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Cigarette smoking and oral microbiota in low-income and African-American populations

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Abstract

Background—Cigarette smoking is a common risk factor for diseases and cancers. Oral microbiota is also associated with diseases and cancers. However, little is known about the impact of cigarette smoking on the oral microbiota, especially among ethnic minority populations.

Methods—We investigated cigarette smoking in relationship with the oral microbiota in a large population of predominately low-income and African-American participants. Mouth rinse samples were collected from 1616 participants within the Southern Community Cohort Study, including 592 current-smokers, 477 former-smokers and 547 never-smokers. Oral microbiota was profiled by 16S ribosomal RNA gene deep sequencing.

Results—Current-smokers showed a different overall microbial composition from formersmokers ($p=6.62\times10^{-7}$) and never-smokers ($p=6.00\times10^{-8}$). The two probiotic genera, *Bifidobacterium* and *Lactobacillus*, were enriched among current-smokers when compared with never-smokers, with Bonferroni-corrected p values ($P_{Bonferroni}$) of 1.28×10^{-4} and 5.89×10^{-7} ,

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respectively. The phylum *Actinobacteria* was also enriched in current-smokers when compared with never-smokers, with a median relative abundance of 12.35% versus 9.36%, respectively, and with a $P_{\text{Bonferroni}}=9.11\times10^{-11}$. In contrast, the phylum *Proteobacteria* was depleted in current smokers ($P_{\text{Bonferroni}}=5.57\times10^{-13}$), with the relative abundance being almost three times that of never-smokers (7.22%) when compared with that of current-mokers (2.47%). Multiple taxa within these two phyla showed differences in abundance/prevalence between current-smokers and neversmokers at $P_{\text{Bonferroni}}<0.05$. The differences in the overall microbial composition and abundance/ prevalence of most taxa were observed among both African-Americans and European-Americans. Meanwhile, such differences were not observed between former-smokers and never-smokers.

Conclusion—Smoking has strong impacts on oral microbial community, which was recovered after smoking cessation.

INTRODUCTION

The human mouth nourishes over 2000 types of microbes, which collectively compose the oral microbiota.¹ Well-balanced oral microbiota maintains oral and systemic health,² while dysbiosis of oral microbiota may lead to diseases.^{3–5}

Cigarette smoking is a common risk factor for many diseases. Various toxicants in cigarette smoke directly contact with oral microbes; thus, long term exposure to smoking toxicants may affect the microbial ecology in oral cavity due to antibiotic effects and oxygen deprivation.⁶ Studies have shown the impact of cigarette smoking on the oral microbiota,^{7–11} though the results were inconsistent across studies. For example, in two previous studies investigating subgingival⁹ and oral wash samples,¹⁰ current-smokers showed a lower microbial diversity than non-smokers. However, in a subsequent study, such a difference was not observed in any of the eight oral sites investigated.¹¹ In addition, most of these studies focused on European-ancestry populations. Studies among ethnic minority populations, for example, African-Americans, are lacking.

In the study presented here, we investigated the impact of cigarette smoking on the oral microbiota using data from 1616 participants (1058 African-Americans and 558 European-Americans) within the Southern Community Cohort Study (SCCS), including 592 current-smokers, 477 former-smokers and 547 never-smokers.

METHODS

Study population

Launched in 2002, the SCCS was designed to investigate health disparities among lowincome populations, with the majority of the study participants being African-American. Detailed descriptions of the SCCS can be accessed elsewhere.¹² Briefly, this study took 7 years to recruit over 85 000 middle-aged adults (40–70 years old) from 12 southeastern US states. In total, ~34 100 participants donated mouth rinse samples during the enrolment. The SCCS was reviewed and approved by review boards at the Vanderbilt University Medical Center and the Meharry Medical College. Written informed consent was provided by all involved individuals.

During enrolment, all participants were requested to complete a comprehensive questionnaire that was designed to collect individuals' personal information, including smoking status. After recruitment, follow-ups were performed through record linkage and surveys via mail or telephone. Major health outcomes were ascertained via linkage with state cancer registries and/or from National Death Index mortality records. Participants included in the present study were selected from four nested case-control studies for incident cases (ascertained during the first follow-up) of upper aerodigestive tract cancer, type 2 diabetes, lung cancer and colorectal cancer (n=1864). All participants were free of diseases when mouth rinse samples were collected. After excluding individuals who did not report a smoking history or disclosed a history of antibiotics usage during the year before their mouth rinse sample donation, the current study included 1616 subjects.

DNA extraction and 16S ribosomal RNA (rRNA) gene sequencing

The Qiagen's QIAmp DNA kit (Qiagen, Germantown, Maryland, USA) was used to extract DNA from mouth rinse samples. The NEXTflex 16S rRNA gene V4 Amplicon-Seq Kit (Bioo Scientific, Austin, Texas, USA) was used to construct the sequencing libraries of the 16S rRNA gene V4 domain. Sequencing was conducted using the Illumina MiSeq 300 (paired-end 150 bp) at the Vanderbilt Technologies for Advanced Genomics Core or using the Illumina Hiseq (paired-end 250 bp) at BGI Americas (Cambridge, Massachusetts, USA). For both sequencing batches, on each 96-well plate, two additional duplicated quality control samples were sequenced. All duplicate samples showed similar microbial profiles: the coefficient of variability for the Faith's Phylogenic Diversity (PD) index (a measurement of microbial community diversity) among the duplicate samples was 0.3%.

Sequencing data processing and quality controls

Raw data from two sequencing batches were processed together by QIIME,¹³ using the closed-reference operational taxonomic unit calling strategy. Taxonomy assignment was conducted using the Human Oral Microbiome Database¹⁴ (HOMD) as reference. In total, 100 153 658 reads (mean \pm SD = 102 302 \pm 77 432; range = (5323–854 744)) were obtained for the 956 samples from the first batch and 30 506 499 reads (mean \pm SD = 47 741 \pm 11 628; range = (20 428–91 660)) were retained for the 660 samples from the second batch.

Statistical analysis

The alpha diversity of each sample was measured by the Faith's PD index. The associations of alpha diversity with potential confounders, including age, sex, race, body mass index (BMI), alcohol consumption, total energy intake, oral health status, disease status at the first follow-up and sequencing batch were estimated through a linear regression analysis. The differences of beta diversity among the three smoking groups were assessed by using MiRKAT¹⁵ V.0.02, based on the weighted UniFrac distance matrix. We also evaluated the differences of beta diversity between current-smokers and non-smokers (including former-smokers and never-smokers).

Cigarette smoking has been associated with weight loss,¹⁶ and recently, multiple animal studies and human clinical trials have reported associations between weight loss and several probiotic bacteria, mainly belonging to the genera *Bifidobacterium* and *Lactobacillus*.^{17–20}

Hence, we compared the prevalence of these two genera, along with the species belonging to them, between current-smokers and never-smokers, between former-smokers and never-smokers, and between current-smokers and non-smokers, through logistic regression analyses.

For other taxa, we focused on four taxonomic levels: phylum, family, genus and species. Similar with the analyses for probiotic taxa, differences of these taxa between current-smokers and never-smokers, between former-smokers and never smokers and between current-smokers and non-smokers were investigated. Based on the relative abundance among never-smokers, taxa were categorised as 'common taxa' (with a median abundance of 0.1%) or 'rare taxa' (with a median abundance of <0.1%) or 'rare taxa' (with a median abundance of <0.1%). For common taxa (five phyla, 15 families, 16 genera and 28 species), relative abundance was normalised by arcsine-square-root transformation, and a linear regression analysis was performed for each taxon to estimate the association of smoking status with the arcsine-square-root transformed taxon relative abundance. For rare taxa, in addition to those probiotic taxa, analyses were limited to those with a prevalence among never-smokers of >30%, including three phyla, 16 families, 35 genera and 98 species. After grouping participants into carriers and non-carriers, a logistic regression analysis was conducted for each taxon to investigate smoking status in association with taxon prevalence.

Among all of the analyses described above, adjustments were made in regression models for potential cofounders, including age, sex, race, BMI, alcohol consumption, total energy intake, oral health status, disease status at the first follow-up and sequencing batch. For each of these covariates, missing data were indicated with a dummy variable and included in regression analyses. Given the intrinsic correlations among taxa from different taxonomic levels, not all association tests were independent. Following Galwey's method,²¹ we estimated the number of independent tests for common taxa and rare taxa (including probiotic taxa) separately using the function 'meff' of the R package 'poolR' (https://rdrr.io/github/ozancinar/poolR/). Among the 64 common taxa and the 152 rare taxa included in the statistical analyses, the independent tests were estimated to be 25 and 69, respectively. For the associations with a Bonferroni-corrected p<0.05, that is, p<2.00×10⁻³ for common taxa and p<7.25×10⁻⁴ for rare taxa, we further performed stratified analyses by race, as well as by sequencing batch, to evaluate the heterogeneity between African-Americans and European-Americans, and between the first and the second sequencing batch. All analyses were carried out using R V.3.3.1.

RESULTS

Characteristics of the study participants

The general profile of study participants' characteristics is shown in table 1. In total, 1616 individuals were included in this study, including 36.6% current-smokers, 29.5% former-smokers and 33.9% never-smokers. Among the African-Americans, 39.1% were current-smokers, 24.9% were former-smokers and 36.0% were never-smokers. Among the European-Americans, a higher percentage of participants were former-smokers (38.4%), with 31.9% being current-smokers and 29.7% never-smokers. Current-smokers tended to have the lowest BMI and never-smokers had the highest BMI. Overall, the study participants

had a very low socioeconomic status. Specifically, ~64% of the current-smokers had an annual household income of less than US\$15 000. Only 65.8% of the study participants had oral health status data, and the majority of them had poor oral health. Specifically, current-smokers had the worst oral health, with ~90% having tooth loss, while ~80% of the non-smokers had tooth loss. We found associations of alpha diversity (Faith's PD index) with race, age, alcohol drinking, tooth loss and sequencing batch at p<0.05.

Current-smokers showed a different overall composition when compared with neversmokers and former-smokers

Differences in beta-diversity (weighted UniFrac matrices) were observed between currentsmokers and never-smokers ($p=6.00\times10^{-8}$), between current-smokers and former-smokers ($p=6.62\times10^{-7}$) and between current-smokers and non-smokers ($p<2.20\times10^{-16}$). Consistently, differences between current-smokers and never-smokers, between currentsmokers and former-smokers and between current-smokers and non-smokers, were observed among African-Americans (p values of 9.72×10^{-4} , 6.93×10^{-3} and 3.55×10^{-4} , respectively), European-Americans (p values of 3.51×10^{-4} , 6.85×10^{-5} and 5.15×10^{-7} , respectively), the first sequencing batch (p values of 9.72×10^{-4} , 9.09×10^{-5} and 1.83×10^{-5} , respectively). However, between former-smokers and never-smokers, no difference was observed either for either combined analyses, or for stratified analyses by race or sequencing batch.

Probiotic bacterial taxa were enriched among current-smokers

At the genus level, both *Bifidobacterium* and *Lactobacillus* were more prevalent among current-smokers (85.6% and 89.4%) than among never-smokers (67.3% and 73.5%), with Bonferroni-corrected p values ($P_{Bonferroni}$) of 1.59×10^{-4} and 1.81×10^{-4} , respectively (table 2). In addition, one species of *Bifidobacterium* and six species of *Lactobacillus* were also enriched in current-smokers when compared with former-smokers and never-smokers. For example, *Bifidobacterium longum* was observed among 67.6% of current-smokers but only 39.7% of never-smokers ($P_{Bonferronf}=1.80 \times 10^{-9}$). The prevalence for *Lactobacillus crispatus* was almost two-fold in current-smokers (61.2%) when compared with that in never-smokers (34.4%), with a $P_{Bonferronf}=1.80 \times 10^{-8}$. Further, all these nine taxa (two genera and seven species) were also significantly more prevalent among current-smokers than among non-smokers. When comparing the former-smokers and never-smokers, none of these probiotic taxa showed a difference.

Actinobacteria were enriched and Proteobacteria were depleted among current-smokers

As shown in table 3, the phylum *Actinobacteria* was enriched in current-smokers, with a median relative abundance of 12.4% in current-smokers and 9.4% in never-smokers $(P_{Bonferronf}=3.24\times10^{-11})$. Within *Actinobacteria*, nine common taxa showed a higher abundance in current-smokers than in never-smokers at $P_{Bonferronf}<0.05$, including two families, three genera and four species (table 3 and online supplementary figure S1). Among them, *Rothia mucilaginosa* showed the strongest enrichment with a $P_{Bonferronf}=1.25\times10^{-8}$. In addition to these common taxa, two rare taxa within *Actinobacteria*, *Bifidobacteriaceae* and *Actinomyces lingnae_(NVP*), showed a higher prevalence in current-smokers than in

never-smokers at $P_{Bonferroni} < 0.05$ (table 4 and online supplementary figure S2). All of these *Actinobacteria* taxa showed a significant differential abundance/prevalence between current-smokers and non-smokers ($P_{Bonferroni} < 0.05$). However, when comparing the former-smokers and never-smokers, none showed a difference (tables 3 and 4).

On the other hand, the phylum *Proteobacteria* was depleted in current-smokers, with the median relative abundance decreased to less than one-third, that is, 2.5% in current-smokers and 7.2% in never-smokers ($P_{Bonferronf}=7.58\times10^{-20}$). In this phylum, nine common taxa showed a lower abundance in current-smokers at $P_{Bonferronf}<0.05$ (table 3 and online supplementary figure S1). Among them, *Neisseriaceae* was the most representative taxon, with the median relative abundance decreased from 1.06% in never-smokers to only 0.06% in current-smokers, which corresponded to a ~18 fold change ($P_{Bonferronf}=1.13\times10^{-23}$). Similarly, within this phylum, seven rare taxa also showed a lower prevalence in current-smokers at $P_{Bonferronf}<0.05$ (table 4 and online supplementary figure S2). For example, the prevalence of *Neisseria oralis* decreased approximately three-fold, which was present in 53.2% of never-smokers but only 19.9% in current-smokers ($P_{Bonferronf}=1.20\times10^{-20}$). Similar to *Actinobacteria*, all of these *Proteobacteria* taxa were also less abundant or prevalent in current-smokers when compared with non-smokers (*P_Bonferronf*<0.05), while no such differences were observed between former-smokers and never-smokers (tables 3 and 4).

Taxa in the phyla *Bacteroidetes*, *Firmicutes* and *Spirochaetes* were also associated with smoking status

Multiple taxa within *Bacteroidetes* and *Firmicutes* also showed a different abundance between current-smokers and never-smokers, as well as between current-smokers and nonsmokers (table 3 and online supplementary figure S1). For example, in *Bacteroidetes*, *Prevotella sp. oral taxon 313* was more abundant while *Flavobacteriaceae* was less abundant among current-smokers. In *Firmicutes, Megasphaera, Megasphaera micronuciformis* and *Streptococcus sp. oral taxon 057* were more abundant among current-smokers, while *Gemella, Streptococcus oligofermentans* and *Streptococcus sp. oral taxon 070* were more abundant among non-smokers. Within these two phyla, a differential prevalence of seven rare taxa was found between current-smokers and non-smokers (table 4 and online supplementary figure S2). In addition, a rare species of the phylum *Spirochaetes, Treponema denticola*, was more prevalent among current-smokers.

Consistent associations of smoking status and oral microbiota between ethnic groups and between sequencing batches

A substantial proportion of the significant associations identified in analyses of all participants (tables 2–4) were consistently observed when stratified by ethnic group or by sequencing batch (online supplementary tables S1–S6). Generally, the associations were much stronger among African-Americans and the first sequencing batch. For example, the probiotic species *Lactobacillus oris* showed a higher prevalence among current-smokers than among never-smokers, with p values of 7.08×10^{-5} in African-Americans and 1.13×10^{-3} in European-Americans (online supplementary tables S1), and p values of 2.19×10^{-6} for the first batch and 6.55×10^{-3} for the second batch (online supplementary table S4). Another example is the common taxa *S. oligofermentans*, which showed higher relative abundance in

current-smokers than in never-smokers with p values of 9.33×10^{-13} in African-Americans and 6.23×10^{-8} in European-Americans (online supplementary table S2), and 1.90×10^{-14} in the first sequencing batch and 1.21×10^{-6} in the second batch (online supplementary table S5). However, we also found that some bacterial taxa showed stronger associations among European-Americans than among African-Americans, for example, *Lactobacillus fermentum* (online supplementary table S1), *R. mucilaginosa* (online supplementary table S2) and *Prevotella nanceiensis* (online supplementary table S3). In addition, several taxa showed stronger associations among the second batch than among the first batch, for example, *B. longum* (online supplementary table S4), *Neisseria pharynges* (online supplementary table S5) and *Kingella denitrificans* (online supplementary table S6). However, a formal test of multiplicative interaction failed to show statistical significance.

DISCUSSION

In this study, we found that among both European-Americans and African-Americans, cigarette smoking impacts overall oral microbial composition, as well as the abundance/ prevalence of multiple microbial taxa, especially for those belonging to the probiotic genera *Bifidobacterium* and *Lactobacillus*, and those within the phyla *Actinobacteria* and *Proteobacteria*. However, these changes may be recovered after smoking cessation.

In addition to smoking status, race, age, alcohol drinking, tooth loss and sequencing batch were also associated with oral microbial richness. The associations of race, age, alcohol drinking and tooth loss with the oral microbiota are consistent with previous studies.^{22–26} We also found significant differences in overall microbial composition between current-smokers and never-smokers, and between current-smokers and non-smokers, but not between current-smokers and former-smokers, which are consistent with results from previous studies.⁹¹⁰

Cigarette smoking has been associated with weight loss or lower BMI, and smoking cessation has been associated with weight gain or higher BMI.²⁷²⁸ In the present study, current-smokers also showed a lower BMI when compared with non-smokers. Interestingly, we found that two probiotic genera, *Bifidobacterium* and *Lactobacillus*, together with seven species belonging to them, were more prevalent among current-smokers when compared with never-smokers and former-smokers. Similar results were reported by two previous studies.⁹¹⁰ For example, in one study, *Bifidobacterium*, *Lactobacillus* and *B. longum* showed a higher abundance among current-smokers.¹⁰ In the other, *L. fermentum*, *L. gasseri* and *L. reuteri* were enriched in current-smokers.⁹

The phylum *Actinobacteria*, along with 11 taxa belonging to it, was enriched in currentsmokers. Among them, *Actinobacteria*, *Atopobium* and *R. mucilaginosa* were consistently reported to be enriched in oral wash samples of current-smokers.¹⁰ *Actinomyces odontolyticus* was observed to be enriched in subgingival samples of current-smokers.⁹ The phylum *Proteobacteria* and 16 taxa within it were depleted among current-smokers. Many of them, including *Proteobacteria*, *Neisseriaceae*, *Burkholderiaceae*, *Neisseria*, *Aggregatibacter*, *Lautropia*, *Kingella*, *Cardiobacterium* and *Neisseria subflava* were consistently reported to be depleted in current-smokers.¹⁰ The depletion of *Neisseria* in

current-smokers was also reported in other three studies.⁸²⁹³⁰ Several in vivo studies suggest that cigarette smoking can inhibit growth of *Neisseria* species.³¹³²

Of the bacterial taxa that were enriched in current-smokers in the present study, several had been associated with risks of various diseases. For example, the common taxa *Actinomyces graevenitzii* was suggested to be involved in pulmonary abscesses in two independent case reports.³³³⁴ Other examples include the probiotic taxa such as *Bifidobacterium* and *Lactobacillus, B. longum, L. fermentum* and *L. reuteri*, which were reported be associated with a decreased risk of obesity prevalence.²⁰ There might be a potential link across smoking, oral probiotic taxa and obesity. *Actinobacteria* and *Actinomyces* were reported to be associated with a decreased risk of type 2 diabetes in our previous study of oral microbiome and type 2 diabetes.³⁵ *T. denticola*, a well-recognised oral pathogen, was found to be associated with a series of periodontal diseases³⁶ and an increased risk of colorectal cancer.⁵

Of the bacterial taxa that were enriched in non-smokers, several had been associated with decreased risks of cancers. For example, the common taxa *Neisseriaceae* and *Neisseria* were previously associated with a decreased risk of esophageal adenocarcinoma.³⁷ Several rare taxa, including *P. nanceiensis, Lachnoanaerobaculum umeaense* and *Lachnospiraceae_[G-2]*, were found to be associated with a decreased risk of esophageal adenocarcinoma or squamous cell carcinoma.³⁷ Another rare taxa, *Kingella*, was associated with a decreased risk of head and neck squamous cell cancer.⁴

Strengths of the present study include a large sample size, which provides higher statistical power compared with previous studies. In addition, most of previous studies focused on European-ancestry populations, while in the present study, the majority of participants were African-American and most of them have low socioeconomic status. Our results not only replicated a considerable proportion of previous findings but went further to compare the associations between African-Americans and European-Americans. Most associations identified in the overall analyses were consistently observed in both ethnic groups and both sequencing batches. Although generally the associations were slightly stronger among African-Americans and the first batch, these differences are not unexpected given the larger sample sizes of these two subsets. The main limitation is that 16S rRNA gene sequencing was used to assess the oral microbiome, which is limited in assessing the species level microbial profile and microbial pathways. Further research using the shotgun metagenomic sequencing technique is needed.

In summary, we demonstrated that among both African-Americans and European-Americans, cigarette smoking has strong impacts on the oral microbial community, which could probably be recovered by smoking cessation.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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REFERENCES

- 1. Warinner C, Rodrigues JFM, Vyas R, et al. Pathogens and host immunity in the ancient human oral cavity. Nat Genet 2014;46.336.44.
- Schwiertz A. Microbiota of the human body: implications in health and disease. New York, NY: Springer Berlin Heidelberg, 2016.
- 3. Fan X, Alekseyenko AV, Wu J, et al. Human oral microbiome and prospective risk for pancreatic cancer: a population-based nested case-control study. Gut 2018;67:120–7. [PubMed: 27742762]
- Hayes RB, Ahn J, Fan X, et al. Association of oral microbiome with risk for incident head and neck squamous cell cancer. JAMA Oncol 2018;4:358–65. [PubMed: 29327043]
- Yang Y, Cai Q, Shu X-O, et al. Prospective study of oral microbiome and colorectal cancer risk in low-income and African American populations. Int J Cancer 2019;144:2381–9. [PubMed: 30365870]
- Macgregor I. Effects of smoking on oral ecology. A review of the literature. Clin Prev Dent 1988;11:3–7.
- 7. Kumar PS, Matthews CR, Joshi V, et al. Tobacco smoking affects bacterial acquisition and colonization in oral biofilms. Infect Immun 2011;79:4730–8. [PubMed: 21859855]
- Morris A, Beck JM, Schloss PD, et al. Comparison of the respiratory microbiome in healthy nonsmokers and smokers. Am J Respir Crit Care Med 2013;187:1067–75. [PubMed: 23491408]
- Mason MR, Preshaw PM, Nagaraja HN, et al. The subgingival microbiome of clinically healthy current and never smokers. Isme J 2015;9:268–72. [PubMed: 25012901]
- Wu J, Peters BA, Dominianni C, et al. Cigarette smoking and the oral microbiome in a large study of American adults. Isme J 2016;10:2435–46. [PubMed: 27015003]
- Yu G, Phillips S, Gail MH, et al. The effect of cigarette smoking on the oral and nasal microbiota. Microbiome 2017;5:3. [PubMed: 28095925]
- Signorello LB, Hargreaves MK, Blot WJ. The southern community cohort study: investigating health disparities. J Health Care Poor Underserved 2010;21(1 Suppl):26–37.
- Caporaso JG, Kuczynski J, Stombaugh J, et al. QIIME allows analysis of high-throughput community sequencing data. Nat Methods 2010;7:335–6. [PubMed: 20383131]
- Chen T, Yu W-H, Izard J, et al. The human oral microbiome database: a web accessible resource for investigating oral microbe taxonomic and genomic information. Database 2010;2010:baq013. [PubMed: 20624719]
- Zhao N, Chen J, Carroll IM, et al. Testing in microbiome-profiling studies with MiRKAT, the microbiome regression-based kernel association test. Am J Hum Genet 2015;96:797–807. [PubMed: 25957468]
- Chiolero A, Faeh D, Paccaud F, et al. Consequences of smoking for body weight, body fat distribution, and insulin resistance. Am J Clin Nutr 2008;87:801–9. [PubMed: 18400700]
- Sáez-Lara M, Robles-Sanchez C, Ruiz-Ojeda F, et al. Effects of probiotics and synbiotics on obesity, insulin resistance syndrome, type 2 diabetes and non-alcoholic fatty liver disease: a review of human clinical trials. Int J Mol Sci 2016;17:928.

- Zarrati M, Salehi E, Mofid V, et al. Relationship between probiotic consumption and IL-10 and IL-17 secreted by PBMCs in overweight and obese people. Iran J Allergy Asthma Immunol 2013;12:404–6. [PubMed: 23996719]
- 19. Kobyliak N, Conte C, Cammarota G, et al. Probiotics in prevention and treatment of obesity: a critical view. Nutr Metab 2016;13:14.
- 20. Yang Y, Cai Q, Zheng W, et al. Oral microbiome and obesity in a large study of low-income and African-American populations. J Oral Microbiol 2019;11:1650597. [PubMed: 31489128]
- Galwey NW. A new measure of the effective number of tests, a practical toolfor comparing families of non-independent significance tests. Genet Epidemiol 2009;33:559–68. [PubMed: 19217024]
- 22. Nasidze I, Li J, Quinque D, et al. Global diversity in the human salivary microbiome. Genome Res 2009;19:636–43. [PubMed: 19251737]
- Gupta VK, Paul S, Dutta C. Geography, ethnicity or Subsistence-Specific variations in human microbiome composition and diversity. Front Microbiol 2017;8:1162. [PubMed: 28690602]
- Lira-Junior R, Åkerman S, Klinge B, et al. Salivary microbial profiles in relation to age, periodontal, and systemic diseases. PLoS One 2018;13:e0189374. [PubMed: 29538390]
- 25. Fan X, Peters BA, Jacobs EJ, et al. Drinking alcohol is associated with variation in the human oral microbiome in a large study of American adults. Microbiome 2018;6:59. [PubMed: 29685174]
- 26. Gholizadeh P, Eslami H, Yousefi M, et al. Role of oral microbiome on oral cancers, a review. Biomed Pharmacother 2016;84:552–8. [PubMed: 27693964]
- Klesges RC, Meyers AW, Klesges LM, et al. Smoking, body weight, and their effects on smoking behavior: a comprehensive review of the literature. Psychol Bull 1989;106:204–30. [PubMed: 2678202]
- Williamson DF, Madans J, Anda RF, et al. Smoking cessation and severity of weight gain in a national cohort. N Engl J Med 1991;324:739–45. [PubMed: 1997840]
- 29. Charlson ES, Chen J, Custers-Allen R, et al. Disordered microbial communities in the upper respiratory tract of cigarette smokers. PLoS One 2010;5:e15216. [PubMed: 21188149]
- 30. Colman G, Beighton D, Chalk AJ, et al. Cigarette smoking and the microbial flora of the mouth. Aust Dent J 1976;21:111–8. [PubMed: 1068665]
- Bardell D. Viability of six species of normal oropharyngeal bacteria after exposure to cigarette smoke in vitro. Microbios 1980;32:7–13.
- 32. Ertel A, Eng R, Smith SM. The differential effect of cigarette smoke on the growth of bacteria found in humans. Chest 1991;100:628–30. [PubMed: 1889244]
- Nagaoka K, Izumikawa K, Yamamoto Y, et al. Multiple lung abscesses caused by Actinomyces graevenitzii mimicking acute pulmonary coccidioidomycosis. J Clin Microbiol 2012;50:3125–8. [PubMed: 22760049]
- 34. Gliga S, Devaux M, Gosset Woimant M, et al. Actinomyces graevenitzii pulmonary abscess mimicking tuberculosis in a healthy young man. Can Respir J 2014;21:e75–7. [PubMed: 25493591]
- Long J, Cai Q, Steinwandel M, et al. Association of oral microbiome with type 2 diabetes risk. J Periodontal Res 2017;52:636–43. [PubMed: 28177125]
- 36. Sela MN. Role of Treponema denticola in periodontal diseases. Crit Rev Oral Biol Med 2001;12:399–413. [PubMed: 12002822]
- Peters BA, Wu J, Pei Z, et al. Oral microbiome composition reflects prospective risk for esophageal cancers. Cancer Res 2017;77:6777–87. [PubMed: 29196415]

What is already known on this subject

- Cigarette smoking has an important impact on the human oral microbiota. However, previous studies were limited by small sample sizes and lack of replication.
- Most of the previous studies only focused on European-ancestry populations, hence the information regarding ethnic minority populations is lacking.

What this study adds

• This investigation gives us information regarding smoking and oral microbiota in low-income and African-American populations.

• We demonstrate that, among both European-Americans and African-Americans, cigarette smoking has considerable impacts on the oral microbial community structure and abundance/prevalence of multiple bacterial taxa, which could probably be recovered by smoking cessation. The associations of cigarette smoking with bacterial taxa has little heterogeneity between African-Americans and European-Americans.

Table 1

Characteristics of participants in two combined studies from the Southern Community Cohort Study

| Age (years) ⁴ 53.8±7.90 59.18±840 55.78±88 Sex (%) Fenale 220 (37.16) 20 (44.03) 55.78±88 Sex (%) Male 372 (62.84) 267 (55.97) 196 (35.83) Race (%) African-American 414 (69.93) 267 (55.97) 196 (35.83) Race (%) African-American 1178 (30.07) 267 (55.97) 196 (35.83) Race (%) African-American 1178 (30.07) 267 (55.97) 381 (69.65) Bace (%) African-American 1178 (30.07) 267 (55.97) 381 (69.65) Race (%) African-American 117 (69.31) 30.32.46.95 31.17±7.43 Annual household income (USS) (%) 26.86.6.41 30.32.46.95 31.17±7.43 Annual household income (USS) (%) 216 (30.73) 78 (16.67) 92 (17.23) Annual household income (USS) (%) 500 and <50 000 64 (10.87) 26 (40.45) Annual household income (USS) (%) 15 000 and <50 000 216 (41.45) 216 (40.45) Acohol consumption ⁴ (%) 100 000 26 (4.24.9) 90 (12.6.9) | Characteristic | Group | Current-smokers (n=592) | Former-smokers (n=477) | Never-smokers (n=547) |
|---|--------------------------------------|---------------------|-------------------------|------------------------|-----------------------|
| Age (years) Internation Internation <thinternation< th=""> <thinternation< th=""></thinternation<></thinternation<> | * | - | 53 18±7 00 | 50 1848 40 | 55 70+0 00 |
| Sex (6) Emale 220 (37.16) 210 (44.03) 331 (64.17) Rate (%) Male 372 (62.34) 267 (55.97) 196 (55.33) Rate (%) African-American 141 (69.93) 267 (55.97) 196 (55.33) Rate (%) African-American 141 (69.93) 263 (55.14) 381 (69.65) Body mass index (BM)* European-American 178 (30.07) 214 (44.86) 166 (30.33) Body mass index (BM)* 25 (50.01) 26 (56.67) 214 (44.86) 216 (40.45) Annual household income (USS) (%) 178 (30.07) 214 (44.86) 31 (56.7) 216 (40.45) Annual household income (USS) (%) 25 (62.67) 20 (32.67) 216 (40.45) 216 (40.45) Annual household income (USS) (%) 26 (50.07) 214 (44.86) 166 (10.43) 216 (40.45) Annual household income (USS) (%) 15 00 and <20 000 | Age (years)" | | 06.1±01.00 | 29.1ŏ±ŏ.42 | ØØ.رØ1.CC |
| Fenale $200(37,16)$ $210(4403)$ $31(6417)$ Race (%) African-American $372(62.84)$ $267(55.97)$ $106(35.83)$ Race (%) African-American $1178(30.07)$ $267(55.97)$ $381(66.65)$ Buop mass index (BM)* Buropean-American $1178(30.07)$ $261(44.86)$ $381(66.65)$ Buop mass index (BM)* Second $26.86:64.41$ 30.32 ± 6.95 31.17 ± 7.43 Annual household income (US\$) (%) (17600) $216(40.45)$ $102(41.45)$ $216(40.45)$ Annual household income (US\$) (%) (1500) $375(6.56)$ 31.72 ± 7.43 $1177(43)$ Annual household income (US\$) (%) 15000 and <20000 $375(6.57)$ $261(4.96)$ $20(40.45)$ Annual household income (US\$) (%) 15000 and <20000 $375(6.57)$ $210(4.45)$ $210(4.45)$ Annual household income (US\$) (%) 15000 and <20000 $261(4.07)$ $210(4.92)$ $210(4.92)$ Annual household income (US\$) (%) 15000 and <20000 21020.37 $281(6.67)$ $210(4.92)$ Annual household income (US\$) (%) | Sex (%) | | | | |
| Mate 372 (62.84) 267 (55.97) 196 (35.83) Race (%) African-American 312 (62.04) 381 (69.65) Body mass index (BM)* African-American 117 (30.07) 265 (55.14) 381 (69.65) Body mass index (BM)* European-American 178 (30.07) 214 (44.86) 166 (30.35) Body mass index (BM)* 25 (56.64) 30.32±6.95 31.17±7.43 Annal household income (USS) (%) 375 (53.67) 194 (41.45) 216 (40.45) Annal household income (USS) (%) 375 (53.67) 78 (16.67) 216 (40.45) Annal household income (USS) (%) 25 (00.37) 78 (16.67) 216 (40.45) Annal household income (USS) (%) 78 (16.67) 78 (16.67) 216 (40.45) Annal household income (USS) (%) 78 (16.71) 78 (16.73) 216 (40.45) Annal household income (USS) (%) 78 (16.71) 78 (16.73) 216 (20.43) Actohol consumption* 26 (00.70) 26 (14.42) 26 (16.43) Alcohol | | Female | 220 (37.16) | 210 (44.03) | 351 (64.17) |
| Race (%) African-American 414 (69 93) 263 (55.14) 81 (6965) Buropean-American 178 (3007) 214 (44.86) 166 (3035) Budy mass index (BMI)* 26.86:6.41 30.32:6.95 31.172:43 Annual household income (USS) (%) 25.86:6.41 30.32:6.05 216 (40.45) Annual household income (USS) (%) 25.000 120 (20.37) 78 (16.67) 216 (40.45) Annual household income (USS) (%) 25.000 120 (20.37) 78 (16.67) 216 (40.45) Annual household income (USS) (%) 25.000 120 (20.37) 78 (16.67) 216 (40.45) Annual household income (USS) (%) 25.000 25 (4.24) 80 (17.09) 216 (40.45) Acobo and <100 000 | | Male | 372 (62.84) | 267 (55.97) | 196 (35.83) |
| | Race (%) | | | | |
| European-American 178 (30.07) 214 (44.86) 166 (30.35) 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 117 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 111 ± 74.35 112 ± 74.35 112 ± 74.35 112 ± 74.35 112 ± 74.35 112 ± 74.35 124 ± 60.34,35 116 ± 74.35 124 ± 60.34,35 | | African-American | 414 (69.93) | 263 (55.14) | 381 (69.65) |
| Body must index (BM)* 26.86 ± 6.41 30.32 ± 6.95 31.17 ± 7.43 Annual household income (USS) (%) < 15000 $375(6.367)$ $194(41.45)$ $216(40.45)$ Annual household income (USS) (%) 15000 and <25000 $375(6.367)$ $194(41.45)$ $216(40.45)$ 15000 and <25000 $64(10.87)$ $78(16.67)$ $92(17.23)$ 25000 and <2000 $64(10.87)$ $90(19.23)$ $92(17.23)$ 15000 and <10000 $25(4.24)$ $80(17.09)$ $89(16.67)$ 10000 $25(4.24)$ $80(17.09)$ $89(16.67)$ Alcohol consumption $f(\%)$ $800(17.09)$ $80(16.67)$ $102(91)$ Alcohol consumption $f(\%)$ $80(17.09)$ $80(16.67)$ $102(2.90)$ Alcohol consumption $f(\%)$ | | European-American | 178 (30.07) | 214 (44.86) | 166 (30.35) |
| Annual household income (US\$) (%) $<15\ 000$ $375\ (63.67)$ $194\ (41.45)$ $216\ (40.45)$ $<15\ 000$ $<15\ 000$ $375\ (63.67)$ $194\ (41.45)$ $216\ (40.45)$ $15\ 000$ $12\ 000$ $64\ (10.87)$ $78\ (16.67)$ $92\ (17.23)$ $25\ 000$ $add <25\ 000$ $64\ (10.87)$ $90\ (19.23)$ $102\ (19.1)$ $25\ 000$ $add <20\ 000$ $64\ (10.87)$ $80\ (17.09)$ $89\ (16.67)$ $26\ 000$ $64\ (10.87)$ $80\ (17.09)$ $80\ (16.67)$ $92\ (17.23)$ Alcohol consumption $\mathring{7}\ (\%)$ $70\ 000$ $5\ (0.85)$ $26\ (5.56)$ $32\ (6.55)$ Alcohol consumption $\mathring{7}\ (\%)$ None $177\ (30.31)$ $24\ (5.20)$ $32\ (6.55)$ Alcohol consumption $\mathring{7}\ (\%)$ None $177\ (30.31)$ $24\ (5.20)$ $34\ (6.33)$ Moderate $87\ (14.90)$ $59\ (12.80)$ $34\ (6.33)$ $166\ (7)\ (20.56)$ $24\ (60.34)$ Ioth loss (\%) $133\ (2.277)$ $28\ (6.7)$ $23\ (6.75)$ $23\ (6.32)$ Ioth loss (\%) $133\ (2.277)$ $28\ (6.7)$ $23\ (4.2.8)$ Ioth loss (\%) | Body mass index $(BMI)^*$ | | 26.86 ± 6.41 | $30.32{\pm}6.95$ | 31.17 ± 7.43 |
| $ \begin{array}{llllllllllllllllllllllllllllllllllll$ | Annual household income (US\$) (%) | | | | |
| | | <15 000 | 375 (63.67) | 194 (41.45) | 216 (40.45) |
| | | 15 000 and <25 000 | 120 (20.37) | 78 (16.67) | 92 (17.23) |
| | | 25 000 and <50 000 | 64 (10.87) | 90 (19.23) | 102 (19.1) |
| | | 50 000 and <100 000 | 25 (4.24) | 80 (17.09) | 89 (16.67) |
| | | 100 000 | 5 (0.85) | 26 (5.56) | 35 (6.55) |
| | Alcohol consumption $\dot{\tau}(\%)$ | | | | |
| | | None | 177 (30.31) | 243 (52.71) | 324 (60.34) |
| $ \begin{array}{llllllllllllllllllllllllllllllllllll$ | | Light | 187 (32.02) | 131 (28.42) | 156 (29.05) |
| | | Moderate | 87 (14.90) | 59 (12.80) | 34 (6.33) |
| Tooth loss (%) None $32 (10.53) = 66 (19.58) = 91 (21.51)$ One to 10 $128 (42.11) = 152 (45.10) = 218 (51.54)$ >10, not all $79 (25.99) = 68 (20.18) = 67 (15.84)$ All $65 (21.38) = 51 (15.13) = 47 (11.11)$ | | Heavy | 133 (22.77) | 28 (6.07) | 23 (4.28) |
| None $32 (10.53)$ $66 (19.58)$ $91 (21.51)$ One to 10 $128 (42.11)$ $152 (45.10)$ $218 (51.54)$ >10, not all $79 (25.99)$ $68 (20.18)$ $67 (15.84)$ All $65 (21.38)$ $51 (15.13)$ $47 (11.11)$ | Tooth loss (%) | | | | |
| | | None | 32 (10.53) | 66 (19.58) | 91 (21.51) |
| >10, not all 79 (25.99) 68 (20.18) 67 (15.84) All 65 (21.38) 51 (15.13) 47 (11.11) | | One to 10 | 128 (42.11) | 152 (45.10) | 218 (51.54) |
| All 65 (21.38) 51 (15.13) 47 (11.11) | | >10, not all | 79 (25.99) | 68 (20.18) | 67 (15.84) |
| | | All | 65 (21.38) | 51 (15.13) | 47 (11.11) |

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 $\overset{f}{\not A}$ lcohol drink, Light, <1 drink/day; Moderate, 1–2 drink/day; Heavy, >2 drink/day.

BMI, body mass index.

Table 2

Higher prevalence of probiotic bacterial taxa among current-smokers than among never-smokers and former-smokers

| | Prevalence | | | ${f P}$ value $^{*}({f P}_{Bonferroni}\dot{f})$ | | |
|---|-----------------------------|-------------------------------|--------------------------|---|--|--|
| Probiotic taxa | Current- smokers (n=592) | Former- smokers (n=477) | Never-smokers (n=547) | Current-smokers versus never- smokers | Former-smokers versus never-smokers | Current-smokers versus non-smokers ‡ |
| Phylum <i>Actinobacteria</i> | | | | | | |
| Genus Bifidobacterium | 85.64% | 71.07% | 67.28% | $2.30 \times 10^{-6} (1.59 \times 10^{-4})$ | 0.88 (1.00) | $2.09 \times 10^{-7} (1.44 \times 10^{-5})$ |
| Species Bifidobacterium longum | 67.57% | 45.07% | 39.67% | $2.61 \times 10^{-11} (1.80 \times 10^{-9})$ | 0.32 (1.00) | $2.91 \times 10^{-12} (2.01 \times 10^{-10})$ |
| Phylum <i>Firmicutes</i> | | | | | | |
| Genus Lactobacillus | 89.36% | 73.38% | 73.49% | $2.62 \times 10^{-6} (1.81 \times 10^{-4})$ | 0.46 (1.00) | $1.15 \times 10^{-7} \ (7.91 \times 10^{-6})$ |
| Species Lactobacillus crispatus | 61.15% | 35.43% | 34.37% | $2.60{	imes}10^{-10}$ ($1.80{	imes}10^{-8}$) | 0.72 (1.00) | 3.85×10^{-13} (2.66×10 ⁻¹¹) |
| Species L. fermentum | 57.60% | 39.83% | 35.65% | $2.09 \times 10^{-6} (1.44 \times 10^{-4})$ | 0.56 (1.00) | 4.59×10^{-7} (3.16×10^{-5}) |
| Species L. gasseri | 72.47% | 56.39% | 53.02% | $6.51{	imes}10^{-6}$ (4.50×10 ⁻⁴) | 0.95 (1.00) | 7.51×10^{-7} (5.18×10^{-5}) |
| Species L. oris | 43.75% | 26.83% | 20.84% | $1.09 \times 10^{-8} (7.54 \times 10^{-7})$ | 0.04 (1.00) | 3.46×10^{-8} (2.39×10 ⁻⁶) |
| Species L. panis | 42.23% | 26.83% | 25.41% | 3.74×10^{-7} (2.58×10 ⁻⁵) | 0.81 (1.00) | $2.33 \times 10^{-7} (1.61 \times 10^{-5})$ |
| Species L. reuteri | 41.55% | 23.27% | 20.84% | $1.65 \times 10^{-8} (1.14 \times 10^{-6})$ | 0.99 (1.00) | 5.36×10^{-10} (3.70×10^{-8}) |
| * P values were calculated by logistic r | regression. Sequencing | g batch as well as oth | ler covariates (age, se | x, race, BMI, alcohol consumption, o | ral health and disease status a | t the first follow-up and total energy |

intake) were adjusted for.

 \dot{f} Bonferroni correction, adjusted for 69 independent tests.

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 $\overset{\sharp}{\star}^{}_{\text{Non-smokers includes former-smokers and never-smokers.}$

BMI, body mass index.

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Table 3

Individual taxa showing a differential relative abundance between current-smokers and never-smokers

| | Median relativ | e abundance | | ${f P}$ value $^{*}(P_{Bonferroni}^{\dagger}\hat{T})$ | | |
|---------------------------------------|--------------------------------|-------------------------------|------------------------------|---|--|--|
| Taxa | Current- smokers (n=592) | Former- smokers (n=477) | Never- smokers (n=547) | Current-smokers versus never- smokers | Former-smokers versus never- smokers | Current-smokers versus non- smokerst [‡] |
| Phylum Actinobacteria | 12.35% | 10.25% | 9.36% | 1.29×10 ⁻¹² (3.24×10 ⁻¹¹) | 0.44 (1.00) | $1.10 \times 10^{-17} (2.75 \times 10^{-16})$ |
| Family Actinomycetaceae | 3.08% | 2.51% | 2.48% | $1.65 \times 10^{-4}(4.11 \times 10^{-3})$ | 0.29~(1.00) | 3.62×10^{-7} (9.06×10 ⁻⁶) |
| Genus Actinomyces | 3.05% | 2.40% | 2.42% | $1.28 \times 10^{-4} (3.19 \times 10^{-3})$ | 0.25 (1.00) | $2.06 \times 10^{-7} (5.14 \times 10^{-6})$ |
| Species Actinomyces graevenitzii | 0.36% | 0.15% | 0.16% | 9.36×10^{-10} (2.34×10 ⁻⁸) | 0.53(1.00) | $6.71 \times 10^{-15} (1.68 \times 10^{-13})$ |
| Species Actinomyces graevenitzii | 1.44% | 0.95% | 0.94% | $2.26 \times 10^{-6} (5.64 \times 10^{-5})$ | 0.91 (1.00) | 2.73×10^{-9} (6.82×10 ⁻⁸) |
| Genus Rothia | 6.64% | 5.50% | 4.81% | $2.76 \times 10^{-9} (6.91 \times 10^{-8})$ | 0.27 (1.00) | $1.77 \times 10^{-12} (4.43 \times 10^{-11})$ |
| Species Rothia mucilaginosa | 5.64% | 4.65% | 3.87% | $5.02 \times 10^{-10} (1.25 \times 10^{-8})$ | 0.11 (1.00) | $2.66 \times 10^{-13} (6.64 \times 10^{-12})$ |
| Family Coriobacteriaceae | 0.13% | 0.09% | 0.12% | $1.24 \times 10^{-5} (3.11 \times 10^{-4})$ | 0.18 (1.00) | $2.39{	imes}10^{-8}$ (5.97 ${	imes}10^{-7}$) |
| Genus Atopobium | 0.13% | 0.09% | 0.12% | $1.98 \times 10^{-5} (4.95 \times 10^{-4})$ | 0.17 (1.00) | $3.90 \times 10^{-8} (9.75 \times 10^{-7})$ |
| Species Atopobium parvulum | 0.11% | 0.08% | 0.10% | $2.21 \times 10^{-5} (5.52 \times 10^{-4})$ | 0.15 (1.00) | $2.22 \times 10^{-8} (5.54 \times 10^{-7})$ |
| Phylum <i>Proteobacteria</i> | 2.47% | 6.22% | 7.22% | 3.03×10^{-21} (7.58×10 ⁻²⁰) | 0.76~(1.00) | $1.91 \times 10^{-21} (4.77 \times 10^{-20})$ |
| Family Neisseriaceae | 0.06% | 1.01% | 1.06% | $4.52 \times 10^{-25} (1.13 \times 10^{-23})$ | 0.88 (1.00) | 1.91×10^{-24} (4.78×10 ⁻²³) |
| Genus Neisseria | 0.05% | 0.87% | 1.01% | $2.74 \times 10^{-24} (6.86 \times 10^{-23})$ | 0.92 (1.00) | $6.28 \times 10^{-24} (1.57 \times 10^{-22})$ |
| Species Neisseria pharyngis | 0.01% | 0.09% | 0.11% | $4.81 \times 10^{-15} (1.20 \times 10^{-13})$ | 0.96~(1.00) | 2.77×10^{-13} (6.94×10 ⁻¹²) |
| Species N. subflava | 0.03% | 0.43% | 0.62% | 3.95×10^{-22} (9.87×10 ⁻²¹) | 0.75 (1.00) | 2.50×10^{-21} (6.24×10 ⁻²⁰) |
| Family Pasteurellaceae | 1.71% | 3.85% | 4.30% | 1.40×10^{-14} (3.49×10 ⁻¹³) | 0.80~(1.00) | 3.76×10^{-15} (9.39×10 ⁻¹⁴) |
| Genus Aggregatibacter | 0.06% | 0.10% | 0.16% | $3.71 \times 10^{-5} (8.43 \times 10^{-4})$ | 0.46(1.00) | $2.71 \times 10^{-4} (6.77 \times 10^{-3})$ |
| Genus <i>Haemophilus</i> | 1.39% | 3.44% | 3.89% | $1.63 \times 10^{-14} \ (4.08 \times 10^{-13})$ | 0.90(1.00) | $2.18 \times 10^{-15} (5.46 \times 10^{-14})$ |
| Species Haemophilus parahaemolyticus | 0.16% | 0.51% | 0.48% | 2.55×10^{-15} (6.37×10 ⁻¹⁴) | 0.36(1.00) | $4.66 \times 10^{-16} (1.16 \times 10^{-14})$ |
| Species H. paraphrohaemolyticus | 1.21% | 2.81% | 3.03% | 9.59×10^{-13} (2.40×10 ⁻¹¹) | 0.66 (1.00) | 3.82×10^{-13} (9.55×10 ⁻¹²) |
| Phylum Bacteroidetes | | | | | | |
| Species Prevotella sp. oral taxon 313 | 4.45% | 2.90% | 2.97% | $5.57 \times 10^{-5} (1.39 \times 10^{-3})$ | 0.37~(1.00) | $3.57 \times 10^{-6} (8.93 \times 10^{-5})$ |
| Family Flavobacteriaceae | 0.04% | 0.10% | 0.10% | $6.67 \times 10^{-5} (1.67 \times 10^{-3})$ | 0.15(1.00) | $5.51 \times 10^{-5} (1.38 \times 10^{-3})$ |
| Phylum <i>Firmicutes</i> | | | | | | |
| Genus Gemella | 1.30% | 1.90% | 2.34% | $4.30 \times 10^{-15} (1.07 \times 10^{-13})$ | 0.02~(0.49) | $1.16 \times 10^{-11} (2.90 \times 10^{-10})$ |

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| | Median relativ | ve abundance | | ${f P}$ value $^{*}(P_{Bonferroni}^{\dagger}^{\dagger})$ | | |
|--|--------------------------------|-------------------------------|------------------------------|--|--|--|
| Taxa | Current- smokers (n=592) | Former- smokers (n=477) | Never- smokers (n=547) | Current-smokers versus never- smokers | Former-smokers versus never- smokers | Current-smokers versus non- smokerst [‡] |
| Species Streptococcus oligofermentans | 0.13% | 0.43% | 0.42% | $1.60 \times 10^{-20} \ (4.00 \times 10^{-19})$ | 0.62 (1.00) | $6.42 \times 10^{-23} (1.60 \times 10^{-21})$ |
| Species Streptococcus sp. oral taxon 057 | 10.36% | 9.01% | 8.07% | 1.59×10^{-10} (3.99×10^{-9}) | 0.07 (1.00) | $3.46 \times 10^{-9} (8.64 \times 10^{-8})$ |
| Species Streptococcus sp. oral taxon 070 | 23.23% | 22.69% | 23.67% | 6.93×10 ⁻⁵ (1.73×10 ⁻³) | 0.43 (1.00) | $1.28 \times 10^{-4} (3.19 \times 10^{-3})$ |
| Genus <i>Megasphaera</i> | 0.26% | 0.10% | 0.12% | $2.05 \times 10^{-8} (5.13 \times 10^{-7})$ | 0.43 (1.00) | $7.29 \times 10^{-12} (1.82 \times 10^{-10})$ |
| Species Megasphaera micronuciformis | 0.24% | 0.10% | 0.11% | $1.95 \times 10^{-7} (4.88 \times 10^{-6})$ | 0.40(1.00) | $1.52 \times 10^{-10} (3.81 \times 10^{-9})$ |
| * P values were calculated by logistic regressi | ion. Sequencing ba | atch as well as othe | r covariates (age, s | ex, race, BMI, alcohol consumption, oral h | nealth and disease status at | the first follow-up and total energy |

intake) were adjusted for.

 $\dot{f}^{\rm L}$ Bonferroni correction, adjusted for 25 independent tests.

 $\overset{4}{\star}$ Non-smokers includes former-smokers and never-smokers.

BMI, body mass index.

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Table 4

Individual taxa showing a differential prevalence between current-smokers and never-smokers

| | Prevalence | | | ${f P}$ value $^{*}(P_{Bonferroni}^{\dagger}\dot{	au})$ | | |
|--|--------------------------------|-------------------------------|------------------------------|---|--|---|
| Taxa | Current- smokers (n=592) | Former- smokers (n=477) | Never- smokers (n=547) | Current-smokers versus never-smokers | Former-smokers versus never-smokers | Current-smokers versus non- smokers [‡] |
| Phylum Actinobacteria | | | | | | |
| Family Biffdobacteriaceae | 95.44% | 89.94% | 84.64% | $7.50 \times 10^{-5} (5.17 \times 10^{-3})$ | 0.15 (1.00) | $1.81 \times 10^{-4} (0.01)$ |
| Species Actinomyces lingnae_(NVP) | 92.40% | 88.05% | 86.65% | $4.52 \times 10^{-6} (3.12 \times 10^{-4})$ | 0.44 (1.00) | $1.00 \times 10^{-6} (6.91 \times 10^{-5})$ |
| Phylum <i>Proteobacteria</i> | | | | | | |
| Family Burkholderiaceae | 42.74% | 63.31% | 69.84% | $1.50 \times 10^{-17} (1.04 \times 10^{-15})$ | 0.05 (1.00) | $5.83 \times 10^{-18} (4.02 \times 10^{-16})$ |
| Genus <i>Lautropia</i> | 41.55% | 62.68% | 69.47% | 4.82×10^{-19} (3.33×10^{-17}) | 0.04(1.00) | $1.29 \times 10^{-19} (8.88 \times 10^{-18})$ |
| Genus Kingella | 67.23% | 79.87% | 85.19% | $3.01 \times 10^{-8} (2.08 \times 10^{-6})$ | 0.08 (0.21) | $1.20 \times 10^{-8} (8.29 \times 10^{-7})$ |
| Species Kingella denitrificans | 24.83% | 45.28% | 46.62% | 6.30×10^{-10} (4.35×10 ⁻⁸) | 0.65 (1.00) | $2.30 \times 10^{-11} (1.59 \times 10^{-9})$ |
| Species K. elongata | 61.99% | 76.94% | 46.62% | $1.68 \times 10^{-7} (1.16 \times 10^{-5})$ | 0.35 (1.00) | 5.08×10^{-9} (3.51×10^{-7}) |
| Species Neisseria oralis | 19.93% | 43.19% | 53.20% | $1.74{\times}10^{-22}$ $(1.20{\times}10^{-20})$ | $5.14{	imes}10^{-4}$ (0.04) | $1.00 \times 10^{-22} (6.96 \times 10^{-21})$ |
| Genus Cardiobacterium | 34.46% | 56.81% | 59.41% | 1.40×10^{-11} (9.67×10 ⁻¹⁰) | 0.43 (1.00) | $2.03 \times 10^{-12} (1.40 \times 10^{-10})$ |
| Phylum <i>Bacteroidetes</i> | | | | | | |
| Species Prevotella nanceiensis | 80.74% | 86.16% | 89.40% | $7.11 \times 10^{-5} (4.91 \times 10^{-3})$ | 0.30 (1.00) | $6.68 \times 10^{-5} (4.61 \times 10^{-3})$ |
| Species Capnocytophaga sputigena | 43.07% | 60.80% | 62.52% | 5.17×10^{-8} (3.57×10^{-6}) | 0.97 (1.00) | $1.20{	imes}10^{-8}$ ($8.28{	imes}10^{-7}$) |
| Phylum <i>Firmicutes</i> | | | | | | |
| Family Lactobacillaceae | 90.37% | 76.73% | 77.15% | $4.45 \times 10^{-5} (3.07 \times 10^{-3})$ | 0.43 (1.00) | $2.99 \times 10^{-6} (2.06 \times 10^{-4})$ |
| Genus Enterococcus | 63.18% | 71.28% | 74.59% | $2.51 \times 10^{-6} (1.73 \times 10^{-4})$ | 0.26 (1.00) | $4.27 \times 10^{-6} (2.95 \times 10^{-4})$ |
| Genus Lachnospiraceae_(G-2) | 51.69% | 52.83% | 61.61% | $1.34 \times 10^{-4} (9.22 \times 10^{-3})$ | $5.30{	imes}10^{-3}$ (0.36) | 7.97×10^{-4} (0.05) |
| Species Lachnoanaerobaculum umeaense | 72.97% | 81.76% | 82.82% | 4.45×10–4 (0.03) | 0.92 (1.00) | $3.80 \times 10^{-5} (2.62 \times 10^{-3})$ |
| Species Eubacterium infirmum | 62.16% | 71.70% | 77.70% | $4.15 \times 10^{-5} (2.88 \times 10^{-3})$ | 0.02 (1.00) | $7.28 \times 10^{-5} (5.02 \times 10^{-3})$ |
| Phylum Spirochaetes | | | | | | |
| Genus Treponema denticola | 69.93% | 49.06% | 54.30% | 4.19×10^{-4} (0.03) | 0.25 (1.00) | $3.15 \times 10^{-5} (2.17 \times 10^{-3})$ |
| * P values were calculated by logistic regression | I. Sequencing batc | h as well as other | covariates (age, sex | ; race, BMI, alcohol consumption, c | oral health and disease status a | t the first follow-up and total energy |

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 \dot{r} Bonferroni correction, adjusted for 69 independent tests.

intake) were adjusted for.

BMI, body mass index.

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