PHILOSOPHICAL TRANSACTIONS B

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Introduction



Cite this article: Ghiselli F, Milani L. 2019 Linking the mitochondrial genotype to phenotype: a complex endeavour. *Phil. Trans. R. Soc. B* **375**: 20190169. http://dx.doi.org/10.1098/rstb.2019.0169

Accepted: 17 October 2019

One contribution of 15 to a theme issue 'Linking the mitochondrial genotype to phenotype: a complex endeavour'.

Subject Areas:

evolution, genetics, molecular biology, synthetic biology, cellular biology, biochemistry

Keywords:

mtDNA genetic variation, heteroplasmy, mitochondrial bottleneck, mitonuclear interactions, mtDNA editing, mitochondrial expression manipulation

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Linking the mitochondrial genotype to phenotype: a complex endeavour

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Finding causal links between genotype and phenotype is a major issue in biology, even more in mitochondrial biology. First of all, mitochondria form complex networks, undergoing fission and fusion and we do not know how such dynamics influence the distribution of mtDNA variants across the mitochondrial network and how they affect the phenotype. Second, the non-Mendelian inheritance of mitochondrial genes can have sex-specific effects and the mechanism of mitochondrial inheritance is still poorly understood, so it is not clear how selection and/or drift act on mtDNA genetic variation in each generation. Third, we still do not know how mtDNA expression is regulated; there is growing evidence for a convoluted mechanism that includes RNA editing, mRNA stability/turnover, post-transcriptional and post-translational modifications. Fourth, mitochondrial activity differs across species as a result of several interacting processes such as drift, adaptation, genotype-by-environment interactions, mitonuclear coevolution and epistasis. This issue will cover several aspects of mitochondrial biology along the path from genotype to phenotype, and it is subdivided into four sections focusing on mitochondrial genetic variation, on the relationship among mitochondria, germ line and sex, on the role of mitochondria in adaptation and phenotypic plasticity, and on some future perspectives in mitochondrial research.

This article is part of the theme issue 'Linking the mitochondrial genotype to phenotype: a complex endeavour'.

1. Mitochondria: powerhouse and beyond

Eukaryotic life is powered by mitochondria, cytoplasmic organelles that originated through a unique endosymbiotic event that changed the history of life on Earth fostering the evolution of multicellular organisms [1,2]. Once a proteobacterium, the mitochondrion retained the bulk of its original biochemical machinery, but its genome (mtDNA) underwent a massive reduction in size, and genes of the ancestral organelle relocated to the nucleus ('endosymbiotic gene transfer' [3]). However, not all the genes moved to the nucleus: of the 1000+ protein-coding genes estimated to have been present in the bacterial ancestor of mitochondria, all known extant mtDNAs retain a subset of 69 ancestral protein-coding genes [4]; animal mtDNAs typically encompass approximately 13 of such genes (but see [5]). Interestingly, the organelles performing oxidative phosphorylation (OXPHOS) retain a genome, and there is a remarkable conservation of genes encoding key OXPHOS subunits in mtDNAs across eukaryotes [4]. Allen [6] postulated that organelle genomes encoding core OXPHOS subunits are necessary for maintaining redox poise (Co-location for Redox Regulation, CoRR; [7]); under this light, the interaction of two or three genomes-or even more in some protists [8]-is unavoidable in eukaryotic cells [9]. The five multi-protein complexes responsible for OXPHOS are composed of subunits encoded by both nuclear and mitochondrial genomes, which need to coevolve despite their markedly different genetic features [10,11]. Because of such differences, and because of some peculiar characteristics of mitochondria, it is quite challenging to reconstruct the evolutionary dynamics and predict the outcomes of mitonuclear interactions and coevolution, and it is particularly difficult to find causal links between

genotype and phenotype in mitochondrial biology [12-14]. First of all, differently from the nuclear genome, mtDNA is subject to non-Mendelian (uniparental) inheritance, and the underlying mechanism is still poorly understood, namely it is not clear how drift and selection act on mtDNA genetic variation each generation (see [15]). Second-albeit with large variation [16]-metazoan mtDNA experiences a higher mutation rate, which results in different evolutionary rates between the two genomes. Third, because of the high mtDNA copy number per cell/organelle, mutations result in a phenotypic effect only when exceeding a threshold level (usually greater than 60% mutant versus wild-type), meaning that alleles can fluctuate at mid-low frequency in the mitochondrial gene pool without being 'visible' to selection [14,17] ('buffering' or 'threshold effect' [15,18]). Fourth, individual mitochondria do not exist as permanently distinct entities, but undergo rapid fission and fusion processes, exchanging proteins, mtDNA and lipids. Fission produces new mitochondria and plays an important role in quality control and selective elimination of damaged organelles. The fission process yields functionally divergent mitochondriawith different membrane potential $(\Delta \psi_m)$ —and depolarized mitochondria are selectively degraded by mitophagy. On the other hand, fusion produces a network whose components share matrix content and electrochemical gradient, and it has been suggested as a mechanism by which mitochondria complement damaged organelles and compensate metabolic deficiencies [17,19-23]. The mtDNA is organized in nucleoids, discrete DNA-protein complexes that are present in multiple copies (hundreds to thousands) per cell. Nucleoids can be segregated across individual organelles in a cell, but, given the dynamic nature of the mitochondrial network, the association between nucleoids and their products, among nucleoids and among products is temporary. Thus, the situation-much more common than once believed [14]-where different mtDNA variants are present in the same individual, a condition called heteroplasmy, is of particular interest. For reasons that are still unknown, heteroplasmy seems to be unfavourable (but see [24,25]) and it has been related to physiological, cognitive and behavioural complications in mice [26], and to human neurodegenerative diseases and common age-related disorders [27]. For the above-mentioned reasons, it is quite difficult to assess the distribution of mtDNA variants across the mitochondrial network, and how phenotype is affected. Indeed, the link between genotype (mtDNA) and phenotype (e.g. OXPHOS activity) depends on the mobility of mtDNA and the diffusion of its products, so such a link can show various degrees of 'leakyness' [17,28]. Understanding the effects of heteroplasmy-and of mitochondrial genetic variability in general-is therefore a complex endeavour.

2. From genotype to phenotype

The life science community is becoming increasingly aware of the great complexity of mitochondrial biology and evolution, a complexity that has been underestimated for a long time. Recently, mitochondrial biology has been getting more attention from scientists across a wide range of disciplines, both basic and applied. Even the mass media have been engaged in mitochondria-related discussion, especially regarding the issue of 'three-parent babies' (mitochondrial replacement therapy for *in vitro* fertilization). On the biomedical side, the central role of mitochondria in a substantial number of cellular processes implies that mitochondrial malfunctions cause a wide typology of diseases. Once considered rare, mitochondrial disease is now thought to affect 1 in 5000 people, making it the second most commonly diagnosed, serious genetic disease after cystic fibrosis (source: Global Mitochondrial Disease Awareness Week website, http:// gmdaw.org/). Linking the mitochondrial genotype with disease, predicting its presence, severity, heritability and finding a therapy is quite a challenging endeavour.

We think this journal issue represents a relevant contribution for multiple fields of study. The link between some of the basic research here reported and future applications might seem far-fetched. We disagree. We are convinced that life sciences have reached a turning point, where new technologies and methods allow us to study a wider range of organisms and to compare the basis of their biological features. Comparative analyses across increasingly large samples of biodiversity are the most powerful approach to understand the evolution and functioning of organisms. The models and approaches described in this issue will contribute to highlighting similarities and differences between known aspects of mitochondrial biology and features of new emerging models that will surely contribute to the overall picture. Indeed, we want to highlight the importance of using comparative methods in a wide range of organisms, and the new models here presented show features that can help in understanding some obscure areas of mitochondrial biology (see [29,30]).

The contributions included are the result of nearly 3 years of interactions and discussions among scientists working in the field of mitochondrial evolutionary biology. Most of the interactions happened during international meetings, the last being the symposium entitled 'Linking the mitochondrial genotype to phenotype: a complex endeavour' at the annual meeting of the Society for Molecular Biology and Evolution, SMBE 2018, Yokohama, Japan (8–12 July 2018). The purpose of this collaborative effort is to provide new perspectives and angles in the field of mitochondrial biology and evolution. We hope that the work of the group of scientists participating in this issue will increase the future contributions from different disciplines of life sciences, encouraging new collaborations and generating discussions.

The issue will cover several aspects of mitochondrial biology along the path from genotype to phenotype, and paying special attention to non-model species. The contributions (table 1) are subdivided into four sections: (i) mitochondrial genetic variation; (ii) the relationship among mitochondria, germ line and sex; (iii) the role of mitochondria in adaptation and phenotypic plasticity; and (iv) some future perspectives on mitochondrial research.

(a) Mitochondrial genetic variation

Genetic variation is the engine of evolution, and this section highlights some focal points about how mitochondrial genetic variation arises and changes within individuals and across generations.

Schaack *et al.* [31] investigate the challenges in estimating mutation rates, given unknowns such as mtDNA effective population size and fixation probability of heteroplasmic mutations. A critical parameter in understanding rates of change is estimating the mitochondrial mutation rate (mtDNA MR). Despite its importance, this kind of estimate is overlooked. Mutation accumulation experiments are demanding and do not help in distinguishing the role of

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Baetis bicaudatus cellular mitochondrial function Drunella coloradensis molecular (mayflies)	Havird <i>et al.</i> [39]	primary	Baetis tricaudatus	organismal	environmentally induced plasticity in	mitochondria of cold-adapted insects were sensitive to even moderate increases in temperature;
		research	Baetis bicaudatus	cellular	mitochondrial function	those living in thermally variable environments had greater thermal tolerance
(mayfiles)			Drunella coloradensis	molecular		
			(mayflies)			

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Table 1. (Continued.)

reference (this issue)	type	organism(s)	scale(s)	evolutionary topic	finding
Elbassiouny <i>et al.</i> [40]	Primary research	Gymnotiformes Mormyroidea (electric fishes)	molecular	molecular evolution of OXPHOS complexes	evidence for convergent patterns of molecular evolution of mitochondrial 0XPH0S genes in two different groups of electrogenic fishes
Mackenzie & Kundariya [41]	Review	plants	organismal organelle	phenotypic plasticity in plants	some of the underlying versatility of plants to adapt to abiotic and biotic stress emerges from neofunctionalization of organelles and organellar proteins
future perspectives					
Klucnika & Ma [42]	Review	animals	organismal molecular	map and edit mtDNA	the efforts in developing systems to induce/increase the basal recombination frequency, and the use of mito-nucleases, could finally lead to the establishment of mitochondria-adapted DNA editing platform
Wallis <i>et al.</i> [43]	Review	mammals	population organismal molecular	tools for manipulating mitochondrial gene expression	mtDNA sequence-specific DNA-binding and RNA-binding proteins tethered to various effector domains show great promise as highly specific, customizable genetic tools for mitochondrial research
Milani & Ghiselli [29]	Opinion piece	model/non-model animals	organismal	perspective in mitochondrial research	the comparative method and new animal models have the potential to address still unanswered biological questions

4

5

mutation from other evolutionary forces. mtDNA MRs depend on the rate of replication errors and unrepaired DNA damage, but since there are multiple copies of the mtDNA genome per mitochondrion and many organelles per cell, the fate of a given mutation also depends on its selective coefficient and selection effectiveness, relative to the likelihood of loss/fixation by genetic drift. Schaack *et al.* [31] review the unique features of the mitochondrial genome that pose a challenge for accurate mutation rate estimation and discuss ways to overcome such challenges and understand mtDNA MR variation within and between individuals, populations and species. They underline that to understand how mtDNA MRs evolve it is essential to extend the analysis to non-model organisms and multiple genotypes per taxon.

Dubie et al. [32] use Caenorhabditis elegans data to discuss proliferation and persistence of spontaneous selfish mitochondrial deletions. Mitochondrial genomes can sustain mutations that are detrimental to individual fitness but that proliferate because of a replicative advantage (hence 'selfish'). Dubie et al. [32] analysed the fitness effects and population dynamics of a mitochondrial genome containing a novel 499 bp deletion in the *ctb-1* gene (Δctb-1). Δctb-1 reached a high heteroplasmic frequency, imposing a significant fitness cost compared to individuals bearing wild-type mitochondria. Deletion-bearing worms were rapidly purged within a few generations when competed against wild-type mtDNA-bearing worms in experimental populations. By contrast, the Δ ctb-1 mitotype was able to persist in large populations comprising heteroplasmic individuals only. The data obtained within experimental lines subjected to severe population bottlenecks indicate a selfish drive. Indeed, levels of mitochondrial heteroplasmy are the product of mutation and selection at different levels of organization, and the use of single individual bottlenecks can eliminate the selection between individuals, revealing the contributions of selection and drift within the germline.

Heteroplasmy is the presence of different mtDNA variants within the same individual. The dynamics of heteroplasmic allele frequency among tissues of the human body is not well understood. Barrett et al. [33] present data supporting a pronounced bottleneck in the mtDNA of human hair. By measuring allele frequency at heteroplasmic sites, Barrett et al. [33] observed high variance in allele frequency among separate hairs from the same individual. These findings have important implications for understanding mtDNA variation across different tissues in the human body occurring during embryonic development and throughout the lifetime. The described population genetic modelling estimated the somatic bottleneck during embryonic follicle development of separate hairs to be much more drastic than somatic bottlenecks for blood and buccal tissues but comparable to the germline bottleneck, and that hair undergoes additional genetic drift before and after the divergence of mtDNA lineages of individual hair follicles. These findings have important implications for our understanding of mtDNA dynamics and also for forensics: heteroplasmic frequency may vary between hairs, and heteroplasmy may be present in the hair sample but absent in another tissue of the same individual.

(b) Mitochondria, germ line and sex

Which mitochondria are inherited from one generation to the next through the germline? Is it a random subset or a selected one? If selection occurs, when and how is it achieved? Strictly maternal inheritance (SMI) entails an asymmetry in the transmission mechanism between sexes: what are the consequences for mitochondrial evolution and for the two sexes?

Knorre [34] reviews the role of mitochondrial dynamics in mtDNA quality control and proposes cases in which mtDNA can evade it. Mitochondria can show different $\Delta \psi_{\rm m}$ on which mitochondrial quality-control mechanisms rely, distinguishing between functional and damaged mitochondria. Nonetheless, mutations that increase $\Delta \psi_m$ can evade such control even being deleterious. Knorre [34] reviews recent findings on intracellular mtDNA quality control by mitophagy and discusses other mechanisms by which the nuclear genome can affect the competition of mtDNA variants in the cell, thus affecting heteroplasmy levels. He also examines the hypothesis that the zygote is the stage at which mtDNA quality control takes place at the intracellular level. Mitochondrial dynamics are required to fulfil multiple functions, but these dynamics can disrupt the genotype-tophenotype linkage at the intracellular level, thus preventing intracellular quality control of mtDNAs. Knorre [34] suggests that this trade-off has been resolved by the evolution of a restriction of intracellular quality control to the germline.

Because of SMI, some authors predict that mitochondrial quality control is less effective in males ('Mother's curse hypothesis', see [44,45]). Bettinazzi et al. [35] use the only known evolutionarily stable exception to SMI to investigate the link between mtDNA variants and sperm performance. Because of the strict maternal inheritance of mitochondria in animals, haplotypes that negatively affect male fertility can become fixed in populations. Doubly uniparental inheritance (DUI) of mitochondria is a stable exception, found so far in 100+ bivalve species that show two mtDNA lineages that evolve independently, transmitted separately, one by oocytes and the other by spermatozoa. Since the two DUI mitochondrial lineages are likely subject to different sexspecific selective pressures, the DUI system is a unique model to evaluate selection on sperm mitochondria for male functions, potentially contributing to male reproductive fitness. This study highlighted a significant divergence in sperm performance and partially in energy metabolism between DUI and SMI species. As sperm mitochondria in DUI species are not an evolutionary dead-end, male-specific energetic adaptations could reflect selection for both fertilization success and male mitotype preservation.

Nagarajan-Radha et al. [36] present new data about how mitochondrial genetic variation exerts sex-specific effects on physiological function. According to the mother's curse hypothesis, maternal inheritance of mitochondria will facilitate the accumulation of mtDNA mutations that are harmful to males but benign/beneficial to females. These male-harming mutations are expected to differ across a population and to cause larger genetic variation and possibly larger phenotypic effects in males and/or have sexually antagonistic effects. Nagarajan-Radha et al. [36] explore signatures of male-bias or sexual antagonism in the metabolic rate by measuring the effects of different mitochondrial haplotypes on the production of carbon dioxide across strains of Drosophila melanogaster, controlling for mass and activity. The study reports sex-specific (male-biased) effects of mtDNA haplotypes on metabolic rate, and a negative intersexual correlation for metabolic rate across haplotypes consistent with the prediction that SMI enabled the accumulation of mutations that increase female fitness, but at the expense of male fitness. The authors highlight the importance

6

to address future research to a broader range of nuclear genetic and environmental contexts and also to other metazoan species.

(c) Mitochondria, adaptation and phenotypic plasticity

Mitochondria have a central role in many fundamental processes of eukaryotic life, well beyond energy production, so it should not be surprising that they have been suggested to be involved in adaptive processes [46,47]. However, the mechanisms underlying mitochondrial-driven adaptation are complex and a subject of debate. This section deals with the contribution of mitochondria and mitonuclear interactions to adaptation, phenotypic plasticity and complex phenotypes.

Rand & Mossman [37] discuss how mitonuclear conflict and cooperation govern the integration of genotypes, phenotypes and environments. The interaction between the mitochondrial and nuclear genomes under changing environments has pervasively influenced organism evolution. Indeed, mitochondria play crucial roles in signalling, altering how nuclear genes are expressed as phenotypes. These interactions are examples of genotype-by-environment (GxE) and gene-by-gene (GxG) interactions, producing context-dependent effects on the link between genotype and phenotype. Mitonuclear interactions have pleiotropic effects across numerous phenotypes and evidence from Drosophila and other organisms shows that mitonuclear interactions are common features of GxE and GxG. Rand & Mossman [37] outline approaches that could help in depicting the phenotypic and fitness landscapes in a nuclear-mitochondrial co-evolved unit and their relation to genetic variation. For example, the population-structure-mitonuclear-coadaptation hypothesis possibly explains why the breakdown of mitonuclear coadaptation is so evident in Tigriopus-highly structured inbred populations-but not in large outbred species such as Drosophila. The authors underline how mitonuclear interactions are important to understand the contextdependent effects underlying the architecture of complex phenotypes.

Camus et al. [38] analysed the impact of mitonuclear interactions on life-history responses to diet in D. melanogaster. Since mitochondria influence resource allocation, severe incompatibilities between mitochondrial and nuclear genomes can have pervasive effects on both fitness and longevity. How milder deficits in mitochondrial function affect life-history trade-offs is less well understood. Camus et al. [38] found that in closely related fly populations (in which the mtDNA genetic distance is similar to that in human populations), mitonuclear interactions do have significant impact on life-history trade-offs, but these effects are not predictable by relatedness and depend on the nuclear background. Camus et al. [38] analysed how mitonuclear interactions affect the trade-off between fecundity and longevity, considering different mitochondrial DNA haplotypes against two contrasting nuclear backgrounds in response to different diets. Mitonuclear interactions had substantial effects on resource allocation and life-history trade-offs in D. melanogaster but did not reflect genetic distance between mitochondrial haplotypes, so their effects are inconsistent, thus not predictable by relatedness. These effects can vary greatly, such as between the two nuclear genotypes used in

this study, thus the authors judge it hazardous to generalize from mtDNA interactions with a single nuclear background.

Havird et al. [39] examined mitochondrial function during thermal acclimation in mayfly larvae (Baetis and Drunella spp.). Modifications in mitochondrial or nuclear-encoded genes can modulate mitochondrial function and underlie environmental adaptation. Environmentally induced plasticity in mitochondrial function is also common, especially in response to thermal acclimation in aquatic systems. Havird et al. [39] examined mitochondrial activity in mayfly larvae from high and low elevation mountain streams during thermal acclimation to ecologically relevant temperatures. They evaluated different respiratory states in isolated mitochondria, and cytochrome oxidase and citrate synthase activities. The data obtained suggest that montane insects may be more vulnerable to rapid climate change. Indeed, mitochondria from samples collected at a low elevation site, with highly variable temperatures, showed greater thermal tolerance than samples from a high elevation site with comparatively stable temperatures, according to predictions of the climate variability hypothesis. The authors discuss how mitochondrial phenotypes are more resilient than whole-organism phenotypes in the face of thermal stress and underline the complex relationships between mitochondrial and organismal genotypes, phenotypes and environmental adaptation.

Elbassiouny et al. [40] discuss adaptations for elevated metabolic burden in electric fishes. Indeed, the interest in understanding molecular adaptations that enable electric fishes to generate and detect electric fields relies also on the extent of energetic burden required that accounts for up to 20% of fish daily energy expenditure. Elbassiouny et al. [40] investigated the molecular evolution of the OXPHOS complexes in the two most diverse clades of weakly electric fishes-South American Gymnotiformes and African Mormyroidea-using codon-based likelihood approaches. From the data obtained, they suggest that the usual strong constraint on mitochondrial OXPHOS variation is significantly reduced in electric compared to non-electric fishes, particularly for some OXPHOS complexes. The results presented are consistent with positive selection on the two fish branches associated with the independent evolutionary origins of electrogenesis, so the authors suggest that adaptive evolution in the OXPHOS machinery may be associated with the evolution of bioelectrogenesis. This evidence corroborates other findings consistent with positive selection associated with major changes in physiology or ecology, such as at the origins of bats and the evolution of powered flight: these studies highlight the utility of comparative analyses to reveal the molecular basis of adaptations that appear to be important in the evolution of novel sensory systems.

Mackenzie & Kundariya [41] review plant adaptation and phenotypic plasticity involving organelle-mediated epigenetic reprogramming. Plants can disperse their progeny to different environments and they can incorporate epigenetics and transgenerational stability, thus allowing a high level of resilience. These genetic network and chromatin features increase acclimation opportunity and allow these sessile organisms to survive environmental change. Interestingly, some of such adaptational versatility of plants arises from neofunctionalization of organelles and organellar proteins. Mackenzie & Kundariya [41] describe plastid specialization and multi-functional organellar protein features that support

7

and enhance plant phenotypic plasticity. Spatio-temporal regulation of plastid composition, unusual inter-organellar protein targeting and retrograde signalling can facilitate multi-functionalization of existing proteins. The process of neofunctionalization of sequences transferred from organelles to the nucleus is discussed, since the evolution of mitochondria and plastids as highly specialized cellular compartments has increased the functional versatility of many nuclear-encoded organellar proteins by virtue of their dual targeting: the redirection of a protein to a new cellular location can indeed influence protein neofunctionalization. The authors also refer to cytoplasmic male sterility (CMS), a mechanism described in over 80 plant species, associated with the expression of novel mitochondrial genes arising from intragenic recombination.

(d) Future perspectives

This section focuses on new and old challenges in mitochondrial biology, reviewing up-and-coming technologies that will improve our ability to study the link between genotype and phenotype by, for example, mtDNA editing and manipulation of mitochondrial gene expression. Finally, we point out the importance of investigating a wider range of biodiversity by enhancing basic, 'curiosity driven' research and applying the comparative approach.

Klucnika & Ma [42] discuss the challenge of mapping and editing animal mitochondrial genomes. Sequence variation among mtDNA haplotypes influences traits such as health and longevity, but also incurable mitochondrial diseases, ageing and cancer. However, significant challenges hamper our ability to precisely map mtDNA variants responsible for traits, and to genetically modify mtDNA. Klucnika & Ma [42] review the efforts in developing systems to map and edit mtDNA, such as how to induce/increase the basal recombination frequency and how to use mito-nucleases to cut endogenous genome and cause their subsequent degradation. The authors also discuss the use of in vitro modified mtDNA directly delivered for transformation-but no mtDNA transformation metazoan system has been established so far-and the use of cell models for creating mutants that would otherwise be homoplasmic lethal at the organismal or tissue level. The impossibility of reliably delivering nucleic acids into animal mitochondria is a huge barrier, also preventing us, for example, from importing RNA for CRISPR-mediated mtDNA editing: the establishment of a mitochondria-adapted CRISPR-Cas9 platform could prompt a revolution in mitochondrial genome engineering and our biological understanding of mitochondria and mtDNA.

Wallis *et al.* [43] review new methods for studying the genotype–phenotype link by manipulating mitochondrial gene expression with engineered proteins. Many genome engineering tools used for nuclear genome modification cannot be used to study mitochondrial genetics owing to the unusual structure and physiology of the mitochondrial

genome. Although challenges in the manipulation of mitochondria persist, new approaches are developed to modify the levels of mutant mammalian mitochondrial DNA and mitochondrial RNAs. Wallis *et al.* [43] review methods such as restriction enzymes targeted to mitochondria (mitoREs), zinc finger proteins fused with a nuclease targeted to mitochondria (mtZFNs), transcription activator-life effectors fused with a nuclease targeted to mitochondria (mitoTALENs) and RNA-binding proteins engineered to target specific mitochondrial RNA—that allow us to manipulate mtDNA, to modulate mitochondrial gene expression and to track and visualize mitochondrial processes, and whose application and study may provide highly specific and customizable genetic tools that could be applied in future therapeutics.

Milani & Ghiselli [29] ponder the potentials of non-model systems in mitochondrial research, highlighting promising candidates. The concept of the model organism is discussed, starting from the words by August Krogh, 1929—'For a large number of problems there will be some animal of choice or a few such animals on which it can be most conveniently studied' ([48], p. 247). Model organisms and inductive reasoning are irreplaceable, but we have to face the problem of overgeneralization. How can we infer general concepts? The role of model organisms in comparative biology is discussed in terms of a model organism-based approach versus comparative method. In doing this, some concepts from philosophy already used in scientific disciplines are used, such as nomothetics, ideographics and an unusual concept of class. Several animals are emerging as models in mitochondrial research: killifish (Fundulus spp. and Nothobranchius furzeri), deer mice (Peromyscus spp.), naked molerats (Heterocephalus glaber), bats of the genus Myotis, the bird Eopsaltria australis, the crustacean Tigriopus californicus and bivalve molluscs are currently used to answer specific biological questions such as the role of mitochondria in ageing and environmental adaptation, mitonuclear interactions and coevolution, genomic conflicts, mitochondrial heteroplasmy and inheritance.

Data accessibility. This article has no additional data.

Authors' contributions. F.G. and L.M. contributed equally to this work. Competing interests. We declare we have no competing interests.

Funding. This work was supported by the Italian Ministry of Education, University and Research (MIUR) SIR Programme grant no. RBSI14G0P5 funded to L.M., MIUR FIR2013 Programme grant no. RBFR13T97A and the Canziani bequest funded to F.G., and 'Ricerca Fondamentale Orientata' (RFO) funding from the University of Bologna to L.M. and F.G.

Acknowledgements. We gratefully thank: all the authors who submitted a contribution for this issue; all the reviewers involved; the Senior Commissioning Editor Helen Eaton for her amazing job in coordinating everything; the Editorial Board for the suggestions to the initial proposal; William F. Martin and John F. Allen for guidance. The involvement of authors in this issue was driven by personal interactions at several international conferences from 2016 to 2018. We apologize to the many outstanding authors in the field of mitochondrial evolutionary biology who were not included.

References

- Lane N, Martin W. 2010 The energetics of genome complexity. *Nature* 467, 929–934. (doi:10.1038/ nature09486)
- Martin WF, Garg S, Zimorski V. 2015 Endosymbiotic theories for eukaryote origin. *Phil. Trans. R. Soc. B* 370, 20140330. (doi:10.1098/rstb.2014.0330)
- Timmis JN, Ayliffe MA, Huang CY, Martin W. 2004 Endosymbiotic gene transfer: organelle genomes forge eukaryotic chromosomes.

Nat. Rev. Genet. 5, 123-135. (doi:10.1038/ nrg1271)

- 4. Sloan DB, Warren JM, Williams AM, Wu Z, Abdel-Ghany SE, Chicco AJ, Havird JC. 2018 Cytonuclear integration and co-evolution. Nat. Rev. Genet. 19, 635-648. (doi:10.1038/s41576-018-0035-9)
- 5. Breton S, Milani L, Ghiselli F, Guerra D, Stewart DT, Passamonti M. 2014 A resourceful genome: updating the functional repertoire and evolutionary role of animal mitochondrial DNAs. Trends Genet. 30, 555-564. (doi:10.1016/j.tig.2014.09.002)
- 6. Allen JF. 1993 Control of gene expression by redox potential and the requirement for chloroplast and mitochondrial genomes. J. Theor. Biol. 165, 609-631. (doi:10.1006/jtbi.1993.1210)
- Allen JF. 2015 Why chloroplasts and mitochondria 7. retain their own genomes and genetic systems: colocation for redox regulation of gene expression. Proc. Natl Acad. Sci. USA 112, 10 231-10 238. (doi:10.1073/pnas.1500012112)
- 8. Archibald JM. 2015 Endosymbiosis and eukaryotic cell evolution. Curr. Biol. 25, R911-R921. (doi:10. 1016/j.cub.2015.07.055)
- 9. Lane N, Martin WFF, Raven JAA, Allen JFF. 2013 Energy, genes and evolution: introduction to an evolutionary synthesis. Phil. Trans. R. Soc. B 368, 20120253. (doi:10.1098/rstb.2012.0253)
- 10. Lane N. 2011 Mitonuclear match: optimizing fitness and fertility over generations drives ageing within generations. Bioessays 33, 860-869. (doi:10.1002/ bies.201100051)
- 11. Wolff JN, Ladoukakis ED, Enríquez JA, Dowling DK. 2014 Mitonuclear interactions: evolutionary consequences over multiple biological scales. Phil. Trans. R. Soc. B 369, 20130443. (doi:10.1098/rstb.2013.0443)
- 12. Aanen DK, Spelbrink JN, Beekman M. 2014 What cost mitochondria? The maintenance of functional mitochondrial DNA within and across generations. Phil. Trans. R. Soc. B 369, 20130438. (doi:10.1098/ rstb.2013.0438)
- 13. Ballard JWO, Melvin RG. 2010 Linking the mitochondrial genotype to the organismal phenotype. Mol. Ecol. 19, 1523–1539. (doi:10.1111/j.1365-294X.2010.04594.x)
- 14. Dowling DK. 2014 Evolutionary perspectives on the links between mitochondrial genotype and disease phenotype. Biochim. Biophys. Acta 1840, 1393–1403. (doi:10.1016/j.bbagen.2013.11.013)
- 15. Milani L, Ghiselli F. 2015 Mitochondrial activity in gametes and transmission of viable mtDNA. Biol. Direct 10, 22. (doi:10.1186/s13062-015-0057-6)
- 16. Allio R, Donega S, Galtier N, Nabholz B. 2017 Large variation in the ratio of mitochondrial to nuclear mutation rate across animals: implications for genetic diversity and the use of mitochondrial DNA as a molecular marker. Mol. Biol. Evol. 34, 2762-2772. (doi:10.1093/molbev/msx197)
- 17. Busch KB, Kowald A, Spelbrink JN. 2014 Quality matters: how does mitochondrial network dynamics and guality control impact on mtDNA integrity? Phil. Trans. R. Soc. B 369, 20130442. (doi:10.1098/rstb.2013.0442)
- 18. Ghiselli F, Milani L, Guerra D, Chang PL, Breton S, Nuzhdin SV, Passamonti M. 2013 Structure, transcription, and variability of metazoan

mitochondrial genome: perspectives from an unusual mitochondrial inheritance system. Genome Biol. Evol. 5, 1535-1554. (doi:10.1093/gbe/evt112)

- 19. Chan DC. 2006 Mitochondria: dynamic organelles in disease, aging, and development. Cell 125, 1241-1252. (doi:10.1016/j.cell.2006.06.010)
- 20. Kim I, Rodriguez-Enriguez S, Lemasters JJ. 2007 Selective degradation of mitochondria by mitophagy. Arch. Biochem. Biophys. 462, 245-253. (doi:10.1016/j.abb.2007.03.034)
- 21. Twig G et al. 2008 Fission and selective fusion govern mitochondrial segregation and elimination by autophagy. EMBO J. 27, 433-446. (doi:10.1038/ sj.emboj.7601963)
- 22. Twig G, Hyde B, Shirihai OS. 2008 Mitochondrial fusion, fission and autophagy as a quality control axis: the bioenergetic view. Biochim. Biophys. Acta 1777, 1092–1097. (doi:10.1016/j.bbabio.2008.05. 001)
- 23. Youle RJ, van der Bliek AM. 2012 Mitochondrial fission, fusion, and stress. Science 337, 1062-1065. (doi:10.1126/science.1219855)
- 24. Ghiselli F et al. 2019 Natural heteroplasmy and mitochondrial inheritance in bivalve molluscs. Integr. Comp. Biol. 59, 1016-1032. (doi:10.1093/icb/icz061)
- 25. Lane N. 2012 The problem with mixing mitochondria. Cell 151, 246-248. (doi:10.1016/j. cell.2012.09.028)
- 26. Sharpley MS et al. 2012 Heteroplasmy of mouse mtDNA is genetically unstable and results in altered behavior and cognition. Cell 151, 333-343. (doi:10. 1016/j.cell.2012.09.004)
- 27. Stewart JB, Chinnery PF. 2015 The dynamics of mitochondrial DNA heteroplasmy: implications for human health and disease. Nat. Rev. Genet. 16, 530-542. (doi:10.1038/nrg3966)
- 28. Kowald A, Kirkwood TBL. 2011 Evolution of the mitochondrial fusion-fission cycle and its role in aging. Proc. Natl Acad. Sci. USA 108, 10 237-10 242. (doi:10.1073/pnas.1101604108)
- 29. Milani L, Ghiselli F. 2019 Faraway, so close. The comparative method and the potential of nonmodel animals in mitochondrial research. Phil. Trans. R. Soc. B 375, 20190186. (doi:10.1098/rstb. 2019.0186)
- 30. Havird JC et al. 2019 Beyond the powerhouse: integrating mitonuclear evolution, physiology, and theory in comparative biology. Integr. Comp. Biol. 59, 856-863. (doi:10.1093/icb/icz132)
- 31. Schaack S, Ho EKH, Macrae F. 2019 Disentangling the intertwined roles of mutation, selection and drift in the mitochondrial genome. Phil. Trans. R. Soc. B 375, 20190173. (doi:10.1098/rstb. 2019.0173)
- 32. Dubie JJ, Caraway AR, Stout MKM, Katju V, Bergthorsson U. 2019 The conflict within: origin, proliferation and persistence of a spontaneously arising selfish mitochondrial genome. Phil. Trans. R. Soc. B 375, 20190174. (doi:10.1098/rstb. 2019.0174)
- 33. Barrett A, Arbeithuber B, Zaidi A, Wilton P, Paul IM, Nielsen R, Makova KD. 2019 Pronounced somatic bottleneck in mitochondrial DNA of human hair.

Phil. Trans. R. Soc. B 375, 20190175. (doi:10.1098/ rstb.2019.0175)

- 34. Knorre DA. 2019 Intracellular quality control of mitochondrial DNA: evidence and limitations. Phil. Trans. R. Soc. B 375, 20190176. (doi:10.1098/rstb. 2019.0176)
- 35. Bettinazzi S, Nadarajah S, Dalpé A, Milani L, Blier PU, Breton S. 2019 Linking paternally inherited mtDNA variants and sperm performance. Phil. Trans. R. Soc. B 375, 20190177. (doi:10.1098/rstb. 2019.0177)
- 36. Nagarajan-Radha V, Aitkenhead I, Clancy DJ, Chown SL, Dowling DK. 2019 Sex-specific effects of mitochondrial haplotype on metabolic rate in Drosophila melanogaster support predictions of the Mother's Curse hypothesis. Phil. Trans. R. Soc. B 375, 20190178. (doi:10.1098/rstb.2019.0178)
- 37. Rand DM, Mossman JA. 2019 Mitonuclear conflict and cooperation govern the integration of genotypes, phenotypes and environments. Phil. Trans. R. Soc. B 375, 20190188. (doi:10.1098/rstb. 2019.0188)
- 38. Camus MF, O'Leary M, Reuter M, Lane N. 2019 Impact of mitonuclear interactions on life-history responses to diet. Phil. Trans. R. Soc. B 375, 20190416. (doi:10.1098/rstb.2019.0416)
- 39. Havird JC, Shah AA, Chicco AJ. 2019 Powerhouses in the cold: mitochondrial function during thermal acclimation in montane mayflies. Phil. Trans. R. Soc. B 375, 20190181. (doi:10.1098/rstb.2019.0181)
- 40. Elbassiouny AA, Lovejoy NR, Chang BSW. 2019 Convergent patterns of evolution of mitochondrial oxidative phosphorylation (OXPHOS) genes in electric fishes. Phil. Trans. R. Soc. B 375, 20190179. (doi:10.1098/rstb.2019.0179)
- 41. Mackenzie SA, Kundariya H. 2019 Organellar protein multi-functionality and phenotypic plasticity in plants. Phil. Trans. R. Soc. B 375, 20190182. (doi:10. 1098/rstb.2019.0182)
- 42. Klucnika A, Ma H. 2019 Mapping and editing animal mitochondrial genomes: can we overcome the challenges? Phil. Trans. R. Soc. B 375, 20190187. (doi:10.1098/rstb.2019.0187)
- 43. Wallis CP, Scott LH, Filipovska A, Rackham O. 2019 Manipulating and elucidating mitochondrial gene expression with engineered proteins. Phil. Trans. R. Soc. B 375, 20190185. (doi:10.1098/rstb.2019.0185)
- 44. Frank SA, Hurst LD, 1996 Mitochondria and male disease. Nature 383, 224. (doi:10.1038/383224a0)
- 45. Gemmell NJ, Metcalf VJ, Allendorf FW. 2004 Mother's curse: the effect of mtDNA on individual fitness and population viability. Trends Ecol. Evol. **19**, 238–244. (doi:10.1016/j.tree.2004.02.002)
- 46. James JE, Piganeau G, Eyre-Walker A. 2016 The rate of adaptive evolution in animal mitochondria. Mol. Ecol. 25, 67-78. (doi:10.1111/mec.13475)
- 47. Meiklejohn CD, Montooth KL, Rand DM. 2007 Positive and negative selection on the mitochondrial genome. Trends Microbiol. 23, 259-263. (doi:10. 1016/j.tig.2007.03.008)
- 48. Krogh A. 1929 The progress of physiology. Am. J. Physiol.-Legacy Content 90, 243-251. (doi:10.1152/ ajplegacy.1929.90.2.243)