

## Research Article

# HAVCR1 Affects the MEK/ERK Pathway in Gastric Adenocarcinomas and Influences Tumor Progression and Patient Outcome

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Received 2 January 2019; Revised 16 August 2019; Accepted 3 October 2019; Published 2 December 2019

Academic Editor: Haruhiko Sugimura

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The hepatitis A virus cellular receptor 1 (HAVCR1) gene as a sensitive and specific biomarker has been reported in various diseases. Especially, HAVCR1 overexpression promotes the development and progression of several human cancers. Hence, we aimed to detect the effects of HAVCR1 on gastric adenocarcinoma (GAC). We first determined the expression of HAVCR1 in GAC tissues compared with normal gastric tissues based on the Cancer Genome Atlas (TCGA) database using bioinformatics analysis methods. Then, we assessed the biological function of HAVCR1 in GAC cells using quantitative real-time reverse transcription-PCR (qRT-PCR), western blot, cell counting kit-8- (CCK-) 8, colony formation assay, wound healing assay, and transwell assay. Our results showed that HAVCR1 expression was upregulated in GAC tissues and positively associated with poor survival. Loss-of-function analyses indicated that knockdown of HAVCR1 inhibited the proliferation, colony formation, migration, and invasion of GAC cells. Furthermore, reduction of HAVCR1 in GAC cells can decrease the expression of phosphorylated MEK/ERK. These findings suggested that HAVCR1 may represent a potential biomarker for GAC prognosis, as well as a novel therapeutic target for GAC treatment.

## 1. Introduction

Gastric cancer is the fourth most common cancer and the second leading cause of cancer mortality in the world [1]. A large majority (approximately 90%) of gastric cancers are gastric adenocarcinomas [1, 2]. A gastric adenocarcinoma (GAC) arises from the glands of the most superficial layer, or the mucosa, of the stomach, which is related with chronic gastritis, high salt intake, *Helicobacter pylori* infection, smoking, and pernicious anemia [2, 3]. Surgical resection is the standard primary therapy for GAC. However, after surgery, the overall 5-year survival rate remains poor because of the high locoregional and distant recurrence rates [3, 4]. To improve the outcome of surgery, adjuvant or neoadjuvant

therapy is frequently used in combination with surgery such as radiotherapy and chemotherapy. However, the addition of radiotherapy and chemotherapy has not been reported to provide any obvious additional benefit [1, 4]. Hence, how to treat GAC effectively has become an urgent task. As target therapy can offer a specific and effective treatment for GAC patients, target therapy has become the research priority of GAC treatment [5, 6]. Therefore, it is critical to choose some effective biomarkers for GAC target therapy.

The hepatitis A virus cellular receptor 1 (HAVCR1) gene maps on the 5q33.2 cytogenetic location, which codes for a type I transmembrane glycoprotein [7, 8]. HAVCR1, also known as human kidney injury molecule-1 (KIM-1) and T-cell immunoglobulin and mucin domain-1 (TIM-1), was

initially described in primate kidney cells [9] and is associated with disease susceptibility [10]. HAVCR1 has been reported as a sensitive and specific biomarker for hepatitis [11, 12] and acute kidney injury [13–15]. According to the previous report of Vila et al., HAVCR1 was firstly proven to inhibit cell differentiation and was highly expressed in clear cell renal cell carcinoma [16]. Moreover, emerging evidence suggested that HAVCR1 was correlated with some aggressive tumors such as renal cell carcinoma [7, 17, 18], human colorectal cancer [10], and ovarian clear cell carcinoma [19]. Consequently, HAVCR1 can be considered as an effective biomarker that is related with tumor development and progression.

In this study, we aimed to explore the expression levels of HAVCR1 in GAC and also explore the prognostic significance and relationship between HAVCR1 expression and GAC patients' clinicopathologic characteristics. We also examined the potential function of HAVCR1 in GAC cells.

## 2. Materials and Methods

**2.1. Database.** The dataset of the gene was from TCGA, which includes 375 samples of GAC tissues and 32 samples of normal gastric tissues. The corresponding full-scale clinical information of 306 GAC patients was also obtained from the TCGA database (<https://cancergenome.nih.gov/>). The data collection processes were in accordance with all laws and regulations.

**2.2. Cell Culture and Transfection.** Human gastric epithelial cell line GES-1 and human gastric cancer cell lines MKN-45 and AGS were purchased from Shanghai Institutes for Biological Sciences (Shanghai, China). Cells were cultured in RPMI-1640 medium supplemented with 10% fetal bovine serum (FBS), 100 U/ml penicillin, and 0.1 mg/ml streptomycin at 37°C in a humid incubator containing 5% CO<sub>2</sub>.

Cells were transfected using Lipofectamine 2000 (Invitrogen Life Technologies, Karlsruhe, Germany) according to the manufacturer's protocol. Two types of siRNAs were used to knockdown the HAVCR1 expression. The siRNA sequences of HAVCR1 are as follows: F: 5'-UCC UUG GUG GGA GAU AGA G-3' for siRNA1 (si-HAVCR1#1), and F: 5'-GAG AAC UCA GGA ACU CUC A-3' for siRNA2 (si-HAVCR1#2). Meanwhile, nonspecific siRNA was used as a negative control group (si-con). The transfected cells were harvested after 48 h of transfection for the following experiments. Transfection efficiency was measured by qRT-PCR and western blot.

**2.3. RNA Extraction and qRT-PCR.** Total RNA was extracted using the Trizol Reagent (Invitrogen, Grand Island, NY, USA) according to the manufacturer's protocol. Samples were reverse transcribed to cDNA using a PrimeScript RT reagent Kit (Takara, Dalian, China). The mRNA expression of HAVCR1 was determined by real-time PCR using SYBR Green (Takara, Dalian, China). The qRT-PCR cycle conditions were 95°C for 5 min, 40 cycles of 95°C for 30 sec, 60°C for 40 sec, and 72°C for 1 min. The results were quantified and normalized using the 2<sup>-ΔΔCt</sup> method relative to GAPDH.

Each assay was independently performed three times. The primers were as follows: HAVCR1 forward, 5'-TGG TGG GAG ATA GAG GAA GCA T-3', and reverse, 5'-GAT CAG CGT TCA GAT CCA GG-3'; GAPDH forward, 5'-GGA GCG AGA TCC CTC CAA AAT-3', and reverse, 5'-GGC TGT TGT CAT ACT TCT CAT GG-3'.

**2.4. Western Blot.** Cells were harvested and treated with lysis buffer (KeyGEN, Nanjing, China) on ice for 30 min. A BCA kit (KeyGEN, Nanjing, China) was used to quantify protein concentration. Equal amounts of protein (20 μg) were resolved on SDS-PAGE and transferred to a PVDF membrane. Membranes were blocked with 5% nonfat milk for 1 h and incubated overnight at 4°C with primary antibodies (dilution 1:1000, Cell Signaling Technology, Inc., Danvers, MA, USA) against MEK (Cat no. 9126), p-MEK (Cat no. 3958), ERK (Cat no. 4695), p-ERK (Cat no. 8544), GAPDH (Cat no. 5174), and HAVCR1 (Cat no. 14971). Subsequently, membranes were washed with TBST and cultured with secondary antibody (anti-rabbit IgG, HRP-conjugated goat anti-rabbit, dilution 1:3000, Cat no. 7074, Cell Signaling Technology, Inc., Danvers, MA, USA) for 2 h at room temperature. The samples were detected by an ECL detection system (Thermo Fisher Scientific). And the intensity of the signal on each membrane was quantified by densitometry (Quantity One software, Bio-Rad). GAPDH was used as control. All experiments were repeated at least three times.

**2.5. Cell Proliferation Assay.** The cell proliferation was determined using CCK-8 according to the manufacturer's protocol. The transfected cells were seeded in 96-well plates at a density 1000 cells per well and determined at 24 h, 48 h, 72 h, and 96 h using a CCK-8 kit. The absorbance was measured at 450 nm using a microplate reader (Thermo Fisher Scientific Microplate Reader, Waltham, Massachusetts, USA). Each experiment was performed independently in triplicate.

**2.6. Colony Formation Assays.** The transfected cells were seeded in six-well plates and incubated in medium with 10% FBS that was replaced every 3 or 4 days. After two weeks, cells were fixed with 4% paraformaldehyde for 30 min and stained with 0.1% crystal violet for 30 min (Sigma-Aldrich, St. Louis, MO, USA). Visible colonies were counted and imaged. Each sample was determined independently at three times.

**2.7. Wound Healing Assay.** The transfected cells were cultured on six-well plates at 37°C in a humid incubator with 5% CO<sub>2</sub> for 24 h, resulting in a monolayer at more than 90% confluence. Then monolayer cells were scraped with a 200 μl standard pipette tip to generate a wound. The cells were washed with fresh medium to remove floating cells. Subsequently, the spread of the wound closure was recorded after 24 h of incubation under an Olympus BX51 microscope (Olympus Corporation, Tokyo, Japan). The experiment was repeated at three times.

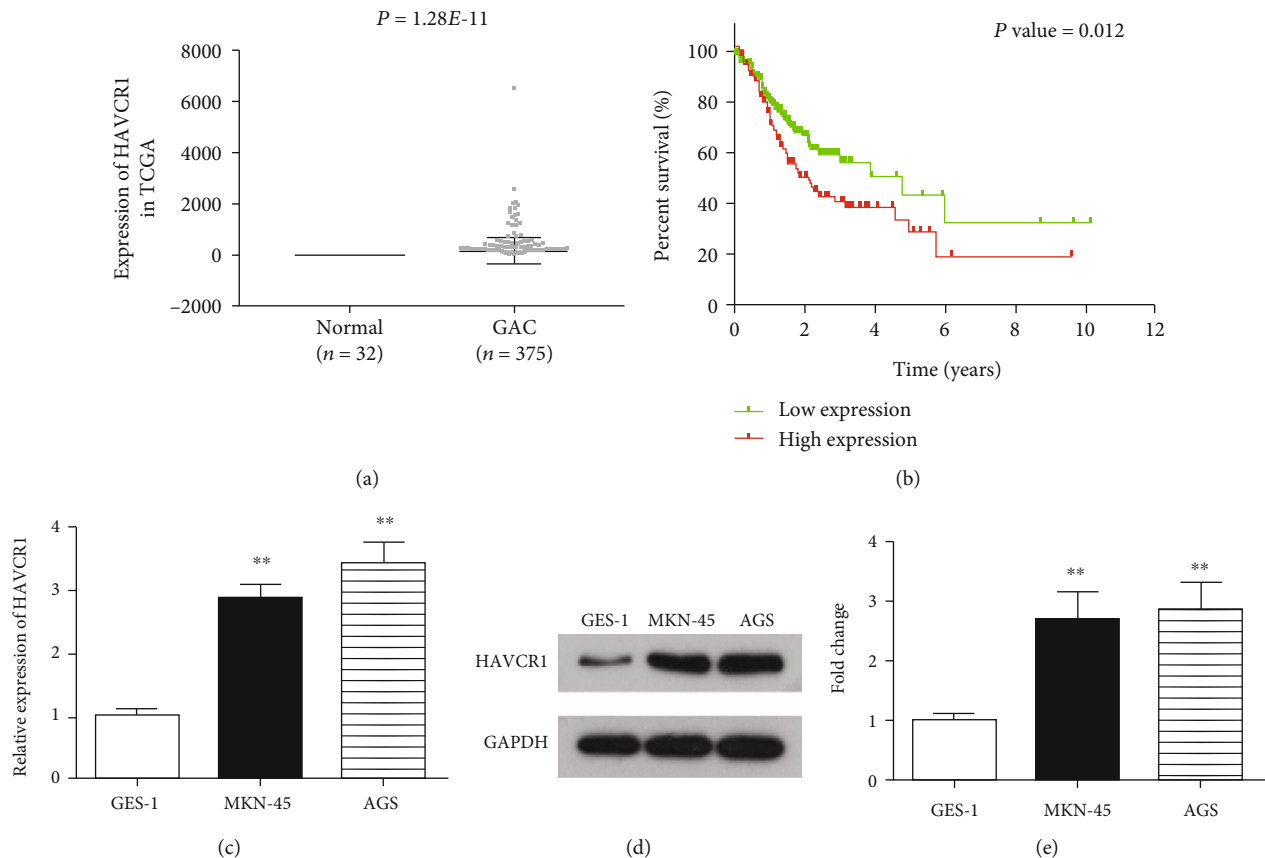


FIGURE 1: HAVCR1 was upregulated in GAC tissues and cell lines, and high HAVCR1 expression was related with poor prognosis. (a) HAVCR1 expression was upregulated in GAC tissues ( $n = 375$ ) compared with normal gastric tissues ( $n = 32$ ). (b) Kaplan-Meier's survival analysis showed that the high regulation of HAVCR1 had a close relationship with the poor outcome of GAC patients,  $p = 0.012$ . (c) QRT-PCR reveals the expression levels of HAVCR1 in GAC cells (MKN-45 and AGS) and in a normal gastric cell line (GES-1). (d) Western blot analysis was performed to detect the expression of HAVCR1. (e) The gray value of protein bands was quantified. \*\* $p < 0.01$  versus GES-1 cells.

**2.8. Transwell Migration and Invasion Assays.** Transwell assays were carried out in 24-well plates using a transwell chamber with/without Matrigel (pore size  $8\mu\text{m}$ , BD Biosciences, Lake Franklin, New Jersey, USA). In the migration assay, cells were suspended in serum-free RPMI 1640 medium and added to the upper chamber at  $1 \times 10^5$  cells/transwell. In the invasion assay, cells suspended in serum-free RPMI 1640 medium were seeded in the upper chamber coated with Matrigel at  $1 \times 10^5$  cells/transwell. In each assay, the lower chamber was filled with 10% FBS RPMI 1640 medium as a chemoattractant. After 24 hours of incubation, the upper surfaces of the transwells were wiped with cotton swabs. The inserts were fixed with paraformaldehyde for 30 min, stained with 0.1% crystal violet for 20 min, and then washed with PBS. Next, the migrating and invading cells were counted and photographed under an Olympus BX51 microscope (Olympus Corp., Shinjuku, Tokyo, Japan) at 200x magnification over five random fields in each well. The experiment was repeated three times.

**2.9. Statistical Analysis.** All values are expressed as the mean  $\pm$  standard deviation (SD) and the data were analyzed by SPSS Statistics software (version 22.0, Chicago, IL, USA).

The relationship between HAVCR1 expression and clinical characteristics was determined by a Chi-square test and one-way ANOVA. The Kaplan-Meier method and the log-rank test were used to generate overall survival curves. The COX regression analysis was used to detect whether HAVCR1 is an independent risk factor for prognosis in GAC cancer. Student's  $t$ -test was used to measure significant differences between two groups. Differences were considered significant for  $p < 0.05$ .

### 3. Results

**3.1. HAVCR1 Expression Was Upregulated in GAC and Correlated with Poor Prognosis.** To identify the potential role of HAVCR1 in GAC, we first assessed HAVCR1 expression in 375 GAC tissues and 32 normal gastric tissues based on The Cancer Genome Atlas (TCGA, <https://cancergenome.nih.gov/>) database. We found that HAVCR1 expression was obviously upregulated in GAC tissues ( $n = 375$ ) compared with normal tissues ( $n = 32$ ;  $p < 0.01$ , Figure 1(a)). Moreover, a Chi-square test revealed that HAVCR1 expression was positively correlated with death ( $p = 0.002$ , Table 1).

TABLE 1: Correlation between HAVCR1 expression and clinicopathological characteristics of GAC patients.

Characteristics	Expression of HAVCR1		<i>p</i> value
	Low	High	
<i>Age</i>			0.067
<60	42	57	
≥60	111	96	
<i>Gender</i>			0.813
Female	56	56	
Male	95	97	
<i>Grade</i>			0.478
G1+G2	60	54	
G3	93	99	
<i>Clinical stage</i>			0.359
I+II	74	66	
III+IV	79	87	
<i>Pathologic-T</i>			0.290
T1+T2	34	42	
T3+T4	119	111	
<i>Pathologic-N</i>			0.711
N0	49	46	
N1	104	107	
<i>Pathologic-M</i>			0.644
M0	144	142	
M1	9	11	
<i>Death</i>			0.002*
Yes	49	75	
No	104	78	

T: tumor status; N: regional lymph node status; M: metastasis status.  
\*Significant correlation.

In addition, after excluding patients without follow-up, 306 GAC patients remained and their full-scale clinical information was extracted from TCGA database. The median expression value was used as a cutoff point, and the GAC patients were divided into two groups: the HAVCR1-high group (>median value;  $n = 153$ ) and the HAVCR1-low group (<median value;  $n = 153$ ). The data were analyzed by the Kaplan-Meier method with a log-rank test, and thus overall survival curves were plotted. As shown in Figure 1(b), compared with the HAVCR1-low group, the HAVCR1-high group showed poor overall survival ( $p = 0.012$ ).

Moreover, the prognostic value of HAVCR1 in GAC patients was evaluated by the Cox regression analysis. As shown in Table 2, univariate analysis showed that HAVCR1 expression ( $p = 0.013$ , HR = 1.58), clinical-stage ( $p = 0.001$ , HR = 1.883), pathologic-N ( $p = 0.005$ , HR = 1.873), and age ( $p = 0.011$ , HR = 1.697) were significantly related to overall survival in GAC patients. Multivariate analysis indicated that HAVCR1 expression ( $p = 0.008$ , HR = 1.633) and age ( $p = 0.002$ , HR = 1.954) could be regarded as independent risk factors for overall survival in GAC patients. Collectively, results showed that HAVCR1 might be a potential biomarker and prognostic predictor for GAC.

**3.2. HAVCR1 Expression Was Upregulated in GAC Cells.** To investigate the effect of HAVCR1 on GAC cells, we first determined the expression of HAVCR1 in different GAC cell lines using quantitative real-time reverse transcription-PCR (qRT-PCR). As shown in Figure 1(c), when compared with the normal gastric cell line (GES-1), HAVCR1 expression was markedly upregulated in GAC cell lines (MKN-45 and AGS), especially the AGS cell line. Moreover, western blotting analysis exhibited that the protein expression of HAVCR1 in MKN-45 and AGS cells was higher than that of GES-1 cells ( $p < 0.01$ , Figures 1(d) and 1(e)). These results confirmed that HAVCR1 expression was highly regulated in GAC cell lines. In order to examine the effects of HAVCR1 in GAC cells, we first knocked down the expression of HAVCR1 using siRNA strategy. QRT-PCR and western blot methods were conducted to detect the silencing efficiency in MKN-45 and AGS cells ( $p < 0.01$ , Figure 2).

**3.3. Downregulation of HAVCR1 Impaired the Proliferation and Colony Formation of GAC Cells.** To explore the effect of HAVCR1 on proliferative and clonogenic abilities of GAC cells, CCK-8 and colony formation assays were performed in MKN-45 and AGS cells. In the CCK-8 assay, the proliferation of MKN-45 and AGS, which were transfected with si-HAVCR1#1 and si-HAVCR1#2, showed an obvious decrease when compared with the negative control group (si-con) ( $p < 0.01$ , Figures 3(a) and 3(b)). The same trends were also observed in the colony formation assay. As shown in Figures 3(c) and 3(d), the number of clones in the si-HAVCR1#1 and si-HAVCR1#2 groups was significantly decreased relative to the si-con group in two GAC cell lines, including MKN-45 and AGS ( $p < 0.01$ ). Therefore, these observations disclosed that knockdown of HAVCR1 played an inhibitory role in the proliferation and colony formation of GAC cells.

**3.4. Reduction of HAVCR1 Attenuated the Migration and Invasion of GAC Cells.** To determine the impact of HAVCR1 on the mobility of GAC cells, a wound healing assay and transwell assays were used for this purpose. For wound-healing analysis in MKN-45 cells, the distances of the wound in the si-HAVCR1#1 and si-HAVCR1#2 groups were larger than that in the si-con group after 24 h of incubation ( $p < 0.01$ , Figure 4(a)). Consistently, the wound closure of the si-HAVCR1#1 and si-HAVCR1#2 groups in AGS cells was wider than that in the si-con group ( $p < 0.01$ , Figure 4(b)). Furthermore, transwell assays in MKN-45 and AGS cells indicated that si-HAVCR1#1 and si-HAVCR1#2 transfection markedly repressed the migration and invasion of GAC cells compared with si-con treatment ( $p < 0.01$ , Figures 4(c) and 4(d)). Taken together, the knockdown of HAVCR1 inhibited the migration and invasion of GAC cells.

**3.5. HAVCR1 Deficiency Affected the Activation of Extracellular Signal-Regulated Kinase (MEK/ERK) Signaling Pathway.** To investigate the potential mechanism of HAVCR1 on the proliferation, colony formation, migration, and invasion of GAC cells, the ERK signaling cascade, as one of the

TABLE 2: Univariate and multivariate analysis of clinic pathologic factors for overall survival in GAC patients.

Variables	<i>p</i> value	Univariate analysis		<i>p</i> value	Multivariate analysis	
		HR	95% CI		HR	95% CI
HAVCR1 expression (high/low)	0.013*	1.580	1.101-2.265	0.008*	1.633	1.134-2.353
Clinical stage (I+II/III+IV)	0.001*	1.883	1.292-2.745	0.092	1.534	0.933-2.523
Pathologic-T (T1+T2/T3+T4)	0.064	1.529	0.975-2.398			
Pathologic-M (M0/M1)	0.126	1.659	0.868-3.173			
Pathologic-N (N0/N1+N2+N3)	0.005*	1.873	1.207-2.906	0.225	1.431	0.802-2.553
Age (<60/≥60)	0.011*	1.697	1.128-2.551	0.002*	1.954	1.292-2.955
Gender (female/male)	0.083	1.405	0.956-2.065			
Grade (G1+G2/G3)	0.124	1.340	0.923-1.946			

HR: hazard ratio; CI: confidence interval. \*Significant correlation.

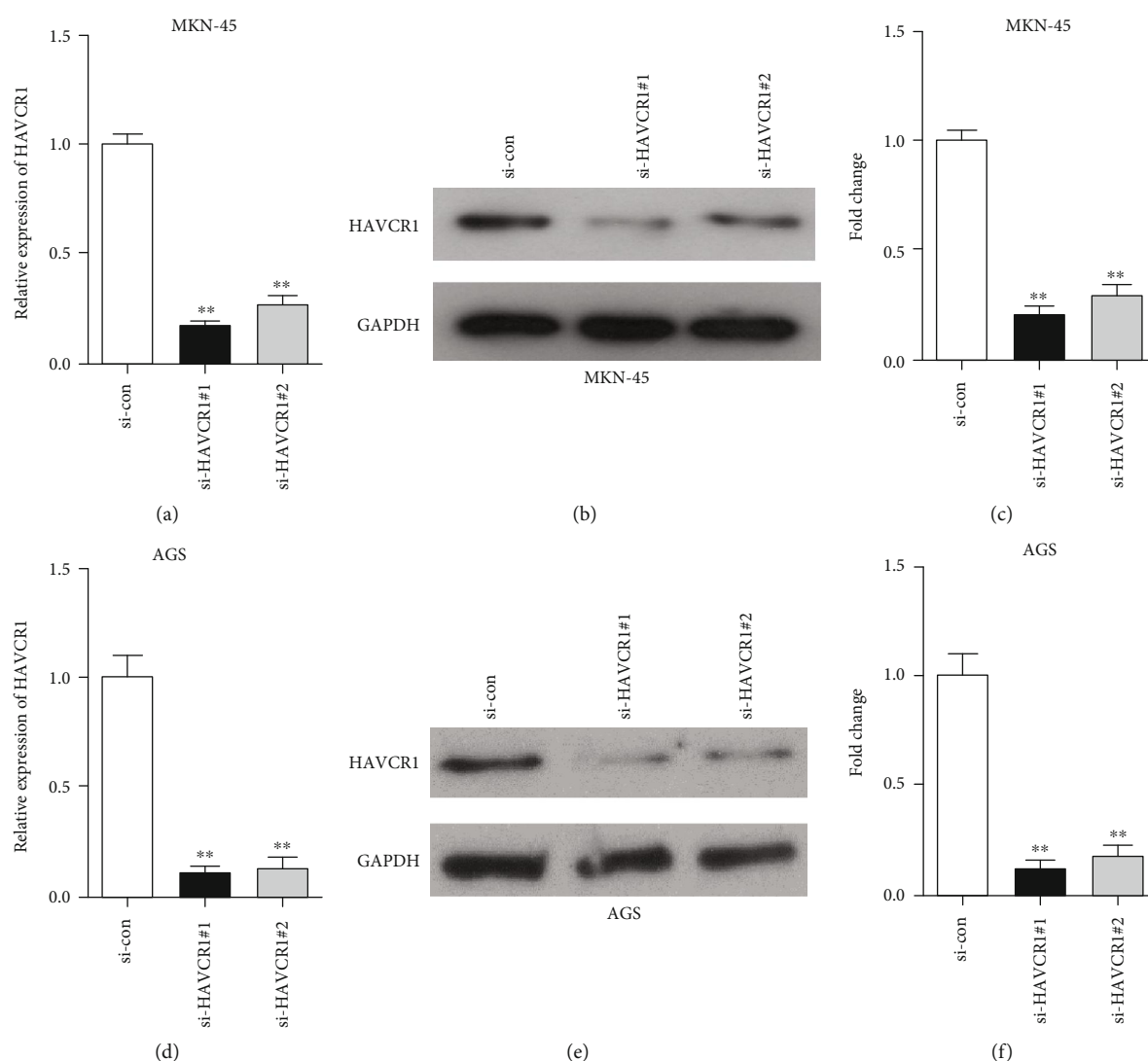


FIGURE 2: HAVCR1 expression was significantly decreased in MKN-45 and AGS cells using siRNA strategy. (a) QRT-PCR was used to assess the HAVCR1 mRNA expression in MKN-45 cells after si-HAVCR1#1 and si-HAVCR1#2 transfection, respectively,  $**p < 0.01$  versus the si-con group. (b and c) The HAVCR1 protein level was evaluated using MKN-45 cells that were transfected with si-HAVCR1#1 or si-HAVCR1#2. The gray value of the protein bands was quantified.  $**p < 0.01$  versus the si-con group. (d) QRT-PCR analysis showed the silencing efficiency of si-HAVCR1#1 and si-HAVCR1#2 in AGS cells,  $**p < 0.01$  versus the si-con group. (e and f) The expression of HAVCR1 in AGS cells was detected by western blot and quantified,  $**p < 0.01$  versus the si-con group.



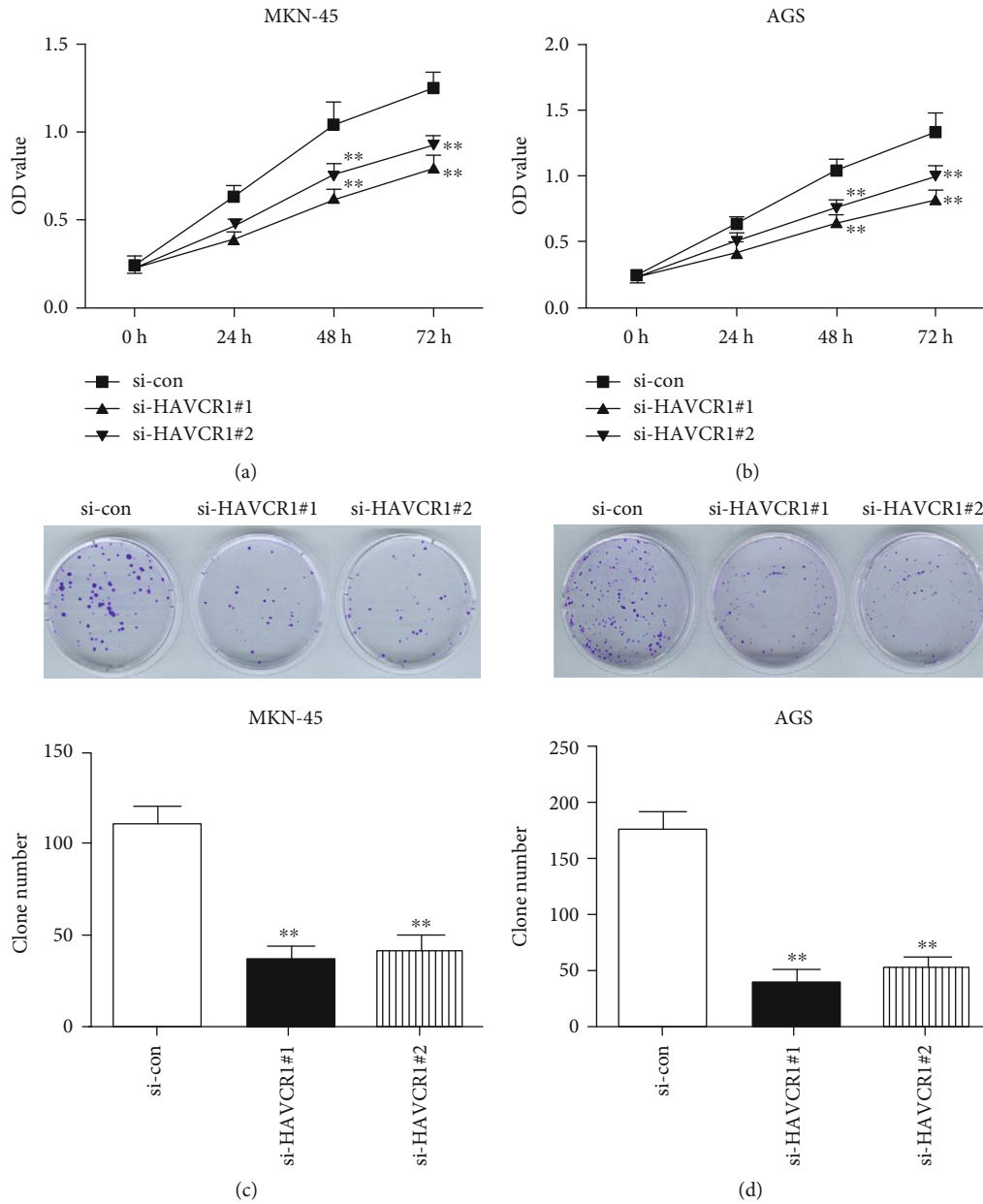


FIGURE 3: Downregulation of HAVCR1 impaired the proliferation and colony formation of GAC cells. (a and b) A CCK-8 assay was used to measure the proliferation of MKN-45 and AGS cells. (c and d) A colony formation assay was conducted in MKN-45 and AGS cells to further determine the impact of HAVCR1 deficiency on clonogenic capability. And the number of clones was counted. \*\* $p < 0.01$  versus the si-con group.

most frequently dysregulated pathways in human cancer, was determined using western blot. Due to the highest silencing efficiency and more obvious effects of si-HAVCR1#1, the subsequent western blotting experiments were implemented by si-HAVCR1#1. After si-HAVCR1#1 transfection, the expression levels of p-MEK and p-ERK in MKN-45 and AGS cells were significantly downregulated compared to the si-con group ( $p < 0.01$ , Figures 5(a)–5(d)), whereas the total MEK and ERK expression levels showed no differences between the si-HAVCR1#1 group and the si-con group ( $p < 0.01$ , Figures 5(a)–5(d)). In summary, HAVCR1 deficiency blocked the activation of the MEK/ERK signaling

pathway, thereby affecting the proliferation, colony formation, migration, and invasion of GAC cells.

#### 4. Discussion

In this study, we first analyzed the expression levels of HAVCR1 in GAC samples based on TCGA database. We found that HAVCR1 expression was upregulated in GAC tissues compared with the normal gastric tissues and positively related with poor prognosis. Cox analysis showed that HAVCR1 might be an independent predictor for prognosis of GAC patients. Moreover, functional experiments revealed

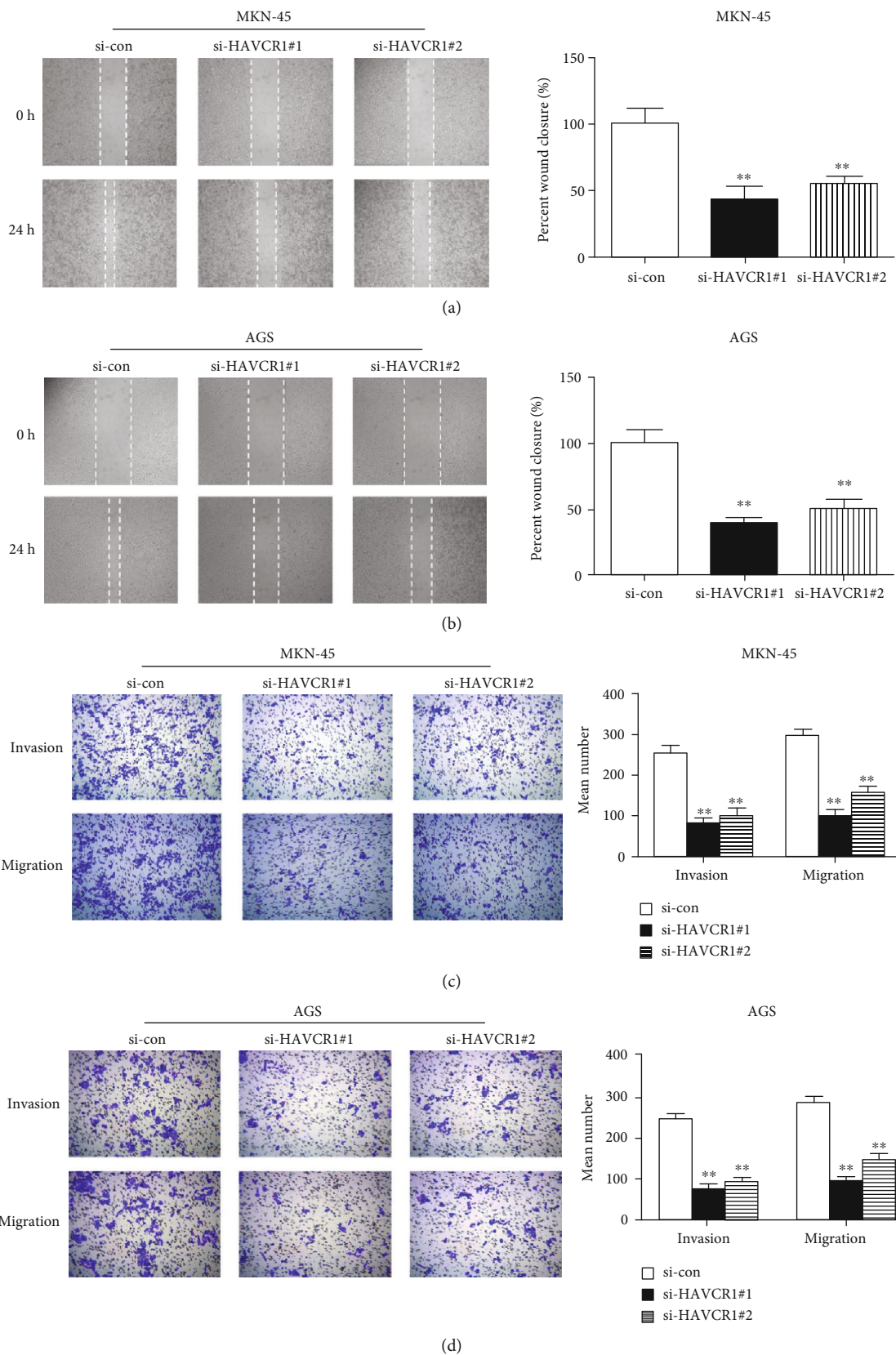


FIGURE 4: Reduction of HAVCR1 attenuated the migration and invasion of GAC cells. A wound healing assay was implemented to assess the migration of (a) MKN-45 cells and (b) AGS cells. And the distance of the wound was also quantified. (c and d) The migration and invasion of GAC cells was further detected by transwell assays.  $**p < 0.01$  versus the si-con group.

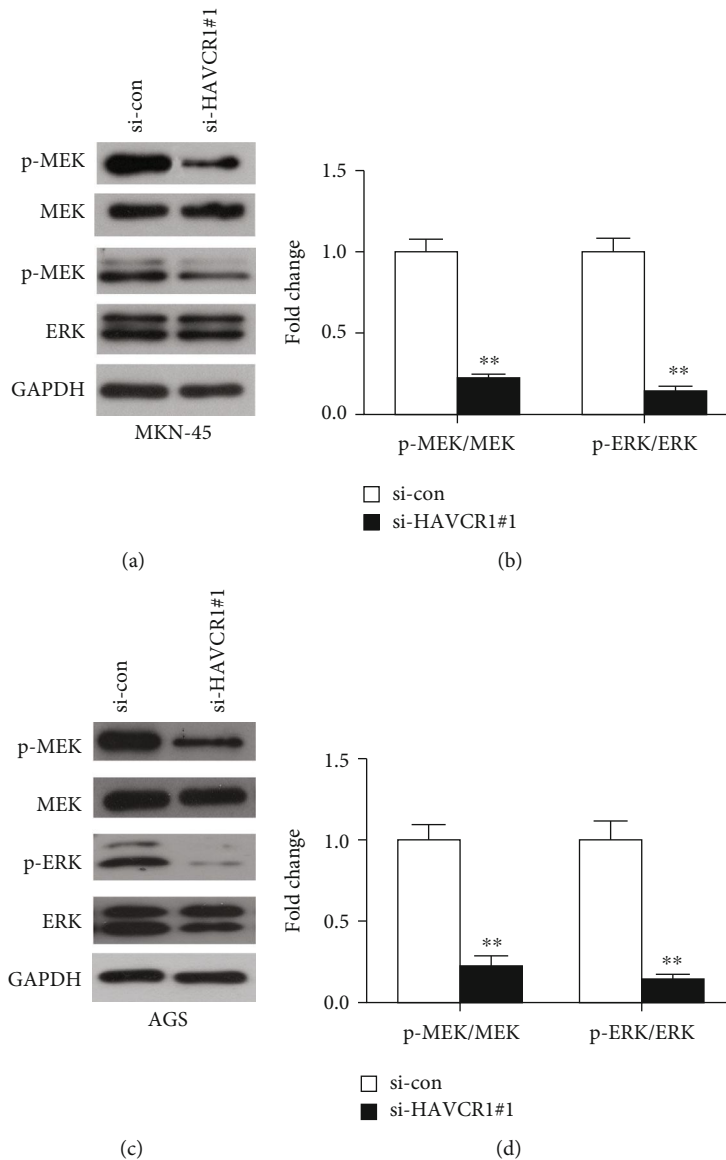


FIGURE 5: Activation of MEK/ERK signaling pathway was associated with HAVCR1 expression. (a) The MEK/ERK related protein levels were examined by western blot analysis in MKN-45 cells after si-HAVCR1#1 transfection. (b) The gray values of protein bands were quantified.  $**p < 0.01$  versus the si-con group. (c) The effects of HAVCR1 deficiency in MEK, p-MEK, ERK, and p-ERK expression levels in AGS cells were measured by western blot. (d) The expression levels of the above proteins were quantified and normalized to GAPDH. One specific siRNA (si-HAVCR1#1) was used as an experimental group because of its higher knockdown efficiency, and nonspecific siRNA was used as a negative control group (si-con).  $**p < 0.01$  versus the si-con group.

that the downregulation of HAVCR1 suppressed the proliferation, colony formation, migration, and invasion of GAC cells via mediating the activation of the MEK/ERK pathway. Hence, HAVCR1 may be considered as a potential biomarker and prognostic predictor for GAC.

HAVCR1 has been reported to be associated with many diseases such as kidney injury [13–15], infection (hepatitis C virus, hepatitis A virus, and ebolavirus) [11, 12, 20, 21], and autoimmune diseases (allergy, asthma, and systemic lupus erythematosus patients) [22–24]. These studies have shown that HAVCR1 has multiple functions in various diseases. Particularly, the role of HAVCR1 as a receptor in hepatocytes has been widely studied. For instance, HAVCR1 can be a facilitator of hepatitis C virus (HCV) entry, which could

stabilize or enhance virus binding to the cell surface membrane and allow the correct virus-receptor positioning for interaction with the main HCV receptors [12]. What's more, HAVCR1 overexpression was observed in renal cell carcinoma [7], human colorectal cancer [10], and ovarian clear cell carcinoma [19], which could promote the development and progression of tumors. Thus, HAVCR1 overexpression may be a diagnostic biomarker for several cancers [10]. According to these findings, we aimed to explore the relationship between HAVCR1 and GAC progression. Based on TCGA database, HAVCR1 expression was upregulated in GAC tissues and correlated with poor survival. Additionally, our experimental results indicated that HAVCR1 deficiency impaired the proliferation, colony formation, migration, and



invasion of GAC cells. Therefore, HAVCR1 might be a potential biomarker for GAC prognosis and an effective therapeutic target for GAC treatment.

The ERK signaling cascade is a central MAPK pathway that regulates a variety of cellular processes such as proliferation, differentiation, survival, migration, and invasion [25–27]. ERK can be activated by various oncogenes and extracellular stimuli, and MEK acts as an upstream key protein of ERK [26, 27]. The ERK signaling pathway has been reported to play an important role in gastric cancer cell growth and metastasis [28–30]. More importantly, Zhang and Cai indicated that HAVCR1 can mediate renal epithelial cell repair through the ERK/MAPK signaling pathway [31]. Hence, in this study, we detected the effect of the downregulation of HAVCR1 on the MEK/ERK pathway in AGS cells. The phosphorylation of MEK and ERK expression levels showed an obvious decrease after the knockdown of HAVCR1. Therefore, we guessed that the effects of HAVCR1 on proliferation, colony formation, migration, and invasion of GAC cells might be associated with the MEK/ERK pathway.

## 5. Conclusions

Our data uncovered that HAVCR1 expression was upregulated in GAC tissues and cell lines, and its high regulation was linked with the poor outcome of GAC patients. Moreover, loss-of-function experiments revealed that the downregulation of HAVCR1 played negative effects on the proliferation, colony formation, migration, and invasion of MKN-45 and AGS cells. The MEK/ERK-related proteins were also decreased after HAVCR1 inhibition. To sum up, HAVCR1 may be a potential biomarker for GAC prognosis, as well as a novel therapeutic target for GAC treatment. Of course, further studies are needed to validate these findings and to clarify the detailed mechanism for the promoting role of HAVCR1 on GAC.

## Data Availability

The data used to support the findings of this study are available from the corresponding author upon request

## Conflicts of Interest

The authors declare no conflict of interests.

## Acknowledgments

The authors acknowledge the support of the Gansu University Research Project (grant no. 2017A-052) and the Lanzhou Chengguan District Science and Technology Research Project (grant no. 2017~3-3).

## References

- [1] E. Van Cutsem, X. Sagaert, B. Topal, K. Haustermans, and H. Prenen, "Gastric cancer," *The Lancet*, vol. 388, no. 10060, pp. 2654–2664, 2016.
- [2] P. Karimi, F. Islami, S. Anandasabapathy, N. D. Freedman, and F. Kamangar, "Gastric cancer: descriptive epidemiology, risk factors, screening, and prevention," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 23, no. 5, pp. 700–713, 2014.
- [3] S. Sandler, "Esophagogastric junction and gastric adenocarcinoma: neoadjuvant and adjuvant therapy, and future directions," *Oncology (Williston Park)*, vol. 28, no. 6, pp. 505–512, 2014.
- [4] E. Niccolai, A. Taddei, D. Prisco, and A. Amedei, "Gastric cancer and the epoch of immunotherapy approaches," *World Journal of Gastroenterology*, vol. 21, no. 19, pp. 5778–5793, 2015.
- [5] S. M. Lim, J. Y. Lim, and J. Y. Cho, "Targeted therapy in gastric cancer: personalizing cancer treatment based on patient genome," *World Journal of Gastroenterology*, vol. 20, no. 8, pp. 2042–2050, 2014.
- [6] C. Duraes, G. M. Almeida, R. Seruca, C. Oliveira, and F. Carneiro, "Biomarkers for gastric cancer: prognostic, predictive or targets of therapy?," *Virchows Archiv*, vol. 464, no. 3, pp. 367–378, 2014.
- [7] T. Cuadros, E. Trilla, E. Sarro et al., "HAVCR/KIM-1 activates the IL-6/STAT-3 pathway in clear cell renal cell carcinoma and determines tumor progression and patient outcome," *Cancer Research*, vol. 74, no. 5, pp. 1416–1428, 2014.
- [8] J. R. García-Lozano, C. Abad, A. Escalera et al., "Identification of HAVCR1 gene haplotypes associated with mRNA expression levels and susceptibility to autoimmune diseases," *Human Genetics*, vol. 128, no. 2, pp. 221–229, 2010.
- [9] G. Kaplan, A. Totsuka, P. Thompson, T. Akatsuka, Y. Moritsugu, and S. M. Feinstone, "Identification of a surface glycoprotein on African green monkey kidney cells as a receptor for hepatitis A virus," *The EMBO Journal*, vol. 15, no. 16, pp. 4282–4296, 1996.
- [10] Y. Wang, T. A. Martin, and W. G. Jiang, "HAVCr-1 expression in human colorectal cancer and its effects on colorectal cancer cells in vitro," *Anticancer Research*, vol. 33, no. 1, pp. 207–214, 2013.
- [11] G. Wojcik, R. Latanich, T. Mosbrugger et al., "Variants in HAVCR1 gene region contribute to hepatitis C persistence in African Americans," *The Journal of Infectious Diseases*, vol. 209, no. 3, pp. 355–359, 2014.
- [12] A. Kachko, M. I. Costafreda, I. Zubkova et al., "Determinants in the Ig Variable domain of human HAVCR1 (TIM-1) are required to enhance hepatitis C virus entry," *Journal of Virology*, vol. 92, no. 6, p. e01742, 2018.
- [13] R. Korbély, J. Wilflingseder, P. Perco et al., "Molecular biomarker candidates of acute kidney injury in zero-hour renal transplant needle biopsies," *Transplant International*, vol. 24, no. 2, pp. 143–149, 2011.
- [14] J. V. Bonventre, "Kidney injury molecule-1: a translational journey," *Transactions of the American Clinical and Climatological Association*, vol. 125, pp. 293–299, 2014.
- [15] K. S. Famulski, D. G. de Freitas, C. Kreepala et al., "Molecular phenotypes of acute kidney injury in kidney transplants," *Journal of the American Society of Nephrology*, vol. 23, no. 5, pp. 948–958, 2012.
- [16] M. R. Vilà, G. G. Kaplan, D. Feigelstock et al., "Hepatitis A virus receptor blocks cell differentiation and is overexpressed in clear cell renal cell carcinoma," *Kidney International*, vol. 65, no. 5, pp. 1761–1773, 2004.
- [17] P. L. Zhang, J. W. Mashni, V. S. Sabbiseti et al., "Urine kidney injury molecule-1: a potential non-invasive biomarker for patients with renal cell carcinoma," *International Urology and Nephrology*, vol. 46, no. 2, pp. 379–388, 2014.

- [18] T. Cuadros, E. Trilla, M. R. Vilà et al., "Hepatitis A virus cellular receptor 1 / kidney injury molecule-1 is a susceptibility gene for clear cell renal cell carcinoma and hepatitis A virus cellular receptor/kidney injury molecule-1 ectodomain shedding a predictive biomarker of tumour progression," *European Journal of Cancer*, vol. 49, no. 8, pp. 2034–2047, 2013.
- [19] F. Lin, P. L. Zhang, X. J. Yang et al., "Human kidney injury molecule-1 (hKIM-1): a useful immunohistochemical marker for diagnosing renal cell carcinoma and ovarian clear cell carcinoma," *The American Journal of Surgical Pathology*, vol. 31, no. 3, pp. 371–381, 2007.
- [20] E. Silberstein, K. Konduru, and G. G. Kaplan, "The interaction of hepatitis A virus (HAV) with soluble forms of its cellular receptor 1 (HAVCR1) share the physiological requirements of infectivity in cell culture," *Virology Journal*, vol. 6, no. 1, p. 175, 2009.
- [21] S. Yuan, L. Cao, H. Ling et al., "TIM-1 acts a dual-attachment receptor for Ebolavirus by interacting directly with viral GP and the PS on the viral envelope," *Protein & Cell*, vol. 6, no. 11, pp. 814–824, 2015.
- [22] L. Chatenoud and J. F. Bach, "Genetic control of hepatitis A severity and susceptibility to allergy," *The Journal of Clinical Investigation*, vol. 121, no. 3, pp. 848–850, 2011.
- [23] J. P. Chen, W. L. Zhao, N. H. He et al., "Association of hepatitis A exposure and TIM-1 with childhood allergic asthma," *The Journal of Asthma*, vol. 49, no. 7, pp. 697–702, 2012.
- [24] H. Yuan, Y. S. Yao, G. M. Chen, J. Sheng, L. Xu, and H. F. Pan, "Decreased serum levels of T-cell immunoglobulin mucin-1 and T-cell immunoglobulin mucin-3 in systemic lupus erythematosus patients," *Journal of Biological Regulators and Homeostatic Agents*, vol. 30, no. 1, pp. 123–129, 2016.
- [25] X. Deschenes-Simard, F. Kottakis, S. Meloche, and G. Ferbeyre, "ERKs in cancer: friends or foes?," *Cancer Research*, vol. 74, no. 2, pp. 412–419, 2014.
- [26] Y. D. Shaul and R. Seger, "The MEK/ERK cascade: from signaling specificity to diverse functions," *Biochimica et Biophysica Acta*, vol. 1773, no. 8, pp. 1213–1226, 2007.
- [27] W. Liu, Z. Zhang, Y. Zhang et al., "HMGB1-mediated autophagy modulates sensitivity of colorectal cancer cells to oxaliplatin via MEK/ERK signaling pathway," *Cancer Biology & Therapy*, vol. 16, no. 4, pp. 511–517, 2015.
- [28] L. Wei, Y. Li, and Z. Suo, "TSPAN8 promotes gastric cancer growth and metastasis via ERK MAPK pathway," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 6, pp. 8599–8607, 2015.
- [29] Z. Lin, C. Zhang, M. Zhang et al., "Targeting cadherin-17 inactivates Ras/Raf/MEK/ERK signaling and inhibits cell proliferation in gastric cancer," *PLoS One*, vol. 9, no. 1, article e85296, 2014.
- [30] J. Qian, X. Kong, N. Deng et al., "OCT1 is a determinant of symbindin-related ERK signalling with independent prognostic significance in gastric cancer," *Gut*, vol. 64, no. 1, pp. 37–48, 2015.
- [31] Z. Zhang and C. X. Cai, "Kidney injury molecule-1 (KIM-1) mediates renal epithelial cell repair via ERK MAPK signaling pathway," *Molecular and Cellular Biochemistry*, vol. 416, no. 1-2, pp. 109–116, 2016.