




ORIGINAL ARTICLE

LIN28A gene polymorphisms modify neuroblastoma susceptibility: A four-centre case-control study

Rui-Xi Hua^{1,2} | Zhenjian Zhuo¹ | Lili Ge³ | Jinhong Zhu^{1,4}  | Li Yuan¹ | Chongfen Chen³ | Jing Liu³ | Jiwen Cheng⁵ | Haixia Zhou⁶ | Jiao Zhang⁷ | Huimin Xia¹  | Xianwei Zhang⁸ | Jing He¹ 

¹Department of Pediatric Surgery, Guangzhou Institute of Pediatrics, Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou, China

²Department of Oncology, The First Affiliated Hospital of Sun Yat-sen University, Guangzhou, China

³Henan Provincial Key Laboratory of Children's Genetics and Metabolic Diseases, Children's Hospital Affiliated to Zhengzhou University, Henan Children's Hospital, Zhengzhou Children's Hospital, Zhengzhou, China

⁴Department of Clinical Laboratory, Biobank, Harbin Medical University Cancer Hospital, Harbin, China

⁵Department of Pediatric Surgery, The Second Affiliated Hospital of Xi'an Jiaotong University, Xi'an, China

⁶Department of Hematology, The Second Affiliated Hospital and Yuying Children's Hospital of Wenzhou Medical University, Wenzhou, China

⁷Department of Pediatric Surgery, The First Affiliated Hospital of Zhengzhou University, Zhengzhou, China

⁸Department of Pediatric Oncologic Surgery, Children's Hospital Affiliated to Zhengzhou University, Henan Children's Hospital, Zhengzhou Children's Hospital, Zhengzhou, China

Correspondence

Jing He, Department of Pediatric Surgery, Guangzhou Institute of Pediatrics, Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, 9 Jinsui Road, Guangzhou 510623, Guangdong, China.
Email: hejing198374@gmail.com

Xianwei Zhang, Department of Pediatric Oncologic Surgery, Children's Hospital Affiliated to Zhengzhou University, Henan Children's Hospital, Zhengzhou Children's Hospital, 33 Longhu Waihuan East Road, Zhengzhou 450018, Henan, China.
Email: zhangxw956658@163.com.

Funding information

Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Grant/Award Number: 2019B030301004; Pearl River S&T Nova Program of Guangzhou, Grant/Award Number: 201710010086

Abstract

Neuroblastoma ranks the most common seen solid tumour in childhood. Overexpression of *LIN28A* gene has been linked to the development of multiple human malignancies, but the relationship between *LIN28A* single nucleotide polymorphisms (SNPs) and neuroblastoma susceptibility is still under debate. Herein, we evaluated the correlation of four potentially functional *LIN28A* SNPs (rs3811464 G>A, rs3811463 T>C, rs34787247 G>A, and rs11247957 G>A) and neuroblastoma susceptibility in 505 neuroblastoma patients and 1070 controls from four independent hospitals in China. The correlation strengths were determined by using odds ratios (ORs) and corresponding 95% confidence intervals (CIs). Among these SNPs, rs34787247 G>A exhibited a significant association with increased susceptibility in neuroblastoma (GA vs GG: adjusted OR = 1.30, 95% CI = 1.03-1.64; AA vs GG: adjusted OR = 2.51, 95% CI = 1.36-4.64, AA/GA vs GG: adjusted OR = 1.42, 95% CI = 1.12-1.80, AA vs GG/GA: adjusted OR = 2.39, 95% CI = 1.29-4.42). Furthermore, the combined analysis of risk genotypes revealed that subjects carrying three risk genotypes (adjusted OR = 1.64, 95% CI = 1.02-2.63) are more inclined to develop neuroblastoma than those without risk genotype, and so do carriers of 1-4 risk

Rui-Xi Hua, Zhenjian Zhuo, and Lili Ge contributed equally to this work.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2019 The Authors. Journal of Cellular and Molecular Medicine published by Foundation for Cellular and Molecular Medicine and John Wiley & Sons Ltd.

genotypes (adjusted OR = 1.26, 95% CI = 1.01-1.56). Stratification analysis further revealed risk effect of rs3811464 G>A, rs34787247 G>A and 1-4 risk genotypes in some subgroups. Haplotype analysis of these four SNPs yields two haplotypes significantly correlated with increased neuroblastoma susceptibility. Overall, our finding indicated that *LIN28A* SNPs, especially rs34787247 G>A, may increase neuroblastoma risk.

KEYWORDS

case-control study, *LIN28A*, neuroblastoma, polymorphism, risk

1 | INTRODUCTION

Neuroblastoma is a solid tumour that predominantly affects infants and young children.¹ It mainly develops from neural crest progenitor cells.^{2,3} Neuroblastoma constitutes about 8%-10% of all paediatric cancers, but disproportionately causes 12%-15% cancer death in children.^{2,4,5} Neuroblastoma displays considerable clinical heterogeneity, ranging from spontaneous recovery to therapy-refractory progression.^{6,7} The distinct difference in survival rate among subgroups was another reflection of such heterogeneity.^{8,9} Patients with neuroblastoma can be classified into three risk groups: low risk, intermediate risk and high risk by using some clinical and biological prognostic factors. Patients with a non-high risk (low and intermediate risk) of neuroblastoma have a long-term survival rate up to of 90% or above, while those with high risk of neuroblastoma only achieve a survival rate as low as 40%.^{10,11}

In recent decades, remarkable advancement has been achieved in comprehending the fundamental aetiology of neuroblastoma.^{12,13} Children's and pregnant women's exposures to many environmental factors were reported to predispose to neuroblastoma, yet the causality could not be finally confirmed.^{14,15} On the other hand, genetic alterations have been shown to be linked to neuroblastoma susceptibility. Mutations in genes *ALK*^{16,17} and *PHOX2B*¹⁸ are frequently observed in hereditary neuroblastoma. Moreover, a number of single nucleotide polymorphisms (SNPs) in association with neuroblastoma predisposition have been identified in genes recently, including *TP53*,¹⁹ *LIN28B*,²⁰ *HACE1*,²⁰ *LMO1*,²¹ *BARD1*,²² *NEFL*²³ and *CDKN1B*.²⁴ Moreover, a fine-mapping analysis of *BARD1* locus (2q35) also identified two independent genome-wide neuroblastoma-associated loci.²⁵ However, the present identified genetic variations could not fully account for the carcinogenesis of neuroblastoma. We are still on the discovery journey of unveiling more causative genetic alterations hidden in the bush.

LIN28 is a conserved RNA-binding protein that plays a significant part in the regulation of cell proliferation, glucose metabolism and pluripotency through interacting with miRNAs.^{26,27} The mammalian genome, *LIN28* gene, encodes two *Lin28* paralogs, *Lin28A* and *Lin28B*.²⁸ *LIN28A* inhibits the maturation process of *let-7* microRNAs and thereby enhances the translation of *let-7* target mRNAs.^{29,30} Briefly, cytoplasmic *LIN28A* induces *pre-let-7* oligo-uridylation

through a TUTase-dependent mechanism.³¹ Such poly-uridylation leads to *pre-let-7* instability and eventually reduces the amount of mature *let-7*.³² *let-7* is a tumour suppressor. Its downregulation promotes tumorigenesis and correlates with poor prognosis.³³ By binding to a variety of mRNA targets, *LIN28A* also has additional functions except for suppressing *let-7* maturation.^{28,34}

Over-activation of the *LIN28A* gene has been observed in various human cancers.^{35,36} The mechanism of the *LIN28A*-mediated tumorigenesis has been extensively investigated. However, the implications of *LIN28A* gene SNPs in neuroblastoma risk remain undiscovered. To determine the relationship between *LIN28A* gene SNPs and neuroblastoma susceptibility, we performed this multi-centre epidemiological study.

2 | MATERIALS AND METHODS

2.1 | Study subjects

The current case-control study included 505 cases and 1070 healthy non-cancer controls, as noted previously (Table S1).³⁷ Cases were newly diagnosed and histologically confirmed with neuroblastoma. Frequency-matched controls on age and sex were recruited from the same residing area as cases. Without no exception, every participant provided his/her necessary written informed consent. Demographic information was gathered up by trained interviewers. The complete criterion for selecting participants was addressed in our previous work.³⁸ This study has gained its approval from the institutional review boards constituted by all participating hospitals including Guangzhou Women and Children's Medical Center, the First Affiliated Hospital of Zhengzhou University, the Second Affiliated Hospital of Xi'an Jiaotong University, and The Second Affiliated Hospital and Yuying Children's Hospital of Wenzhou Medical University.

2.2 | Polymorphism selection and genotyping

We retrieved four SNPs with potential function in the *LIN28A* gene from the dbSNP database and SNPinfo software.³⁹ Selection

criteria were briefly described below: (a) the minor allele frequency reported in HapMap was >5% for Chinese Han subjects; (b) putative functional potentials SNPs located in the 5'-flanking region, exon, 5'-untranslated region (UTR) and 3' UTR, which might affect transcription activity or binding capacity of the microRNA binding site; and (c) SNPs in low linkage disequilibrium (LD) with each other ($R^2 < 0.8$). There was no significant LD ($R^2 < 0.8$) among these four SNPs of *LIN28A* ($R^2 = 0.183$ between rs3811464 and rs3811463, $R^2 = 0.009$ between rs3811464 and rs34787247, $R^2 = 0.054$ between rs3811464 and rs11247957; $R^2 = 0.03$ between rs3811463 and rs34787247, $R^2 = 0.052$ between rs3811463 and rs11247957; $R^2 = 0.002$ between rs34787247 and rs11247957) (Figure S1). The locations of these SNPs in the *LIN28A* are as below: rs3811464 G>A in the upstream, rs3811463 T>C, rs34787247 G>A, and rs11247957 G>A are all in the 3' UTR. More detailed selection standards were exhibited at our previous work.³⁸ DNA was isolated from the blood sample using a TIANamp Blood DNA Kit (TianGen Biotech Co. Ltd.). Then, the DNA was further performed to genotype using the TaqMan methodology instructed by the manufacturers.⁴⁰⁻⁴² Negative controls (water samples) were used to ensure genotyping preciseness. A repeated genotyping of 10% randomly selected sample was also conducted in all plates with concordance rates of 100%.

2.3 | Statistical analysis

The χ^2 test was applied to test the difference in the distributions of subject characteristics between the cases and controls. A goodness-of-fit chi-squared test was adopted to find whether there exists Hardy-Weinberg equilibrium (HWE) among controls. Logistic regression analysis was applied to detect any association with neuroblastoma risk, with the crude and adjusted odds ratios (ORs) and 95% confidence intervals (CIs). The adjusted ORs were adjusted for age and gender. We determined the risk genotypes for each SNP based on its association with neuroblastoma risk. If a genotype of a SNP was shown to increase neuroblastoma risk (OR > 1), the genotype was regarded as a risk genotype. Carriers of 3 risk genotypes represented those carrying three risk genotypes of the four SNPs, while 1-4 risk genotypes represented those carrying 1-4 risk genotypes.⁴³ The stratification analyses were also performed to identify the associations by age, gender, sites of origins and clinical stages. Moreover, a combination of rs3811464 G>A, rs3811463 T>C, rs34787247 G>A, rs11247957 G>A was regarded as a haplotype. Unphased genotype data were used to identify haplotype frequencies and individual haplotypes. Logistic regression analyses also help to obtain haplotype frequencies and distinct haplotypes, with the adjustment for gender and age.^{44,45} The haplotype of the highest rate was used as the reference group to calculate ORs for haplotype associated with neuroblastoma risk. We set $P < .05$ as a significant borderline for

all tests. We used SAS 9.1 (SAS Institute, Cary, NC) to compute all statistics.

3 | RESULTS

3.1 | The relationship between *LIN28A* SNPs and neuroblastoma susceptibility

The association of all variant genotypes of the four *LIN28A* SNPs (rs3811464 G>A, rs3811463 T>C, rs34787247 G>A, rs11247957 G>A) with neuroblastoma risk is shown in Table 1 for combined subjects and in Table S2 for divided subjects. All these SNPs in controls were in accordance with HWE (all with an HWE $P > .05$). In the single-locus analysis, only one variant, rs34787247 G>A, in the *LIN28A* gene could significantly influence neuroblastoma susceptibility. Carriers of rs34787247A allele showed increased susceptibility to neuroblastoma (GA vs GG: adjusted OR = 1.30, 95% CI = 1.03-1.64, $P = .027$; AA vs GG: adjusted OR = 2.51, 95% CI = 1.36-4.64, $P = .003$; AA/GA vs GG: adjusted OR = 1.42, 95% CI = 1.12-1.80, $P = .004$; AA vs GG/GA: adjusted OR = 2.39, 95% CI = 1.29-4.42, $P = .006$). The rs3811464 GA/AA, rs3811463 TC/CC, rs34787247 GA/AA and rs11247957 GA/AA are treated as risk genotypes. Then, we analysed the combined effect of risk genotypes and observed that participants with 3 or 1-4 risk genotypes experienced a 1.64-fold (adjusted OR = 1.64, 95% CI = 1.02-2.63, $P = .04$) and 1.26-fold (adjusted OR = 1.26, 95% CI = 1.01-1.56, $P = .04$) increase in the risk of developing neuroblastoma, respectively.

3.2 | Stratification analysis

Table 2 displays the contents of the association between *LIN28A* gene polymorphisms and susceptibility to neuroblastoma in certain groups separated by age, gender, sites of origins and clinical stages. We detected the rs3811464 AA genotypes carriers were more likely to have increased neuroblastoma risk in the subgroup of tumours in retroperitoneal (adjusted OR = 2.23, 95% CI = 1.04-4.81, $P = .041$). As for rs34787247 polymorphism, compared to its GG genotype, stronger risk effect of GA/AA genotypes was found among children ≤ 18 month (adjusted OR = 1.51, 95% CI = 1.04-2.19, $P = .031$), >18 month (adjusted OR = 1.38, 95% CI = 1.01-2.87, $P = .042$), males (adjusted OR = 1.47, 95% CI = 1.08-2.01, $P = .015$) and patients at clinical stages of III + IV (adjusted OR = 1.43, 95% CI = 1.05-1.97, $P = .025$). Besides, the combined analysis stated that the 1-4 risk genotypes had an enhanced neuroblastoma risk in the patients with tumour in retroperitoneal (adjusted OR = 1.44, 95% CI = 1.003-2.07, $P = .048$) and subgroup at early clinical stages I + II + 4S (adjusted OR = 1.44, 95% CI = 1.08-1.91, $P = .014$).

TABLE 1 Association between *LIN28A* gene polymorphisms and neuroblastoma susceptibility

Genotype	Cases (N = 505)	Controls (N = 1070)	P ^a	Crude OR (95% CI)	P	Adjusted OR (95% CI) ^b	p ^b
	No. (%)	No. (%)					
rs3811464 G>A (HWE = 0.063)							
GG	359 (71.09)	790 (73.83)		1.00		1.00	
GA	123 (24.36)	250 (23.36)		1.10 (0.87-1.39)	.440	1.09 (0.87-1.38)	.452
AA	23 (4.55)	30 (2.80)		1.71 (0.99-2.96)	.057	1.70 (0.98-2.96)	.058
Additive			.115	1.17 (0.96-1.43)	.116	1.17 (0.96-1.43)	.114
Dominant	146 (28.91)	280 (26.17)	.253	1.15 (0.91-1.45)	.253	1.15 (0.91-1.45)	.250
Recessive	482 (95.45)	1040 (97.20)	.072	1.65 (0.95-2.88)	.075	1.66 (0.95-2.88)	.075
rs3811463 T>C (HWE = 0.530)							
TT	364 (72.08)	785 (73.36)		1.00		1.00	
TC	127 (25.15)	260 (24.30)		1.08 (0.86-1.36)	.528	1.07 (0.85-1.35)	.558
CC	14 (2.77)	25 (2.34)		1.24 (0.64-2.39)	.531	1.25 (0.64-2.41)	.514
Additive			.530	1.07 (0.87-1.31)	.530	1.07 (0.87-1.32)	.525
Dominant	141 (27.92)	285 (26.64)	.592	1.07 (0.84-1.35)	.592	1.07 (0.84-1.35)	.591
Recessive	491 (97.23)	1045 (97.66)	.603	1.19 (0.61-2.31)	.604	1.20 (0.62-2.33)	.591
rs34787247 G>A (HWE = 0.390)							
GG	353 (69.90)	821 (76.73)		1.00		1.00	
GA	130 (25.74)	229 (21.40)		1.30 (1.03-1.65)	.026	1.30 (1.03-1.64)	.027
AA	22 (4.36)	20 (1.87)		2.52 (1.37-4.66)	.003	2.51 (1.36-4.64)	.003
Additive			.0006	1.42 (1.16-1.74)	.0007	1.42 (1.16-1.74)	.0007
Dominant	152 (30.10)	249 (23.27)	.004	1.42 (1.12-1.80)	.004	1.42 (1.12-1.80)	.004
Recessive	483 (95.64)	1050 (98.13)	.004	2.39 (1.29-4.42)	.006	2.39 (1.29-4.42)	.006
rs11247957 G>A (HWE = 0.554)							
GG	481 (95.25)	1032 (96.45)		1.00		1.00	
GA	24 (4.75)	38 (3.55)		1.38 (0.83-2.32)	.219	1.39 (0.83-2.33)	.218
AA	0 (0.00)	0 (0.00)		/	/	/	/
Additive			.253	1.36 (0.80-2.29)	.254	1.36 (0.81-2.30)	.249
Dominant	24 (4.75)	38 (3.55)	.253	1.36 (0.80-2.29)	.254	1.36 (0.81-2.30)	.249
Combined effect of risk genotypes ^c							
0	192 (38.02)	465 (43.46)		1.00		1.00	
1	198 (39.21)	404 (37.76)		1.14 (0.93-1.39)	.204	1.14 (0.93-1.39)	.213
2	82 (16.24)	156 (14.58)		1.22 (0.92-1.63)	.173	1.21 (0.91-1.62)	.188
3	31 (6.14)	44 (4.11)		1.64 (1.02-2.62)	.041	1.64 (1.02-2.63)	.040
4	2 (0.40)	1 (0.09)		4.64 (0.42-51.33)	.210	4.54 (0.42-50.24)	.217
Trend			.010	1.17 (1.04-1.32)	.010	1.17 (1.04-1.32)	.010
0	192 (38.02)	465 (43.46)		1.00		1.00	
1-4	313 (61.98)	605 (56.54)	.041	1.25 (1.01-1.56)	.041	1.26 (1.01-1.56)	.040

Abbreviations: CI, confidence interval; OR, odds ratio.

Significance of bold values are the P values less than 0.05 or the 95% CIs excluded 1.

^a χ^2 test for genotype distributions between neuroblastoma patients and controls.

^bAdjusted for age and gender.

^cRisk genotypes were rs3811464 GA/AA, rs3811463 TC/CC, rs34787247 GA/AA and rs11247957 GA/AA.

3.3 | *LIN28A* haplotype analysis

We further determined whether the haplotypes of the four *LIN28A* SNPs were linked to neuroblastoma risk. As shown in Table 3, the

haplotype consisting of wild-type alleles (GTGG) was defined as the reference group. We detected a significant elevated neuroblastoma risk in subjects with haplotypes GTAG (adjusted OR = 1.35, 95% CI = 1.07-1.72, $P = .012$) and ACAG (adjusted OR = 3.20, 95% CI = 1.41-7.25, $P = .005$).

TABLE 2 Stratification analysis of risk genotypes with neuroblastoma susceptibility

Variables	rs3811464 (cases/ controls)		P ^a	rs34787247 (cases/ controls)		P ^a	Combined risk geno- types (case/controls)			P ^a		
	GG/GA	AA		AOR (95% CI) ^a	GG		GA/AA	AOR (95% CI) ^a	0		1-4	AOR (95% CI) ^a
Age, month												
≤18	181/413	8/12	1.50 (0.60-3.75)	.381	126/319	63/106	1.51 (1.04-2.19)	.031	67/186	122/239	1.42 (0.99-2.02)	.055
>18	301/627	15/18	1.74 (0.87-3.51)	.120	227/502	89/143	1.38 (1.01-1.87)	.042	125/279	191/366	1.17 (0.89-1.54)	.271
Gender												
Females	203/439	10/9	2.41 (0.97-6.04)	.060	149/340	64/108	1.35 (0.94-1.95)	.105	81/200	132/248	1.31 (0.94-1.83)	.109
Males	279/601	13/21	1.34 (0.66-2.72)	.415	204/481	88/141	1.47 (1.08-2.01)	.015	111/265	181/357	1.21 (0.91-1.61)	.184
Sites of origin												
Adrenal gland	167/1040	6/30	1.28 (0.52-3.13)	.593	124/821	49/249	1.31 (0.91-1.88)	.147	69/465	104/605	1.17 (0.84-1.62)	.359
Retroperitoneal	138/1040	9/30	2.23 (1.04-4.81)	.041	105/821	42/249	1.33 (0.90-1.95)	.152	51/465	96/605	1.44 (1.003-2.07)	.048
Mediastinum	129/1040	6/30	1.67 (0.68-4.11)	.262	94/821	41/249	1.43 (0.96-2.12)	.078	55/465	80/605	1.13 (0.78-1.62)	.528
Others	40/1040	2/30	1.67 (0.38-7.27)	.494	27/821	15/249	1.84 (0.96-3.51)	.066	15/465	27/605	1.38 (0.73-2.63)	.326
Clinical stages												
I + II + 4s	237/1040	13/30	1.93 (0.99-3.75)	.054	177/821	73/249	1.35 (1.00-1.84)	.054	87/465	163/605	1.44 (1.08-1.91)	.014
III + IV	222/1040	10/30	1.59 (0.76-3.31)	.216	162/821	70/249	1.43 (1.05-1.97)	.025	97/465	135/605	1.07 (0.80-1.43)	.646

Abbreviations: AOR, adjusted odds ratio; CI, confidence interval.

Significance of bold values are the P values less than 0.05 or the 95% CIs excluded 1.

^aAdjusted for age and gender, omitting the corresponding stratify factor.

TABLE 3 Association between inferred haplotypes of *LIN28A* gene and neuroblastoma susceptibility

Haplotypes ^a	Cases (N = 1010)	Controls (N = 2140)	Crude OR (95% CI)	P	Adjusted OR ^b (95% CI)	p ^b
	No. (%)	No. (%)				
GTGG	642 (63.56)	1467 (68.55)	1.00		1.00	
GTGA	2 (0.20)	0 (0.00)	/	/	/	/
GTAG	129 (12.77)	218 (10.19)	1.35 (1.07-1.71)	.013	1.35 (1.07-1.72)	.012
GTAA	1 (0.10)	0 (0.00)	/	/	/	/
GCGG	54 (5.35)	123 (5.75)	1.00 (0.72-1.40)	.985	1.01 (0.72-1.40)	.977
GCGA	2 (0.20)	3 (0.14)	1.52 (0.25-9.14)	.645	1.51 (0.25-9.04)	.655
GCAG	10 (0.99)	18 (0.84)	1.27 (0.58-2.77)	.548	1.27 (0.58-2.76)	.551
GCAA	1 (0.10)	1 (0.05)	2.29 (0.14-36.59)	.559	2.29 (0.14-36.71)	.558
ATGG	64 (6.34)	124 (5.79)	1.18 (0.86-1.62)	.306	1.18 (0.86-1.62)	.304
ATGA	0 (0.00)	0 (0.00)	/	/	/	/
ATAG	17 (1.68)	21 (0.98)	1.85 (0.97-3.53)	.062	1.85 (0.97-3.53)	.062
ATAA	0 (0)	0 (0.00)	/	/	/	/
ACGG	56 (5.54)	121 (5.65)	1.06 (0.76-1.47)	.740	1.06 (0.76-1.47)	.740
ACGA	16 (1.58)	33 (1.54)	1.11 (0.61-2.03)	.740	1.11 (0.61-2.04)	.728
ACAG	14 (1.39)	10 (0.47)	3.20 (1.41-7.24)	.005	3.20 (1.41-7.25)	.005
ACAA	2 (0.20)	1 (0.05)	4.57 (0.41-50.49)	.215	4.54 (0.41-50.19)	.217

Abbreviations: CI, confidence interval; OR, odds ratio.

Significance of bold values are the P values less than 0.05 or the 95% CIs excluded 1.

^aThe haplotype order was rs3811464, rs3811463, rs34787247 and rs11247957.

^bObtained in logistic regression models with adjustment for age and gender.

4 | DISCUSSION

At the present, there remain many hidden genetic factors in association with neuroblastoma risk to be discovered to fill up the knowledge gaps. Thus, the identification of more polymorphisms is needed to unearth the full range of neuroblastoma susceptibility variations. Herein, we undertook a four-centre case-control study to investigate the role of *LIN28A* polymorphisms on neuroblastoma risk in Chinese children. We are the pioneer in unveiling the association of the rs34787247A allele with an elevated neuroblastoma risk in a Chinese population.

LIN28A gene resides on chromosome 1p36.11. Several lines of evidence suggested the roles of *LIN28A* gene polymorphisms in cancer risk. Zhang et al⁴⁶ found *LIN28B* rs221636 could decrease the risk of oral cavity cancer in a study of 384 cases and 731 healthy controls, including six SNPs in *let-7/LIN28* gene. Nevertheless, they failed to detect the association of *LIN28A* rs4659441 and rs3811463 with oral cavity cancer risk. Permut-Wey et al⁴⁷ observed the predisposing role of rs12728900 and rs11247946 in *LIN28A* on epithelial ovarian cancer in European ancestry. Sung et al⁴⁸ carried out two genome-wide association studies in East Asia, 5066 breast cancer cases and 4337 controls recruited from Chinese and Koreans. They reported that the 237 SNPs located in microRNA biogenesis pertinent pathway genes had no significant association with breast cancer risk, including SNPs of *LIN28A* (rs11247954, rs12728900, rs3811463, rs4274112, rs4659441, rs6598964, and rs6683792).

Chen et al⁴⁹ determined the effect of SNPs of *LIN28* gene on the breast cancer risk. In analysing five SNPs (rs12122703 A>G, rs3811464 G>A, rs11247955 G>A, rs3811463 T>C, rs6697410 T>G) in *LIN28*, they successfully identified rs3811463 and rs6697410 to be linked to breast cancer risk using a hospital-based case-control study in 1004 cases and 1296 controls. They further conducted a community-based validation study using 511 cases and 645 controls. They validated that the rs3811463-C allele predisposed to an increased risk of breast cancer. Further functional experiments suggested that the rs3811463C allele located near the *let-7* binding site of the *LIN28* gene. This variant attenuated *let-7*-induced degradation of *LIN28* mRNA, leading to enhanced levels of LIN28 protein, which could, in turn, decrease mature *let-7* level, finally alter breast cancer risk. The quite association results of *LIN28A* SNPs reported by studies implied that the effects of *LIN28A* on cancer risk would be modified by many factors like ethnicities, sample sizes and cancer types. Therefore, discovering the function of *LIN28A* SNPs on a particular type of cancer and specific ethnicity is of great necessity.

Given *LIN28A*'s vital role in malignancies, we undertook this first epidemiological study to outline the correlation of *LIN28A* polymorphisms and neuroblastoma risk in a Chinese population. Despite the abundance of reports on *LIN28A* gene variation and cancers, investigations of contribution of *LIN28A* SNPs to neuroblastoma cancer risks were scarce. Our results thoroughly showed rs34787247A allele could contribute to an increased neuroblastoma risk. Moreover, we also observed an enhanced risk of neuroblastoma in subjects

carrying 3 or 1-4 risk genotypes. However, no significant relationship of the rest three variants rs3811464, rs3811463 and rs11247957 was detected.

We carried out a pioneering study on the association between *LIN28A* gene SNPs and susceptibility to neuroblastoma. Limitations also existed. Firstly, the sample size is not large enough to generate reliable statistics. Some of the results might be merely fortuitous events, particularly the stratification analysis. Secondly, we examined four SNPs in this research. More potential neuroblastoma risk-associated SNPs in the *LIN28A* gene await to be explored. Thirdly, although the participants were enrolled from four different cities, findings from the restricted Chinese population could not be extrapolated to other ethnicities directly. Lastly, environmental factors were not considered in this study.

In all, we presented a multi-centre case-control study in Chinese children. For the first time, our findings unveiled a contributing role of *LIN28A* gene SNPs in neuroblastoma risk. In the future, an integrative analysis, covering more profound and specific factors, with environmental factors, genetic-environmental interaction, should be carried out to unearth the aetiology of neuroblastoma.

ACKNOWLEDGEMENTS

This study was supported by grants from the Pearl River S&T Nova Program of Guangzhou (No: 201710010086) and Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease (No: 2019B030301004). The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

CONFLICT OF INTEREST

The authors declare no competing financial interests.

AUTHOR'S CONTRIBUTIONS

All authors contributed significantly to this work. RX Hua, L. Yuan, C. Chen, J. Liu, J. Cheng, H. Zhou, J. Zhang, and J. He performed the research study and collected the data; J. He analysed the data; H. Xia, X. Zhang and J. He designed the research study; RX Hua, Z. Zhuo and L. Ge wrote the paper; J Zhu and J. He prepared all the Tables. All authors reviewed the manuscript. In addition, all authors have read and approved the manuscript.

ORCID

Jinhong Zhu  <https://orcid.org/0000-0002-0408-3101>

Huimin Xia  <https://orcid.org/0000-0002-0103-1672>

Jing He  <https://orcid.org/0000-0002-1954-2892>

DATA AVAILABILITY STATEMENT

All the data were available upon request.

REFERENCES

- Nagaich U. Neuroblastoma cancer in children. *J Adv Pharm Technol Res.* 2019;10:45.

- Newman EA, Abdessalam S, Aldrink JH, et al. Update on neuroblastoma. *J Pediatr Surg.* 2019;54:383-389.
- Cheung NK, Dyer MA. Neuroblastoma: developmental biology, cancer genomics and immunotherapy. *Nat Rev Cancer.* 2013;13:397-411.
- Maris JM, Hogarty MD, Bagatell R, Cohn SL. Neuroblastoma. *Lancet.* 2007;369:2106-2120.
- Meena JP, Gupta AK. Neuroblastoma in a developing country: miles to go. *Indian J Pediatr.* 2019;86:403-405.
- Meany HJ. Non-high-risk neuroblastoma: classification and achievements in therapy. *Children (Basel).* 2019;6:E5.
- Morgenstern DA, Bagatell R, Cohn SL, et al. The challenge of defining "ultra-high-risk" neuroblastoma. *Pediatr Blood Cancer.* 2019;66:e27556.
- Matthay KK, Maris JM, Schleiermacher G, et al. Neuroblastoma. *Nat Rev Dis Primers.* 2016;2:16078.
- Westermann F, Schwab M. Genetic parameters of neuroblastomas. *Cancer Lett.* 2002;184:127-147.
- MacFarland S, Bagatell R. Advances in neuroblastoma therapy. *Curr Opin Pediatr.* 2019;31:14-20.
- Bagley AF, Grosshans DR, Philip NV, et al. Efficacy of proton therapy in children with high-risk and locally recurrent neuroblastoma. *Pediatr Blood Cancer.* 2019;66(8):e27786.
- Barr EK, Applebaum MA. Genetic predisposition to neuroblastoma. *Children (Basel).* 2018;5:E119.
- Zhong X, Liu Y, Liu H, Zhang Y, Wang L, Zhang H. Identification of potential prognostic genes for neuroblastoma. *Front Genet.* 2018;9:589.
- Cook MN, Olshan AF, Guess HA, et al. Maternal medication use and neuroblastoma in offspring. *Am J Epidemiol.* 2004;159:721-731.
- Menegaux F, Olshan AF, Neglia JP, Pollock BH, Bondy ML. Day care, childhood infections, and risk of neuroblastoma. *Am J Epidemiol.* 2004;159:843-851.
- Ogawa S, Takita J, Sanada M, Hayashi Y. Oncogenic mutations of ALK in neuroblastoma. *Cancer Sci.* 2011;102:302-308.
- Aygun Z, Batur S, Emre S, Celkan T, Ozman O, Comunoglu N. Frequency of ALK and GD2 expression in neuroblastoma. *Fetal Pediatr Pathol.* 2019;38:326-334.
- Trochet D, Bourdeaut F, Janoueix-Lerosey I, et al. Germline mutations of the paired-like homeobox 2B (PHOX2B) gene in neuroblastoma. *Am J Hum Genet.* 2004;74:761-764.
- Diskin SJ, Capasso M, Diamond M, et al. Rare variants in TP53 and susceptibility to neuroblastoma. *J Natl Cancer Inst.* 2014;106:dju047.
- Diskin SJ, Capasso M, Schnepf RW, et al. Common variation at 6q16 within HACE1 and LIN28B influences susceptibility to neuroblastoma. *Nat Genet.* 2012;44:1126-1130.
- Oldridge DA, Wood AC, Weichert-Leahey N, et al. Genetic predisposition to neuroblastoma mediated by a LMO1 super-enhancer polymorphism. *Nature.* 2015;528:418-421.
- Capasso M, Devoto M, Hou C, et al. Common variations in BARD1 influence susceptibility to high-risk neuroblastoma. *Nat Genet.* 2009;41:718-723.
- Capasso M, Diskin S, Cimmino F, et al. Common genetic variants in NEFL influence gene expression and neuroblastoma risk. *Cancer Res.* 2014;74:6913-6924.
- Capasso M, McDaniel LD, Cimmino F, et al. The functional variant rs34330 of CDKN1B is associated with risk of neuroblastoma. *J Cell Mol Med.* 2017;21:3224-3230.
- Cimmino F, Avitabile M, Diskin SJ, et al. Fine mapping of 2q35 high-risk neuroblastoma locus reveals independent functional risk variants and suggests full-length BARD1 as tumor-suppressor. *Int J Cancer.* 2018;143:2828-2837.
- Shyh-Chang N, Daley GQ. Lin28: primal regulator of growth and metabolism in stem cells. *Cell Stem Cell.* 2013;12:395-406.

27. Kim JD, Toda C, Ramirez CM, Fernandez-Hernando C, Diano S. Hypothalamic ventromedial Lin28a enhances glucose metabolism in diet-induced obesity. *Diabetes*. 2017;66:2102-2111.
28. Moss EG, Lee RC, Ambros V. The cold shock domain protein LIN-28 controls developmental timing in *C. elegans* and is regulated by the lin-4 RNA. *Cell*. 1997;88:637-646.
29. Newman MA, Thomson JM, Hammond SM. Lin-28 interaction with the Let-7 precursor loop mediates regulated microRNA processing. *RNA*. 2008;14:1539-1549.
30. Rybak A, Fuchs H, Smirnova L, et al. A feedback loop comprising lin-28 and let-7 controls pre-let-7 maturation during neural stem-cell commitment. *Nat Cell Biol*. 2008;10:987-993.
31. Heo I, Joo C, Kim YK, et al. TUT4 in concert with Lin28 suppresses microRNA biogenesis through pre-microRNA uridylation. *Cell*. 2009;138:696-708.
32. Heo I, Joo C, Cho J, Ha M, Han J, Kim VN. Lin28 mediates the terminal uridylation of let-7 precursor MicroRNA. *Mol Cell*. 2008;32:276-284.
33. Takamizawa J, Konishi H, Yanagisawa K, et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res*. 2004;64:3753-3756.
34. Yu L, Jiang Y, Tang B. Lin28a functionally modulates bupivacaine-induced dorsal root ganglion neuron apoptosis through TrkA activation. *Biomed Pharmacother*. 2018;98:63-68.
35. Viswanathan SR, Powers JT, Einhorn W, et al. Lin28 promotes transformation and is associated with advanced human malignancies. *Nat Genet*. 2009;41:843-848.
36. Wang YC, Chen YL, Yuan RH, et al. Lin-28B expression promotes transformation and invasion in human hepatocellular carcinoma. *Carcinogenesis*. 2010;31:1516-1522.
37. Cheng J, Zhuo Z, Xin Y, et al. Relevance of XPD polymorphisms to neuroblastoma risk in Chinese children: a four-center case-control study. *Aging (Albany NY)*. 2018;10:1989-2000.
38. He J, Wang F, Zhu J, et al. Association of potentially functional variants in the XPG gene with neuroblastoma risk in a Chinese population. *J Cell Mol Med*. 2016;20:1481-1490.
39. Zhuo Z, Fu W, Liu J, et al. LIN28A gene polymorphisms confer Wilms tumour susceptibility: a four-centre case-control study. *J Cell Mol Med*. 2019;23:7105-7110.
40. Chen X, Wang Y, Cheng K, et al. Genetic variants in the regulatory region of SLC10A1 are not associated with the risk of hepatitis B virus infection and clearance. *Infect Genet Evol*. 2016;44:495-500.
41. Chang J, Tian J, Zhu Y, et al. Exome-wide analysis identifies three low-frequency missense variants associated with pancreatic cancer risk in Chinese populations. *Nat Commun*. 2018;9:3688.
42. Chang J, Zhong R, Tian J, et al. Exome-wide analyses identify low-frequency variant in CYP26B1 and additional coding variants associated with esophageal squamous cell carcinoma. *Nat Genet*. 2018;50:338-343.
43. Hua RX, Zhuo ZJ, Zhu J, et al. Association between genetic variants in the XPG gene and gastric cancer risk in a Southern Chinese population. *Aging (Albany NY)*. 2016;8:3311-3320.
44. Lin DY, Zeng D, Millikan R. Maximum likelihood estimation of haplotype effects and haplotype-environment interactions in association studies. *Genet Epidemiol*. 2005;29:299-312.
45. Spinka C, Carroll RJ, Chatterjee N. Analysis of case-control studies of genetic and environmental factors with missing genetic information and haplotype-phase ambiguity. *Genet Epidemiol*. 2005;29:108-127.
46. Zhang Y, Zhu L, Wang R, et al. Genetic variants in let-7/Lin28 modulate the risk of oral cavity cancer in a Chinese Han population. *Sci Rep*. 2014;4:7434.
47. Permeth-Wey J, Kim D, Tsai YY, et al. Ovarian Cancer Association C. LIN28B polymorphisms influence susceptibility to epithelial ovarian cancer. *Cancer Res*. 2011;71:3896-3903.
48. Sung H, Zhang B, Choi JY, et al. Common genetic variants in the microRNA biogenesis pathway are not associated with breast cancer risk in Asian women. *Cancer Epidemiol Biomarkers Prev*. 2012;21:1385-1387.
49. Chen AX, Yu KD, Fan L, et al. Germline genetic variants disturbing the Let-7/LIN28 double-negative feedback loop alter breast cancer susceptibility. *PLoS Genet*. 2011;7:e1002259.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Hua R-X, Zhuo Z, Ge L, et al. LIN28A gene polymorphisms modify neuroblastoma susceptibility: A four-centre case-control study. *J Cell Mol Med*. 2020;24:1059-1066. <https://doi.org/10.1111/jcmm.14827>