimated 2.3 million Americans are exposed to cSiO₂ occupationally.⁶ Exposure to cSiO₂ is critical to the development of the human disease silicosis, which presents as chronic inflammation with severe fibrosis in the lungs and culminates in the loss of lung function.⁷ High levels of cSiO₂ and other silicates are present in the lungs of miners exposed to high levels of coal dust containing cSiO₂, indicating that the dust cannot be cleared from the lungs following chronic exposure.⁸ Clearance of inhaled cSiO₂ from the lung has also been shown to be extremely slow in rodents,⁹ thus repeated inhalation of cSiO₂ activates a persistent inflammatory response in the lungs.¹⁰ Continued over-activation of immune cells coupled with the release of cellular material following cSiO₂-induced cell death results in production of antibodies against host antigens, contributing to the development of systemic autoimmunity.^{11,12}

The etiological linkage between cSiO₂ exposure and the development of autoimmunity is supported by human epidemiological studies, as reviewed by Parks et al.¹³ The review's authors concluded that existing data met the Sir Bradford Hill criteria for establishing causality¹⁴; specifically, there was strength and consistency in the association between cSiO₂ exposure and lupus. This was observed across different study designs and populations, there was an obvious exposure-response gradient, and epidemiological findings are consistent with mechanistic studies in animals. More recently, a National Institute for Environmental Health Sciences expert panel evaluated the role of environmental factors in autoimmune disease and concluded that cSiO₂ contributes to multiple autoimmune diseases, including lupus, rheumatoid arthritis, systemic sclerosis, and ANCA vasculitis.³

Omega-3 HUFAs counter inflammation and autoimmunity

Consumption of omega-3 highly unsaturated fatty acids (HUFAs) has been shown to ameliorate chronic inflammatory and autoimmune diseases.^{15–17} The major omega-3 HUFAs eicosapentaenoic acid (EPA), docosapentaenoic acid (DPA), and docosahexaenoic acid (DHA) are abundant in oily fish. Since the Western diet is lacking in fish and, in general, marine fatty acids,^{18,19} the most effective way to increase omega-3 HUFAs is through dietary supplementation.^{20,21} Indeed, after multivitamins, fish oil is the second most widely used supplement in the US.²² Crude fish oil or semi-purified mixtures of EPA, DPA, and DHA have been utilized in clinical trials for cardiovascular disease, inflammatory diseases, and autoimmune diseases,²³ including lupus (reviewed elsewhere²⁴). While there have been many positive findings, there are also inconclusive results, leaving many open questions about the dose required to see beneficial effects as well as the mechanisms behind omega-3 HUFAs' protective actions.

Preclinical evidence that dietary DHA consumption prevents cSiO₂-triggered lupus

To investigate contributions of the exposome to onset and progression of lupus, we have employed the NZBWF1 mouse, a widely used animal model that mimics human lupus.^{25,26} As in humans, lupus development in NZBWF1 mice primarily occurs in females. These mice develop robust autoantibody responses and lupus nephritis around 32 weeks of age and typically will die within a year. Intriguingly, when these mice are intranasally instilled with cSiO₂, they develop lupus about 3 months earlier than vehicle-treated animals.¹² Our lab has focused on understanding the early inflammatory events that lead to cSiO₂-triggered onset of lupus in this model, as well as the potential to block cSiO₂-triggered inflammation using dietary intervention with the omega-3 HUFA DHA.^{12,27,28} In our most recently published study, 6-week old female NZBWF1 mice were introduced to diets supplemented with DHA-enriched algal oil, providing 0, 0.4 or 1% DHA per day, which are calorically equivalent to human doses of 0, 2 or 5 grams/day, respectively.²⁹ These mice were intranasally instilled with 1 mg cSiO₂ at 8, 9, 10, and 11 weeks of age. Cohorts were then sacrificed at 1, 5, 9, and 13 weeks after the final instillation and bronchoalveolar lavage fluid (BALF), lungs, kidneys, and blood were collected for analysis. Mice exposed to cSiO₂ developed glomerulonephritis at 13 weeks post-instillation, but this response was blocked by DHA feeding.

DHA's protective effects against cSiO₂-induced kidney inflammation were specifically linked to reduced pathogenesis in the lung.²⁹ Cell counts from the BALF of cSiO₂-treated NZBWF1 mice revealed a robust increase in immune cells at 9 and 13 weeks post-instillation, with the main cell types being macrophages and neutrophils; at 13 weeks, lymphocytes were also present in the BALF. cSiO₂-triggered elevations of these cells were significantly suppressed by feeding DHA, both at the low and high doses. Relatedly, following H&E staining, we observed perivascular and peribronchiolar lymphoid cell infiltration in cSiO₂-instilled mice, suggesting the development of ectopic lymphoid structures (ELS).^{12,30,31} This was confirmed by immunohistochemical staining for T cells, B

cells, and follicular dendritic cells. The infiltrating lymphoid cells clearly adopted a germinal center-like formation, with lymphoid cells surrounding aggregated follicular dendritic cells and yielding IgG- producing plasma cells.³² Organized lymphoid structures in the lungs of mice have also been termed inducible bronchus-associated lymphoid tissue, or iBALT, as they develop only in response to environmental exposures or infections.^{33,34} Intriguingly, DHA supplementation impeded formation of cSiO₂-induced ectopic lymphoid structures and germinal centers in the lung at all time points.

In lupus and other autoimmune diseases, ELS can serve as a platform for the development of autoantibodies.^{31,35} Using an autoantigen microarray, we identified numerous autoantibodies in the BALF and plasma that were upregulated in response to cSiO₂, beginning at 5 weeks post-instillation and becoming highly robust at 9 and 13 weeks.³⁶ These included IgG, IgM, and IgA isotypes. In mice that consumed diets supplemented with DHA, cSiO₂-induced autoantibody production was suppressed. Importantly, known human lupus markers, namely anti-DNA, anti-RNP, anti-SM, anti- histone, anti-dsDNA, anti-Ro/SSA, anti-La/SSB, and anti-complement autoantibodies,³⁷ were suppressed by feeding DHA.

Type 1 interferon response is induced by cSiO₂ but suppressed by DHA supplementation.

Recently, we analyzed gene expression in the lungs and the kidneys using a platform targeting 770 immune-related genes to assess the effects cSiO₂ exposure and DHA consumption on mRNA signatures over time in female NZBWF1 mice (publication in press).³⁸ These analyses were performed on samples from the experiment detailed above, as well as in animals sacrificed either one day following a single cSiO₂ exposure or one day after the fourth exposure. Just 1 day after the last of four weekly cSiO₂ doses, genes related to inflammation as well as innate and adaptive immunity were dramatically increased in lungs of animals fed the control diet but were significantly reduced in mice fed DHA diets. Importantly, mRNA signatures in lungs of cSiO₂-treated mice fed the control diet over 13 weeks reflected progressive amplification of type 1 interferon (IFN)- and chemokine-related gene pathways, in addition to other immune-related pathways. At all timepoints, these lung signatures were suppressed in mice fed DHA.

IFN signaling is strongly implicated in the development of autoimmune diseases, with type 1 IFN – and more specifically IFN α – being associated with the development of lupus.³⁹ Early studies using the lupus-prone NZBWF1 mouse model showed that induction of type I IFN using polyinosinic:polycytidylic acid accelerated the disease onset.⁴⁰ In 1979, it was reported that patients with autoimmune disease had increased levels of circulating IFN, and in lupus patients both type I and type II IFN were observed.⁴¹ In the late 1990s and early 2000s, there were multiple reports that patients administered IFN α to treat certain malignancies and hepatitis C developed lupus-like symptoms, further strengthening the connection between type I IFN and the development of lupus.^{42,43,44} In 2003, Baechler et al. showed that expression of IFN target genes was closely correlated with disease activity and severity.⁴⁵ Of high relevance to our model, it was recently reported that cSiO₂-induced

cell death and self-dsDNA release could induce a type I IFN response in mice via the cytosolic receptor STING (stimulator of interferon genes).^{46,47} These findings were extended to humans, showing that patients with silicosis had an increase in circulating dsDNA as well as activation of the pathway sensing DNA and inducing a type I IFN response in the lung tissue.⁴⁶

Proposed role for type 1 IFN in cSiO₂-induced autoimmunity

When cSiO₂ is inhaled it can induce cell death (e.g. apoptosis, pyroptosis, and necroptosis) in multiple cell types in the lung, including alveolar macrophages and neutrophils (Figure 1). If dead cells are not properly cleared, cellular debris accrues. Of note, impaired efferocytosis (i.e. phagocytosis of dead/dying cells) is implicated in lupus and other autoimmune diseases, resulting in insufficient clearance of cellular debris, including host nucleic acids.⁴⁸ In addition, cSiO₂ has been shown to induce formation of neutrophil extracellular traps via a mechanism known as NETosis, leading to the extrusion of nuclear contents that further contribute to extracellular DNA accumulation.⁴⁹ While it is well established that viral nucleic acids can trigger an IFN response, endogenous nucleic acids released by dying and NETosing cells can as well.^{50,51} Many cell types can recognize double stranded and single stranded nucleic acids through various receptors, including toll like receptors and cytosolic nucleic acid sensors such as STING,⁴⁷ ultimately leading to the release of type I interferons. In lupus, one of the key cell types implicated in this response is the plasmacytoid dendritic cell (pDC), which primarily releases IFN α .³⁹

Based on published studies and our findings, we propose that the type I IFN response plays a central role in cSiO₂-triggered autoimmunity. IFN α can act on multiple cell types and many of its actions promote a feed-forward loop to drive the type I IFN response. For example, IFN α can induce additional NETosis⁵² and can also act on macrophages, leading to the release of proinflammatory chemokines and cytokines that stimulate antigen presentation and T and B cell activation. Additionally, IFN α can act directly on myeloid dendritic cells to promote antigen presentation. In the case of cSiO₂-triggered autoimmunity, host antigens are likely to be presented due to the impaired clearance of cellular debris. IFN α can further promote B cell maturation into plasma cells. In lupus, these may be autoreactive plasma cells producing autoantibodies against host antigens, including host DNA. These DNA-containing immune complexes can be recognized by pDCs, leading to additional release of IFN α .⁵³

Of high relevance to human autoimmunity, IFN α elicits the release of B-cell activating factor (BAFF) from multiple cell types.^{54,55} The main role of BAFF is to promote B-cell maturation, increasing the release of autoantibodies and driving this feed-forward cycle. BAFF is the target of the drug Belimumab (BenlystaTM), which was approved by the FDA in 2011, making it the first drug in 52 years approved to treat lupus in adults.⁵⁶ In April 2019, Benlysta was also approved for treatment of pediatric lupus.⁵⁷ We have found that cSiO₂ induces elevated BAFF in the BALF and plasma of NZBWF1 mice and that elevation of this critical B-cell maturation factor is dose-dependently suppressed by DHA supplementation.²⁸

Can omega-3 HUFA supplementation be used as a personalized nutrition tool against lupus?

cSiO₂ treatment markedly increases expression of several cytokines and receptors that are pathogenic in the progression of lupus. Importantly, dietary DHA reversed this effect (Table 1).^{28,29} Relevant to the treatment of lupus, several of the cytokines identified in our preliminary and published studies are pharmacological targets for drugs already approved or currently in clinical trials for the treatment of lupus and other autoimmune diseases.^{53,56,58–62} Notably, IFN α is a target of ongoing lupus clinical trials using multiple strategies to impede different steps of the IFN pathway, as reviewed previously.⁵³ Many of the therapies in development are monoclonal antibodies, which are extremely expensive.⁶³ For example, a Belimumab dosing regimen costs approximately \$35,000 per year, placing a substantial financial burden on uninsured patients and their families.^{56,64} While omega-3 HUFA supplementation might not eliminate the need for these drugs, it has the potential to interfere with many of these targets simultaneously for less than few dollars a day, reducing the need for high doses of a drug over prolonged periods of time.

Recently, a population study conducted under the aegis of the Michigan Lupus Epidemiology & Surveillance (MILES) Program reported that higher dietary intake of omega-3 fatty acids, and lower dietary omega-6:omega-3 ratios, were favorably associated with patient-reported outcomes, particularly self-reported lupus activity and sleep quality.⁴ To harness the protective effects of DHA and other omega-3 HUFAs, it is important to understand how they act. When consumed, fatty acids are incorporated into the cell membrane,²¹ and the resulting levels of omega-3 and omega-6 HUFAs in the membrane can have a direct effect on inflammatory pathways in the cell. One way that membrane HUFAs can influence inflammatory responses is by impacting lipid raft formation. It has been reported that DHA and EPA can impede lipid raft formation and prevent the aggregation and activation of transmembrane receptors involved in inflammatory signaling pathways.⁶⁵ HUFAs can also be cleaved from the membrane, both extracellularly and intracellularly by phospholipases to generate free HUFAs.^{66,67} Free EPA and DHA have been shown to activate transmembrane receptors involved in blocking pro-inflammatory signaling pathways^{68,69} and are also reported to be ligands for PPAR γ , which can inhibit NF-kB dependent transcription.^{70,71} Additionally, both EPA and DHA can be metabolized to downstream specialized pro-resolving mediators, known as SPMs, which include resolvins, protectins, maresins, and anti-inflammatory epoxide metabolites.^{72,73} These have also been shown to inhibit inflammatory signaling pathways^{74,75} and promote both phagocytosis of bacteria and efferocytosis of dead cells.^{76,77} Finally, EPA and DHA can directly compete with omega-6 HUFAs for incorporation into the cell membrane and for metabolism to downstream eicosanoids. This is of biological importance, as downstream metabolites of the omega-6 HUFA arachidonic acid (i.e. prostaglandins, thromboxanes, and leukotrienes) are well known to promote an inflammatory response.⁷⁸ Accordingly, altering the HUFA balance of the membrane to favor omega-3 HUFAs can have a direct impact on promoting resolution over inflammation.

Because the levels of omega-6 and omega-3 HUFAs may have profound and counteracting effects on inflammation, resolution, and disease status, it is critical to gauge their content in the phospholipid membrane. Assessing the phospholipid content of one's red blood cells as a biomarker of HUFA status in tissues is now quite simple with several companies providing relatively inexpensive, at-home finger prick tests.⁷⁹ From just one drop of blood collected and stabilized on filter paper, dozens of major fatty acids, including omega-3 and -6 HUFAs, can be quantified. We favor using the omega-3 HUFA score over other alternative approaches to reflect the omega-3 content in the red blood cells (RBCs) of our mice. The HUFA score represents the major omega-3 HUFAs as a percent of total HUFAs,⁸⁰ while other widely used indices present EPA and DHA as a percent of total fatty acids. There are a few advantages to presenting the omega-3 content using the HUFA score. First, HUFAs have similar chemical properties and will be modified – or degraded – at similar rates. Therefore, the HUFA score should remain relatively constant despite methodological differences.⁸¹ Second, the major omega-6 and omega-3 HUFAs are the primary substrates for known proinflammatory and anti-inflammatory lipid mediators.^{21,81} Limiting analysis to the HUFA pool allows the clinician to better estimate potential competition between omega-6- and omega-3-derived metabolites. Finally, the HUFA score is predictable from dietary fat intake, which would be important for developing a personalized nutritional intervention.⁸²

In a preliminary investigation,⁸³ using data from a previously published study that assessed dose response of DHA on cSiO₂-triggered lupus in NZBWF1 mice,²⁸ we showed that an increased omega-3 HUFA score in RBCs was associated with decreased autoimmune biomarkers, such as anti-dsDNA IgG antibodies and BAFF. The HUFA score was also negatively correlated with B and T cell infiltration in the lungs and levels of pro-inflammatory cytokines and chemokines, such as TNF- α and MCP-1, respectively. Similar associations were found between these endpoints and HUFA scores in the lung, spleen, and kidney. The high omega-3 HUFA scores (~70%) associated with reduced autoimmune disease in mice are similar to those reported for humans living in communities predominately consuming fish and other marine fats.⁸⁴ This is substantially higher than HUFA scores observed in individuals consuming a Western diet (~30%),⁸² which consists of foods rich in omega-6 fatty acids, such as vegetable oils, poultry, and beef.⁸⁵ These findings suggest that the omega-3 content of the RBCs, which is decidedly influenced by diet, may be leveraged to ameliorate the inflammatory status of patients with lupus and other autoimmune diseases. These personalized interventions are in step with the goals of the NIH's Precision Medicine Initiative, which defines precision medicine as “an innovative approach to disease prevention and treatment that takes into account individual differences in people's genes, environments, and lifestyles”.⁸⁶

Conclusion

Taken together, intranasal exposure of lupus-prone mice to cSiO₂ induces a type I IFN gene signature that precedes ELS induction in the lung, with lymphoid cell infiltration occurring as early as 1 week post-instillation and worsening over the course of the disease. ELS neogenesis evokes vigorous elevations of diverse autoantibodies in the BALF and plasma that form immune complexes with their cognate antigens and deposit in the kidneys, ultimately resulting in glomerulonephritis. When animals are fed DHA, the IFN-related

genes are suppressed, ELS development is suppressed, autoantibodies are reduced, and animals present much less severe glomerulonephritis (Figure 2). Because DHA consumption results in an enrichment of omega-3 HUFAs in the phospholipids, measuring the HUFA score of patients could be ideal for developing strategies to effectively combat inflammatory and autoimmune diseases. While it is critical to validate these preclinical findings in humans, our assessment provides insight into the range of omega-3 HUFA scores that might be required in humans to achieve protective effects. Ultimately, a precision health strategy for an individual would require at least two tactics to potentiate the omega-3 HUFA score: 1) increasing omega-3 HUFA intake by dietary supplementation and 2) reducing consumption of foods that containing high amounts of omega-6 HUFAs. This strategy may protect and benefit patients with lupus or other autoimmune diseases, individuals genetically predisposed to these diseases, or individuals environmentally exposed to cSiO₂ and other autoimmune disease triggers.

Acknowledgments

We thank Dr. Melissa Bates for her exceptional animal studies, which provide the underpinning of this review and the current work in our lab. We also thank Dr. Abby Benninghoff and Dr. Lichchavi Rajasinghe for their research insights, data analysis and technical support.

Funding

Research was funded by NIH ES027353 (JJP, JRH), NIEHS Training Grant T32ES007255 (KAW), Ruth L. Kirschstein Individual Predoctoral NRSA F31ES030593 (KAW), Lupus Foundation of America (KAW, JJP), the Dr. Robert and Carol Deibel Family Endowment (JJP).

References

1. Dorner T, Furie R. Novel paradigms in systemic lupus erythematosus. *Lancet* 2019;393(10188): 2344–2358. doi: 10.1016/S0140-6736(19)30546-X [PubMed: 31180031]
2. Durcan L, O'Dwyer T, Petri M. Management strategies and future directions for systemic lupus erythematosus in adults. *Lancet* 2019;393(10188):2332–2343. doi: 10.1016/S0140-6736(19)30237-5 [PubMed: 31180030]
3. Parks CG, Miller FW, Pollard KM, et al. Expert panel workshop consensus statement on the role of the environment in the development of autoimmune disease. *Int J Mol Sci* 2014;15(8):14269–14297. doi: 10.3390/ijms150814269 [PubMed: 25196523]
4. Charoenwoodhipong P, Harlow SD, Marder W, et al. Dietary omega polyunsaturated fatty acid intake and patient-reported outcomes in systemic lupus erythematosus: The Michigan Lupus Epidemiology & Surveillance (MILES) Program. *Arthritis Care & Research*(ja). doi: 10.1002/acr.23925
5. Parks CG, Cooper GS, Nylander-French LA, et al. Occupational exposure to crystalline silica and risk of systemic lupus erythematosus: a population-based, case-control study in the southeastern United States. *Arthritis Rheum* 2002;46(7):1840–1850. doi: 10.1002/art.10368 [PubMed: 12124868]
6. Workers' exposure to respirable crystalline silica: Final rule overview. Occupational Safety and Health Administration; 2016 <https://www.osha.gov/Publications/OSHA3683.pdf>. Accessed June 25, 2019.
7. Leung CC, Yu IT, Chen W. Silicosis. *Lancet* 2012;379(9830):2008–2018. doi: 10.1016/S0140-6736(12)60235-9 [PubMed: 22534002]
8. Cohen RA, Petsonk EL, Rose C, et al. Lung pathology in U.S. coal workers with rapidly progressive pneumoconiosis implicates silica and silicates. *Am J Respir Crit Care Med* 2016;193(6):673–680. doi: 10.1164/rccm.201505-1014OC [PubMed: 26513613]

9. Kawasaki H A review of the fate of inhaled alpha-quartz in the lungs of rats. *Inhal Toxicol* 2019;1–10. doi: 10.1080/08958378.2019.1597218
10. Kawasaki H A mechanistic review of silica-induced inhalation toxicity. *Inhalation Toxicology* 2015;27:363–377. doi: 10.3109/08958378.2015.1066905 [PubMed: 26194035]
11. Brown JM, Pfau JC, Holian A. Immunoglobulin and lymphocyte responses following silica exposure in New Zealand mixed mice. *Inhal Toxicol* 2004;16(3):133–139. doi: 10.1080/08958370490270936 [PubMed: 15204774]
12. Bates MA, Brandenberger C, Langohr I, et al. Silica triggers inflammation and ectopic lymphoid neogenesis in the lungs in parallel with accelerated onset of systemic autoimmunity and glomerulonephritis in the lupus-prone NZBWF1 mouse. *PLoS ONE* 2015;10(5):e0125481. doi: 10.1371/journal.pone.0125481 [PubMed: 25978333]
13. Parks CG, Cooper GS. Occupational exposures and risk of systemic lupus erythematosus: a review of the evidence and exposure assessment methods in population- and clinic-based studies. *Lupus* 2006;15(11):728–736. doi: 10.1177/0961203306069346 [PubMed: 17153843]
14. Hill AB. The environment and disease: Association or causation? *Proc R Soc Med* 1965;58:295–300. <https://www.ncbi.nlm.nih.gov/pubmed/14283879>. Accessed May 29, 2019. [PubMed: 14283879]
15. Gioxari A, Kaliora AC, Marantidou F, Panagiotakos DP. Intake of omega-3 polyunsaturated fatty acids in patients with rheumatoid arthritis: A systematic review and meta-analysis. *Nutrition* 2018;45:114–124 e114. doi: 10.1016/j.nut.2017.06.023 [PubMed: 28965775]
16. Calder PC. Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. *Biochim Biophys Acta* 2015;1851(4):469–484. doi: 10.1016/j.bbali.2014.08.010 [PubMed: 25149823]
17. Pestka JJ. n-3 polyunsaturated fatty acids and autoimmune-mediated glomerulonephritis. *Prostaglandins Leukot Essent Fatty Acids* 2010;82(4–6):251–258. doi: 10.1016/j.plefa.2010.02.013 [PubMed: 20189790]
18. Papanikolaou Y, Brooks J, Reider C, Fulgoni VL 3rd. U.S. adults are not meeting recommended levels for fish and omega-3 fatty acid intake: results of an analysis using observational data from NHANES 2003–2008. *Nutr J* 2014;13:31. doi: 10.1186/1475-2891-13-31 [PubMed: 24694001]
19. Thuppal SV, von Schacky C, Harris WS, et al. Discrepancy between knowledge and perceptions of dietary omega-3 fatty acid intake compared with the omega-3 index. *Nutrients* 2017;9(9). doi: 10.3390/nu9090930
20. Micha R, Khatibzadeh S, Shi P, et al. Global, regional, and national consumption levels of dietary fats and oils in 1990 and 2010: a systematic analysis including 266 country-specific nutrition surveys. *BMJ* 2014;348:g2272. doi: 10.1136/bmj.g2272 [PubMed: 24736206]
21. Browning LM, Walker CG, Mander AP, et al. Incorporation of eicosapentaenoic and docosahexaenoic acids into lipid pools when given as supplements providing doses equivalent to typical intakes of oily fish. *Am J Clin Nutr* 2012;96(4):748–758. doi: 10.3945/ajcn.112.041343 [PubMed: 22932281]
22. Clarke TC, Black LI, Stussman BJ, Barnes PM, Nahin RL. Trends in the use of complementary health approaches among adults: United States, 2002–2012. *Natl Health Stat Report* 2015(79):1–16. doi:
23. Calder PC. Omega-3 fatty acids and inflammatory processes: from molecules to man. *Biochem Soc Trans* 2017;45(5):1105–1115. doi: 10.1042/BST20160474 [PubMed: 28900017]
24. Akbar U, Yang M, Kurian D, Mohan C. Omega-3 fatty acids in rheumatic diseases: A critical review. *J Clin Rheumatol* 2017;23(6):330–339. doi: 10.1097/RHU.0000000000000563 [PubMed: 28816722]
25. Dubois EL, Horowitz RE, Demopoulos HB, Teplitz R. NZB/NZW mice as a model of systemic lupus erythematosus. *JAMA* 1966;195(4):285–289. <https://www.ncbi.nlm.nih.gov/pubmed/4159181>. Accessed Jan 24, 2019. [PubMed: 4159181]
26. Crampton SP, Morawski PA, Bolland S. Linking susceptibility genes and pathogenesis mechanisms using mouse models of systemic lupus erythematosus. *Dis Model Mech* 2014;7(9):1033–1046. doi: 10.1242/dmm.016451 [PubMed: 25147296]

27. Pestka JJ, Vines LL, Bates MA, He K, Langohr I. Comparative effects of n-3, n-6 and n-9 unsaturated fatty acid-rich diet consumption on lupus nephritis, autoantibody production and CD4+ T cell-related gene responses in the autoimmune NZBWF1 mouse. *PLoS One* 2014;9(6):e100255. doi: 10.1371/journal.pone.0100255 [PubMed: 24945254]
28. Bates MA, Brandenberger C, Langohr II, et al. Silica-triggered autoimmunity in lupus-prone mice blocked by docosahexaenoic acid consumption. *PLoS One* 2016;11(8):e0160622. doi: 10.1371/journal.pone.0160622 [PubMed: 27513935]
29. Bates MA, Akbari P, Gilley KN, et al. Dietary docosahexaenoic acid prevents silica-induced development of pulmonary ectopic germinal centers and glomerulonephritis in the lupus-prone NZBWF1 mouse. *Front Immunol* 2018;9:2002. doi: 10.3389/fimmu.2018.02002 [PubMed: 30258439]
30. Bates MA P, Gilley KN., Jackson-Humbles DN, Wagner JG, Li N, Kopec AK, Wierenga KM, Brandenberger C, Holian A, Benninghoff AD, Harkema JR, and Pestka JJ Dietary docosahexaenoic acid prevents silica-induced development of pulmonary ectopic germinal centers and glomerulonephritis in the lupus-prone NZBWF1 mouse. *Frontiers Immunol* (in press) 2018 doi:
31. Corsiero E, Nerviani A, Bombardieri M, Pitzalis C. Ectopic lymphoid structures: Powerhouse of autoimmunity. *Front Immunol* 2016;7:430. doi: 10.3389/fimmu.2016.00430 [PubMed: 27799933]
32. Nacionales DC, Weinstein JS, Yan XJ, et al. B cell proliferation, somatic hypermutation, class switch recombination, and autoantibody production in ectopic lymphoid tissue in murine lupus. *J Immunol* 2009;182(7):4226–4236. doi: 10.4049/jimmunol.0800771 [PubMed: 19299721]
33. Moyron-Quiroz JE, Rangel-Moreno J, Kusser K, et al. Role of inducible bronchus associated lymphoid tissue (iBALT) in respiratory immunity. *Nat Med* 2004;10(9):927–934. doi: 10.1038/nm1091 [PubMed: 15311275]
34. Hwang JY, Randall TD, Silva-Sanchez A. Inducible Bronchus-Associated Lymphoid Tissue: Taming Inflammation in the Lung. *Front Immunol* 2016;7:258. doi: 10.3389/fimmu.2016.00258 [PubMed: 27446088]
35. Jones GW, Jones SA. Ectopic lymphoid follicles: inducible centres for generating antigen-specific immune responses within tissues. *Immunology* 2016;147(2):141–151. doi: 10.1111/imm.12554 [PubMed: 26551738]
36. Rajasinghe L, Li Q, Bates M, Akbari P, Harkema J, Pestka J. Docosahexaenoic acid (DHA) suppresses broad spectrum of pathogenic autoantibodies elicited in murine model of lupus flaring (OR12–03-19). *Current developments in nutrition* 2019;3(Suppl 1). doi: 10.1093/cdn/nzz049.OR12-03-19
37. Li QZ, Zhou J, Wandstrat AE, et al. Protein array autoantibody profiles for insights into systemic lupus erythematosus and incomplete lupus syndromes. *Clin Exp Immunol* 2007;147(1):60–70. doi: 10.1111/j.1365-2249.2006.03251.x [PubMed: 17177964]
38. Benninghoff AD, Bates MA, Wierenga KA, et al. Docosahexaenoic acid consumption impedes early interferon- and chemokine-related gene expression during silica-triggered flaring of murine lupus. *Front Immunol* In Press
39. Crow MK, Olfieriev M, Kirou KA. Type I interferons in autoimmune disease. *Annu Rev Pathol* 2019;14:369–393. doi: 10.1146/annurev-pathol-020117-043952 [PubMed: 30332560]
40. Steinberg AD, Baron S, Talal N. The pathogenesis of autoimmunity in New Zealand mice, I. Induction of antinucleic acid antibodies by polyinosinic-polycytidylic acid. *Proc Natl Acad Sci U S A* 1969;63(4):1102–1107. doi: 10.1073/pnas.63.4.1102 [PubMed: 5307809]
41. Hooks JJ, Moutsopoulos HM, Geis SA, Stahl NI, Decker JL, Notkins AL. Immune interferon in the circulation of patients with autoimmune disease. *N Engl J Med* 1979;301(1):5–8. doi: 10.1056/NEJM197907053010102 [PubMed: 449915]
42. Gota C, Calabrese L. Induction of clinical autoimmune disease by therapeutic interferon-alpha. *Autoimmunity* 2003;36(8):511–518. <https://www.ncbi.nlm.nih.gov/pubmed/14984028>. Accessed Apr 15, 2019. [PubMed: 14984028]
43. Rizvi R, Hojjati M. Interferon-alpha induced lupus in a patient with chronic hepatitis C virus. *J Clin Rheumatol* 2011;17(3):152–153. doi: 10.1097/RHU.0b013e31821557e7 [PubMed: 21441813]

44. Ronnblom LE, Alm GV, Oberg KE. Autoimmunity after alpha-interferon therapy for malignant carcinoid tumors. *Ann Intern Med* 1991;115(3):178–183. doi: 10.7326/0003-4819-115-3-178 [PubMed: 2058872]
45. Baechler EC, Batliwalla FM, Karypis G, et al. Interferon-inducible gene expression signature in peripheral blood cells of patients with severe lupus. *Proc Natl Acad Sci U S A* 2003;100(5):2610–2615. doi: 10.1073/pnas.0337679100 [PubMed: 12604793]
46. Benmerzoug S, Rose S, Bounab B, et al. STING-dependent sensing of self-DNA drives silica-induced lung inflammation. *Nat Commun* 2018;9(1):5226. doi: 10.1038/s41467-018-07425-1 [PubMed: 30523277]
47. Paludan SR, Bowie AG. Immune sensing of DNA. *Immunity* 2013;38(5):870–880. doi: 10.1016/j.immuni.2013.05.004 [PubMed: 23706668]
48. Abdolmaleki F, Farahani N, Gheibi Hayat SM, et al. The Role of Efferocytosis in Autoimmune Diseases. *Front Immunol* 2018;9:1645. doi: 10.3389/fimmu.2018.01645 [PubMed: 30083153]
49. Rizzi M, Camiato F, Tonello S, et al. Charged molecular silica trigger in vitro NETosis in human granulocytes via both oxidative and autophagic pathways. *Eur Rev Med Pharmacol Sci* 2018;22(20):7058–7068. doi: 10.26355/eurrev_201810_16178 [PubMed: 30402874]
50. McNab F, Mayer-Barber K, Sher A, Wack A, O’Garra A. Type I interferons in infectious disease. *Nat Rev Immunol* 2015;15(2):87–103. doi: 10.1038/nri3787 [PubMed: 25614319]
51. Crow MK, Ronnblom L. Type I interferons in host defence and inflammatory diseases. *Lupus Sci Med* 2019;6(1):e000336. doi: 10.1136/lupus-2019-000336 [PubMed: 31205729]
52. Garcia-Romo GS, Caielli S, Vega B, et al. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med* 2011;3(73):73ra20. doi: 10.1126/scitranslmed.3001201
53. Chasset F, Arnaud L. Targeting interferons and their pathways in systemic lupus erythematosus. *Autoimmun Rev* 2018;17(1):44–52. doi: 10.1016/j.autrev.2017.11.009 [PubMed: 29108825]
54. Jacob N, Guo S, Mathian A, et al. B Cell and BAFF dependence of IFN-alpha-exaggerated disease in systemic lupus erythematosus-prone NZM 2328 mice. *J Immunol* 2011;186(8):4984–4993. doi: 10.4049/jimmunol.1000466 [PubMed: 21383240]
55. Lopez P, Scheel-Toellner D, Rodriguez-Carrio J, Caminal-Montero L, Gordon C, Suarez A. Interferon-alpha-induced B-lymphocyte stimulator expression and mobilization in healthy and systemic lupus erythematosus monocytes. *Rheumatology (Oxford)* 2014;53(12):2249–2258. doi: 10.1093/rheumatology/keu249 [PubMed: 24942493]
56. Lamore R 3rd, Parmar S, Patel K, Hilas O Belimumab (benlysta): a breakthrough therapy for systemic lupus erythematosus. *P T* 2012;37(4):212–226. <https://www.ncbi.nlm.nih.gov/pubmed/22593633>. Accessed Apr 20, 2018. [PubMed: 22593633]
57. FDA approves first treatment for pediatric patients with lupus. *Arnold N*; 2019 <https://www.fda.gov/news-events/press-announcements/fda-approves-first-treatment-pediatric-patients-lupus>. Accessed Apr 26, 2019.
58. Sheppard M, Laskou F, Stapleton PP, Hadavi S, Dasgupta B. Tocilizumab (Actemra). *Hum Vaccin Immunother* 2017;13(9):1972–1988. doi: 10.1080/21645515.2017.1316909 [PubMed: 28841363]
59. Pelechas E, Voulgari PV, Drosos AA. Sirukumab: a promising therapy for rheumatoid arthritis. *Expert Opin Biol Ther* 2017;17(6):755–763. doi: 10.1080/14712598.2017.1315099 [PubMed: 28374627]
60. Blair HA, Deeks ED Infliximab Biosimilar (CT-P13; Infliximab-dyyb): A review in autoimmune inflammatory diseases. *BioDrugs* 2016;30(5):469–480. doi: 10.1007/s40259-016-0193-2 [PubMed: 27650650]
61. Menne J, Eulberg D, Beyer D, et al. C-C motif-ligand 2 inhibition with emapticap pegol (NOX-E36) in type 2 diabetic patients with albuminuria. *Nephrol Dial Transplant* 2017;32(2):307–315. doi: 10.1093/ndt/gfv459 [PubMed: 28186566]
62. Chatenoud L CD3-specific antibody-induced active tolerance: from bench to bedside. *Nat Rev Immunol* 2003;3(2):123–132. doi: 10.1038/nri1000 [PubMed: 12563296]
63. Shaughnessy AF. Monoclonal antibodies: magic bullets with a hefty price tag. *BMJ* 2012;345:e8346. doi: 10.1136/bmj.e8346 [PubMed: 23236036]

64. Panopalis P, Yazdany J, Gillis JZ, et al. Health care costs and costs associated with changes in work productivity among persons with systemic lupus erythematosus. *Arthritis Rheum* 2008;59(12): 1788–1795. doi: 10.1002/art.24063 [PubMed: 19035422]
65. Wong SW, Kwon MJ, Choi AM, Kim HP, Nakahira K, Hwang DH. Fatty acids modulate Toll-like receptor 4 activation through regulation of receptor dimerization and recruitment into lipid rafts in a reactive oxygen species-dependent manner. *J Biol Chem* 2009;284(40):27384–27392. doi: 10.1074/jbc.M109.044065 [PubMed: 19648648]
66. Norris PC, Dennis EA. Omega-3 fatty acids cause dramatic changes in TLR4 and purinergic eicosanoid signaling. *Proc Natl Acad Sci U S A* 2012;109(22):8517–8522. doi: 10.1073/pnas.1200189109 [PubMed: 22586114]
67. Norris PC, Gosselin D, Reichart D, Glass CK, Dennis EA. Phospholipase A2 regulates eicosanoid class switching during inflammasome activation. *Proc Natl Acad Sci U S A* 2014;111(35):12746–12751. doi: 10.1073/pnas.1404372111 [PubMed: 25139986]
68. Li X, Yu Y, Funk CD. Cyclooxygenase-2 induction in macrophages is modulated by docosahexaenoic acid via interactions with free fatty acid receptor 4 (FFA4). *FASEB J* 2013;27(12):4987–4997. doi: 10.1096/fj.13-235333 [PubMed: 24005906]
69. Yan Y, Jiang W, Spinetti T, et al. Omega-3 fatty acids prevent inflammation and metabolic disorder through inhibition of NLRP3 inflammasome activation. *Immunity* 2013;38(6):1154–1163. doi: 10.1016/j.immuni.2013.05.015 [PubMed: 23809162]
70. Ricote M, Glass CK. PPARs and molecular mechanisms of transrepression. *Biochim Biophys Acta* 2007;1771(8):926–935. doi: 10.1016/j.bbali.2007.02.013 [PubMed: 17433773]
71. Chang HY, Lee HN, Kim W, Surh YJ. Docosahexaenoic acid induces M2 macrophage polarization through peroxisome proliferator-activated receptor gamma activation. *Life Sci* 2015;120:39–47. doi: 10.1016/j.lfs.2014.10.014 [PubMed: 25445227]
72. Serhan CN. Systems approach to inflammation resolution: identification of novel anti-inflammatory and pro-resolving mediators. *J Thromb Haemost* 2009;7 Suppl 1:44–48. doi: 10.1111/j.1538-7836.2009.03396.x [PubMed: 19630766]
73. Ostermann AI, Schebb NH. Effects of omega-3 fatty acid supplementation on the pattern of oxylipins: a short review about the modulation of hydroxy-, dihydroxy-, and epoxy-fatty acids. *Food Funct* 2017;8(7):2355–2367. doi: 10.1039/c7fo00403f [PubMed: 28682409]
74. Sham HP, Walker KH, Abdunour RE, et al. 15-epi-Lipoxin A4, Resolvin D2, and Resolvin D3 Induce NF-kappaB Regulators in Bacterial Pneumonia. *J Immunol* 2018;200(8):2757–2766. doi: 10.4049/jimmunol.1602090 [PubMed: 29523657]
75. Titos E, Rius B, Lopez-Vicario C, et al. Signaling and immunoresolving actions of resolvin D1 in inflamed human visceral adipose tissue. *J Immunol* 2016;197(8):3360–3370. doi: 10.4049/jimmunol.1502522 [PubMed: 27647830]
76. Chiang N, Fredman G, Backhed F, et al. Infection regulates pro-resolving mediators that lower antibiotic requirements. *Nature* 2012;484(7395):524–528. doi: 10.1038/nature11042 [PubMed: 22538616]
77. Fredman G, Hellmann J, Proto JD, et al. An imbalance between specialized pro-resolving lipid mediators and pro-inflammatory leukotrienes promotes instability of atherosclerotic plaques. *Nat Commun* 2016;7:12859. doi: 10.1038/ncomms12859 [PubMed: 27659679]
78. Lands B Highly unsaturated fatty acids (HUFA) mediate and monitor food's impact on health. *Prostaglandins Other Lipid Mediat* 2017;133:4–10. doi: 10.1016/j.prostaglandins.2017.05.002 [PubMed: 28535956]
79. Johnston DT, Deuster PA, Harris WS, Macrae H, Dretsch MN. Red blood cell omega-3 fatty acid levels and neurocognitive performance in deployed U.S. Servicemembers. *Nutr Neurosci* 2013;16(1):30–38. doi: 10.1179/1476830512Y.0000000025 [PubMed: 22748167]
80. Stark KD. The percentage of n-3 highly unsaturated fatty acids in total HUFA as a biomarker for omega-3 fatty acid status in tissues. *Lipids* 2008;43(1):45–53. doi: 10.1007/s11745-007-3128-3 [PubMed: 17985169]
81. Lands B, Bibus D, Stark KD. Dynamic interactions of n-3 and n-6 fatty acid nutrients. *Prostaglandins Leukot Essent Fatty Acids* 2018;136:15–21. doi: 10.1016/j.plefa.2017.01.012 [PubMed: 28189338]

82. Strandjord SE, Lands B, Hibbeln JR. Validation of an equation predicting highly unsaturated fatty acid (HUFA) compositions of human blood fractions from dietary intakes of both HUFAs and their precursors. *Prostaglandins Leukot Essent Fatty Acids* 2018;136:171–176. doi: 10.1016/j.plefa.2017.03.005 [PubMed: 28390839]
83. Wierenga KA, Bates MA, Lock AL, et al. Percent n-3 in highly unsaturated fatty acids (HUFAs) is predictive of disease outcomes in environmental toxicant-triggered autoimmunity. In *The Toxicologist*. Vol 168. *Toxicol Sci* 168(1): Society of Toxicology, 2019; 2019.
84. Yamada T, Strong JP, Ishii T, et al. Atherosclerosis and omega-3 fatty acids in the populations of a fishing village and a farming village in Japan. *Atherosclerosis* 2000;153(2):469–481. <https://www.ncbi.nlm.nih.gov/pubmed/11164437>. Accessed June 12, 2019. [PubMed: 11164437]
85. USDA Food Composition Database. United States Department of Agriculture; 2018 <https://ndb.nal.usda.gov/ndb/nutrients/report/nutrientsfrm?max=25&offset=0&totCount=0&nutrient1=675&nutrient2=620&nutrient3=&subset=0&sort=c&measureby=g>. Accessed July 1, 2019.
86. FACT SHEET: President Obama’s Precision Medicine Initiative. The White House; 2015 <https://obamawhitehouse.archives.gov/the-press-office/2015/01/30/fact-sheet-president-obama-s-precision-medicine-initiative>. Accessed August 6, 2019.

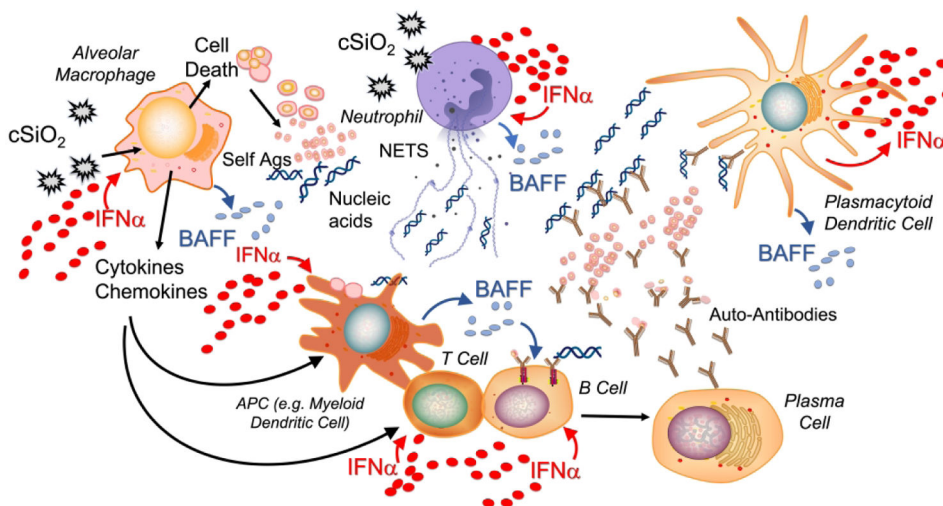


Figure 1: Putative role for type 1 IFN in cSiO₂-induced lupus.

When inhaled, cSiO₂ induces cell death in multiple cell types present in the lung (e.g. neutrophils and macrophages) leading to the extracellular release of host nucleic acids. cSiO₂ also stimulates release of neutrophil extracellular traps (NETs) which contribute to the accumulation of extracellular DNA. Extracellular nucleic acids, either viral or innate, induce the release of type I IFN from many immune cell types. In lupus, plasmacytoid dendritic cells (pDCs) are recognized as a primary producer of IFN α , the major type I IFN. Extracellular DNA along with other uncleared cellular debris can be presented by antigen presenting cells, promoting the development of autoreactive B and T cells. This process is enhanced directly by IFN α and by other cytokines and chemokines, such as B-cell activating factor (BAFF), released from immune cells in response to IFN α . Autoreactive plasma cells will produce autoantibodies against host antigens, forming DNA-containing immune complexes that can be recognized by pDCs to further drive this cycle. *Adapted from Chasset F & Arnaud L, 2018.*⁵⁰

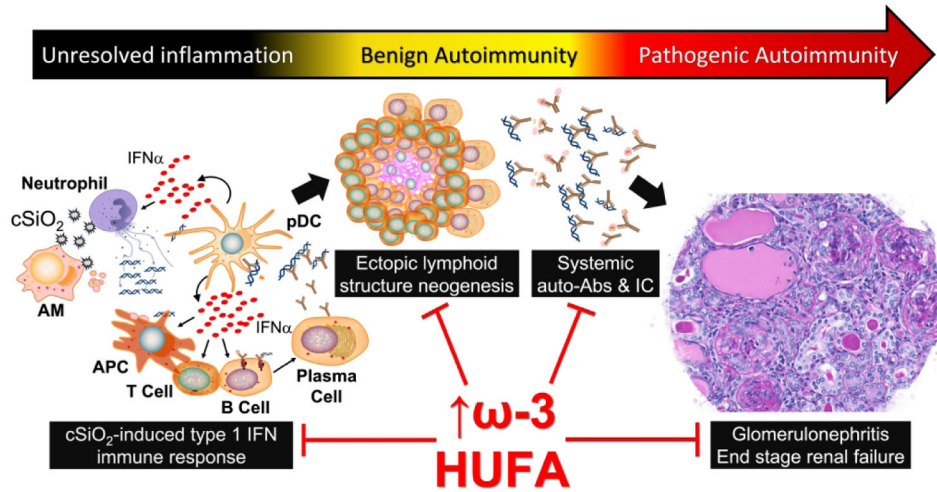


Figure 2: cSiO₂-induced autoimmune pathogenesis is correlated with the omega-3 HUFA score. The cSiO₂-induced type I IFN gene signature promotes ectopic lymphoid structure neogenesis in the lungs, and production of pathogenic autoantibodies which form immune complexes (IC) with host antigens. This culminates in systemic autoimmunity and glomerulonephritis. When mice are fed diets rich in DHA, autoimmune pathogenesis is suppressed. Improved disease status and decreased inflammatory endpoints are associated with an increase in omega-3 HUFA in the cell phospholipid membrane. Biomarkers that evaluate membrane fatty acids, such as the HUFA score, are crucial to developing omega-3 supplementation strategies to prevent or ameliorate inflammatory and autoimmune disease.

Table 1.

Silica enhances (↑) and DHA suppresses (↓) many inflammatory endpoints targeted by drugs either approved or in clinical trials for treatment of autoimmune disease.

Target	Silica	DHA	Treatment	Drug
IFN α	↑	↓	IFN α mAb	Rontalizumab, Sifalimumab ⁵⁰
			IFNAR1 mAb	Anifrolumab ⁵⁰
			IFN α vaccine	Interferon- α -kinoid (IFN-K) ⁵⁰
BAFF	↑	↓	BAFF mAb	Belimumab (Benlysta TM) ⁵³
IL-6	↑	↓	IL-6R mAb	Tocilizumab ⁵⁵
			IL-6 mAb	Sirukumab ⁵⁶
TNF- α	↑	↓	TNF- α mAb	Infliximab ⁵⁷
MCP-1	↑	↓	Neutralize MCP-1	Emapticap ⁵⁸
CD3	↑	↓	CD3 mAb	Muromonab-CD3 ⁵⁹

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript