

Synthesis of Silver Nanoparticles Using a Novel Cyanobacteria *Desertifilum* sp. extract: Their Antibacterial and Cytotoxicity Effects

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Reham Samir Hamida ¹
Nabila Elsayed Abdelmeguid¹
Mohamed Abdelaal Ali²
Mashaal Mohammed
Bin-Meferij³
Mahmoud Ibrahim Khalil ⁴

¹Molecular Biology Unit, Department of Zoology, Faculty of Science, Alexandria University, Alexandria, Egypt;

²Biotechnology Unit, Department of Plant Production, College of Food and Agriculture Science, King Saud University, Riyadh, Saudi Arabia; ³Department of Biology, College of Science Princess Nourah bint Abdulrahman University, Riyadh, Saudi Arabia; ⁴Department of Biological Sciences, Faculty of Science, Beirut Arab University, Beirut, Lebanon

Background: The emergence of multi drug-resistant (MDR) bacterial infections and cancer has necessitated the development and discovery of alternative eco-safe antibacterial and anticancer agents. Biogenic fabrication of metallic nanoparticles is an emerging discipline for production of nanoproducts that exert potent anticancer and antibacterial activity, and do not suffer from the limitations inherent in physicochemical synthesis methods.

Methodology: In this study, we isolated, purified, and characterized a novel cyanobacteria extract (*Desertifilum* IPPAS B-1220) to utilize in biofabrication of silver nanoparticles (D-SNPs). D-SNPs were produced by adding *Desertifilum* extract to silver nitrate solution under controlled conditions. Biofabrication of D-SNPs was confirmed using a UV-Vis spectrophotometer. The resultant D-SNPs were characterized using XRD, FTIR, SEM, and TEM. The toxicity of D-SNPs against five pathogenic bacteria and three cancer cell lines (MCF-7, HepG2, and Caco-2) was evaluated.

Results: Formation of D-SNPs was indicated by a color change from pale yellow to dark brown. The peak of the surface plasmon resonance of the D-SNPs was at 421 nm. The XRD detected the crystallinity of D-SNPs. FTIR showed that polysaccharides and proteins may have contributed to the biofabrication of D-SNPs. Under SEM and TEM, the D-SNPs were spherical with diameter ranges from 4.5 to 26 nm. The D-SNPs significantly suppressed the growth of five pathogenic bacteria, and exerted cytotoxic effects against MCF-7, HepG2, and Caco-2 cancer cells with IC₅₀ values of 58, 32, and 90 µg/mL, respectively.

Conclusion: These findings showed for the first time the potentiality of novel cyanobacteria strain *Desertifilum* IPPAS B-1220 to fabricate small SNPs that acted as potent anticancer and antibacterial material against different cancer cell lines and pathogenic bacterial strains. These findings encourage the researchers to focus on cyanobacteria in general and especially *Desertifilum* sp. IPPAS B-1220 for synthesizing different NPs that opening the window for new applications.

Keywords: nanotechnology, eco-friendly, D-SNPs, physicochemical, cytotoxic activity

Introduction

Nanoscience is developing rapidly, and many beneficial nanoproducts have been produced to benefit society.¹ Metal nanoparticles (MNPs) exhibit unique optical and magnetic properties, have high surface area to volume ratios, and have low melting points and excellent mechanical strength, which allows for use of MNPs in medicine and industry.² Moreover, these properties provide benefits in bio-imaging,³ diagnostics, and drug delivery.⁴ Among MNPs, silver nanoparticles (SNPs) have

Correspondence: Mashaal Mohammed Bin-Meferij; Mahmoud Ibrahim Khalil
Tel +966 554477376; +961 81982014
Email mmbinmufayrij@pnu.edu.sa;
m.khalil@bau.edu.lb

received the most attention due to their unique physico-chemical and biological properties.⁵

Different physicochemical routes have been used to produce nanoparticles (NPs) with a variety of shapes and sizes, but the production of toxic by-products has limited use of these NPs.⁶ Advances in NP synthesis and increased use of these particles in many fields have led researchers to evaluate mechanisms to improve the biocompatibility of NPs.^{7,8} In addition, increased use of NPs has resulted in interest in the development of green approaches for the fabrication of NPs. Biological synthesis has several advantages including reduced production of toxic by-products, improved stability, and reduced toxicity against healthy cells.^{9,10} Furthermore, green synthesis is a low-cost approach, which allows for the rapid formation of efficient NPs.¹¹ Also, comparing with the traditional methods, green methods enable the fabrication of different shapes of NPs that are distinguished with unique features under mild pressure and temperature conditions.^{9,11} Several green chemistry methods have been developed for the fabrication of NPs using plant extracts,¹² algae,¹³ fungi,¹⁴ and bacteria.¹⁵ Cyanobacteria are excellent bio-systems for the intracellular or extracellular production of NPs. However, the biofabrication of metallic NPs using cyanobacteria has not been well characterized.^{16,17} Husain et al¹⁸ used the marine cyanobacterium *Microchaete NCCU-342* to fabricate AgNO₃ into smaller SNPs. Cepoi et al¹⁹ suggested that *Spirulina Platensis* and *Nostoc linckia* had potential for biofabrication of NPs because their cells contain numerous bioactive compounds. *Desertifilum* sp. was initially collected from biological crusts and identified as a filamentous cyanobacterial genus. *Desertifilum* sp. are oscillatocean cyanobacteria, which have filaments without differentiation of heterocytes and akinetes.²⁰ These oscillatocean cells contain several bio-compounds such as pigments, proteins, and polysaccharides that can represent as potent reducing and capping materials during the formation of NPs.²¹

Overuse of antibiotics for the treatment of infectious diseases has contributed to the emergence of multidrug-resistant (MDR) bacteria.²² This issue has spurred the development of alternative neutral antibacterial agents with unique properties.²³ Advances in nanotechnology have allowed for the production of MNPs, including SNPs, that exhibit low toxicity against healthy human cells and high bactericidal activity. Silver nanoparticles have potential as an alternative to antibacterial drugs due to promising effects against MDR bacteria in vivo and in vitro.^{24,25} Silver nanoparticles have

been shown to exert a significant inhibitory activity toward gram-negative and gram-positive bacteria due to their small sizes and large surface area to volume ratios, which facilitate the extensive interactions between SNPs and bacteria.²⁶

Currently, the scientific community is progressively developing to develop and discover novel therapeutic agents to treat cancer that is considered as the second leading cause of death worldwide.^{27,28} Cancer is a complex genetic disorder characterized by uncontrolled and abnormal cell growth.²⁹ Chemotherapy and radiation are the primary treatments for tumors, but these therapies cause significant damage to normal cells.³⁰ Advances in nanotechnology have allowed for development of MNPs that can be used for detection, prevention, and inhibition of cancer with low toxicity against normal cells.³ Aygun et al³¹ showed that after treatment MDA-MB-231 and HeLa cancer cells with Pt NPs, a significant cellular growth inhibition was observed. Bin-Meferij et al³² showed that SNPs synthesized using *Nostoc* sp. exhibited significant cytotoxicity against Caco-2 cells. Silver nanoparticles fabricated using *Lyngbya majuscula* showed antiproliferative activity against leukemia cell lines (K562, MOLT3, and REH).¹⁶ Recent publication showed that biogenic SNPs exhibited higher anticancer activity against MCF-7 and lower toxicity than 5-fluorouracil against normal cells.³³

In this study, a novel cyanobacteria strain, *Desertifilum* IPPAS B-1220, was isolated, purified, cultured and genetically identified to use for green fabrication of silver NPs from silver nitrate. The resultant D-SNPs were physico-chemical characterized and their bactericidal and antitumor activity was screened using five pathogenic bacterial strains and three cancer cell lines (MCF-7, HepG2, and Caco-2).

Materials and Methods

Materials

Silver nanoparticles were synthesized using *Desertifilum* sp. extracts. Silver nitrate (AgNO₃) and cell culture materials were purchased from Sigma-Aldrich (St. Louis, MO, USA). Caco-2, MCF-7, and HepG2 were obtained from the Medical Research Institute (Alexandria, Egypt).

Cyanobacteria Isolation and Culture Conditions

Desertifilum sp. was previously isolated from a soil sample in Borg-El Arab city, Egypt, and purified by serial dilution as described by Bolch et al.³⁴ Ten microliters of each

diluted isolate was plated on BG11 nutrient agar plates and incubated at 25°C. The cultured strains were examined periodically using light and inverted microscopes to ensure that the cultured biomasses were pure.

The purified *Desertifilum* colonies were transferred to sterile 500 mL flasks containing BG-11 media (pH 7). The cultures were grown under a cool white fluorescent lamp that provided a light intensity of 2000 ± 200 Lux over a 12 h light/dark cycle at room temperature for 2 weeks.

Identification of *Desertifilum* IPPAS B-1220

sp

Morphological Examination

Light and inverted microscopes (Optika, Ponteranica BG, Italy) were used to identify the morphological features of the pure colonies, including shape, structure, and color. Taxonomic identification was performed based on morphology.³⁵

Molecular Identification

DNA Extraction

Genomic DNA of the selected strain was extracted and purified as previously described by Singh et al.³⁶ The DNA concentration was determined using a spectrophotometer (Jenway, Staffordshire, OSA, UK) at 260 nm. Agarose gel electrophoresis (0.8%; ReadyAgarose™ Precast Gel System Bio-Rad Laboratories, Inc., California, USA) was used to evaluate the integrity of the extracted genomic DNA.

Polymerase Chain Reaction and Sequencing Analyses

Polymerase Chain Reaction (PCR) was used to determine the quality of the extracted genomic DNA template, and for 16S rDNA amplification using species-specific primers. Polymerase chain reaction was performed using a thermal cycler (Multigene Optimax, Labnet International, Inc., Edison, NJ, USA). The full length 1500 bp fragment of the 16S rRNA gene from the genome of the selected cyanobacterial isolate was amplified using PCR.³⁷

The forward primer was 5'-AGAGTTTGATCMTGG CTCAG-3' (position 8 in the 16S rRNA gene according to *E. coli* numbering) and the reverse primer was 3'TACGGY ACCTTGTTACGACTT-5' (position 1514 in the 16S rRNA gene according to *E. coli* numbering). The thermal program was as follows: 4 min at 95°C followed by 30 cycles at 94°C for 1 min each (denaturation), then 55°C for 1 min (primer annealing), 72°C for 2 min (extension), and 10 min at 72°C. For standard PCR, the annealing temperature was set at 55°C. Following completion of the PCR reaction, a portion of the

PCR product was quantified using agarose gel electrophoresis.³⁸ The PCR products were purified to remove unincorporated nucleotides and excess primers using a PCR purification kit (Qiagen PCR Purification Kit, Hilden, Germany). The PCR product was mixed with 5 volumes of phosphate buffer (included in the kit), and the mixture was loaded onto a column. After centrifuging for 30 s at 13,000 rpm, the column was washed with 0.75 mL of TE buffer and centrifuged for 1 min to dry. The DNA was eluted using 50 µL of elution buffer. Gel electrophoresis was used to confirm successful DNA isolation, and the amplified DNA was stored in nuclease-free water and sequenced using an ABI 3730 DNA sequencer (Thermo Fisher Scientific, Waltham, MA, USA).

Phylogenetic Analysis

Sequence similarity was analyzed using a BLAST search (<http://www.ncbi.nlm.nih.gov/>) after multiple alignments with sequences of closely related species. Comparative analysis was done utilizing CLUSTAL-W (<https://www.genome.jp/tools-bin/clustalw>) in the Bioedit program. BLAST and MEGA software were used to assess the similarity of the phylogenetic tree to other cyanobacteria species.

Preparation of Cyanobacteria Extracts

Desertifilum biomasses were harvested after 15 days by centrifugation at 4000 rpm for 20 min, and the pellets were washed at least twice with distilled water to remove any trace elements. The pellets were freeze-dried, and the dried pellets were crushed using a pestle and mortar to obtain a fine powder. An aqueous cyanobacteria extract was prepared by mixing 15 mg of fine powder with 15 mL of distilled water, then incubated at 40°C for 1 day. After 24 h, the aqueous extract was filtered using a Whatman filter No. 42 (Camlab, Cambridge, United Kingdom) and stored in a refrigerator at 4°C until use.

Production of SNPs by Cyanobacteria

Green synthesis of SNPs was performed by adding 90 mL of 1 mM aqueous AgNO₃ to 10 mL of cyanobacteria filtrate at room temperature under direct illumination (2000 ± 200 Lux) for 24 h. A negative control was prepared by dissolving 90 mL of 1 mM aqueous AgNO₃ in 10 mL of pure distilled water without cyanobacterial biomass. After 24 h, the mixture was centrifuged at 10,000 rpm for 15 min at 10°C, and the resultant pellets were washed with 10 mL of distilled water at least 3 times to remove any free biomass remnants or uncapped ligands. The pellets were spread on a sterilized

petri dish and dried in an oven at 30°C for 1 day. The dried D-SNPs were scraped using a scalpel, and the fine powder was collected and stored in a sterile tube for analysis.

Characterization of D-SNPs

UV-Visible Spectroscopy

During the biological synthesis, and after 24 h, an aliquot of biofabricated D-SNPs was withdrawn and monitored using UV-VIS spectroscopy (UV1800 PC spectrophotometer, Shimadzu, Japan) across the wavelength range of 300 nm to 800 nm at a resolution of 1 nm.

X-Ray Diffraction

X-ray diffraction analysis (XRD) (X-ray 7000, Shimadzu, Japan) was used to evaluate the crystallinity of the bio-synthesized SNP powder. The sample was drop-coated onto a silica plate by applying many layers of small amounts of D-SNPs on with intermittent drying. This resulted in a thick coat of SNPs, which was evaluated from 0° to 80° (2 θ) using Cu K α radiation generated at 30 kV and 30 mA with scan speed 4 deg/min.

Fourier-Transform Infrared Spectroscopy

The functional groups and composition of D-SNPs were determined using FTIR (Shimadzu, Japan). The D-SNP powders were diluted in potassium bromide (in the ratio 1:100) and the FT-IR instrument was operated in diffuse reflectance mode (DRS-800). Spectra were collected from 400 to 4000 cm⁻¹ at a resolution of 4 cm⁻¹. Functional groups were identified using reference spectra.

Scanning Electron Microscopy

Scanning electron microscopy (SEM) (Jeol, Tokyo, Japan) at 20 kV was used to characterize the shape and size of the resultant SNPs. The samples were coated with gold using a sputter coater (model: S150B, Edwards High Vacuum Ltd., England) to prevent the build-up of local electrical charges.

Transmission Electron Microscopy

Transmission electron microscopy (TEM, Jeol) was used to capture high-resolution, two-dimensional images of the biosynthesized SNPs. The morphology of the SNPs was visualized using TEM operated at an accelerating voltage of 200 kV. Samples were prepared by adding a drop of SNP suspension onto a carbon-coated copper grid and allowed to dry under an infrared lamp prior to analysis.

Cell Culture

Three different cancer cell lines were used; MCF-7 breast cancer cells, HepG2 liver cancer cells, and Caco-2 colorectal adenocarcinoma cells. Cells were maintained in Dulbecco's Modified Eagle's Minimum Essential Medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 50 U/mL penicillin and streptomycin. The cells were incubated in a humidified atmosphere containing 5% CO₂ at 37°C. Confluent cells were passaged using trypsin-EDTA.

D-SNP Cytotoxicity

The cytotoxicity of D-SNPs was screened against MCF-7, HepG2, and Caco-2 cells using the MTT assay as mentioned by Bin-Meferij et al.³² In brief, the malignant cells were cultured in 96-well plates at a density of 7 × 10³ cells/well. After reaching confluency, the cells were subjected to increasing concentrations of 0.22- μ m filtrated D-SNPs (31.25–1000 μ g/mL). After 24 h, the DMEM medium was discarded and replaced with 100 μ L of fresh culture medium, and 10 μ L of 12 mM MTT stock solution was added to each well. Ten microliters of 12 mM MTT stock solution was added to 100 μ L of medium as a negative control. The cells were incubated for 4 h at 37°C, then 100 μ L/well DMSO was added, and the solutions were thoroughly mixed to dissolve the formazan crystals. Absorbance (*A*) was measured at 570 nm. The viability percentage was calculated using the formula:

$(A_{\text{Treated}} - A_{\text{Blank}}) / (A_{\text{Control}} - A_{\text{Blank}}) \times 100$. The half-maximal growth inhibitory concentration (IC₅₀) was determined using a sigmoidal curve.

Antibacterial Activity of D-SNPs

Microbial Strains and Culture

Five bacterial strains of gram-negative and gram-positive bacteria were obtained from King Khalid University hospital laboratory and the Department of Microbiology, King Saud University, Riyadh, Saudi Arabia. These strains included *Bacillus cereus* ATCC 10876, *Pseudomonas aeruginosa* ATCC 27853, *Bacillus cereus* ATCC 10876, *Bacillus subtilis* ATCC 6633, *Shigella flexneri*, and *Salmonella enterica* (clinical isolates). The five pathogens were grown in nutrient broth (Scharlab, S.L, SPAIN) at 37°C overnight and maintained through continuous sub-culturing in broth and on solid media.

Bactericidal Activity of D-SNPs Using Agar Well Diffusion Method

To evaluate the antibacterial activity of D-SNPs, 4 mL of bacterial saline mixture was mixed with 50 mL of warm

nutrient agar (Merck, Darmstadt, Germany), and the mixture was poured into sterile plates and incubated for up to 18 h at 37°C. Four 8-mm wells were bored in each agar plate using a cork borer. One hundred microliters of D-SNPs suspension (1.54 mg/mL) was poured into three of the wells using a micropipette, and the blank solvent was used as a negative control. Other plates were treated with silver nitrate and streptomycin as a positive control. This experiment was repeated in a triplicate, and the mean and the standard error of the mean were calculated. The treated bacterial plates were incubated for 24 h at 37°C. The inhibitory activity of D-SNPs against bacteria was assessed by determining the diameter of the inhibition zone (IZ) in mm using a transparent ruler.

Statistical Analysis

All data are presented as a mean of three independent replicates (mean \pm SEM), one-way analysis of variance (ANOVA) was performed for statistical analyses utilizing Prism 8.3 software (GraphPad Software Inc., La Jolla, CA, USA). Data were considered statistically significant at $P < 0.01$ and $P < 0.001$. For D-SNPs analysis, Origin8 (OriginLab Corporation, Northampton, MA, USA) and ImageJ (National Institutes of Health, Bethesda, MD, USA) were used.

Results and Discussion

Our hypothesis was “A novel cyanobacteria *Desertifilum sp.* will be biofabricate an ecofriendly silver nanoparticle that will have a biological activity against cancer cells and bacteria”

Desertifilum sp. showed a green or bright blue-green non-branching filamentous structure under a light microscope (Figure 1). Inverted light micrographs showed that the filaments were thin and long (more than 500 μm), as well as their widths ranged from 4 to 7 μm with an average width of 5.9 μm . Also, the filaments were present alone or organized in separate bundles or spiral trichomes of a dense mesh (Figure 2A). The filaments had apical ends as well as these filaments were motile non-constructed or slightly constricted at cross-walls (Video 1), the trichomes appear narrow at the end and were enclosed by a hyaline sheath firm (Figure 2B).³⁹

The resultant amplicon (1.5 kb) of the *Desertifilum sp.* is shown in Figure 3. The phylogenetic tree of *Desertifilum sp.* is shown in Figure 4. The AM2 strain was 99% similar to *Desertifilum* IPPAS B-1220.⁴⁰ *Desertifilum* IPPAS B-1220 sequence was deposited in GenBank as MN629136.

To our knowledge, no previous studies have reported the use of *Desertifilum sp.* for the biosynthesis of nano-metals.

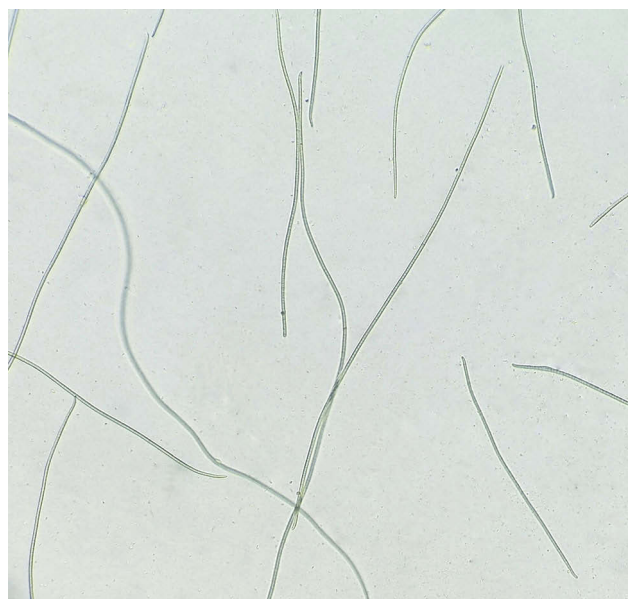


Figure 1 Low-power light micrograph of *Desertifilum sp.*
Note: Filamentous appearance of *Desertifilum sp.*

Therefore, our study is the first to use this microorganism for the green synthesis of SNPs. The reduction process of AgNO_3 to D-SNPs was characterized by a change in color from pale yellow to dark brown.⁴¹ The color change was due to the vibrations of surface plasmons in the resultant NPs.⁴² The D-SNPs had a peak at 421 nm (Figure 5), which indicated that D-SNPs were successfully synthesized from AgNO_3 . Sharpening of the SPR peak width is an indicator of nanometal size and monodispersity, and absorbance across the range of 320–580 nm is typical for SNPs.^{43,44} Ali et al⁴⁵ found that the absorption peak of biogenic SNPs synthesized by *Oscillatoria* absorbed at 450 nm. Husain et al⁴⁶ showed that SNPs synthesized from *Oscillatoria sp.* have an absorption peak at 485 nm. Furthermore, O-SNPs had a surface plasmon peak at 426 nm.⁴⁷ Accordingly, the lower wavelength value indicated that smaller sized spherical NPs were formed,⁴³ which suggested that *Desertifilum sp.* may produce particularly small nanoscale particles. Looking into the literature, our results indicated that *Desertifilum sp.* may be a promising candidate for biosynthesis SNPs.

The differences in SPR values may have been due to the different species of cyanobacteria used to produce SNPs,⁴⁸ and the mechanisms of interaction between the cyanobacteria and NPs. Until now, the exact mechanism of green synthesis of NP is poorly understood, El-Batal et al⁴⁹ reported that an NADH-dependent reductase was critical in SNP biofabrication. El-Naggar et al⁵⁰ also revealed that cellular molecules such as proteins, enzymes, amino acids,

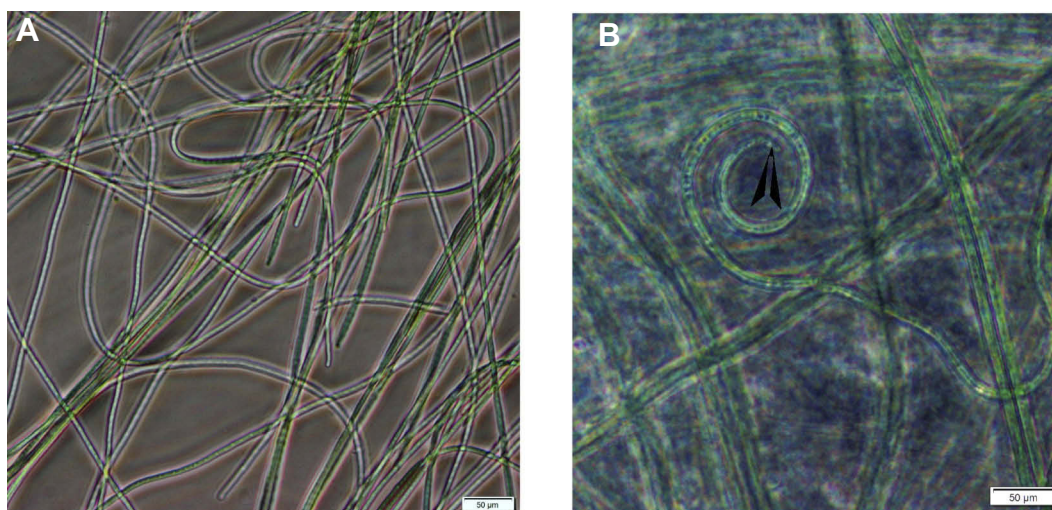


Figure 2 Inverted light micrographs of *Desertifilum* sp.

Notes: (A) The filamentous structure of *Desertifilum* sp. (B) The apical end (blackhead arrow) of a *Desertifilum* sp. filament. Scale bar = 50 µm.

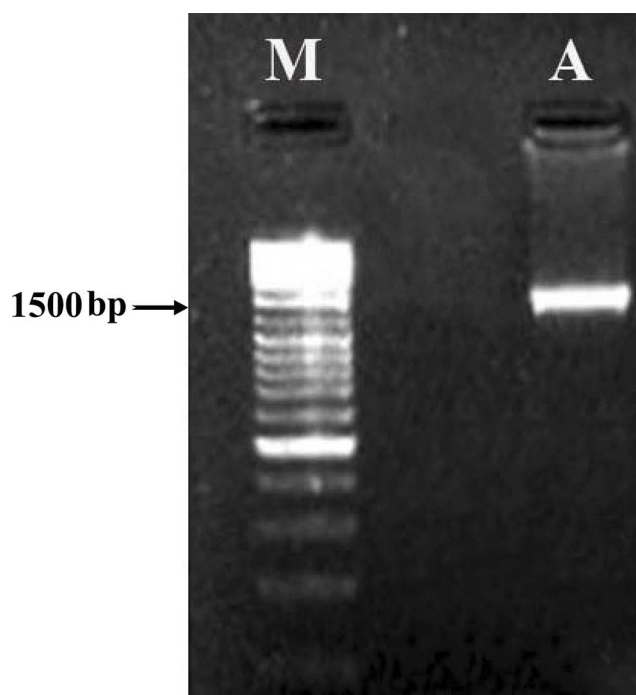


Figure 3 Electrophoresis of the resultant amplicon in 1% agarose gel.

Notes: (A) 16S rDNA amplified using PCR from the genomic DNA of *Desertifilum* sp. (M) Molecular marker.

carbohydrates, and photosynthetic pigments have been implicated in the fabrication process of silver ions.

Mahdiah et al⁵¹ suggested that the biofabrication of NPs is a two-step process. The first step involves adherence of SNPs to algal cells through electrostatic attraction between positively charged metal ions and negatively charged carboxylate ions on the algal surface. The second step involves the

reduction of metal ions to MNPs by secreted reductase enzyme by the algal cells. The current data (Figure 5) may be interpreted based on that the cyanobacteria biomolecules such as proteins, enzymes, metabolites reduce silver nitrate into SNPs and/or phycobilisomes that mediated conformational changes to absorb light energy for initiating photosynthesis, this light absorption allows to transport the electrons from ground state to an excited state.⁵² So, under illumination may be silver nitrate incorporated into SNPs due to electrons moving between energy levels.

The range of 0° to 80° with 2θ values of 29.7 ,^{53,54} 38.4 ,⁵⁵ 44.6 ,⁵⁶ 64.7 ,⁵⁷ and 77.7 ,⁵⁵ respectively, was associated with (310), (111), (200), (220), and (311) planes of silver (Figure 6). Each spectrum was compared with the available standard from the Joint Committee on Powder Diffraction Standards, and the biogenic D-SNPs were crystalline. The Scherrer equation, $D = (k\lambda)/(\beta \cos \theta)$, was used to determine the crystallite size of the most intense peak at 2θ of 29.7 . The crystallite size of the D-SNPs was 19.4 nm. Furthermore, interplanar spacing was 0.3 nm, as determined using the Bragg equation, $d = \lambda/2 \sin \theta$.

Analysis using Fourier transform infrared (FTIR) showed that bioactive groups surrounded the biogenic silver nanoparticles. The FTIR spectrum of SNPs had six peaks at 601.81 cm^{-1} , 1042.35 cm^{-1} , 1626.05 cm^{-1} , 2353.23 cm^{-1} , and 3453.72 cm^{-1} for SNPs (Figure 7). The major peaks at 1042.35 , 1626.05 , and 3453.72 cm^{-1} indicated that the amides on the SNP surface might have included peptides and proteins present in the cell extract. Spectra of SNPs showed a strong broad absorption peak at

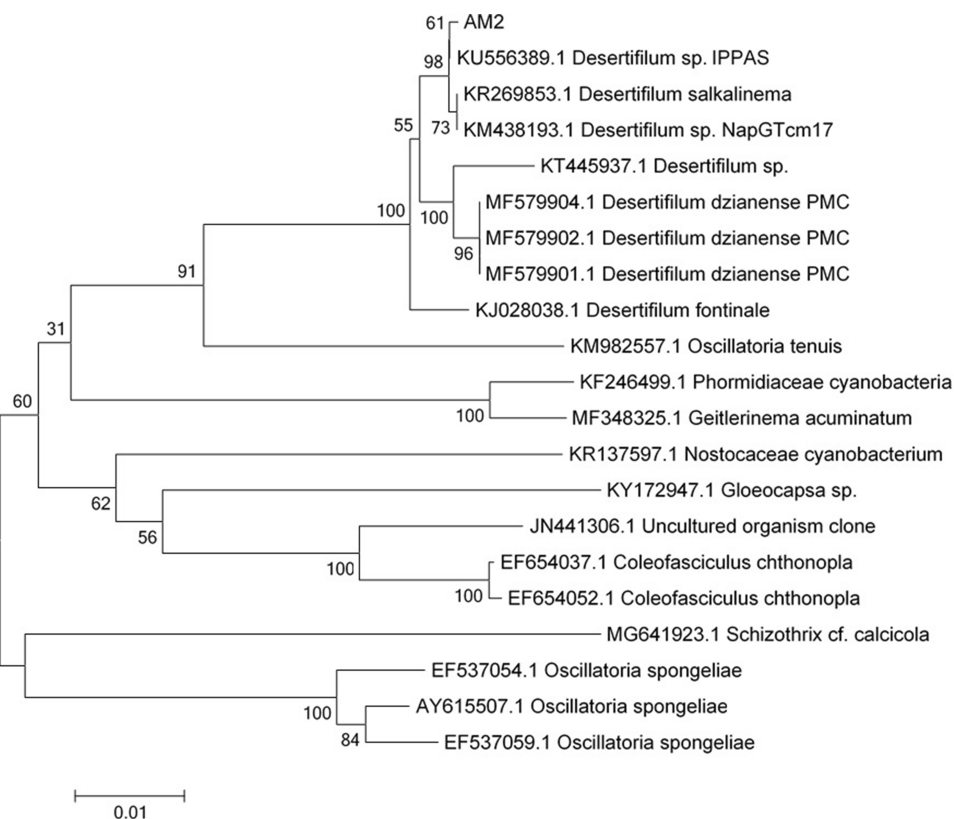


Figure 4 The phylogenetic tree of *Desertifilum* sp.

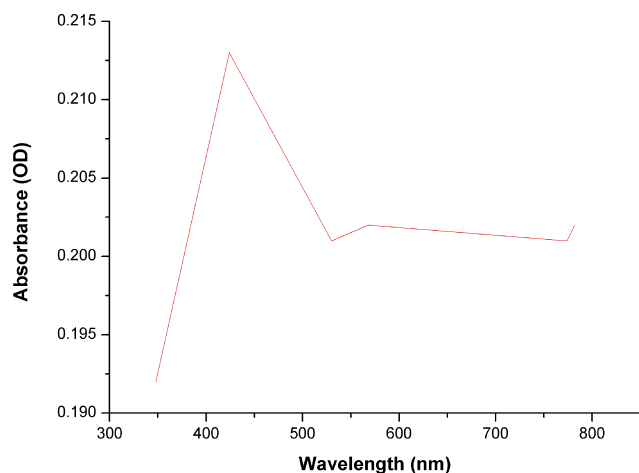


Figure 5 Ultraviolet-visible absorption spectra of D-SNPs.

Note: Plasmon resonance of D-SNPs at 421 nm.

Abbreviation: D-SNPs, silver nanoparticles synthesized by *Desertifilum* sp.

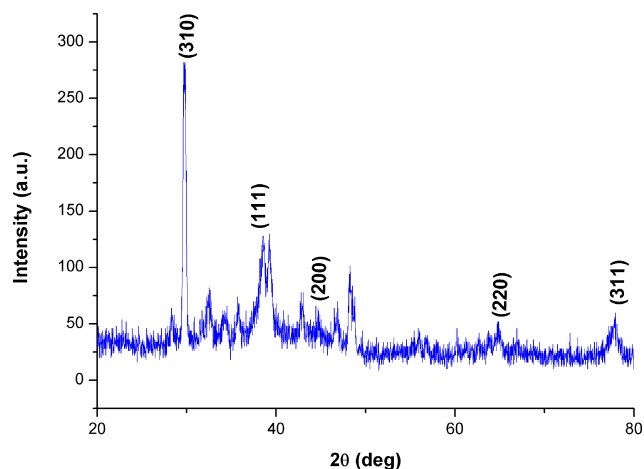


Figure 6 XRD pattern of D-SNPs.

Abbreviations: XRD, x-ray diffraction; D-SNPs, silver nanoparticles synthesized by *Desertifilum* sp.

3453.72 cm^{-1} , which is characteristic of O-H stretching vibration of the polysaccharide and N-H stretching vibration of proteins. A peak at 2353.23 cm^{-1} corresponded to C≡C stretching modes of vibration in alkyne groups. The absorption peak at 1626.05 cm^{-1} was assigned to bending N-H stretching vibration of amine groups of proteins and

C=C stretching vibrations of alkenes. The band observed at 1042.35 cm^{-1} corresponded to the C-N stretching vibration of amines of proteins and C-O stretching vibration of vinyl ethers. The band at 601.81 cm^{-1} represented stretching of C-Br compounds. Based on the FTIR data, the presence of amide bonds of proteins and the hydroxyl

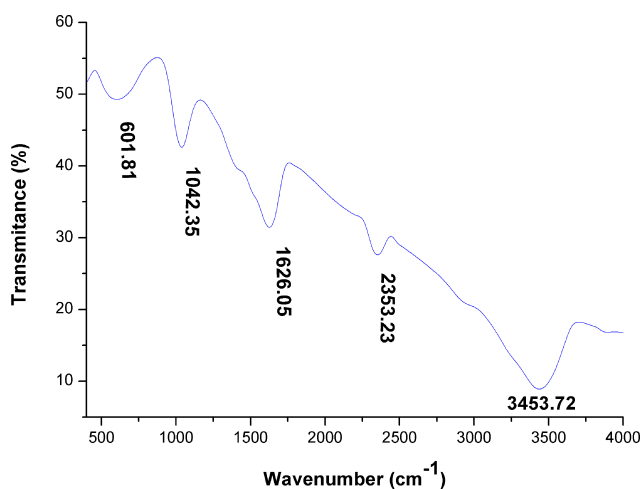


Figure 7 FTIR spectra of green SNPs synthesized using *Desertifilum* sp.
Abbreviations: FTIR, Fourier-transform infrared spectroscopy; SNPs, silver nanoparticles.

group of polysaccharides indicated that the proteins and polysaccharides that surrounded silver may be responsible for the reduction of silver ion into SNPs and resulted in the formation of a capping layer around SNPs to prevent agglomeration and thereby the particles are stabilized. Our findings agreed with those of El-Naggar et al⁵⁰ and Hamouda et al⁴⁷ who revealed that biomolecules present in cell extracts could reduce Ag^+ ions. Ahmad et al⁵⁸ suggested that nitrate reductase may be a critical molecule in the reduction process of Ag^+ ion to Ag^0 by cyanobacteria grown on media containing NO_3^- .

Scanning electron micrographs showed that D-SNPs were roughly spherical and exhibited some agglomeration (Figure 8A). The size range of SNPs produced by *Desertifilum* sp. ranged from 22 to 40 nm with an average size of 31 nm (Figure 8B). Rasheed et al⁵⁹ showed that

SNPs fabricated using *Convolvulus arvensis* extract were spherical in shape and had an average size of 60 nm. These results indicated that *Desertifilum* sp. may be a promising microorganism for producing smaller SNPs.

Analysis using TEM showed that D-SNPs were spherical and were well dispersed without significant agglomeration. Particles ranged in size from 4.5 nm to 26 nm (Figure 9). The size range values agreed with the crystal sizes determined using XRD. Green SNPs were previously fabricated using *Nostoc* sp.,⁶⁰ *Synechococcus* sp.,¹⁷ *Anabaena* sp.,⁶¹ *Limnothrix* sp., *Synechocystis* sp. 48–3, *Anabaena doliolum* sp.⁶² and *Nostoc commune* sp.⁶³ with sizes ranging from 51 to 100, 15.2 to 266.7, 24.13, 31.86, 14.64, 10 to 50, and 15 to 54 nm, respectively. Yousefzadi et al⁶⁴ reported that SNPs formed by the seaweed *Enteromorpha flexuosa* were 2–32 nm. The variations in sizes of SNPs produced by the same phylum may have been due to differences in silver reducing efficiency. Previous studies and our findings indicated that *Desertifilum* sp. has the potential to produce smaller SNPs.

This report demonstrated for the first time the anticancer activity of D-SNPs synthesized using cell extracts from *Desertifilum* sp. We evaluated the cytotoxic effects of D-SNPs against three human cell lines (MCF-7, HepG2, and Caco-2) using the MTT assay. The data revealed that D-SNPs significantly decreased the proliferation of the three different cancer cell lines in a dose-dependent manner compared with untreated cells (Figure 10A). The IC_{50} values of D-SNPs against HepG2, MCF-7, and Caco-2 cells were 32, 58, and 90 $\mu\text{g}/\text{mL}$, respectively (Figure 10B). The current data proved the efficiency of D-SNPs as an anticancer agent against different cell lines. The mechanisms by which SNPs inhibit cancer

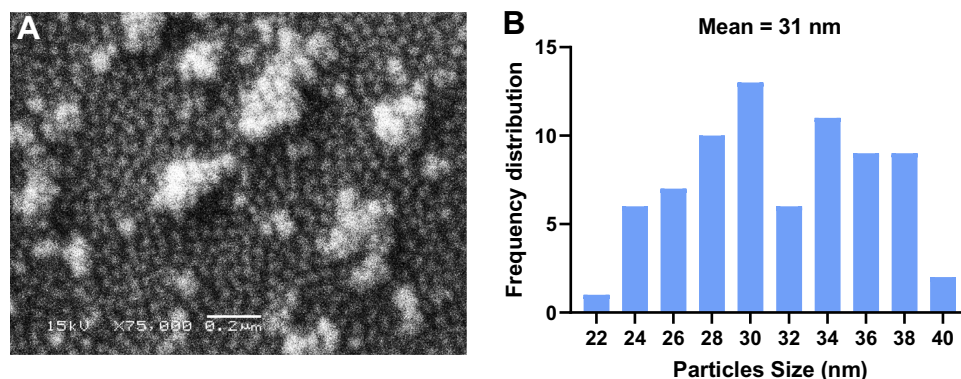


Figure 8 SEM micrographs of D-SNPs.
Notes: (A) Spherical shape of D-SNPs, with some aggregation. Scale bar, 0.2 μm . (B) The frequency distribution of D-SNPs, average size 31 nm.
Abbreviations: SEM, scanning electron microscope; D-SNPs, silver nanoparticles fabricated using *Desertifilum* sp.

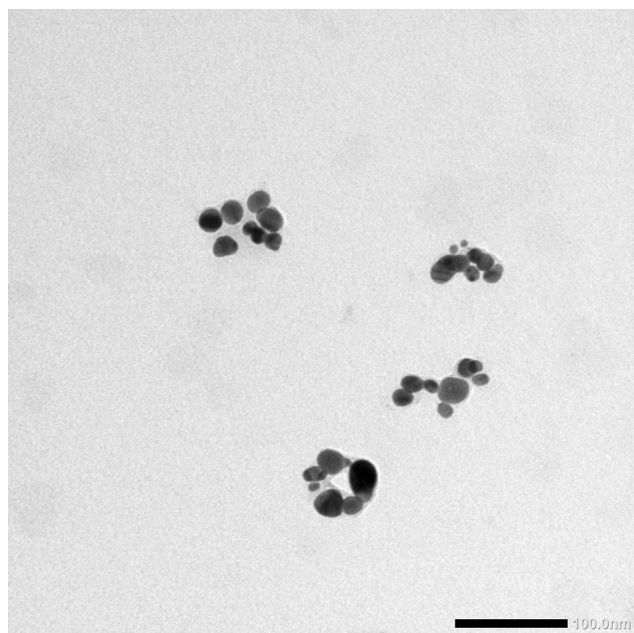


Figure 9 TEM examination of D-SNPs.

Notes: TEM micrograph demonstrating the spherical shape and distribution of D-SNPs with a size range from 4.5 to 26 nm. Scale bar, 100 nm.

Abbreviations: TEM, transmission electron microscope, D-SNPs, silver nanoparticles fabricated using *Desertifilum* sp.

cell growth are poorly understood. However, a number of studies have shown that SNPs induced reactive oxygen species production, resulting in apoptosis.^{65,66} The current results showed that D-SNPs induced the greatest toxic effects against HepG2 cells. The higher sensitivity of HepG2 cells to D-SNPs may have been due to charge and the type of biomolecules surrounding D-SNPs. Baetke et al and El Nagar et al reported that SNP toxicity depended on size, concentration, and surface composition.^{4,33} Netala et al⁶⁷ reported that surface charge and small size enhanced diffusion and dispersion into the tumor matrix. Bhatnagar et al⁶⁸ showed that SNPs fabricated using extracellular pigment from *Talaromyces purpurogenus* have a significant cytotoxicity effect against liver cancer cell line. Abaza et al⁶⁹ showed that Cs/PVA/Ag nanocomposites caused in killing 50% of HepG2 cells at 43.7 $\mu\text{g/mL}$. Based on these data, D-SNPs may be a potent antitumor agent against HepG2 with low IC_{50} . Ranijitham et al⁷⁰ reported that SNPs induced cytotoxicity in MCF-7 cells in a dose–response manner. Ramar et al⁷¹ showed that SNPs synthesized using fruits of *Solanum trilobatum* have an anticancer activity against MCF-7 cell line with IC_{50} equal to 300 $\mu\text{g/mL}$. This result indicates that D-SNPs may be a more efficient antitumor agent against MCF-7 cells. Martins et al⁷² showed that a polyelectrolyte complex containing SNPs exerted cytotoxic effects against Caco-2 cells, but not healthy African green monkey cells

(VERO cells), at concentrations above 100 $\mu\text{g/mL}$. Comparing the chemically synthesized SNPs,^{69,72} D-SNPs showed a potent anticancer activity with low IC_{50} values and may be an alternative material to treat cancer.

This report evaluated for the first time the antibacterial activity of SNPs biosynthesized using *Desertifilum* sp. cellular extracts against different pathogenic bacteria strains. The antibacterial properties of D-SNPs against five bacterial strains (*Bacillus cereus* ATCC 10876, *Pseudomonas aeruginosa* ATCC 27853, *Shigella flexneri*, *Salmonella enterica* (clinical isolate), and *Bacillus subtilis* ATCC 6633) were evaluated using the agar well-diffusion assay. The data showed that D-SNPs induced varying degrees of toxicity against the tested bacteria (Figure 11). In addition, D-SNPs suppressed bacterial growth to a greater extent than silver nitrate, which was used as a positive control (Figure 12). As expected, the antibiotic streptomycin showed the highest antibacterial activity against five selected pathogens followed by D-SNPs (Table 1). Our results showed that D-SNPs inhibited the growth of *Shigella flexneri* to the greatest extent, as evidenced by an inhibition zone (IZ) diameter of 22.7 ± 0.3 while streptomycin resulted in IZ diameter equal to 26.3 ± 0.5 . Also, for *S. enterica* the diameter of IZ measured after treating with D-SNPs was 16.7 ± 0.3 however the IZ diameter of streptomycin was 18.4 ± 0.4 . These results are interesting since the values of IZ diameter caused by D-SNPs are nearly equals to the IZ value caused by streptomycin.

Although the D-SNPs have a potent inhibitory effect against *B. cereus* and *B. subtilis* with IZ diameter values of 17.3 ± 0.3 and 16.3 ± 0.3 , respectively, the streptomycin was the most antibacterial agent that caused an IZ with a diameter of 29.5 ± 1.5 and 24.9 ± 0.5 , respectively. D-SNPs inhibited the growth of *Pseudomonas aeruginosa* to a lesser degree, as evidenced by an inhibition zone diameter of 15 ± 0.0 (Table 1). In contrast, *Pseudomonas aeruginosa* were the most bacteria sensitive to AgNO_3 treatment with IZ diameter of 14.7 ± 0.3 , this value approximately the same as the IZ diameter resulted from treating the same bacteria with D-SNPs (Table 1). *B. subtilis* were the most bacteria that resisted the inhibitory action of silver nitrate with an IZ diameter of 9.5 ± 0.06 .

These data indicate that D-SNPs have a significant antibacterial effect against gram-negative and gram-positive bacteria. Paul et al⁷³ reported that SNPs synthesized extracellularly using *Pseudomonas aeruginosa* exerted cytotoxic activity against *Shigella flexneri* and *Bacillus subtilis*, as evidenced by inhibition zone diameters of 16.0 and

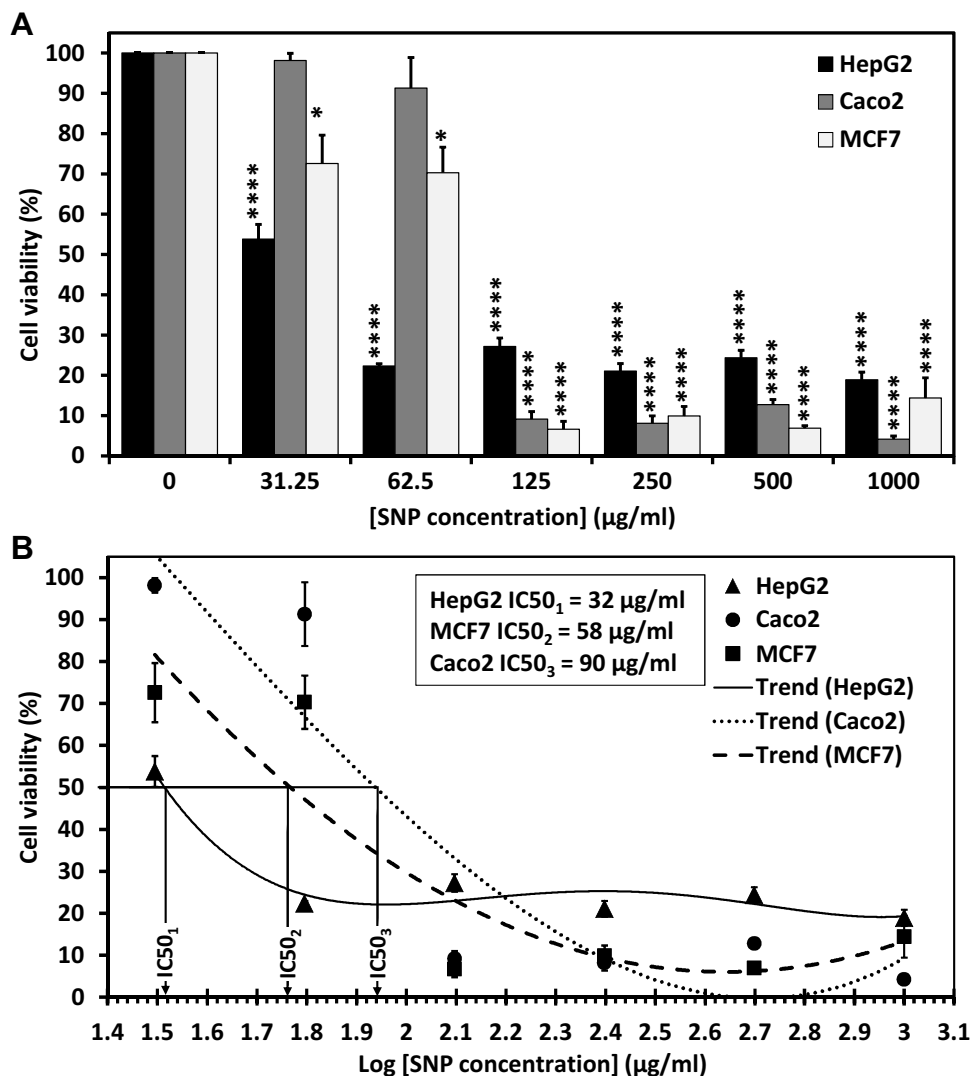


Figure 10 Dose-dependent growth inhibition of MCF-7, HepG2, and Caco-2 cells using D-SNPs.

Notes: (A) The antiproliferative effect of D-SNPs against MCF-7, HepG2, and Caco-2 cell lines was expressed as the percent viable cells. (B) MCF-7, HepG2, and Caco-2 cell viability percentage and the log concentration of D-SNPs. Data are presented as the mean \pm SEM; P values were calculated versus control cells: **** $P < 0.001$ and * $P < 0.01$.

Abbreviations: D-SNPs, silver nanoparticles formed using *Desertifilum* sp.; SEM, standard error of the mean.

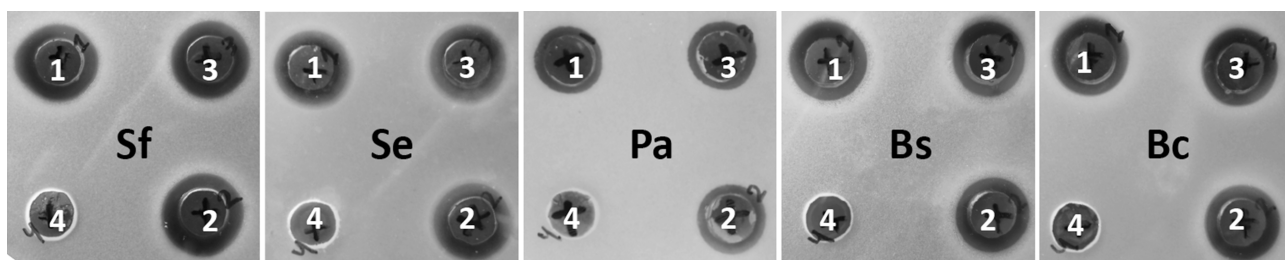


Figure 11 Antibacterial activity of D-SNPs against five pathogenic bacteria strains.

Notes: (Sf) *Shigella flexneri*, (Se) *Salmonella enterica*, (Pa) *Pseudomonas aeruginosa*, (Bs) *Bacillus subtilis*, and (Bc) *Bacillus cereus*. The number 1, 2, and 3 referred to triplicate experiments with D-SNPs, and 4 was distilled water as a negative control.

Abbreviation: D-SNPs, silver nanoparticles synthesized by *Desertifilum* sp.

17.6 mm, respectively. El Nagar et al³³ showed that the diameter of the inhibition zone of *Pseudomonas aeruginosa* treated with biogenic SNPs was 10 mm, and silver nitrate

(5 mM) did not exert any antibacterial activity. Allafchian et al⁷⁴ reported that SNPs synthesized using *Phlomis* leaf extract inhibited the growth of *Bacillus cereus*, with an

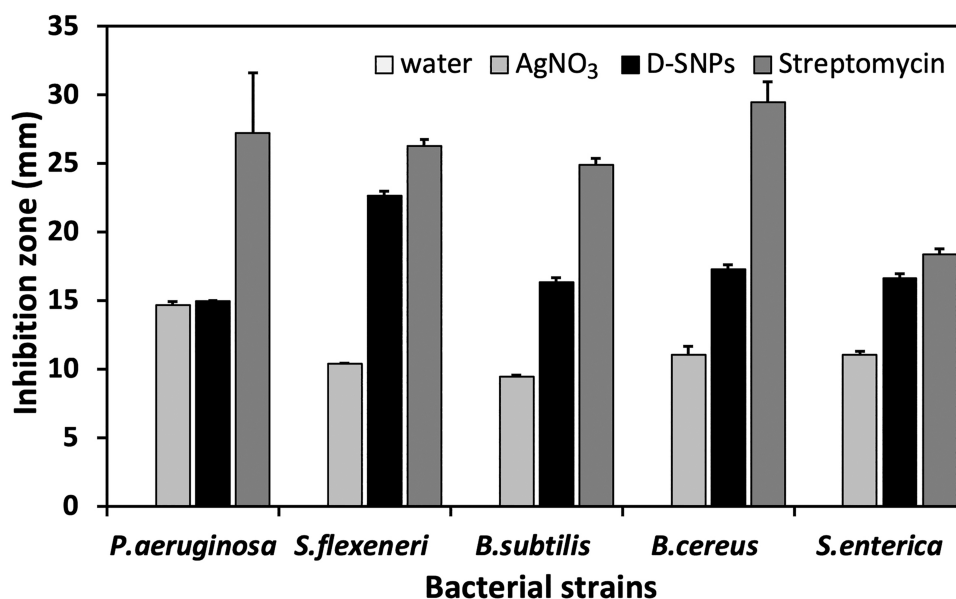


Figure 12 Antibacterial activity of D-SNPs.

Notes: Antibacterial activity of D-SNPs, streptomycin, AgNO₃, and distilled H₂O against *Pseudomonas aeruginosa*, *Shigella flexneri*, *Bacillus subtilis*, *Bacillus cereus*, and *Salmonella enterica*. The data are expressed as the mean ± SEM.

Abbreviation: D-SNPs, silver nanoparticles synthesized using *Desertifilum* sp.

inhibition zone diameter of 12.1 mm. In contrast, the treatment of *Bacillus cereus* with silver nitrate resulted in an inhibition zone diameter of 9.5 mm. Growth of *Salmonella enterica* was suppressed following treatment with SNPs fabricated using an aqueous extract of seaweed *E. compressa*, with an inhibition zone diameter of 10.5 mm (Table 2).⁷⁵

The highest toxic effects of D-SNPs against *Shigella flexneri* may have been due to the thinner cell wall of these gram-negative bacteria, allowing for better penetration of D-SNPs.⁷⁶ Our results suggested that D-SNPs exhibited superior antibacterial effects due to the presence of surface biomolecules and small particle size.⁷⁴ Romero et al⁷⁷ explained that the

Table 1 Agar Well-Diffusion Assay

Treatment	<i>P. aeruginosa</i>	<i>S. flexneri</i>	<i>B. subtilis</i>	<i>B. cereus</i>	<i>S. enterica</i>
*Bacterial strains					
Distilled H ₂ O	NA	NA	NA	NA	NA
AgNO ₃	14.67 ± 0.29	10.4 ± 0.06	9.5 ± 0.06	11.07 ± 0.64	11.07 ± 0.27
D-SNPs	15.0 ± 0.00	22.67 ± 0.33	16.33 ± 0.33	17.33 ± 0.33	16.67 ± 0.33
Streptomycin	27.2 ± 4.40	26.3 ± 0.48	24.9 ± 0.45	29.5 ± 1.45	18.4 ± 0.44

Notes: The inhibition zone diameter (mm) in response to D-SNPs, streptomycin, AgNO₃, and distilled H₂O treatments against *P. aeruginosa*, *S. flexneri*, *B. subtilis*, *B. cereus*, and *S. enterica*. * The five pathogens were grown in nutrient broth (Scharlab, S.L, SPAIN) at 37°C overnight and maintained through continuous sub-culturing in broth and on solid media.

Abbreviations: D-SNPs, silver nanoparticles synthesized by *Desertifilum* sp.; AgNO₃, silver nitrate; H₂O, water.

Table 2 The Antibacterial Activity of Different Biogenic SNPs Against Different Bacteria Strains

Reducing Agents	Size of SNPs (nm)	Target Bacteria	IZ Diameter (mm)	Citation
<i>Pseudomonas aeruginosa</i>	50–85	<i>Shigella flexneri</i> , <i>Bacillus subtilis</i>	16.0, 17.6	73
<i>Nostoc linckia</i>	9.39–25.89	<i>Pseudomonas aeruginosa</i>	10	33
<i>Phlomis leaf extract</i>	19–30	<i>Bacillus cereus</i>	12.1	74
Seaweed <i>E. compressa</i> extract	4–24	<i>Salmonella enterica</i>	10.5	75

biocompounds that surround the NP surface, charge and size of these particles play an important role in the attraction between bacterial cells and NPs and penetration of bacterial cell wall. Other publications are in accordance with our finding in which NPs with size less than 10 nm have the ability to penetrate the bacterial cell wall and interact with the bacterial biomolecule such as DNA, protein, lipids and sometimes with respiratory enzymes cause liberating of ROS such as hydrogen peroxide (H_2O_2), hydroxyl (OH^\cdot) and superoxide ($O_2^{\cdot-}$) radicals that increase oxidative stress and damage nucleic acid and protein leading to death.⁷⁸ Recent research by Sondi and Salopek-Sondi referred that the accumulation of SNPs (12 nm) on the cell wall of *E. coli* results in pit formation. Those pits cause a loss of outer membrane integrity, resulting in the release of cellular materials such as LPS molecules and proteins, and causing eventual cell death.⁷⁹ The current findings indicated that D-SNPs may be potent antibacterial agents with a highly inhibitory effect against different pathogenic bacteria and may be used as alternative antibiotics to antibiotics that can be included in several products such as medical dressings and textile.

Conclusions

Cyanobacteria are an ancient group of photoautotrophic prokaryotes that contain valuable biomolecules such as pharmaceuticals, pigments, and proteins. *Desertifilum* sp. IPPAS B-1220 is a novel member of cyanobacteria that contain several biocompounds that can participate in SNP fabrication. In this study, we isolated, purified, cultured and molecular identified a novel cyanobacteria species (*Desertifilum* sp. IPPAS B-1220) and used their aqueous extract to synthesize silver nanoparticles. The biogenic SNPs detailed in this study were characterized using UV-Vis, FT-IR, XRD, SEM, and TEM. Physicochemical analyses showed that the D-SNPs were spherical and ranged in size from 4.5 to 26 nm. The presence of amides and hydroxyl groups indicated that proteins and polysaccharides were important factors in the biosynthesis of D-SNPs. The green D-SNPs produced in this study exerted potent antibacterial effects against gram-positive and gram-negative bacteria. Moreover, D-SNPs exerted significant cytotoxicity against MCF-7, HepG2, and Caco2. Our study showed that this approach was simple, green, efficient, and resulted in the production of SNPs at ambient temperature without the use of any toxic reducing or dispersing agents. Moreover, SNPs fabricated using *Desertifilum* sp. IPPAS B-1220 may be a promising antibacterial and anticancer agent that can be used in the medical and pharmaceutical industries. Further investigations are needed to demonstrate the optimal

conditions to produce SNPs using *Desertifilum* sp. IPPAS B-1220. Additionally, future studies are needed to explain the biofabrication process using cyanobacteria and to explore the mechanism of action of biogenic SNPs against both bacteria and cancer cell lines.

Abbreviations

MNPs, metallic nanoparticles; NPs, nanoparticles; D-SNPs, silver nanoparticles synthesized by *Desertifilum* sp; SNPs, silver nanoparticles; $AgNO_3$, silver nitrate; PCR, polymerase chain reaction; nm, nanometer; min, minute; μL , microliter; mg, milligram; mL, milliliter; μg , microgram; mM, millimolar; h, hour; cm^{-1} , inverse centimeter; rpm, revolutions per minute; SEM, scanning electron microscope; TEM, transmission electron microscope; DLS, dynamic light scattering; FTIR, Fourier-transform infrared spectroscopy; XRD, X-ray diffraction; kV, kilovolt; kb, kilobase; SPR, surface plasmon resonance; ELISA, enzyme-linked immunosorbent assay; UV-vis, ultraviolet-visible spectroscopy; IZ, inhibition zone; platinum nanoparticles, Pt NPs.

Ethics Approval and Informed Consent

The current study was performed in accordance with the Research Ethical Committee guidelines published by the National Health and Medical Research Council and the Ministry of Health and Population in Egypt. The Department of Zoology (Science Faculty, Alexandria University) and the Department of Cancer Management and Research (Medical Research Institute, Alexandria University) granted permission for this work.

Data Sharing Statement

The data supporting this article are available in [Figures 1–12](#). The data sets analyzed in the present study are available from the corresponding author upon reasonable request.

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Author Contributions

All authors contributed to data analysis, drafting, or revising the manuscript. All authors approved the final manuscript, and agree to be accountable for all aspects of the work.

Disclosure

This research was included in a request for patent 765805 from the Egyptian patent office, Academy of Scientific Research and Technology (ASRT), Egypt, submitted on 12/3/2019, titled “Biosynthesis of SNPs for the first time using new species of cyanobacteria *Desertifilum IPPAS B-1220* and its tumor activity” <http://www.egypo.gov.eg/>. The authors report no other conflicts of interest in this work.

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