REVIEW ARTICLE

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Heart failure with preserved ejection fraction: present status and future directions

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Abstract

The clinical importance of heart failure with preserved ejection fraction (HFpEF) has recently become apparent. HFpEF refers to heart failure (HF) symptoms with normal or near-normal cardiac function on echocardiography. Common clinical features of HFpEF include diastolic dysfunction, reduced compliance, and ventricular hypokinesia. HFpEF differs from the better-known HF with reduced ejection fraction (HFrEF). Despite having a "preserved ejection fraction," patients with HFpEF have symptoms such as shortness of breath, excessive tiredness, and limited exercise capability. Furthermore, the mortality rate and cumulative survival rate are as severe in HFpEF as they are in HFrEF. While beta-blockers and renin-angiotensin-aldosterone system modulators can improve the survival rate in HFrEF, no known therapeutic agents show similar effectiveness in HFpEF. Researchers have examined molecular events in the development of HFpEF using small and middle-sized animal models. This review discusses HFpEF with regard to etiology and clinical features and introduces the use of mouse and other animal models of human HFpEF.

Heart failure

The heart supplies oxygen and nutrients to the body through the blood circulation. Rhythmic cardiac motion is required to achieve this function. Cardiac activity can roughly be divided into two phases: diastole (or relaxation) and systole (or contraction)¹. During diastole, the ventricle rapidly increases in volume, with an abrupt decline in intracardiac pressure². When ventricular pressure becomes equal to atrial pressure, the valve between the atrium and ventricle opens. At that moment, blood from a large vein fills the ventricle in early diastole; then, forceful contraction of the atrium moves a small amount of blood into the ventricle. Systole follows diastole. After filling, the ventricle begins to contract. When the contractile pressure exceeds that in a large artery, blood in the ventricle is pushed into the artery. Normal performance in both diastole and systole is required for normal cardiac function².

Heart failure (HF) implies an inadequate blood supply³ and is exemplified by systolic HF⁴. The major

characteristic of systolic HF is decreased contractile function^{3,4}. A patient with HF symptoms and an ejection fraction <40% on echocardiography is diagnosed with systolic HF. Systolic HF is known as HF with reduced ejection fraction (HFrEF)^{3,4}. The most common cause of HFrEF is a loss of effective left ventricular myocardium due to myocardial infarction, which is an ischemic event⁵. Reduced circulation triggers compensatory mechanisms⁶. Decreased perfusion in the kidney results in activation of the renin-angiotensin-aldosterone system (RAAS)⁷. At the same time, increased residual blood in the chamber at end-systole stresses the ventricular wall; this promotes neurohormonal activation and leads to eccentric heart remodeling⁸. Because of its central role in the development of HFrEF, targeted therapy against the RAAS (i.e., the use of an angiotensin-converting enzyme inhibitor, type 1 angiotensin receptor blocker, or aldosterone antagonist) is effective in HFrEF patients^{3,4}. Betaadrenoceptor regulators are standard therapies for HFrEF, as these agents significantly reduce cardiac burden and automaticity^{3,4}.

Many patients with typical HF symptoms, such as shortness of breath, limited exercise capacity, or cough,

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have a normal ejection fraction (>50%) on echocardiography, indicating that systolic function is normal. These patients are diagnosed with HF with preserved ejection fraction (HFpEF). HFrEF refers to systolic HF, but HFpEF does not imply diastolic HF.

Heart failure with a preserved ejection fraction

The prevalence of HFpEF has increased in the last two decades, and approximately half of all HF patients are diagnosed with HFpEF9. HFpEF is more common in women, while HFrEF is more common in men¹⁰. While many patients have been categorized as having "HFpEF," their individual characteristics are heterogeneous¹¹. Big data analysis of the HFpEF cohort has revealed several risk factors. Shah et al. identified the clinical features of HFpEF: 68% were women, 90% had hypertension, 70% had obesity, 62% had hyperlipidemia, and 52% had diabetes mellitus (DM)¹². Another research group further supported this finding. Kao et al. clarified the phenotypes of HFpEF: 59% were women, 100% had DM, 84% had hyperlipidemia, and 75% had obesity (>30 kg/m² body mass index) and hypertension¹³. Both studies consistently confirmed common cardiometabolic features: women with hypertension, obesity, DM, or hyperlipidemia (Fig. 1) predominated^{12,13}.

Commonly, the systolic function in HFpEF remains within the normal range, but the diastolic function is variable^{3,4}. HFpEF is not equivalent to diastolic dysfunction or diastolic HF. Diastolic dysfunction can be defined as a functional defect in relaxation, ventricular filling, or distensibility¹⁴. Diastolic dysfunction indicates abnormal

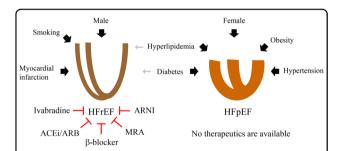


Fig. 1 Comorbidity associated with each type of heart failure. HFrEF or systolic heart failure is more common in males. Well-known underlying risk factors include smoking and myocardial death due to

infarction. However, HFpEF is more common in females. Comorbidities include obesity, hypertension, hyperlipidemia, and diabetes mellitus. While beta-blockers, ACEi/ARB agents, ARNI, ivabradine, and MRAs reduce mortality in HFrEF, no medications are available for HFpEF. ACEi, angiotensin-converting enzyme inhibitor; ARB, angiotensin II type 1 receptor blocker; ARNI, angiotensin receptor neprilysin inhibitor; HFrEF, heart failure with reduced ejection fraction; HFpEF, heart failure with preserved ejection fraction; MRA, mineralocorticoid receptor antagonist. Arrow widths reflect the association and relevance of hazard ratios. The red blunted lines show effective inhibition of disease progression

ventricular mechanical ability. HFpEF is a clinical term used to indicate HF with normal contractile function but without any consideration of diastolic function⁴. Diastolic dysfunction itself is a kind of benign aging process and can be observed in the absence of HF symptoms¹⁵. Although HFpEF and diastolic heart failure are not synonymous, many clinical features overlap^{3,12}. Chronic sustained diastolic dysfunction is a definite risk factor for HFpEF¹⁶.

Another type of HF with subnormal ejection fraction has recently been described, with 40% < ejection fraction < 50%. This ejection fraction is midway between that of a normal and failing heart and has been defined as HF with mid-range ejection fraction (HFmrEF)⁴. Clinically, HFmrEF shares features of both HFpEF and HFrEF¹⁷. In the absence of a cohort study for HFmrEF, the management approaches vary. Some clinicians treat HFmrEF and HFrEF similarly, while others treat these as cases of borderline HFpEF¹⁸. Some experts argue that HFmrEF represents a transition between HFpEF and HFrEF¹⁹, but the evidence is not convincing. The symptoms and hospitalization patterns resemble those of HFpEF, while clinical features, prognosis, and outcomes of HFmrEF are similar to those of HFrEF^{4,20}.

Pharmacological interventions for heart failure with a preserved ejection fraction

Pharmacological treatment for HFrEF is well established. Beta-adrenoceptor blockers, RAAS inhibitors, and mineralocorticoid receptor blockers can reduce morbidity and mortality in HFrEF patients^{3,4}. Large-scale clinical trials have been instituted to identify effective drugs based on the risk factors for HFpEF^{21–23}, some of which are ongoing.

The beta (β)-adrenergic system is primarily located in the heart (β 1) and smooth muscle (β 2). When endogenous catecholamines activate the β 1 signaling cascade, the heart contracts more rapidly and forcefully²⁴. However, aberrant activation of the neurohormonal system worsens cardiac remodeling in HFrEF, implying that β -adrenergic blockage is beneficial²⁵. The β -blockers allow the heart to rest through a significant reduction in cardiac load. Various studies using β -blockers have been conducted using different patient enrollment criteria. However, the overall outcome of β -blocker trials revealed no survival benefit for HFpEF patients²². HFmrEF patients showed better survival than HFpEF patients with β -blocker treatment²⁰.

The RAAS controls both blood pressure and the circulating volume of blood by regulating vascular tone and sodium reabsorption, respectively²⁶. When the baroreceptors in the carotid sinus detect falling blood pressure or when the filtration flow rate in the kidney stimulates the macula densa, the homeostatic system in the body activates the RAAS²⁷. The main active component of the

RAAS is angiotensin II. Hence, the administration of an angiotensin-converting enzyme inhibitor (ACEi) to block the generation of the active form of angiotensin from angiotensin I, or the use of an angiotensin receptor blocker (ARB) or AT1 receptor blocker, can inhibit HF progression²⁸. Although ACEi/ARB agents are effective in HFrEF, clear evidence was not observed in HFpEF²².

Aldosterone, an endogenous mineralocorticoid hormone, accelerates myocardial remodeling, especially after an ischemic event. Mineralocorticoid receptor blockers alleviate myocardial fibrosis and extracellular matrix deposition²⁹. Pfeffer et al. reported that aldosterone antagonists could be beneficial for HFpEF in the Treatment of Preserved Cardiac Function Heart Failure with an Aldosterone Antagonist (TOPCAT) clinical trial in the United States³⁰. Beldhuis et al. warned that the prescription of an aldosterone antagonist should be avoided in HFpEF with decreased renal function³¹. However, cautious interpretation of the TOPCAT study is required. Other than in patients in the United States, aldosterone antagonists have shown no benefits. Furthermore, no positive outcome was found in the analysis of summary data from multiple facilities³⁰. More detailed and longterm studies involving different countries or races are needed.

The absence of clear evidence of benefit from β-blockers, RAAS inhibitors, or aldosterone antagonists prompted numerous clinical trials. Physicians chose medicines according to the risk factors for HFpEF to perform these clinical trials. Nitric oxide directly stimulates and relaxes vascular smooth muscle³². The use of a nitric oxide donor is the most useful strategy for the reduction in peripheral vascular resistance, i.e., afterload. Redfield et al. performed a trial using isosorbide mononitrate, an oral long-acting nitric oxide derivative, and found that exercise capacity was not improved²³. The Inorganic Nitrite Delivery to Improve Exercise Capacity in HFpEF (INDIE-HFpEF) clinical trial enrolled additional patients. Disappointingly, inorganic nitrite failed to improve exercise capacity³³. Further, sildenafil, a phosphodiesterase-5 inhibitor, was evaluated. Redfield et al., in the Phosphodiesterase-5 Inhibition to Improve Clinical Status and Exercise Capacity in Diastolic Heart Failure (RELAX) trial, reported that sildenafil treatment failed to improve exercise capacity or clinical status³⁴. Several trials with limited numbers of patients showed promising outcomes with the use of lipid-lowering statins³⁵. Confirmation would require a well-designed clinical trial with adequate power. In summary, clearly effective medication treatment for HFpEF remains undefined.

Recently, several therapeutics have been developed for HFrEF. The negative inotropic agent ivabradine was first approved in angina³⁶, and its indication has expanded to

chronic HFrEF^{37,38}. Ivabradine significantly reduced the risk of hospitalization in HF patients who were tolerant to beta-blockers and had a left ventricular ejection fraction of <35% and a heart rate that exceeded 70 beats per minute³⁸. A randomized, double-blinded, placebocontrolled trial to assess the effectiveness of ivabradine in HFpEF was completed (EDIFY trial). Unfortunately, the reduction in heart rate by ivabradine failed to improve the clinical outcome of HFpEF³⁹.

Angiotensin receptor neprilysin inhibitor (ARNI) is a combination drug that includes conventional angiotensin blockers and neprilysin inhibitors. Although neprilysin degrades several vasoactive peptides, the natriuretic peptide is a notable substrate. Compared with ACEi or ARB, ARNI reduced the rate of cardiovascular death or hospitalization by 20%⁴⁰. A randomized clinical trial on HFpEF (PARAGON-HF; NCT01920711) with ARNI has completed patient enrollment. The PARAGON-HF trial will determine the usefulness of the combination drug ARNI in HFpEF⁴¹.

It is noteworthy to observe the clinical outcome of the sodium-glucose co-transporter-2 (SGLT2) inhibitor trial. Originally, SGLT2 inhibitors were used for the treatment of type II DM. SGLT2 inhibitors interfere with sodium-glucose-linked co-transporter 2 in renal proximal convoluted tubules, which then reduces glucose reuptake⁴². A notable clinical outcome was observed in the EMPA-REG trial: cardiovascular events were reduced when the SGLT2 inhibitor was combined with conventional treatment regimens⁴³. New clinical trials such as DELIVER (NCT03619213), EMPEROR-Preserved (NCT03057951), and PRESERVED-HF (NCT03030235) are ongoing to determine the therapeutic potential of SGLT2 in HFpEF patients^{44,45}.

Research using an experimental rodent model for human heart failure with a preserved ejection fraction

Model validation: cardiac function measured on echocardiography

Clinically, HFpEF and diastolic HF are not synonymous⁴⁶. A significant proportion of HFpEF patients have normal diastolic function and normal cardiac geometry. However, there are more patients with diastolic dysfunction and ventricular hypertrophy than there are patients without diastolic dysfunction. For this reason, an experimental model of human HFpEF generally requires an investigation of diastolic dysfunction, ventricular hypertrophy, interstitial fibrosis, or exercise intolerance⁴⁷.

Echocardiography is widely utilized to assess diastolic function. Ventricular filling is divided into two phases. Ventricular relaxation in early diastole induces rapid and passive ventricular filling, whereas atrial contraction in late diastole adds a small amount of filling to the

ventricle¹. Doppler images record two prominent peaks, which are the E (ventricular relaxation) and A (atrial contraction) waves. As the E wave primarily reflects diastolic properties, a decreased E wave indicates diastolic dysfunction⁴⁸. A significant amount of residual blood in the atrium due to impaired diastolic function augments the atrial contribution to ventricular filling, demonstrated by an increased A wave⁴⁸. A decreased E/A ratio reflects diastolic dysfunction⁴⁸.

When the atrial pressure is further increased, the E wave is increased, but the E/A ratio paradoxically returns to normal. This phenomenon is called a pseudonormal filling pattern^{48,49}. For differential diagnosis, the direct measurement of ventricular movement is required. The tissue Doppler imaging mode directly records the mitral valve annulus movement. The two phases of mitral valve motion show waveforms similar to the E and A waves in tissue Doppler mode: the E' wave represents early diastole, and the A' wave represents atrial contraction⁴⁸. Because it directly represents ventricular function, the E' wave is further diminished in the pseudonormal period^{48,49}. Therefore, the E/E' ratio is markedly increased in diastolic dysfunction (Fig. 2). An increased E/E' ratio is one of the criteria used in the differential diagnosis⁴.

Model validation: heart failure symptoms with anatomical changes

A rodent model of HFpEF represents cardiac hypertrophy with or without interstitial fibrosis⁴⁷. Experimental cardiac hypertrophy is associated with increased cardiac mass and wall thickness. Simply weighing the intact heart or dividing its weight by total body weight indicates the hypertrophic phenotype⁵⁰. In addition to concentric hypertrophy, which is usual in HFpEF, a heart with normal intracardiac dimensions is commonly observed⁵¹. Hence, the measurement of wall thickness is mandatory

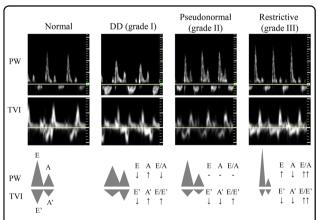


Fig. 2 Echocardiographic findings and schematic demonstrations according to the severity of diastolic dysfunction. DD, diastolic dysfunction; PW, pulse wave; TVI, tissue velocity image

to demonstrate hypertrophy. Many studies have used left ventricular free wall thickness to document hypertrophy. Moreover, cardiac perivascular fibrosis and interstitial fibrosis present as hypertrophy⁵². Because a major component of cardiac fibrosis is the presence of collagen in the extracellular matrix, Masson's trichrome or Picrosirius red staining can demonstrate its severity^{53,54}. Chronic diastolic dysfunction results in the retrograde build-up of blood, with predominant accumulation in the lung⁵⁵. As in the hypertrophied heart, the engorgement of tissue with blood results in weight gain. Increased lung weight is considered to represent blood congestion and implies diastolic HF⁵⁵. In addition to anatomical differences, a notable symptom of HF is limited exercise capacity²². To assess locomotor ability in rodents, a treadmill or rotarod performance test is used^{56,57}. Locomotor ability, however, can be affected by high brain function⁵⁸. For this reason, decreased exercise capacity is not definitive but only supportive of the HFpEF phenotype.

Animal model: comorbidity

To date, the treatment for HFpEF remains unclear ¹⁴. A detailed study using an appropriate animal model enables drug development. Big data analysis has identified several comorbid factors: DM, obesity, hypertension, hyperlipidemia, and female sex ^{12,13}. Several established rodent models of HFpEF are listed in Table 1. Because of convenience, rodents are widely used to assess cardiac function and HF phenotype.

Many research groups have used rats for hypertension research. Both systolic and diastolic pressures are sensitive to volume overload, the use of hypertensive agonists, and increased salt ingestion⁵⁹. The spontaneously hypertensive rat (SHR) strain is widely used for human hypertension research⁶⁰. Starting at <12 months of age, the SHR strain shows the gradual development of concentric hypertrophy with preserved contractility^{60,61}. However, the strain develops cardiac decompensation, with left ventricular dilatation and reduced ejection fraction after 18 months⁶², a phenomenon rarely observed in human HFpEF. The SHR model closely resembles human HFpEF, but the phenotypic onset time is relatively late. It is difficult to maintain a group of these rats for ~1 year, making the use of the SHR strain expensive and time consuming.

In addition, the deoxycorticosterone acetate (DOCA)-salt model is an acceptable alternative for hypertension research⁶⁰. Young adult Wistar rats (~8 weeks of age) undergo unilateral nephrectomy and are given drinking water with 1% sodium chloride. Starting 1 week after nephrectomy, the rats received subcutaneous injections of DOCA (25 mg in 0.4 mL of dimethylformamide every 4 days) for 28 days. The rats develop concentric hypertrophy, fibrosis, and hypertension⁶³. Moreover, the DOCA-salt model can be used in mice, but blood pressure

Table 1 Rodent models for human HFpEF research

Species	Comorbidity	Strain	Manipulation	Phenotypes
Rat	Hypertension	SHR		Hypertension and hypertrophy in 12 months HFpEF in 18 months
	Hypertension	Wistar	DOCA salt Unilateral nephrectomy 1% NaCl drinking water	Hypertrophy Severe hypertension
	Hypertension	DSS	4–8% NaCl chow	Severe hypertension Diastolic heart failure
Mouse	Hypertension	C57BL/6	DOCA salt Unilateral nephrectomy 1% NaCl drinking water	Mild hypertension Hypertrophy
	Hypertension	C57BL/6	Aldosterone Unilateral nephrectomy 1% NaCl drinking water	Hypertrophy Fibrosis Diastolic dysfunction
	Hypertension	C57BL/6	Pressure overload TAC AoB	Hypertrophy and fibrosis Diastolic dysfunction → systolic dysfunction
	Diabetes mellitus	db/db		Insulin resistance Hypertrophy Diastolic dysfunction
	Obesity	ob/ob		Hypertrophy Diastolic dysfunction
	Obesity + hypertension	C57BL/6	High-fat diet + L-NAME	Hypertrophy Diastolic dysfunction Pulmonary congestion
	Aging	SAMP		Diastolic dysfunction

The black arrow indicates the transition

AoB aortic banding, DOCA deoxycorticosterone acetate, DSS Dahl salt-sensitive, L-NAME L-NG-nitroarginine methyl ester, SAMP senescence-accelerated mouse prone 8, SHR spontaneous hypertensive rat, TAC transverse aortic constriction

elevation is limited⁶⁴. Thus, it is not clear whether the DOCA-salt model truly represents hypertension-induced HFpEF.

The Dahl salt-sensitive (DSS) rat with high-salt chow is considered a noninvasive and short-term HFpEF model. DSS rats fed 4–8% salty chow for 2 months develop typical HFpEF phenotypes^{65,66}. The heart shows concentric hypertrophy, interstitial fibrosis, and diastolic dysfunction. The DSS with a high-salt diet model shows prolonged action potential, resulting in polymorphic ventricular tachyarrhythmia⁶⁵. The DSS model, however, develops severe hypertension (>175 mmHg), which is uncommon in humans⁶⁷. To accurately reflect human HFpEF, the DSS model would require the use of an antihypertensive drug.

Although rats are acceptable models for research on HFpEF, mice are easier to work with than rats. There is no significant change in blood pressure in mice compared with rats. Therefore, most acceptable hypertension models using rats are not comparable to those using mice. The ability to perform large-scale experiments with genetically modified animals and access to an animal bank is easier to accomplish with mice than with rats.

The DOCA model is applicable to both mice and rats⁶⁴. DOCA salt combined with 1% sodium chloride causes hypertension in rats. Blood pressure is mildly increased or unchanged in mice, although concentric hypertrophy and interstitial fibrosis are observed. Unilateral nephrectomy in mice treated with DOCA salt leads to diastolic dysfunction. Unilateral nephrectomy was performed on C57BL/6 mice, followed by implantation of a DOCA pellet releasing 0.7 mg/day for 3 weeks; in addition, the mice were given 1% sodium chloride drinking water⁶⁸. The DOCA model showed no difference in heart rate or ejection fraction. DOCA reduced exercise capacity, but pulmonary congestion was uncommon⁶⁹.

In addition, in place of DOCA, infusion with d-aldosterone induces HFpEF^{70} . Although applicable in

rats, mice are considered more suitable for this model. C57BL/6 mice underwent implantation of d-aldosterone osmotic pumps (0.30 µg/h) after unilateral nephrectomy and were given 1% sodium chloride drinking water for 30 days⁷¹. Both systolic and diastolic pressure were elevated, but the heart rate was not changed. Moreover, systolic function was preserved. The mice developed concentric hypertrophy, diastolic dysfunction, pulmonary edema, and locomotor impairment⁷². Humans with HFpEF show high levels of aldosterone. The elevation of serum creatinine and albuminuria was detected in aldosterone model mice, similar to the findings in humans with HFpEF. Molecular events in the aldosterone infusion model were comparable to those in humans with HFpEF⁴. Hence, the aldosterone model represents both the clinical phenotype of HFpEF and the molecular events⁷³.

Partial ligation or constriction of a large artery induces an abrupt and severe increase in afterload⁷⁴. Constriction can be applied at the aortic root, transverse aorta, descending aorta, or even abdominal aorta⁷⁵. The aortic root and transverse aorta are frequently selected. The constriction of a large artery generates a steep increase in blood pressure just proximal to the point of manipulation⁷⁶. However, the segment distal to the constriction maintains normal blood pressure. Hence, the arterial constriction model resembles hypertension-based HFpEF, but the effect is limited to the heart. Aortic root constriction or aortic banding (AoB) usually results in eccentric hypertrophy due to intracardiac volume overload with excessive pulmonary edema⁵⁵. Massive interstitial fibrosis accompanies hypertrophy, which interferes with ventricular movement. Although mice with AoB transiently maintain physical hemodynamics, undergo aberrant decompensatory remodeling⁷⁷. Prolonged AoB exacerbates systolic dysfunction, ultimately leading to HFrEF. The transition from HFpEF to HFrEF is rarely observed in humans⁴, and the findings after prolonged AoB do not correspond to clinical outcomes. Transverse aortic constriction (TAC) represents partial ligation between the brachiocephalic artery and left common carotid artery. TAC induces less severe vascular resistance and intraventricular volume overload than AoB due to the preservation of the brachiocephalic artery. TAC induces concentric hypertrophy as well as eccentric hypertrophy with interstitial fibrosis. Moreover, prolonged TAC progresses to systolic heart failure. Hence, diastolic dysfunction induced by pressure overload fails to elicit human HFpEF⁷⁸.

Since obesity and DM are notable comorbidities in HFpEF⁷⁹, genetically modified db/db^{80} or ob/ob^{81} mice are widely used for cardiometabolic research. The db/db mouse has mutant leptin receptors that induce aberrant obesity. This leads to spontaneous insulin resistance, which is comparable to that in human type 2 DM.

Without further administration of agonists, the *db/db* mouse shows diastolic dysfunction with concentric hypertrophy. The left ventricular ejection fraction remains in the normal range. Locomotor ability is associated with the severity of diastolic dysfunction⁸². The *ob/ob* mouse lacks leptins. Because of polyphagia, obesity and type 2 DM develop spontaneously. Disease progression in *ob/ob* mice resembles that in human type 2 DM. However, the use of the *ob/ob* mouse has some limitations. Leptin governs appetite control in addition to having a cardio-protective role. A loss of leptin impairs contraction-relaxation coupling in cardiomyocytes. Furthermore, leptin deficiency in human obesity is rare, meaning that the *ob/ob* mouse model is not appropriate for the study of human HFpEF⁸³.

A noninvasive and nongenetically modified model of HFpEF was recently reported⁸⁴. Schiattarella et al. considered common comorbidities in human HFpEF and created a model that combined hyperlipidemia and hypertension. They administered a high-fat diet with a nitric oxide synthase inhibitor ad libitum. After 5 weeks, they observed significant impairment of diastolic function, with pulmonary congestion and exercise intolerance. At 15 weeks, significant signs and symptoms of HFpEF had developed⁸⁴. This model mimics human pathophysiology, implying its suitability for use in research.

In addition, age is an important risk factor for HFpEF¹². The relative life span in rodents is even shorter than in humans, but "old" age in mice is only 18 months. It is difficult to maintain stable conditions during long-term rearing in a research facility. Therefore, the inbred senescence-accelerated mouse prone (SAMP) strain is useful to reduce the time course of aging⁸⁵. Spontaneous acceleration of aging in the SAMP strain phenotype begins as early as postnatal 6 months. The SAMP8 strain shows cardiac hypertrophy with fibrosis as well as diastolic dysfunction. Systolic function is well preserved. The SAMP8 mouse carries more comorbidities associated with HFpEF than do other mice⁸⁶. A detailed study using SAMP8 mice is required in the future.

Future perspective and conclusion

Mortality and morbidity in HFpEF have continuously increased for more than two decades^{3,4}. Furthermore, approximately half of newly diagnosed HF patients have preserved ejection fraction. The social cost of HFpEF has greatly increased, but no verified treatment regimen is available⁴. This brief review discussed the clinical trials and rodent models used for human HFpEF research. Disappointingly, several treatments that proved effective for HFrEF failed to improve the survival rate or even quality of life in HFpEF patients⁹. A limited number of Asian studies described the therapeutic potential of lipid-lowering statins, but the power of a randomized,

multirace trial is required for verification³⁵. In the TOP-CAT study, the efficacy of spironolactone was unclear³⁰. This mineralocorticoid receptor blocker reduced mortality only in patients enrolled in the United States, implying that genetic background could be a confounding factor in HFpEF. Detailed and homogeneous grouping of HFpEF patients could resolve this question.

Rats and mice have been used for the study of human HFpEF. Most rodent HFpEF models show concentric or eccentric hypertrophy, interstitial fibrosis, and diastolic dysfunction. For the assessment of underlying molecular events, vascular inflammation, endothelial dysfunction, or microvascular uncoupling was suggested⁴⁷. Cumulative data revealed that pharmacological intervention for cardiac hypertrophy ameliorates cardiac fibrosis accompanying the ventricular stiffness that results in diastolic dysfunction⁸⁷. Histone deacetylase (HDAC) inhibitors are considered promising for the treatment of cardiac hypertrophy and therefore diastolic dysfunction 50,53,88-90. In a rodent model, cardiac hypertrophy was completely blocked when HDAC inhibitors were concurrently administered 50,88-90. Moreover, our group showed that HDAC inhibitors could induce the regression of preexisting hypertrophy and fibrosis⁹¹. HDAC inhibitors should be considered novel therapeutic agents for HFpEF. HDAC inhibitors were first developed as an anticancer agent, which suggests that using HDAC inhibitors as HFpEF modulators may have cytotoxic side effects. In fact, notable cytotoxicity-associated adverse reactions of HDAC inhibitors such as SAHA include hair loss, diarrhea, peripheral numbness, or tingling sensation⁹². Furthermore, our group demonstrated vascular calcification as a potential side effect of HDAC inhibitors⁹³. Since DM is one of the notable comorbidities of HFpEF, using HDAC inhibitors to treat vulnerable patients should be avoided. In addition, immune modulators may be useful in HFpEF. Hulsmans et al. reported the contribution of macrophages to diastolic dysfunction⁹⁴. In fact, a clinical trial using interleukin-1B, the Interleukin-1 Blockade in Heart Failure with Preserved Ejection Fraction (HFpEF): A Randomized Placebo-controlled Double-Blinded Study (D-HART2), has been completed and the results have been reported⁹⁵. While systemic inflammation was significantly improved, survival benefits were not observed. The power of the D-HART2, however, was relatively small, which was a weak point of the study⁹⁵. Thus, another immune modulator should be considered. Endothelial dysfunction failed to produce nitric oxide or improvement in vascular elasticity. Restoration of either endothelial nitric oxide synthase activity or cGMP activity was attempted. Organic nitrate donors and inorganic nitrate suppliers showed no benefit for exercise intolerance^{23,33}.

Physicians have focused on symptom control due to the lack of therapeutic agents^{3,4}. Numerous clinical trials have been instituted, but these require long-term follow-up. To correctly understand HFpEF, an animal study of HFpEF should be conducted in parallel with a human study. Promising outcomes from animal research should be considered when developing treatment regimens in humans. Combined comorbidity is common in humans with HFpEF, making the development of a rodent model difficult. It is noteworthy that a rodent model with more than two comorbidities in a single mouse has been generated⁸⁴. Nonetheless, understanding HFpEF in a rodent model could aid in the understanding of human HFpEF.

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Conflict of interest

The authors clearly declare that they have no conflict of interest.

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References

- Bombardini, T. Myocardial contractility in the echo lab: molecular, cellular and pathophysiological basis. Cardiovasc Ultrasound 3, 27 (2005).
- Fukuta, H. & Little, W. C. The cardiac cycle and the physiologic basis of left ventricular contraction, ejection, relaxation, and filling. *Heart Fail Clin.* 4, 1–11 (2008).
- Yancy, C. W. et al. 2017 ACC/AHA/HFSA focused update of the 2013 ACCF/ AHA guideline for the management of heart failure: a report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. Circulation 136, e137–e161 (2017)
- 4. Ponikowski, P. et al. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC). Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. Eur. J. Heart Fail. 18, 891–975 (2016).
- Gerber, Y. et al. Mortality associated with heart failure after myocardial infarction: a contemporary community perspective. Circ. Heart Fail. 9, e002460 (2016).
- Burchfield, J. S., Xie, M. & Hill, J. A. Pathological ventricular remodeling: mechanisms: part 1 of 2. Circulation 128, 388–400 (2013).
- Shearer, F., Lang, C. C. & Struthers, A. D. Renin-angiotensin-aldosterone system inhibitors in heart failure. Clin. Pharm. Ther. 94, 459–467 (2013).
- Borlaug, B. A. & Redfield, M. M. Diastolic and systolic heart failure are distinct phenotypes within the heart failure spectrum. *Circulation* 123, 2006–2013 (2011).
- Dunlay, S. M., Roger, V. L. & Redfield, M. M. Epidemiology of heart failure with preserved ejection fraction. *Nat. Rev. Cardiol.* 14, 591–602 (2017).
- Mohammed, S. F. et al. Comorbidity and ventricular and vascular structure and function in heart failure with preserved ejection fraction: a community-based study. Circ. Heart Fail. 5, 710–719 (2012).
- van Heerebeek, L. & Paulus, W. J. Understanding heart failure with preserved ejection fraction: where are we today? Neth. Heart J. 24, 227–236 (2016).

- 12. Shah, S. J. et al. Phenomapping for novel classification of heart failure with preserved ejection fraction. *Circulation* **131**, 269–279 (2015).
- Kao, D. P. et al. Characterization of subgroups of heart failure patients with preserved ejection fraction with possible implications for prognosis and treatment response. Eur. J. Heart Fail. 17, 925–935 (2015).
- 14. Tibrewala, A. & Yancy, C. W. Heart failure with preserved ejection fraction in women. *Heart Fail Clin.* **15**, 9–18 (2019).
- Burlew, B. S. Diastolic dysfunction in the elderly-the interstitial issue. Am. J. Geriatr. Cardiol. 13, 29–38 (2004).
- Lam, C. S. et al. Cardiac structure and ventricular-vascular function in persons with heart failure and preserved ejection fraction from Olmsted County, Minnesota. Circulation 115, 1982–1990 (2007).
- Luscher, T. F. Heart failure subgroups: HFrEF, HFmrEF, and HFpEF with or without mitral regurgitation. Eur. Heart J. 39, 1–4 (2018).
- 18. Lopatin, Y. Heart failure with mid-range ejection fraction and how to treat it. *Card. Fail Rev.* **4**, 9–13 (2018).
- Tsuji, K. et al. Characterization of heart failure patients with mid-range left ventricular ejection fraction-a report from the CHART-2 Study. Eur. J. Heart Fail. 19. 1258–1269 (2017).
- Cleland, J. G. F. et al. Beta-blockers for heart failure with reduced, mid-range, and preserved ejection fraction: an individual patient-level analysis of doubleblind randomized trials. Eur. Heart J. 39, 26–35 (2018).
- Campbell, R. T. et al. What have we learned about patients with heart failure and preserved ejection fraction from DIG-PEF, CHARM-preserved, and I-PRE-SERVE? J. Am. Coll. Cardiol. 60, 2349–2356 (2012).
- Redfield, M. M. Heart failure with preserved ejection fraction. N. Engl. J. Med. 375, 1868–1877 (2016).
- Redfield, M. M. et al. Isosorbide mononitrate in heart failure with preserved ejection fraction. N. Engl. J. Med. 373, 2314–2324 (2015).
- Johnson, M. The beta-adrenoceptor. Am. J. Respir. Crit. Care Med. 158, S146–S153 (1998).
- Bristow, M. R. beta-adrenergic receptor blockade in chronic heart failure. Circulation 101, 558–569 (2000).
- Sancho, J., Re, R., Burton, J., Barger, A. C. & Haber, E. The role of the reninangiotensin-aldosterone system in cardiovascular homeostasis in normal human subjects. *Circulation* 53, 400–405 (1976).
- Patel, S., Rauf, A., Khan, H. & Abu-Izneid, T. Renin-angiotensin-aldosterone (RAAS): the ubiquitous system for homeostasis and pathologies. *Biomed. Pharmacother.* 94, 317–325 (2017).
- 28. Unger, T. & Li, J. The role of the renin-angiotensin-aldosterone system in heart failure. *J. Renin Angiotensin Aldosterone Syst.* **5**(Suppl 1), S7–S10 (2004).
- Zannad, F. Aldosterone and heart failure. Eur. Heart J. 16(Suppl N), 98–102 (1995)
- Pfeffer, M. A. et al. Regional variation in patients and outcomes in the Treatment of Preserved Cardiac Function Heart Failure With an Aldosterone Antagonist (TOPCAT) trial. Circulation 131, 34–42 (2015).
- Beldhuis, I. E. et al. Efficacy and safety of spironolactone in patients with HFpEF and chronic kidney disease. JACC Heart Fail. 7, 25–32 (2019).
- Messina, A. G. et al. The effect of nitrous oxide on left ventricular pump performance and contractility in patients with coronary artery disease: effect of preoperative ejection fraction. *Anesth. Analg.* 77, 954–962 (1993).
- Borlaug, B. A. et al. Effect of inorganic nitrite vs placebo on exercise capacity among patients with heart failure with preserved ejection fraction: the INDIE-HFpEF randomized clinical trial. JAMA 320, 1764–1773 (2018).
- Redfield, M. M. et al. Effect of phosphodiesterase-5 inhibition on exercise capacity and clinical status in heart failure with preserved ejection fraction: a randomized clinical trial. JAMA 309, 1268–1277 (2013).
- Marume, K et al. Effect of statins on mortality in heart failure with preserved ejection fraction without coronary artery disease—report from the JASPER study. Circulation J. 83, 357–367 (2019).
- Fox, K. et al. Ivabradine for patients with stable coronary artery disease and leftventricular systolic dysfunction (BEAUTIFUL): a randomised, double-blind, placebo-controlled trial. *Lancet* 372, 807–816 (2008).
- Koruth, J. S., Lala, A., Pinney, S., Reddy, V. Y. & Dukkipati, S. R. The clinical use of ivabradine. J. Am. Coll. Cardiol. 70, 1777–1784 (2017).
- Swedberg, K. et al. Ivabradine and outcomes in chronic heart failure (SHIFT): a randomised placebo-controlled study. Lancet 376, 875–885 (2010).
- Komajda, M. et al. Effect of ivabradine in patients with heart failure with preserved ejection fraction: the EDIFY randomized placebo-controlled trial. Eur. J. Heart Fail. 19, 1495–1503 (2017).

- McMurray, J. J. et al. Angiotensin-neprilysin inhibition versus enalapril in heart failure. N. Engl. J. Med. 371, 993–1004 (2014).
- Solomon, S. D. et al. Angiotensin receptor neprilysin inhibition in heart failure with preserved ejection fraction: rationale and design of the PARAGON-HF trial. JACC Heart Fail. 5, 471–482 (2017).
- 42. Vallon, V. The mechanisms and therapeutic potential of SGLT2 inhibitors in diabetes mellitus. *Annu Rev. Med.* **66**, 255–270 (2015).
- Zinman, B. et al. Empagliflozin, cardiovascular outcomes, and mortality in type 2 diabetes. N. Enal. J. Med. 373, 2117–2128 (2015).
- McMurray, J. J. V. et al. A trial to evaluate the effect of the sodium-glucose cotransporter 2 inhibitor dapagliflozin on morbidity and mortality in patients with heart failure and reduced left ventricular ejection fraction (DAPA-HF). Eur. J. Heart Fail. 21, 665–675 (2019).
- Elgendy, I. Y., Mahtta, D. & Pepine, C. J. Medical therapy for heart failure caused by ischemic heart disease. *Circulation Res.* 124, 1520–1535 (2019).
- Borlaug, B. A. The pathophysiology of heart failure with preserved ejection fraction. Nat. Rev. Cardiol. 11, 507 (2014).
- Valero-Munoz, M., Backman, W. & Sam, F. Murine models of heart failure with preserved ejection fraction: a "fishing expedition". JACC Basic Transl. Sci. 2, 770–789 (2017).
- Nagueh, S. F. et al. Recommendations for the evaluation of left ventricular diastolic function by echocardiography: an update from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. J. Am. Soc. Echocardiogr. 29, 277–314 (2016).
- Mottram, P. M. & Marwick, T. H. Assessment of diastolic function: what the general cardiologist needs to know. *Heart* 91, 681–695 (2005).
- Eom, G. H. et al. Casein kinase-2alpha1 induces hypertrophic response by phosphorylation of histone deacetylase 2 S394 and its activation in the heart. Circulation 123, 2392–2403 (2011).
- Heinzel, F. R., Hohendanner, F., Jin, G., Sedej, S. & Edelmann, F. Myocardial hypertrophy and its role in heart failure with preserved ejection fraction. *J. Appl Physiol.* 119, 1233–1242 (2015).
- Ho, C. Y. et al. Myocardial fibrosis as an early manifestation of hypertrophic cardiomyopathy. N. Engl. J. Med. 363, 552–563 (2010).
- Yoon, S. et al. PP2A negatively regulates the hypertrophic response by dephosphorylating HDAC2 S394 in the heart. Exp. Mol. Med. 50, 83 (2018).
- Hadi, A. M. et al. Rapid quantification of myocardial fibrosis: a new macrobased automated analysis. Cell Oncol. (Dordr.). 34, 343–354 (2011).
- Chen, Y. et al. Left ventricular failure produces profound lung remodeling and pulmonary hypertension in mice: heart failure causes severe lung disease. Hypertension 59, 1170–1178 (2012).
- Bull, M. et al. Alternative splicing of titin restores diastolic function in an HFpEFlike genetic murine model Ttn^{DLAjan}. Circulation Res. 119, 764–772 (2016).
- Roos, K. P. et al. Hypertrophy and heart failure in mice overexpressing the cardiac sodium-calcium exchanger. J. Card. Fail. 13, 318–329 (2007).
- 58. Deacon, R. M. Measuring motor coordination in mice. J. Vis. Exp. e2609 (2013).
- Doggrell, S. A. & Brown, L. Rat models of hypertension, cardiac hypertrophy and failure. *Cardiovascular Res.* 39, 89–105 (1998).
- Horgan, S., Watson, C., Glezeva, N. & Baugh, J. Murine models of diastolic dysfunction and heart failure with preserved ejection fraction. J. Card. Fail 20, 984–995 (2014).
- Pfeffer, J. M., Pfeffer, M. A., Fishbein, M. C. & Frohlich, E. D. Cardiac function and morphology with aging in the spontaneously hypertensive rat. *Am. J. Physiol.* 237, H461–H468 (1979).
- Damatto, R. L. et al. Heart failure-induced skeletal myopathy in spontaneously hypertensive rats. Int J. Cardiol. 167, 698–703 (2013).
- Matsumura, Y. et al. Enhanced blood pressure sensitivity to DOCA-salt treatment in endothelin ET(B) receptor-deficient rats. Br. J. Pharmacol. 129, 1060–1062 (2000).
- Mohammed, S. F. et al. Mineralocorticoid accelerates transition to heart failure with preserved ejection fraction via "nongenomic effects". Circulation 122, 370–378 (2010).
- Cho, J. H. et al. Delayed repolarization underlies ventricular arrhythmias in rats with heart failure and preserved ejection fraction. *Circulation* 136, 2037–2050 (2017)
- Jeong, M. Y. et al. Histone deacetylase activity governs diastolic dysfunction through a nongenomic mechanism. Sci. Transl. Med. 10, eaao0144 (2018).
- Doi, R. et al. Development of different phenotypes of hypertensive heart failure: systolic versus diastolic failure in Dahl salt-sensitive rats. J. Hypertens. 18, 111–120 (2000).

- Grobe, J. L. et al. Angiotensinergic signaling in the brain mediates metabolic effects of deoxycorticosterone (DOCA)-salt in C57 mice. *Hypertension* 57, 600–607 (2011).
- Bowen, T. S. et al. High-intensity interval training prevents oxidant-mediated diaphragm muscle weakness in hypertensive mice. FASEB J. 31, 60–71 (2017).
- Sam, F. et al. Mice lacking osteopontin exhibit increased left ventricular dilation and reduced fibrosis after aldosterone infusion. Am. J. Hypertens. 17, 188–193 (2004)
- Tanaka, K. et al. Effects of adiponectin on calcium-handling proteins in heart failure with preserved ejection fraction. Circ. Heart Fail. 7, 976–985 (2014).
- Wilson, R. M., De Silva, D. S., Sato, K., Izumiya, Y. & Sam, F. Effects of fixed-dose isosorbide dinitrate/hydralazine on diastolic function and exercise capacity in hypertension-induced diastolic heart failure. *Hypertension* 54, 583–590 (2009).
- Mohammed, S. F. et al. Coronary microvascular rarefaction and myocardial fibrosis in heart failure with preserved ejection fraction. *Circulation* 131, 550–559 (2015).
- Beetz, N. et al. Ablation of biglycan attenuates cardiac hypertrophy and fibrosis after left ventricular pressure overload. J. Mol. Cell. Cardiol. 101, 145–155 (2016).
- Merino, D. et al. Experimental modelling of cardiac pressure overload hypertrophy: Modified technique for precise, reproducible, safe and easy aortic arch banding-debanding in mice. Sci. Rep. 8, 3167 (2018).
- Mohammed, S. F. et al. Variable phenotype in murine transverse aortic constriction. Cardiovasc Pathol. 21, 188–198 (2012).
- Boluyt, M. O. et al. Heart failure after long-term supravalvular aortic constriction in rats. Am. J. Hypertens. 18, 202–212 (2005).
- Dunlay, S. M., Roger, V. L., Weston, S. A., Jiang, R. & Redfield, M. M. Longitudinal changes in ejection fraction in heart failure patients with preserved and reduced ejection fraction. *Circ. Heart Fail.* 5, 720–726 (2012).
- Lavie, C. J., Milani, R. V. & Ventura, H. O. Obesity and cardiovascular disease: risk factor, paradox, and impact of weight loss. J. Am. Coll. Cardiol. 53, 1925–1932 (2009).
- Chen, H. et al. Evidence that the diabetes gene encodes the leptin receptor: identification of a mutation in the leptin receptor gene in db/db mice. Cell 84, 491–495 (1996).
- 81. Edvell, A. & Lindstrom, P. Initiation of increased pancreatic islet growth in young normoglycemic mice (Umeå +/?). Endocrinology 140, 778–783 (1999).

- Ostler, J. E. et al. Effects of insulin resistance on skeletal muscle growth and exercise capacity in type 2 diabetic mouse models. Am. J. Physiol. Endocrinol. Metab. 306, E592–E605 (2014).
- Clement, K. Genetics of human obesity. C. R. Biol. 329, 608–622 (2006). discussion 653-605.
- Schiattarella, G. G. et al. Nitrosative stress drives heart failure with preserved ejection fraction. Nature 568, 351–356 (2019).
- Zhang, L. et al. Changes of brain activity in the aged SAMP mouse. Biogerontology 8, 81–88 (2007).
- Reed, A. L. et al. Diastolic dysfunction is associated with cardiac fibrosis in the senescence-accelerated mouse. Am. J. Physiol. Heart Circulatory Physiol. 301, H824–H831 (2011).
- 87. Schiattarella, G. G. & Hill, J. A. Inhibition of hypertrophy is a good therapeutic strategy in ventricular pressure overload. *Circulation* **131**, 1435–1447 (2015).
- Eom, G. H. et al. Regulation of acetylation of histone deacetylase 2 by p300/ CBP-associated factor/histone deacetylase 5 in the development of cardiac hypertrophy. Circulation Res. 114, 1133–1143 (2014).
- Yoon, S. et al. Inhibition of heat shock protein 70 blocks the development of cardiac hypertrophy by modulating the phosphorylation of histone deacetylase 2. Cardiovasc. Res. cvy317–cvy317 (2018). [Epub ahead of print].
- Kong, Y. et al. Suppression of class I and II histone deacetylases blunts pressure-overload cardiac hypertrophy. Circulation 113, 2579–2588 (2006).
- Kee, H. J. et al. Inhibition of histone deacetylation blocks cardiac hypertrophy induced by angiotensin II infusion and aortic banding. *Circulation* 113, 51–59 (2006).
- Subramanian, S., Bates, S. E., Wright, J. J., Espinoza-Delgado, I. & Piekarz, R. L. Clinical toxicities of histone deacetylase inhibitors. *Pharmaceuticals* 3, 2751–2767 (2010).
- Kwon, D. H. et al. MDM2 E3 ligase-mediated ubiquitination and degradation of HDAC1 in vascular calcification. *Nat. Commun.* 7, 10492 (2016).
- Hulsmans, M. et al. Cardiac macrophages promote diastolic dysfunction. J. Exp. Med. 215, 423–440 (2018).
- Van Tassell, B. W. et al. Interleukin-1 blockade in heart failure with preserved ejection fraction: rationale and design of the Diastolic Heart Failure Anakinra Response Trial 2 (D-HART2). Clin. Cardiol. 40, 626–632 (2017).