# *Original Article* Cytotoxic T lymphocyte-associated antigen-4 gene polymorphisms and biliary atresia susceptibility in Chinese children

Jia Liu, Yifan Yang, Rui Dong, Chao Zheng, Jiahao Pei, Gong Chen, Zhen Shen, Yanlei Huang, Shan Zheng

*Department of Pediatric Surgery, Children's Hospital of Fudan University, and Key Laboratory of Neonatal Disease, Ministry of Health, Shanghai, China*

Received October 17, 2016; Accepted October 27, 2016; Epub May 1, 2018; Published May 15, 2018

Abstract: Biliary atresia (BA) is a devastating liver disease of complex pathogenesis in neonates, characterized by an inflammatory and fibrosing obstruction of extrahepatic bile ducts. Cytotoxic T lymphocyte-associated antigen-4 (CTLA4) is expressed on the surface of a subset of regulatory T cells (Treg) and down regulates the human immune response. To investigate the possible association between CTLA4 gene polymorphisms and BA susceptibility, we conducted a case-control study in the Chinese children. Three single nucleotide polymorphisms (SNPs) in the CLTA4 gene (rs231725, rs231775 and rs3087243) were genotyped in 113 BA patients and 133 healthy controls. The statistical analysis revealed no significant difference between BA patients and healthy controls in allele or genotype frequencies (rs231725, P = 0.2718, OR = 0.814, 95% CI = 0.564-1.175; rs231775, P = 0.1599, OR = 1.316, 95% CI = 0.897-1.931; rs3087243, P = 0.0572, OR = 1.582, 95% CI = 0.984-2.543), neither in the distribution of haplotypes of these CTLA4 gene SNPs. The result of our study is the first one to provide the evidence that there is no significant association between CLTA4 gene polymorphisms and BA susceptibility in Chinese children.

Keywords: Biliary atresia, cytotoxic T lymphocyte-associated antigen-4, single nucleotide polymorphism

#### Introduction

Biliary atresia (BA) is a major cause of liver transplantation in children. Characterized by progressive obliteration of the extrahepatic biliary system and fibrosis in liver, it will lead to fatal consequences if there is no prompt and proper treatment [1, 2]. The incidence of BA varies geographically and has a slight female predominance, while in Asia it is estimated to be 1 in 5000, higher than in western countries [2, 3]. Although surgical treatment such as Kasai hepatoportoenterostomy can remove the bile duct remnants and improve short-term outcome, many children suffered liver disease progression to end-stage cirrhosis, which indicates a multifaceted etiology of this disease [4].

In the past decades, many relative hypotheses of BA etiology have been established, which can be summarized to genetic causes, environmental factors and immune dysregulation [1, 3, 5]. Recently, studies investigating single nucleotide polymorphisms (SNPs) with BA susceptibility are gaining much more focus. Several genes have been discovered a significant association with BA, including adiponectin, adducin 3 (ADD3), X-prolyl aminopeptidase P1 (XPNP-EP1) and intercellular adhesion molecule-1 (ICAM-1), showing that genetic factors may play important rules in BA etiology [1, 2, 5-7].

As a promising candidate, the CTLA4 gene has been studied for genetic susceptibility to many autoimmune diseases. Cytotoxic T lymphocyteassociated antigen-4 (CTLA4) is a protein receptor that plays an important role in immune regulation by down-regulating human T lymphocyte immune response [8]. Constitutively expressed on a subset of regulatory T cells (Treg), CTLA4 plays a distinct role in Treg generation, function and homeostasis [9-11]. In BA, deficits in Treg quantity and/or function would disrupt immunologic balance and deteriorate bile duct injury [5]. These findings suggest that CTLA4 gene may be implicated in the pathogenesis of BA.

<b>SNP</b>	Common name	Allele major/minor	Gene location	P value for HWE
rs231725	۰	A/G	3'UTR	0.935
rs231775	49AG	G/A	Exon 1	0.886
rs3087243	CT60	G/A	3'UTR	0.999

Table 1. Three SNPs selected for the study

3'UTR, 3'untranslated region.

Table 2. Oligonucleotide sequences used for genotyping

<b>SNP</b>	Primers	Sequences
rs231725	First	5'-ACGTTGGATGTCCCAAATTTTGCCTCCACC-3'
	Second	5'-ACGTTGGATGGAGGTGAAACCAAGTATAGC-3'
	Extension	5'-AACAAAAGAGGAGAAATCCA-3'
rs231775	First	5'-ACGTTGGATGATTTCAGCGGCACAAGGCTC-3'
	Second	5'-ACGTTGGATGCACTCACCTTTGCAGAAGAC-3'
	<b>Extension</b>	5'-GGAGCAGCTGAACCTGGCT-3'
rs3087243	First	5'-ACGTTGGATGTACCTGCAAGTCATTCTTGG-3'
	Second	5'-ACGTTGGATGCCTGTGTTAAACAGCATGCC-3'
		Extension 5'-CCACCACTATTTGGGATATAAC-3'

Therefore, we established a case-control study to investigate the possible association between CTLA4 SNPs and BA susceptibility.

### Subjects and methods

#### *Subjects*

This study was approved by the ethics committee of the Children's Hospital of Fudan University. Written informed consent was obtained from all subjects' parents or legal guardians before blood samples collection.

BA patients. For this case-control study, we analyzed a total of 113 unrelated Chinese children with BA (70 boys and 43 girls) from the Children's Hospital of Fudan University (Shanghai, China). These BA children were diagnosed by exploratory laparotomy with operative cholangiography between August 2014 and July 2015. The age (mean  $\pm$  standard deviation) at the time of the operation of these patients was 68.1 ± 20.7 days (range 23-163 days).

Healthy controls. 133 unrelated healthy Chinese children (85 boys and 48 girls) from the Department of Pediatrics were recruited randomly as the healthy controls. All these healthy children were of similar age with the case group and did not show any signs of BA or other liver disease.

## *Genotyping*

Genomic DNA from patients and controls was extracted from whole blood using the TIANamp Blood DNA Kit (Tiangen, Beijing, China), according to the manufacturer's protocol. The three observed SNPs (rs231725, rs231775 and rs308- 7243) were selected based on previous reports [9, 10], with minor allele frequencies >5% according to the National Center for Biotechnology Information SNP Database (dbSNP) (http://www.ncbi. nlm.nih.bov/SNP/). These SNPs were all located in the CTLA4 gene, SNPs rs231725 and rs3087243 in the 3'-untranslated region (3'- UTR), and SNP rs231775 in exon 1 (Table 1). Primers for PCR and single-base extension were designed by the Assay Designers software,

version 3.0 (Sequenom, San Diego, CA, USA), and synthesized by Benegene Biotech (Shanghai, China; Table 2). All genotyping was performed using the MassARRAY on a matrixassisted laser desorption ionization-time of flight mass spectrometry platform and analyzed using the MassARRAY Typer software, version 3.4 (Sequenom). The concentration of DNA samples should be more than 10 ng/ul, and OD 260/280 should between 1.6-2.0 before the analysis. Negative and positive control samples were used as quality controls to monitor and control all the process.

## *Statistical analysis*

The Hardy-Weinberg equilibrium (HWE) test was done for each SNP for the case and control groups. The significance in allele and genotype frequencies between BA children and healthy controls was assessed using the χ2 test. A *p* value of less than 0.05 was considered to be statistically significant. The odds ratio (OR) and 95% confidence intervals (CI) were also calculated. Those statistical analysis were performed by the SPSS 18.0 program (SPSS Inc., Chicago, IL, USA). The haplotype frequencies of CTLA4 were estimated using the Haploview 4.2 program (http://www.broad.mit.edu/mpg/ haploview/).

	Case, $n$ $%$	Control, $n$ $%$	P value	OR (95% CI)
A	137 (0.606)	174 (0.654)	0.2718	$0.814(0.564 - 1.175)$
G	89 (0.394)	92 (0.346)		
A	77 (0.341)	75 (0.282)	0.1599	1.316 (0.897-1.931)
G	149 (0.659)	191 (0.718)		
A	46 (0.204)	37 (0.139)	0.0572	1.582 (0.984-2.543)
G	180 (0.796)	229 (0.861)		

Table 3. Allele frequencies of the three SNPs in the CTLA4 gene of patients with BA and control group

OR, odds ratio; CI, confidence interval.

Table 4. Genotype frequencies of the three SNPs in the CTLA4 gene of patients with BA and control group

o - - - -					
Genotype		Case, $n$ $%$	Control, $n$ $%$	P value	OR (95% CI)
rs231725	AG	59 (0.522)	56 (0.421)		
	AA	39 (0.345)	59 (0.444)		
	GG	15(0.133)	18 (0.135)	0.2435	
	AA+AG	98 (0.867)	115 (0.865)	0.9525	1.023 (0.490-2.135)
rs231775	AG	55 (0.487)	51(0.383)		
	AA	11 (0.097)	12 (0.090)		
	GG	47 (0.416)	70 (0.526)	0.2112	
	AG+GG	102 (0.903)	121 (0.910)	0.8484	$0.92(0.389 - 2.172)$
rs3087243	AG	38 (0.336)	31(0.233)		
	AA	4(0.035)	3(0.023)		
	GG	71 (0.628)	99 (0.744)	0.1448	
	AG+GG	109 (0.965)	130 (0.977)	0.5461	$0.629(0.138-2.871)$

OR, odds ratio; CI, confidence interval; HWE, Hardy-Weinberg equilibrium.

#### Table 5. CTLA4 haplotypes in patients with BA and control group



## **Results**

In total, three SNPs in the CTLA4 gene of 246 subjects (113 BA patients and 133 healthy controls) were genotyped. The three examined SNPs in both groups were in HWE, as showed in Table 1. And the minor allele frequencies of all SNPs were greater than 5%. The allele and genotype frequencies of the three SNPs in the CTLA4 gene are listed in Tables 3 and 4. We found no significant differences between BA patients and healthy controls ( $rs231725$ ,  $P =$ 0.2718, OR = 0.814, 95% CI = 0.564-1.175; rs231775, P = 0.1599, OR = 1.316, 95% CI = 0.897-1.931; rs3087243, P = 0.0572, OR = 1.582, 95% CI = 0.984-2.543). Neither when

we compare the combination of the homozygous and heterozygous genotypes of the major allele with other genotypes in each of the three SNPs. Distributions of haplotypes were also observed for the three SNPs (Table 5), which showed no significant difference between BA patients and healthy controls, either.

## **Discussion**

On the basis of former findings, we proposed the hypothesis that CTLA4 polymorphisms may be associated with BA susceptibility. However, the three CTLA4 SNPs analyzed in our casecontrol study demonstrated no significant association with BA.

The etiology of BA is multifactorial and unclear. Among those possible explanations, it is widely acknowledged that genetic variation and immune dysregulation are likely to play strong roles [3, 5]. In recent years, studies investigating genetic variants as susceptibility factors for BA are garnering interest. Some studies discovered a significant association between BA and

2848 Int J Clin Exp Pathol 2018;11(5):2846-2851

SNP rs17095355 on chromosome 10q24, located between XPNPEP1 and ADD3 genes, suggesting that SNPs in XPNPEP1 and ADD3 may serve as a BA susceptibility factor or disease modifier [2, 3, 5-7]. Meanwhile, other variations such as adiponectin SNP (+276G/T, rs1501299), ITGB2 3'-UTR+145C/A and the deletion on chromosome 2q37.3 are also conferred increased susceptibility to BA [1, 12, 13]. More significant, some of these BA-susceptible genes are involved in immune regulations, which indicates that genetic polymorphisms may be associated with the development of inflammation and fibrosis in the affected liver and lead to BA susceptibility [1].

Evidenced by the infiltration of inflammatory cells in liver and the injury of bile duct, the immune dysregulation is also a central pathogenesis of BA [1, 3, 5, 14-19]. Cholangiocytes initially respond to cellular injury through proinflammatory factors and develop fibrosis progression and biliary tree obliteration when the patient's immune system is imbalanced, resulting in liver fibrosis and extrahepatic bile ducts atresia [2]. Treg is an important factor in balancing the immune response and can prevent activation of autoreactive T cells. Deficiency of Treg quantity and/or function would weaken inflammation or autoimmunity inhibition and result in an exaggerated inflammatory response leading to bile duct injury [5, 19]. Previous studies have shown that dysregulation of Tregs is present in murine BA and diminished Treg function may be implicated in the BA etiology [20-22].

CTLA4 and CD28 are two potential targets for Treg manipulation [10]. Encoded by CTLA4 gene located on human chromosome 2q33, CTLA4 is a glycoprotein receptor expressed on the surface of a subset of Treg. It is a homologous molecule of CD28 with an opposing role. The CD28 promotes T cell activation, while CTLA4 competitively binds to B7 ligands (CD80 and CD86) on the surface of antigen-presenting cells (APCs) and delivers inhibitory signals [23, 24]. Tregs capture B7 ligands from APCs, which requires the help of CTLA4 on its surface [25, 26]. In vitro and in vivo studies have shown CTLA4 mutations are associated with Treg frequency and function [11, 26, 27]. Therefore, CTLA4 is a key factor in Treg homeostasis and immune tolerance, while the abnormal expression of CTLA4 can induce autoimmune diseases [11].

Furthermore, the three CTLA4 SNPs selected in our study have been demonstrated to have an association with liver damage and some autoimmune diseases [8, 23, 24, 28-30]. The rs-231725 was proved to be significantly associated with primary biliary cirrhosis (PBC) [28, 31-34]. The rs231775 (+49A/G) is related to threonine or alanine dimorphism, which can reduce the inhibitory function of CTLA4 and contributes to autoimmune diseases such as Grave's disease (GD), type 1 diabetes, PBC and systemic lupus erythematous disease [24, 28, 29, 35]. The rs3087243 (CT60) has been reported to influence the production of the soluble isoform of CTLA4 (sCTLA4, secreted by resting T cells and can suppress T-cell activation) [8, 29, 35] and significantly correlated with GD and Hashimoto's thyroiditis [23, 30, 35]. Therefore, we proposed a potential association between the three SNPs with BA susceptibility.

However, our study have some limitations. First, the small sample size may be not enough to discover positive results. Then, multiple comparisons between different subgroups such as gender and geographical regions have not been investigated. Moreover, the limited selection of SNPs in our study may be not widely cover the susceptible genes.

In conclusion, our case-control study is the first one to indicate that there is no significant association between the CTLA4 SNPs (rs231725, rs231775 and rs3087243) and BA susceptibility in Chinese children. Elucidating the role of genetic polymorphisms may provide a better understanding of the etiology and develop new therapeutic strategies both in prevention and treatment. We advocate for further studies with bigger populations, and wider range of CTLA4 SNPs on the associations between BA and CTLA4.

## Acknowledgements

This study received financial support from National Key Clinical Specialty Construction Programs of China (2014-2016), Shanghai 'Non key-in-key discipline' Clinical medical centers (2014-2016), Shanghai Hospital Development Center (SHDC12014106), National Natural Science Foundation of China (no. 81370472, no. 81300517, no. 81401243 and no. 81500394), Shanghai City Health Bureau for Youth Scientific Fund Project (no. 20134y100), Shanghai Rising-Star Program (A type) (no. 15QA1400800) and The Science Foundation of Shanghai (no. 13ZR1451800, no. 14ZR1404000, and no. 14411969860).

#### Disclosure of conflict of interest

None.

Address correspondence to: Dr. Shan Zheng, Department of Pediatric Surgery, Children's Hospital of Fudan University, and The Key Laboratory of Neonatal Disease, Chinese Ministry of Health, 399 Wan Yuan Road, Shanghai 201102, China. Tel: +8618017591199; Fax: +86-021-64931912; E-mail: [szheng@shmu.edu.cn](mailto:szheng@shmu.edu.cn)

#### References

- [1] Udomsinprasert W, Tencomnao T, Honsawek S, Anomasiri W, Vejchapipat P, Chongsrisawat V and Poovorawan Y. +276 G/T single nucleotide polymorphism of the adiponectin gene is associated with the susceptibility to biliary atresia. World J Pediatr 2012; 8: 328-334.
- [2] Garcia-Barcelo MM, Yeung MY, Miao XP, Tang CS, Cheng G, So MT, Ngan ES, Lui VC, Chen Y, Liu XL, Hui KJ, Li L, Guo WH, Sun XB, Tou JF, Chan KW, Wu XZ, Song YQ, Chan D, Cheung K, Chung PH, Wong KK, Sham PC, Cherny SS and Tam PK. Genome-wide association study identifies a susceptibility locus for biliary atresia on 10q24.2. Hum Mol Genet 2010; 19: 2917- 2925.
- [3] Asai A, Miethke A and Bezerra JA. Pathogenesis of biliary atresia: defining biology to understand clinical phenotypes. Nat Rev Gastroenterol Hepatol 2015; 12: 342-352.
- [4] Bijl EJ, Bharwani KD, Houwen RH and de Man RA. The long-term outcome of the Kasai operation in patients with biliary atresia: a systematic review. Neth J Med 2013; 71: 170-173.
- [5] Feldman AG and Mack CL. Biliary atresia: cellular dynamics and immune dysregulation. Semin Pediatr Surg 2012; 21: 192-200.
- [6] Cheng G, Tang CS, Wong EH, Cheng WW, So MT, Miao X, Zhang R, Cui L, Liu X, Ngan ES, Lui VC, Chung PH, Chan IH, Liu J, Zhong W, Xia H, Yu J, Qiu X, Wu XZ, Wang B, Dong X, Tou J, Huang L, Yi B, Ren H, Chan EK, Ye K, O'Reilly PF, Wong KK, Sham PC, Cherny SS, Tam PK and Garcia-Barcelo MM. Common genetic variants regulating ADD3 gene expression alter biliary atresia risk. J Hepatol 2013; 59: 1285- 1291.
- [7] Zeng S, Sun P, Chen Z, Mao J, Wang J, Wang B and Liu L. Association between single nucleotide polymorphisms in the ADD3 gene and susceptibility to biliary atresia. PLoS One 2014; 9: e107977.
- [8] de Jong VM, Zaldumbide A, van der Slik AR, Laban S, Koeleman BP and Roep BO. Variation in the CTLA4 3'UTR has phenotypic consequences for autoreactive T cells and associates with genetic risk for type 1 diabetes. Genes Immun 2016; 17: 75-78.
- [9] Fan LY, Tu XQ, Cheng QB, Zhu Y, Feltens R, Pfeiffer T and Zhong RQ. Cytotoxic T lymphocyte associated antigen-4 gene polymorphisms confer susceptibility to primary biliary cirrhosis and autoimmune hepatitis in Chinese population. World J Gastroenterol 2004; 10: 3056- 3059.
- [10] Kavanagh B, O'Brien S, Lee D, Hou Y, Weinberg V, Rini B, Allison JP, Small EJ and Fong L. CTLA4 blockade expands FoxP3+ regulatory and activated effector CD4+ T cells in a dose-dependent fashion. Blood 2008; 112: 1175-1183.
- [11] Tang AL, Teijaro JR, Njau MN, Chandran SS, Azimzadeh A, Nadler SG, Rothstein DM and Farber DL. CTLA4 expression is an indicator and regulator of steady-state CD4+ FoxP3+ T cell homeostasis. J Immunol 2008; 181: 1806- 1813.
- [12] Zhao R, Song Z, Dong R, Li H, Shen C and Zheng S. Polymorphism of ITGB2 gene 3'-UTR+145C/A is associated with biliary atresia. Digestion 2013; 88: 65-71.
- [13] Leyva-Vega M, Gerfen J, Thiel BD, Jurkiewicz D, Rand EB, Pawlowska J, Kaminska D, Russo P, Gai X, Krantz ID, Kamath BM, Hakonarson H, Haber BA and Spinner NB. Genomic alterations in biliary atresia suggest region of potential disease susceptibility in 2q37.3. Am J Med Genet A 2010; 152A: 886-895.
- [14] Mack CL, Feldman AG and Sokol RJ. Clues to the etiology of bile duct injury in biliary atresia. Semin Liver Dis 2012; 32: 307-316.
- [15] Bessho K and Bezerra JA. Biliary atresia: will blocking inflammation tame the disease? Annu Rev Med 2011; 62: 171-185.
- [16] Petersen C and Davenport M. Aetiology of biliary atresia: what is actually known? Orphanet J Rare Dis 2013; 8: 128.
- [17] Bezerra JA, Tiao G, Ryckman FC, Alonso M, Sabla GE, Shneider B, Sokol RJ and Aronow BJ. Genetic induction of proinflammatory immunity in children with biliary atresia. Lancet 2002; 360: 1653-1659.
- [18] Mack CL, Falta MT, Sullivan AK, Karrer F, Sokol RJ, Freed BM and Fontenot AP. Oligoclonal expansions of CD4+ and CD8+ T-cells in the target organ of patients with biliary atresia. Gastroenterology 2007; 133: 278-287.
- [19] Brindley SM, Lanham AM, Karrer FM, Tucker RM, Fontenot AP and Mack CL. Cytomegalovirus-specific T-cell reactivity in biliary atresia at the time of diagnosis is associated with defi-

cits in regulatory T cells. Hepatology 2012; 55: 1130-1138.

- [20] Li K, Zhang X, Tang ST, Yang L, Cao GQ, Li S and Yang DH. gammadelta T cells and Foxp3(+) Treg cells infiltration in children with biliary atresia and its significance. Int J Clin Exp Med 2015; 8: 18512-18517.
- [21] Tucker RM, Feldman AG, Fenner EK and Mack CL. Regulatory T cells inhibit Th1 cell-mediated bile duct injury in murine biliary atresia. J Hepatol 2013; 59: 790-796.
- [22] Miethke AG, Saxena V, Shivakumar P, Sabla GE, Simmons J and Chougnet CA. Post-natal paucity of regulatory T cells and control of NK cell activation in experimental biliary atresia. J Hepatol 2010; 52: 718-726.
- [23] Fang W, Zhang Z, Zhang J, Cai Z, Zeng H, Chen M and Huang J. Association of the gene CT60/ rs3087243 single-nucleotide polymorphisms with Graves' disease. Biomed Rep 2015; 3: 691-696.
- [24] Chen Z, Fei M, Fu D, Zhang L, Ma Y, Wang Y, Zhang F, Xia Q and Wang X. Association between cytotoxic T lymphocyte antigen-4 polymorphism and type 1 diabetes: a meta-analysis. Gene 2013; 516: 263-270.
- [25] Matheu MP, Othy S, Greenberg ML, Dong TX, Schuijs M, Deswarte K, Hammad H, Lambrecht BN, Parker I and Cahalan MD. Imaging regulatory T cell dynamics and CTLA4-mediated suppression of T cell priming. Nat Commun 2015; 6: 6219.
- [26] Kuehn HS, Ouyang W, Lo B, Deenick EK, Niemela JE, Avery DT, Schickel JN, Tran DQ, Stoddard J, Zhang Y, Frucht DM, Dumitriu B, Scheinberg P, Folio LR, Frein CA, Price S, Koh C, Heller T, Seroogy CM, Huttenlocher A, Rao VK, Su HC, Kleiner D, Notarangelo LD, Rampertaap Y, Olivier KN, McElwee J, Hughes J, Pittaluga S, Oliveira JB, Meffre E, Fleisher TA, Holland SM, Lenardo MJ, Tangye SG and Uzel G. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. Science 2014; 345: 1623-1627.
- [27] Schubert D, Bode C, Kenefeck R, Hou TZ, Wing JB, Kennedy A, Bulashevska A, Petersen BS, Schaffer AA, Gruning BA, Unger S, Frede N, Baumann U, Witte T, Schmidt RE, Dueckers G, Niehues T, SeneviratneS, Kanariou M, Speckmann C, Ehl S, Rensing-Ehl A, Warnatz K, Rakhmanov M, Thimme R, Hasselblatt P, Emmerich F, Cathomen T, Backofen R, Fisch P, Seidl M, May A, Schmitt-Graeff A, Ikemizu S, Salzer U, Franke A, Sakaguchi S, Walker LS, Sansom DM and Grimbacher B. Autosomal dominant immune dysregulation syndrome in humans with CTLA4 mutations. Nat Med 2014; 20: 1410-1416.
- [28] Juran BD, Atkinson EJ, Schlicht EM, Fridley BL and Lazaridis KN. Primary biliary cirrhosis is associated with a genetic variant in the 3' flanking region of the CTLA4 gene. Gastroenterology 2008; 135: 1200-1206.
- [29] Joshita S, Umemura T, Yoshizawa K, Katsuyama Y, Tanaka E, Nakamura M, Ishibashi H and Ota M. Association analysis of cytotoxic Tlymphocyte antigen 4 gene polymorphisms with primary biliary cirrhosis in Japanese patients. J Hepatol 2010; 53: 537-541.
- [30] Zhao SX, Pan CM, Cao HM, Han B, Shi JY, Liang J, Gao GQ, Peng YD, Su Q, Chen JL, Zhao JJ and Song HD. Association of the CTLA4 gene with Graves' disease in the Chinese Han population. PLoS One 2010; 5: e9821.
- [31] Li Q, Wang B, Pan F, Zhang R, Xiao L, Guo H, Ma S and Zhou C. Association between cytotoxic T-lymphocyte antigen 4 gene polymorphisms and primary biliary cirrhosis in Chinese population: data from a multicenter study. J Gastroenterol Hepatol 2013; 28: 1397-1402.
- [32] Li M, Zheng H, Li T, Gao P, Zhang XL and Liu DW. Cytotoxic T-lymphocyte associated antigen-4 gene polymorphisms and primary biliary cirrhosis: a systematic review. J Gastroenterol Hepatol 2012; 27: 1159-1166.
- [33] Aiba Y, Nakamura M, Joshita S, Inamine T, Komori A, Yoshizawa K, Umemura T, Horie H, Migita K, Yatsuhashi H, Nakamuta M, Fukushima N, Saoshiro T, Hayashi S, Kouno H, Ota H, Muro T, Watanabe Y, Nakamura Y, Komeda T, Shimada M, Masaki N, Komatsu T, Yagura M, Sugi K, Koga M, Tsukamoto K, Tanaka E and Ishibashi H. Genetic polymorphisms in CTLA4 and SLC4A2 are differentially associated with the pathogenesis of primary biliary cirrhosis in Japanese patients. J Gastroenterol 2011; 46: 1203-1212.
- [34] Donaldson P, Veeramani S, Baragiotta A, Floreani A, Venturi C, Pearce S, Wilson V, Jones D, James O, Taylor J, Newton J and Bassendine M. Cytotoxic T-lymphocyte-associated antigen-4 single nucleotide polymorphisms and haplotypes in primary biliary cirrhosis. Clin Gastroenterol Hepatol 2007; 5: 755-760.
- [35] Jaiswal PK, Singh V and Mittal RD. Cytotoxic T lymphocyte antigen 4 (CTLA4) gene polymorphism with bladder cancer risk in North Indian population. Mol Biol Rep 2014; 41: 799-807.