



Glioblastoma Cellular Origin and the Firework Pattern of Cancer Genesis from the Subventricular Zone

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Glioblastoma (GBM) is a disease without any definite cure. Numerous approaches have been tested in efforts to conquer this brain disease, but patients invariably experience recurrence or develop resistance to treatment. New surgical tools, carefully chosen samples, and experimental methods are enabling discoveries at single-cell resolution. The present article reviews the cell-of-origin of isocitrate dehydrogenase (IDH)-wildtype GBM, beginning with the historical background for focusing on cellular origin and introducing the cancer genesis patterned on firework. The authors also review mutations associated with the senescence process in cells of the subventricular zone (SVZ), and biological validation of somatic mutations in a mouse SVZ model. Understanding GBM would facilitate research on the origin of other cancers and may catalyze the development of new management approaches or treatments against IDH-wildtype GBM.

Key Words : Cancer genesis · Cell-of-origin · Clock-like mutation · Firework pattern · Glioblastoma · Subventricular zone.

INTRODUCTION

Isocitrate dehydrogenase (IDH)-wildtype glioblastoma (GBM) is a disease with a dismal prognosis, a distinction that has led to GBM being dubbed the “emperor of all cancers”. The disease progression is rapid, and even with standard therapy⁵³⁾ and supra-total resection⁵²⁾, the median survival of patients from diagnosis is approximately 14–20 months^{53,63)}.

There are continuing efforts by researchers in diverse disci-

plines, including epidemiology and molecular biology, to identify the cause of GBM. Epidemiological research on GBM suggests numerous risk factors that should be taken into account, such as irradiation of the head^{18,23,46,50,55)} and the allergic history of patient⁴⁾. Biological approaches for determining the cause of GBM include attempts to identify molecular signatures in patient samples (e.g., DNA mutations)⁶¹⁾, the use of *in vivo* mouse models⁵⁸⁾, and investigation of progenitor cells of GBM^{3,13,43)}.

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This article reviews the proof-of-concept that GBM originates in the subventricular zone (SVZ) from the microscale perspective such as a molecular biology with a cellular level resolution. The first sections present a brief overview of the disease and recapitulate the importance of mutations in the development of GBM. The remaining sections describe the cell-of-origin of GBM and the cancer genesis patterned on firework from SVZ⁴³ and discuss a new hypothesis driving GBM research.

HISTORY : PROGRESS TOWARD THE STEM CELL ORIGIN OF GBM

There are plenty of discussions and arguments about the existence of the cell-of-origin from the early era of neuro-oncology whether the disease originates from embryonic cells or dedifferentiated cells⁵¹. The brain cancer model induced from the mutation of specific cells of brain is blurring the gap between those two sides² by leading an edge of this field with a subsequent drug discovery targeting GBM⁵⁸.

The cell-of-origin of GBM has been suspected to have characteristics of a stem cell, and this cell type has also been regarded as the source of tumor recurrence²². There have been many trials conducted to isolate such treatment-resistant insidious cells from the tumor tissues^{3,34,41}. Those cells were labeled as tumorspheres^{27,35,36} and their behaviors and characteristics have been validated from multiple sources^{14,25,30,39,42,49,64}. As the patient-derived tumor cells are heterogeneous in the sample dimension (patient age, the size and location of the sample in the original tumor mass, sample control, purity of samples, or previous treatments), the mutation-induced tumor model has been accepted as another robust and predictable platform in the repeated experiments and clearly demonstrates the consequences of mutation^{1,20,43} better than the orthotopic xenograft by primary tumorsphere^{31,35,38,42}.

An epiphany came from the experience of 5-aminolevulinic acid (5-ALA) fluorescence without any tumor cells in the tumor specimen by pathologic examinations⁴⁵. During surgery to remove GBM, high fluorescence of 5-ALA was found in the walls of ventricles⁴⁵, which are non-coincidentally exposed after supra-total resection (or a so-called planned lobectomy)^{52,53}. Moon et al.⁴⁵ described interesting patterns of 5-ALA fluorescence in the ventricular wall of brain tumor patients.

What they found the most interesting was the ventricular walls with positive 5-ALA fluorescence without magnetic resonance imaging enhancement and no tumor cells in the pathology, a finding that perplexed surgeons⁴⁵.

Concurrence of 5-ALA positive tissues with no tumor cells in the ventricle sample⁴⁵ helped and guided us to find the origin of GBM in the *in vivo* model⁴³. This 5-ALA glittering tumor negative samples from the ventricular wall or the SVZ has a pattern of mutations that has not been previously discovered. By comparing those mutation patterns, a novel way of understanding the disease could be revealed. The tumor-private mutations in the tumor tissue might facilitate the development of a new targeted therapy. In addition, the anatomical location of those cells would mature the Big Bang theory^{7,60} into the Firework theory of subventricular abnormal cells⁴³.

ETIOLOGY

Epidemiologic studies have linked a number of pollutants to the incidence of GBM⁴⁸. Ionizing radiation to the head at a younger age increases the risk of glioma and meningioma^{8,23,46,50,55}. An allergic or atopic disease has been associated with a lower risk of GBM, and consistently with lower-grade glioma⁴. DNA mutations and its consequent syndromes increase the risk of acquiring GBM^{32,48}, such as Turcot syndrome (from mismatch repair-deficiency)¹⁵, Li-Fraumeni syndrome (from the germline mutation of *TP53*)³⁷ and neurofibromatosis type 1 (from *NFI* gene)^{26,59}.

However, there are additional unproven and controversial risk factors. One example is non-ionizing radiation from cellular phones. Studies have not shown a consistent increase in the risk of glioma²⁴. Another example is Radon exposure which reported to be associated with the risk of glioma in a Danish cohort⁹, but neither radon exposure nor background radiation was associated with glioma incidence in a cohort of British children²⁸. Inconsistent results have been obtained with respect to the association of GBM development with exposure to air pollutants or particulate matter, owing in part to difficulties in measuring exposure history⁵. There is hardly any or no evidence linking insecticides, rubber processing, pesticides, farming, solvents, metal fumes, or jet engine manufacturing to the incidence of glioma^{48,54}.

IDENTIFICATION OF SOMATIC MUTATIONS IN SVZ STEM CELLS

Two senescence-associated mutational patterns were found in GBM : mutational signatures 1 and 5⁴³). These two types dominated the mutation profile of SVZ samples of the astrocytic ribbon (SVZ[‡] in Fig. 1A, 33% [7.8 genes out of 23 total genes] and 45% [10.4 genes out of 23 total genes] for signature 1 and 5 respectively in the SVZ samples)⁴³).

The average number of genes for mutation signature 1 jumped up from 7.8 genes (SVZ) to 81.9 genes in tumor samples of the Severance GBM cohort (10.5 fold increase)⁴³). While the fold change of average number of genes that contribute to mutation signature 5 (fold 0.71, calculated from 10.4 genes of SVZ to 7.4 genes of the tumor) remains relatively stable as that of other signatures (fold 1.18, calculated from 4.8 genes of SVZ

to 5.7 genes in the tumor)⁴³). These findings suggest that GBM arises from the senescence process, especially if we focus on the number of genes of mutational signature 1 and 5 from SVZ to tumor, or alternatively from other causes that are associated with age.

DIRECTION OF CLONAL EVOLUTION

Accumulation of mutations in both the stem cells and the tumor region raises questions regarding the origin of the mutations. Are mutations in both tumors and SVZ related? Are they coincidental? Do mutations in the SVZ reflect contamination from the tumor? To deconvolute this complex pattern of mutations, the authors compared shared mutations (i.e., those in common to both the SVZ and tumor) and tumor-ex-

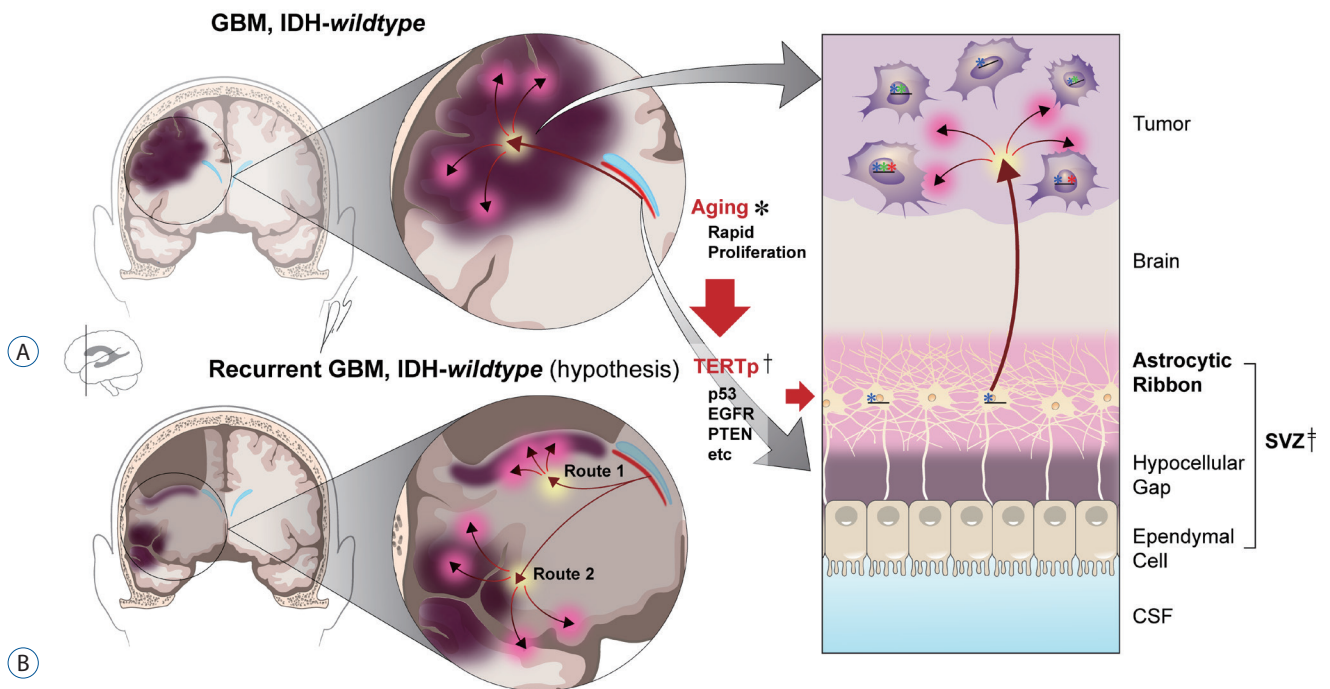


Fig. 1. Cell-of-origin of IDH-wildtype GBM and GBM genesis mechanism. A : Illustration of IDH-wildtype GBM genesis from the SVZ⁴³). Left : gross structure of the tumor in the brain. Red arrow (yellow background in the arrowhead) indicates migrating GBM origin cell from SVZ (red line under the sky blue lateral ventricle) to the hemisphere. Red arrow with a pink background in the arrowhead indicates the explosion (or evolutionary increase in heterogeneity) of tumor cells that creates the tumor bulk. Right : schematic overview of tumorigenesis from the SVZ to the cortex (below → top). Some portion of cells in the astrocytic band in the SVZ (†) acquire mutations (illustrated with asterisks with blue in the SVZ; additional mutations are depicted in cells of the GBM in green and red asterisks) as an incidental consequence of senescence (*), with mutations in the *TERT* promoter serving as the possible initial step (†), followed by mutations in *TP53*, *PTEN* and *EGFR*, among others. B : Illustration of the hypothetical SVZ origin of IDH-wildtype GBM recurrence via route 1 or 2. Route 1 indicates same site recurrence, while route 2 indicates different site recurrence from the GBM cell-of-origin in the SVZ. GBM : glioblastoma, IDH : isocitrate dehydrogenase, TERTp : promoter of telomerase reverse transcriptase (Gene symbol : TERT), p53 : tumor protein p53 (Gene symbol : TP53), EGFR : epidermal growth factor receptor, PTEN : phosphatase and tensin homolog, CSF : cerebrospinal fluid, SVZ : subventricular zone.

clusive mutations (i.e., those not found in the SVZ)⁴³. There were several shared mutations in the tumor-free SVZ and GBM bulk tumor, including those in the *TERT* promoter⁴³. An analysis of variant allele frequencies (VAFs) and single-cell sequencing results revealed a unidirectional pattern from SVZ to bulk tumor. A two-dimensional VAF plot in a previous report⁴³ showed unique patterns of shared mutations between the tumor and the SVZ, with low-level driver mutations being located near the y-axis or tumor-axis. However, the tumor-contaminated SVZ-tumor pair showed more genes that are spread along the identity line as opposed to the uncontaminated SVZ-tumor pair⁴³ (Fig. 1A).

Single-cell sequencing of the DNA of bulk tumors also validated the result showing directionality from the SVZ to the tumor⁴³. Twenty-four clones were derived from one IDH-wildtype GBM patient (GBM245)⁴³. The genes on the table of Lee et al.⁴³ are subset of those genes of directionality. This surprising pattern of mutation was later confirmed from the intraventricular injection of plasmids to induce mutation in the SVZ^{16,43}.

PROGRESSION OF MUTATIONS IN GBM FROM THE SVZ

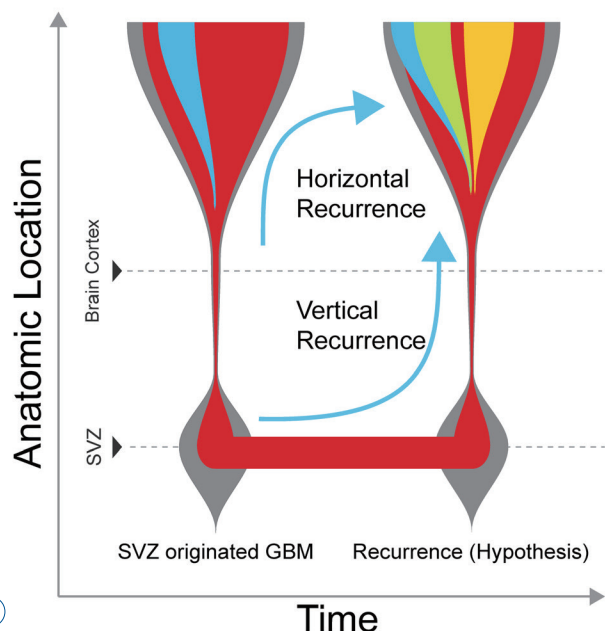
Mutations oftentimes start with *TERT* promoter mutations in cells of the astrocytic ribbon in SVZ under the ventricle ($TERT^p^+$ in Fig. 1A). With additional mutations in oncogenes, the mutated cells from SVZ migrate to the cortex and manifest as the GBM (Fig. 1A, right)⁴³. The authors searched for mutations in triple-matched samples of tumor-free SVZ, GBM tumor, and normal brain cortex (or blood)⁴³. The authors have hypothesized that the SVZ, which includes neural stem cells, is suspected of being the origin of tumors⁴³; mutations then progress from the SVZ to the tumor mass. In addition to this directionality, the number of mutations increases dramatically after the cell-of-origin reaches the cortex. As described in the previous study of Lee et al.⁴³, about 20 different somatic mutations of SVZ that contribute to the mutational patterns increased to around 100 mutations in the tumor. Furthermore, mutational signature 1 dominated most mutations in tumors, whereas mutational signature 5 was preserved

Firework pattern GBM genesis (metaphor)



A

Firework pattern GBM genesis (scheme)



B

Fig. 2. Firework pattern of IDH-wildtype GBM genesis. A : Artistic illustration of IDH-wildtype GBM originating from the SVZ (colored in gold). Each firework trail corresponds to a different cancer clone. In this metaphorical depiction, the SVZ is represented as a cannon on the ground, denoting the starting point of GBM genesis. B : Conceptual illustration of the time line of the genesis of the firework pattern of IDH-wildtype GBM. Horizontal recurrence (or classical model) : GBM recurs from SVZ (red) → tumor (blue) → recurrence (green), vertical recurrence (hypothetical model) : primary GBM originates from SVZ (red) → tumor (blue), and it recurs from SVZ (red) → recurrent tumor (orange). GBM : glioblastoma, SVZ : subventricular zone, IDH : isocitrate dehydrogenase.

at about the same level between the SVZ and the tumor⁴³). This pattern is slightly different version of the Big Bang concept and, considering the anatomic location of the progression, the authors refer to it as a “Firework” pattern (Fig. 2).

BIOLOGICAL VALIDATION

Deep under the ependymal layer of the SVZ is a ribbon-like feature called the astrocytic ribbon⁵⁷ (SVZ^z in Fig. 1A). This area is populated by neural stem cells in both mouse and human brains⁶⁵. Using molecular techniques described below, we introduced three types of mutations into the population of cells in the SVZ. These mutation-bearing cells migrated to the cortex, ultimately forming high-grade gliomas⁴³ (Fig. 1A), presenting with firework-like GBM genesis (Fig. 2A).

A mass of GBM is created in the cortex by the mutation in the SVZ cells⁴³. Crossbred mice (postnatal day 2–3 pups) carrying LSL-tdTomato and LSL-EGFRviii⁷¹ were used for these experiments⁴³. A Cre-containing CRISPR/Cas9 plasmid was electroporated into one side of the ventricle in the brain to knock out *Pten* and *Trp53* (mouse analog of human *TP53*) genes using single guide RNA (sgRNA) for these genes. After injection into the ventricular space with a sharp needle, the electroporation device was applied to deliver the plasmid into the SVZ⁴³. In the presence of the mutations, brain tumors developed in 90% of mice (9/10), with 67% of tumors developing in a region distant from the mutation-arising SVZ, and these mice showed a median survival of around 20 weeks⁴³.

Tumorigenesis, or Cancer genesis from the triple mutations marks the important step in the research of GBM⁶¹. By this finding⁴³, more subtle nature of IDH-wildtype GBM can be revealed in the subsequent researches.

HYPOTHESIS : RECURRENCE OF GBM FROM THE SVZ

One of the next steps in the research of IDH-wildtype GBM should be related with the recurrence pattern of GBM (The hypothesis on the recurrence of GBM). Careful selection of samples without tumor in the SVZ is the beginning point of this research. Recent article by Watts group⁶¹ is one example of selecting the tumor-invaded SVZ (median tumor content

22.1%, rather than tumor-free SVZ) by the IDH-wildtype GBM and its results and implications should be carefully contemplated in the research of vertical recurrence from the cell of SVZ (Fig. 2B).

GBM recurs after irradiation of tumor margins at varying depth⁶⁹. Although irradiating the SVZ appears to have a prognostic benefit for GBM patients^{29,47}, it does not cure these patients or prevent exacerbation of symptoms. There are at least two scenarios for recurrence of IDH-wildtype GBM. In the first case, GBM-initiating cells already present in the parenchyma of the brain restart the tumor after a dormant period (Fig. 2B, horizontal recurrence). In the second, dormant cells in the SVZ re-migrate to the tumor bed area, where they create the recurrent tumor (Fig. 1B, the only hypothesis in this article; Fig. 2B, vertical recurrence). We hope to resolve the second phenomenon using tumor-free SVZ samples from GBM patients.

FUTURE : DEVELOPMENT OF THERAPEUTICS TARGETING THE SVZ

Many previous attempts to treat GBM have failed and the list harbors 5-fluorouracil and methotrexate¹⁰. A breakthrough in treating GBM came with the application of alkylating agents^{18,62,67}. Nitrosoureas proved the most effective at the time, and it became the most commonly used drugs for the management of brain tumors, including GBM^{56,70}. Their antitumor effects were widely accepted and even led to the development of an implantable chemotherapeutic system impregnated with the nitrosourea derivative carmustine¹¹.

However, these nitrosoureas had toxicity issues, and temozolomide subsequently gained popularity because of its limited adverse effects compared with earlier alkylating agents⁶. But even temozolomide proved to be ineffective in treatment of recurrent GBM similarly to other alkylating agents^{33,44,66}. And the therapeutic efficacy of other novel options is limited in Bevacizumab^{19,21,68}, IDH-targeting drugs¹² and immunotherapy^{17,40}.

After the repeated efforts to treat the tumor itself, the origin of GBM emerged as the next generation target. Ultimately, *in vitro* and *in vivo* models will accelerate the screening for drugs that can preemptively target the initial process of cancer genesis with firework pattern from SVZ.

CONCLUSION

Mutated neural stem cells, that are present in the astrocytic ribbon in the SVZ, may migrate to the brain cortex and generate IDH-wildtype GBM with/or without any molecular or environmental cue⁴³). While this concept has been demonstrated scientifically, elucidating the detailed mechanism underlying the GBM genesis process will require additional investigation. Our ongoing research efforts would focus on identifying the cell-of-origin to develop a potential treatment option for IDH-wildtype GBM, the “Emperor of cancer”.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

INFORMED CONSENT

This type of study does not require informed consent.

AUTHOR CONTRIBUTIONS

Conceptualization : SJY, JHM, EHK, JHC, JHL, SGK

Data curation : SJY, SGK

Funding acquisition : SGK

Methodology : SJY, HJK, JHL, RJC, JKS, JHL, SGK

Project administration : SGK

Visualization : SJY, DSJ, SGK

Writing - original draft : SJY, SGK

Writing - review & editing : SJY, JP, HJK, JHL, EJ, RJC, JKS, JHM, EHK, JHC, JHL, SGK

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References

- Alcantara Llaguno S, Chen J, Kwon CH, Jackson EL, Li Y, Burns DK, et al. : Malignant astrocytomas originate from neural stem/progenitor cells in a somatic tumor suppressor mouse model. **Cancer Cell** **15** : 45-56, 2009
- Alcantara Llaguno S, Sun D, Pedraza AM, Vera E, Wang Z, Burns DK, et al. : Cell-of-origin susceptibility to glioblastoma formation declines with neural lineage restriction. **Nat Neurosci** **22** : 545-555, 2019
- Alcantara Llaguno SR, Wang Z, Sun D, Chen J, Xu J, Kim E, et al. : Adult lineage-restricted cns progenitors specify distinct glioblastoma subtypes. **Cancer Cell** **28** : 429-440, 2015
- Amirian ES, Zhou R, Wrensch MR, Olson SH, Scheurer ME, Il'yasova D, et al. : Approaching a scientific consensus on the association between allergies and glioma risk: a report from the glioma international case-control study. **Cancer Epidemiol Biomarkers Prev** **25** : 282-290, 2016
- Andersen ZJ, Pedersen M, Weinmayr G, Stafoggia M, Galassi C, Jørgensen JT, et al. : Long-term exposure to ambient air pollution and incidence of brain tumor: the european study of cohorts for air pollution effects (escape). **Neuro Oncol** **20** : 420-432, 2018
- Bae SH, Park MJ, Lee MM, Kim TM, Lee SH, Cho SY, et al. : Toxicity profile of temozolomide in the treatment of 300 malignant glioma patients in korea. **J Korean Med Sci** **29** : 980-984, 2014
- Barthel FP, Wesseling P, Verhaak RGW : Reconstructing the molecular life history of gliomas. **Acta Neuropathol** **135** : 649-670, 2018
- Braganza MZ, Kitahara CM, Berrington de González A, Inskip PD, Johnson KJ, Rajaraman P : Ionizing radiation and the risk of brain and central nervous system tumors: a systematic review. **Neuro Oncol** **14** : 1316-1324, 2012
- Bräuner EV, Andersen ZJ, Andersen CE, Pedersen C, Gravesen P, Ulbak K, et al. : Residential radon and brain tumour incidence in a danish cohort. **PLoS One** **8** : e74435, 2013
- Bredel M : Anticancer drug resistance in primary human brain tumors. **Brain Res Brain Res Rev** **35** : 161-204, 2001
- Brem H, Piantadosi S, Burger PC, Walker M, Selker R, Vick NA, et al. : Placebo-controlled trial of safety and efficacy of intraoperative controlled delivery by biodegradable polymers of chemotherapy for recurrent gliomas. The polymer-brain tumor treatment group. **Lancet** **345** : 1008-1012, 1995
- Calvert AE, Chalastanis A, Wu Y, Hurley LA, Kouri FM, Bi Y, et al. : Cancer-associated idh1 promotes growth and resistance to targeted therapies in the absence of mutation. **Cell Rep** **19** : 1858-1873, 2017
- Chen J, McKay RM, Parada LF : Malignant glioma: lessons from genomics, mouse models, and stem cells. **Cell** **149** : 36-47, 2012
- Choi J, Lee JH, Koh I, Shim JK, Park J, Jeon JY, et al. : Inhibiting stem-

- ness and invasive properties of glioblastoma tumorsphere by combined treatment with temozolomide and a newly designed biguanide (hl156a). **Oncotarget 7** : 65643-65659, 2016
15. Dipro S, Al-Otaibi F, Alzahrani A, Ulhaq A, Al Shail E : Turcot syndrome: a synchronous clinical presentation of glioblastoma multiforme and adenocarcinoma of the colon. **Case Rep Oncol Med 2012** : 720273, 2012
 16. Fernández ME, Croce S, Boutin C, Cremer H, Raineteau O : Targeted electroporation of defined lateral ventricular walls: a novel and rapid method to study fate specification during postnatal forebrain neurogenesis. **Neural Dev 6** : 13, 2011
 17. Filley AC, Henriquez M, Dey M : Recurrent glioma clinical trial, checkmate-143: the game is not over yet. **Oncotarget 8** : 91779-91794, 2017
 18. Fine HA : The basis for current treatment recommendations for malignant gliomas. **J Neurooncol 20** : 111-120, 1994
 19. Franceschi E, Lamberti G, Paccapelo A, Di Battista M, Genestreti G, Minichillo S, et al. : Third-line therapy in recurrent glioblastoma: is it another chance for bevacizumab? **J Neurooncol 139** : 383-388, 2018
 20. Friedmann-Morvinski D, Bushong EA, Ke E, Soda Y, Marumoto T, Singer O, et al. : Dedifferentiation of neurons and astrocytes by oncogenes can induce gliomas in mice. **Science 338** : 1080-1084, 2012
 21. Gilbert MR, Dignam JJ, Armstrong TS, Wefel JS, Blumenthal DT, Vogelbaum MA, et al. : A randomized trial of bevacizumab for newly diagnosed glioblastoma. **N Engl J Med 370** : 699-708, 2014
 22. Goffart N, Kroonen J, Rogister B : Glioblastoma-initiating cells: relationship with neural stem cells and the micro-environment. **Cancers (Basel) 5** : 1049-1071, 2013
 23. Inskip PD, Sigurdson AJ, Veiga L, Bhatti P, Ronckers C, Rajaraman P, et al. : Radiation-related new primary solid cancers in the childhood cancer survivor study: comparative radiation dose response and modification of treatment effects. **Int J Radiat Oncol Biol Phys 94** : 800-807, 2016
 24. INTERPHONE Study Group : Brain tumour risk in relation to mobile telephone use: results of the interphone international case-control study. **Int J Epidemiol 39** : 675-694, 2010
 25. Jeong H, Park J, Shim JK, Lee JE, Kim NH, Kim HS, et al. : Combined treatment with 2'-hydroxycinnamaldehyde and temozolomide suppresses glioblastoma tumorspheres by decreasing stemness and invasiveness. **J Neurooncol 143** : 69-77, 2019
 26. Jeong TS, Yee GT : Glioblastoma in a patient with neurofibromatosis type 1: a case report and review of the literature. **Brain Tumor Res Treat 2** : 36-38, 2014
 27. Kang SG, Cheong JH, Huh YM, Kim EH, Kim SH, Chang JH : Potential use of glioblastoma tumorsphere: clinical credentialing. **Arch Pharm Res 38** : 402-407, 2015
 28. Kendall GM, Little MP, Wakeford R, Bunch KJ, Miles JC, Vincent TJ, et al. : A record-based case-control study of natural background radiation and the incidence of childhood leukaemia and other cancers in great britain during 1980-2006. **Leukemia 27** : 3-9, 2012
 29. Khalifa J, Tensaouti F, Lusque A, Plas B, Lotterie JA, Benouaich-Amiel A, et al. : Subventricular zones: new key targets for glioblastoma treatment. **Radiat Oncol 12** : 67, 2017
 30. Kim EH, Lee JH, Oh Y, Koh I, Shim JK, Park J, et al. : Inhibition of glioblastoma tumorspheres by combined treatment with 2-deoxyglucose and metformin. **Neuro Oncol 19** : 197-207, 2017
 31. Kim S, Alexander CM : Tumorsphere assay provides more accurate prediction of in vivo responses to chemotherapeutics. **Biotechnol Lett 36** : 481-488, 2014
 32. Kinnersley B, Mitchell JS, Gousias K, Schramm J, Idbaih A, Labussière M, et al. : Quantifying the heritability of glioma using genome-wide complex trait analysis. **Sci Rep 5** : 17267, 2015
 33. Kitange GJ, Carlson BL, Schroeder MA, Grogan PT, Lamont JD, Decker PA, et al. : Induction of mgmt expression is associated with temozolomide resistance in glioblastoma xenografts. **Neuro Oncol 11** : 281-291, 2009
 34. Kong BH, Moon JH, Huh YM, Shim JK, Lee JH, Kim EH, et al. : Prognostic value of glioma cancer stem cell isolation in survival of primary glioblastoma patients. **Stem Cells Int 2014** : 838950, 2014
 35. Kong BH, Park NR, Shim JK, Kim BK, Shin HJ, Lee JH, et al. : Isolation of glioma cancer stem cells in relation to histological grades in glioma specimens. **Childs Nerv Syst 29** : 217-229, 2013
 36. Kong BH, Shin HD, Kim SH, Mok HS, Shim JK, Lee JH, et al. : Increased in vivo angiogenic effect of glioma stromal mesenchymal stem-like cells on glioma cancer stem cells from patients with glioblastoma. **Int J Oncol 42** : 1754-1762, 2013
 37. Kratz CP, Achatz MI, Brugières L, Frebourg T, Garber JE, Greer MC, et al. : Cancer screening recommendations for individuals with li-fraumeni syndrome. **Clin Cancer Res 23** : e38-e45, 2017
 38. Kwak J, Shim JK, Kim DS, Lee JH, Choi J, Park J, et al. : Isolation and characterization of tumorspheres from a recurrent pineoblastoma patient: feasibility of a patient-derived xenograft. **Int J Oncol 49** : 569-578, 2016
 39. Laks DR : **The assessment of glioblastoma tumorspheres reveals molecular determinants of proliferation and therapeutic response**. Ann Arbor : University of California, 2015, p245
 40. Lamberti G, Franceschi E, Brandes AA : The burden of oncology promises not kept in glioblastoma. **Future Neurol 13** : 1-4, 2018
 41. Lathia JD, Mack SC, Mulkearns-Hubert EE, Valentim CL, Rich JN : Cancer stem cells in glioblastoma. **Genes Dev 29** : 1203-1217, 2015
 42. Lee CH, Yu CC, Wang BY, Chang WW : Tumorsphere as an effective in vitro platform for screening anti-cancer stem cell drugs. **Oncotarget 7** : 1215-1226, 2015
 43. Lee JH, Lee JE, Kahng JY, Kim SH, Park JS, Yoon SJ, et al. : Human glioblastoma arises from subventricular zone cells with low-level driver mutations. **Nature 560** : 243-247, 2018
 44. Lee SY : Temozolomide resistance in glioblastoma multiforme. **Genes Dis 3** : 198-210, 2016
 45. Moon JH, Kim SH, Shim JK, Roh TH, Sung KS, Lee JH, et al. : Histopathological implications of ventricle wall 5-aminolevulinic acid-induced fluorescence in the absence of tumor involvement on magnetic resonance images. **Oncol Rep 36** : 837-844, 2016
 46. Neglia JP, Robison LL, Stovall M, Liu Y, Packer RJ, Hammond S, et al. :

- New primary neoplasms of the central nervous system in survivors of childhood cancer: a report from the childhood cancer survivor study. **J Natl Cancer Inst** **98** : 1528-1537, 2006
47. Nourallah B, Digpal R, Jena R, Watts C : Irradiating the subventricular zone in glioblastoma patients: is there a case for a clinical trial? **Clin Oncol (R Coll Radiol)** **29** : 26-33, 2017
 48. Ostrom QT, Bauchet L, Davis FG, Deltour I, Fisher JL, Langer CE, et al. : The epidemiology of glioma in adults: a "state of the science" review. **Neuro Oncol** **16** : 896-913, 2014
 49. Park J, Shim JK, Kang JH, Choi J, Chang JH, Kim SY, et al. : Regulation of bioenergetics through dual inhibition of aldehyde dehydrogenase and mitochondrial complex I suppresses glioblastoma tumorspheres. **Neuro Oncol** **20** : 954-965, 2018
 50. Pearce MS, Salotti JA, Little MP, McHugh K, Lee C, Kim KP, et al. : Radiation exposure from CT scans in childhood and subsequent risk of leukaemia and brain tumours: a retrospective cohort study. **Lancet** **380** : 499-505, 2012
 51. Ramon Y Cajal Agüeras S : Pío del río-hortega: a visionary in the pathology of central nervous system tumors. **Front Neuroanat** **10** : 13, 2016
 52. Roh TH, Kang SG, Moon JH, Sung KS, Park HH, Kim SH, et al. : Survival benefit of lobectomy over gross-total resection without lobectomy in cases of glioblastoma in the noneloquent area: a retrospective study. **J Neurosurg**, 2019 [Epub ahead of print]
 53. Roh TH, Park HH, Kang SG, Moon JH, Kim EH, Hong CK, et al. : Long-term outcomes of concomitant chemoradiotherapy with temozolomide for newly diagnosed glioblastoma patients: a single-center analysis. **Medicine (Baltimore)** **96** : e7422, 2017
 54. Ruder AM, Carreón T, Butler MA, Calvert GM, Davis-King KE, Waters MA, et al. : Exposure to farm crops, livestock, and farm tasks and risk of glioma: the upper midwest health study. **Am J Epidemiol** **169** : 1479-1491, 2009
 55. Sadetzki S, Chetrit A, Freedman L, Stovall M, Modan B, Novikov I : Long-term follow-up for brain tumor development after childhood exposure to ionizing radiation for tinea capitis. **Radiat Res** **163** : 424-432, 2005
 56. Salzman M : Survival in glioblastoma: historical perspective. **Neurosurgery** **7** : 435-439, 1980
 57. Sanai N, Tramontin AD, Quiñones-Hinojosa A, Barbaro NM, Gupta N, Kunwar S, et al. : Unique astrocyte ribbon in adult human brain contains neural stem cells but lacks chain migration. **Nature** **427** : 740-744, 2004
 58. Shi Y, Lim SK, Liang Q, Iyer SV, Wang HY, Wang Z, et al. : Gboxin is an oxidative phosphorylation inhibitor that targets glioblastoma. **Nature** **567** : 341-346, 2019
 59. Shibahara I, Sonoda Y, Suzuki H, Mayama A, Kanamori M, Saito R, et al. : Glioblastoma in neurofibromatosis 1 patients without idh1, braf v600e, and tert promoter mutations. **Brain Tumor Pathol** **35** : 10-18, 2018
 60. Sottoriva A, Kang H, Ma Z, Graham TA, Salomon MP, Zhao J, et al. : A big bang model of human colorectal tumor growth. **Nat Genet** **47** : 209-216, 2015
 61. Spiteri I, Caravagna G, Cresswell GD, Vatsiou A, Nichol D, Acar A, et al. : Evolutionary dynamics of residual disease in human glioblastoma. **Ann Oncol** **30** : 456-463, 2019
 62. Stenning SP, Freedman LS, Bleehen NM : An overview of published results from randomized studies of nitrosoureas in primary high grade malignant glioma. **Br J Cancer** **56** : 89-90, 1987
 63. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al. : Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. **N Engl J Med** **352** : 987-996, 2005
 64. Sung KS, Shim JK, Lee JH, Kim SH, Park S, Roh TH, et al. : Success of tumorsphere isolation from who grade IV gliomas does not correlate with the weight of fresh tumor specimens: an immunohistochemical characterization of tumorsphere differentiation. **Cancer Cell Int** **16** : 75, 2016
 65. Tabata H, Yoshinaga S, Nakajima K : Cytoarchitecture of mouse and human subventricular zone in developing cerebral neocortex. **Exp Brain Res** **216** : 161-168, 2012
 66. Tseng WL, Hsu HH, Chen Y, Tseng SH : Tumor recurrence in a glioblastoma patient after discontinuation of prolonged temozolomide treatment. **Asian J Neurosurg** **12** : 727-730, 2017
 67. Walker MD, Green SB, Byar DP, Alexander E Jr, Batzdorf U, Brooks WH, et al. : Randomized comparisons of radiotherapy and nitrosoureas for the treatment of malignant glioma after surgery. **N Engl J Med** **303** : 1323-1329, 1980
 68. Wenger KJ, Wagner M, You SJ, Franz K, Harter PN, Burger MC, et al. : Bevacizumab as a last-line treatment for glioblastoma following failure of radiotherapy, temozolomide and lomustine. **Oncol Lett** **14** : 1141-1146, 2017
 69. Wernicke AG, Smith AW, Taube S, Mehta MP : Glioblastoma: radiation treatment margins, how small is large enough? **Pract Radiat Oncol** **6** : 298-305, 2016
 70. Wilson CB, Gutin P, Boldrey EB, Drafts D, Levin VA, Enot KJ : Single-agent chemotherapy of brain tumors. A five-year review. **Arch Neurol** **33** : 739-744, 1976
 71. Zhu H, Acquaviva J, Ramachandran P, Boskovitz A, Woolfenden S, Pfannl R, et al. : Oncogenic egfr signaling cooperates with loss of tumor suppressor gene functions in gliomagenesis. **Proc Natl Acad Sci U S A** **106** : 2712-2716, 2009