



RESEARCH ARTICLE

REVISED **Genome-wide CRISPR screening identifies new regulators of glycoprotein secretion [version 2; peer review: 2 approved]**

Stephanie Popa ¹, Julien Villeneuve¹, Sarah Stewart ¹, Esther Perez Garcia², Anna Petrunkina Harrison², Kevin Moreau¹

¹University of Cambridge Metabolic Research Laboratories, Wellcome Trust-Medical Research Council Institute of Metabolic Science, University of Cambridge, Cambridge, CB2 0QQ, UK

²NIHR Cambridge BRC Cell Phenotyping Hub, Department of Medicine, University of Cambridge, Cambridge, CB2 0QQ, UK

v2 **First published:** 09 Aug 2019, 4:119 (<https://doi.org/10.12688/wellcomeopenres.15232.1>)
Latest published: 16 Jan 2020, 4:119 (<https://doi.org/10.12688/wellcomeopenres.15232.2>)

Abstract

Background: The fundamental process of protein secretion from eukaryotic cells has been well described for many years, yet gaps in our understanding of how this process is regulated remain.

Methods: With the aim of identifying novel genes involved in the secretion of glycoproteins, we used a screening pipeline consisting of a pooled genome-wide CRISPR screen, followed by secondary siRNA screening of the hits to identify and validate several novel regulators of protein secretion.

Results: We present approximately 50 novel genes not previously associated with protein secretion, many of which also had an effect on the structure of the Golgi apparatus. We further studied a small selection of hits to investigate their subcellular localisation. One of these, GPR161, is a novel Golgi-resident protein that we propose maintains Golgi structure via an interaction with golgin A5.

Conclusions: This study has identified new factors for protein secretion involved in Golgi homeostasis.

Keywords

secretion, CRISPR screen, Golgi, endoplasmic reticulum

Open Peer Review

Reviewer Status

	Invited Reviewers	
	1	2
version 2 (revision) 16 Jan 2020	 report	 report
version 1 09 Aug 2019	 report	 report

- Hesso Farhan** , University of Oslo, Oslo, Norway
- David Stephens** , University of Bristol, Bristol, UK

Any reports and responses or comments on the article can be found at the end of the article.

Corresponding author: Kevin Moreau (kevin.moreau@astrazeneca.com)

Author roles: **Popa S:** Conceptualization, Formal Analysis, Funding Acquisition, Investigation, Methodology, Validation, Visualization, Writing – Original Draft Preparation; **Villeneuve J:** Investigation, Methodology, Validation, Visualization, Writing – Review & Editing; **Stewart S:** Data Curation, Funding Acquisition, Writing – Review & Editing; **Perez Garcia E:** Methodology, Writing – Review & Editing; **Petrunkina Harrison A:** Funding Acquisition, Methodology, Writing – Review & Editing; **Moreau K:** Conceptualization, Formal Analysis, Funding Acquisition, Investigation, Methodology, Project Administration, Resources, Supervision, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: This work was supported by a Wellcome Trust PhD studentship for S.J.P. [109152], a Wellcome Trust Strategic Award [100574], the Medical Research Council Metabolic Diseases Unit [MRC_MC_UU_12012/5], the Isaac Newton Trust/Wellcome Trust ISSF/ University of Cambridge joint research grant for J.V. and K.M., Biotechnology and Biological Science Research Council Future Leader Fellowship for S.E.S and the National Institute for Health Research for E.P.G. and A.P.H.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2020 Popa S *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Popa S, Villeneuve J, Stewart S *et al.* **Genome-wide CRISPR screening identifies new regulators of glycoprotein secretion [version 2; peer review: 2 approved]** Wellcome Open Research 2020, 4:119 (<https://doi.org/10.12688/wellcomeopenres.15232.2>)

First published: 09 Aug 2019, 4:119 (<https://doi.org/10.12688/wellcomeopenres.15232.1>)

REVISED Amendments from Version 1

This version includes modifications suggested by the reviewers, which was essentially text corrections and additional discussion of the data with previous literature.

Any further responses from the reviewers can be found at the end of the article

Introduction

Protein secretion is a fundamental and well-known process in cell biology, in which proteins are transported from the endoplasmic reticulum (ER) to the Golgi via coat protein-coated vesicles and subsequently to the plasma membrane¹⁻³. The localisation and activity of proteins involved in this secretory pathway must be tightly regulated to ensure correct spatio-temporal distribution of membranes and cargo proteins along the pathway. While the molecular machinery of secretion is relatively well understood, our knowledge remains incomplete, particularly regarding the regulation of protein secretion. For example, evidence suggests that multiple trafficking routes for transport within the Golgi exist⁴. The complex process of sorting proteins at the Golgi into vesicles for their correct destinations also requires further investigation⁵⁻⁷.

Recently, genome-wide CRISPR screening has emerged as a powerful strategy to identify novel gene functions; this type of screening has the advantages of being unbiased and more reliable than previously-used methods, in terms of introducing genetic mutations⁸. As such, it provides an opportunity to uncover new information about the regulation of protein secretion. Pooled CRISPR screening has been used for a wide range of applications, including the investigation of drug-resistance mechanisms in cancer cells⁹, the genetics of pluripotency¹⁰, autophagy regulators^{11,12} and host factors required for viral infection¹³. We previously demonstrated that unbiased pooled genome-wide CRISPR screening could effectively reveal key players required for glycoprotein secretion, using galectin-3 retention at the cell surface to assess glycoprotein secretion¹⁴. Galectin-3 is a cytosolic protein that is secreted without entering the conventional secretory pathway for its export to the extracellular space. Vesicular and non-vesicular modes of secretion have been proposed, but the precise series of events involved in the unconventional secretion of galectin-3 remain ill defined¹⁵. Once outside the cell, galectin-3 binds to β -galactosides resident on the cell surface¹⁶ and can therefore be used as an indirect measure of glycoprotein transport to the cell surface via the ER-Golgi secretory pathway. Using a binding partner of glycans as a readout, rather than one specific glycoprotein, allows regulators of general glycoprotein secretion to be discovered. It is also possible that the hits identified may not exclusively regulate glycoprotein secretion but may be more general regulators of more protein secretion; further experiments would be required to make this distinction.

Here, we devised a powerful experimental pipeline, using an improved pooled genome-wide CRISPR screen, followed by two arrayed secondary screening methods using siRNA knock-down to identify new factors involved in glycoprotein secretion and Golgi apparatus architecture. Combining these methods

allows the speed and reduced cost of pooled CRISPR screening to be taken together with the advantages of an arrayed siRNA screen in which genotype and phenotype remain linked¹⁷. Using this approach, we were able to validate 55 novel hits that are important for glycoprotein secretion. We found that many of these hits are also important for maintenance of the Golgi architecture. One of these hits, GPR161, is a novel Golgi-localised protein that decreases glycoprotein secretion and disrupts the Golgi structure.

Methods**Cell culture**

HeLa cells, suspension HeLa cells expressing Cas9 (sHeLa-Cas9) and HeLa cells expressing horseradish peroxidase fused to a signal sequence to direct its secretion (HeLa-ss-HRP) were cultured in 10% culture medium: 10% (v/v) FBS, 100 U Penicillin / 0.1 mg ml⁻¹ streptomycin, 2 mM L-glutamine in high glucose DMEM, in 5% CO₂ at 37°C. See [Table 1](#) for a list of all reagents used and their sources.

CRISPR screen: transduction with Brunello library

24 h before transduction, suspension HeLa cells were passaged. The lentiviral Brunello library, which targets 19,114 genes in the human genome with four unique guides per gene¹⁸, was used to transduce 1×10^8 suspension HeLa cells expressing Cas9 (sHeLa-Cas9) in polybrene media (10 μ g ml⁻¹ polybrene, 10% (v/v) FBS, 100 U Penicillin / 0.1 mg ml⁻¹ streptomycin, 2 mM L-glutamine in high glucose DMEM (Sigma, D6546)) at an MOI of 0.50 by spinoculation at 1000 \times g for 30 min at 20°C. This gave a transduction efficiency of 40% and therefore an overall percentage of 31% cells with one guide per cell, as calculated using the Poisson distribution for the probability that a cell will be infected with at least one virus, $1 - P(0, \text{MOI})$ ¹⁹. Cells were incubated for 24 h at 37°C, 5% CO₂ before passage into puromycin media (1 μ g ml⁻¹ puromycin, 10% (v/v) FBS, 100 U Penicillin / 0.1 mg ml⁻¹ streptomycin, 2 mM L-glutamine in high glucose DMEM). Cells were cultured in puromycin media for seven days to retain only transduced cells. The transduction efficiency was measured as the percentage of live cells after 48 h in puromycin media.

CRISPR screen: cell sorting by flow cytometry

Cells were washed once with DMEM, incubated with 50 μ g ml⁻¹ anti-galectin-3 antibody conjugated to Alexa Fluor 647 for 30 min at 4°C, then washed again. Approximately 3×10^7 cells were sorted at the NIHR Cambridge BRC cell phenotyping hub on a BD FACS AriaIII and FACS Aria Fusion (using a 100 μ m Nozzle and run at a pressure of 25 psi); Alexa Fluor 647 fluorescence was detected with a 670/30 BP detector on the AriaIII and Aria Fusion cell sorters. Target cell population to be sorted was gated based on the lowest anti-galectin-3-AF647 fluorescent signal, using Purity as sort precision mode. After cell sorting, cells were cultured in 20% serum media for 24–48 h, then cultured in culture media. When the population had expanded to 5×10^7 cells, two samples were taken; 5×10^6 cells were frozen for guide sequencing, and 1×10^6 were assessed by flow cytometry. Gating leniency was decreased with progressive rounds, as shown in [Figure 1](#). After the third sort, two distinct negative populations were observed, so a fourth sort was performed to separate these two populations,

Table 1. List of reagents.

Reagent or resource	Source	Identifier
Antibodies		
Rat anti-galectin-3-647 mAbFlow cytometry concentration 50 µg ml ⁻¹	BioLegend	125408, RRID: AB_1186110
Sheep antiTGN46 pAb, IF concentration 5 µg ml ⁻¹	BioRad	AHP500, RRID: AB_324049
Mouse anti-GM130 mAb, IF concentration 25 µg ml ⁻¹	BD Biosciences	610822, RRID: AB_398141
Mouse anti-Sec31 mAb, IF concentration 25 µg ml ⁻¹	BD Biosciences	612351, RRID: AB_39971
Mouse anti-MHC-I mAb, IF concentration 20 µg ml ⁻¹	ThermoFisher	MA1-70111, RRID: AB_1076705
Rabbit anti-calnexin pAb, IF concentration 5 µg ml ⁻¹	Abcam	ab22595, RRID: AB_2069006
Rabbit anti-GPR161 pAb, IF concentration 10 µg ml ⁻¹	MyBioSource	MBS719938, RRID: AB_2801289
Rabbit anti-TMEM220 pAb, IF concentration 100 µg ml ⁻¹	Aviva Systems Biology	ARP44467_P050, RRID: AB_2801328
Rabbit anti-Golgin A5 pAb, IF concentration 10 µg ml ⁻¹	GeneTex	GTX104255, RRID: AB_2037117
Donkey anti-sheep IgG-568 pAb, IF concentration 4 µg ml ⁻¹	ThermoFisher	A21099, RRID: AB_2535753
Goat anti-mouse IgG-488 pAb, IF concentration 4 µg ml ⁻¹	ThermoFisher	A11001, RRID: AB_2534069
Goat anti-rabbit IgG-568 pAb, IF concentration 4 µg ml ⁻¹	ThermoFisher	A11011, RRID: AB_143157
Mouse anti-FLAG M2 mAb, IF concentration 0.5 µg ml ⁻¹	Sigma	F1804, RRID: AB_262044
Cell lines		
HeLa	ATCC	CCL-2
HeLa-HRP	Gift from Vivek Malhotra	As described ²⁰
sHeLa (HeLa S3)	Gift from Paul Lehner	Sigma, 87110901
sHeLa-Cas9	In house production	As described ²¹
Genome-wide screen library		
Lentiviral Brunello	David Root and John Doench, Addgene	73178-LV
Plasmids		
Mouse-GPR161-FLAG	Insight Biotechnology LTD	MR224248
Human-TMEM220-FLAG	Genescript USA INC	OHu14539
Human-COL14A1-FLAG	Strattech Scientific LTD	HG13622-CF-SIB
Mouse-FAM98B-FLAG	Insight Biotechnology LTD	MR206836
Mouse-FAM102B-FLAG	Insight Biotechnology LTD	MR212923
Mouse-MXRA7-FLAG	Insight Biotechnology LTD	MR218215
Software		
MaGECK	https://sourceforge.net/p/mageck/wiki/Home/	Version 0.5.7
MaGECK-vispr	https://bitbucket.org/liulab/mageck-vispr	Version 0.5.3
GraphPad PRISM	GraphPad	Version 7
GO term mapper	https://go.princeton.edu/cgi-bin/GOTermMapper	Accessed 7 th Jan 2019
CellProfiler	http://cellprofiler.org/releases/	Version 3.1.5
CellProfiler Analyst	http://cellprofiler.org/releases/	Version 2.2.1
Fiji	https://imagej.net/Fiji/Downloads	ImageJ-win64, downloaded 25 th May 2018
R	https://www.r-project.org/	Version 3.5.1

Reagent or resource	Source	Identifier
Reagents		
High glucose Dulbecco's Modified Eagle Medium (DMEM)	Molecular Probes	D6546
Hank's Balanced Salt Solution (HBSS)	ThermoFisher	14025092
Fetal bovine serum (FBS)	ThermoFisher	10270-106
L-glutamine	Sigma	G7513
Penicillin/streptomycin	Sigma	P0781
Polybrene	Merck	TR-1003-G
Dulbecco's Modified Eagle Medium (DMEM)	Life Technologies	21063-029
Propidium iodide	Biologend	421301
Gentra Puregene Cell kit	Qiagen	158388
dNTP	ThermoFisher	R0191
ExTaq	Takara	RR001B
AMPure XP magnetic beads	Beckman Coulter	A63880
White 96 well CulturPlate	PerkinElmer	6005680
96 well ViewPlate	PerkinElmer	6005182
PBS	ThermoFisher	D8537
ProLong with DAPI	Invitrogen	P36962
Paraformaldehyde	Sigma	158127
TransIT 2020	Mirus	MIR 5405
Lipofectamine 2000	ThermoFisher	12566014
Phalloidin Green	ThermoFisher	A12379
ProLong Gold with DAPI	ThermoFisher	P36935
Tween 20	Sigma	P1379
ECL Western Blotting Detection Reagent	Amersham	RPN2106
Westar XLS100	Cyanagen	XLS100
Restore Plus	ThermoFisher	46430
Primers and siRNA		
P5 primer mix	Integrated DNA Technologies	See <i>Extended data</i> , E2 ²²
P7 primers	Integrated DNA Technologies	See <i>Extended data</i> , E3 ²²
siGenome smart pool plate	Dharmacon	G-CUSTOM-384690 (see <i>Extended data</i> , E4) ²²

gating each population around a 5–10% peak. For flow cytometry of 1×10^6 cells after cell sorting, cells were immunostained as above, with an additional propidium iodide stain for 5 min, and analysed on an Accuri™ C6 (BD Biosciences) equipped with lasers providing 488 nm and 640 nm excitation sources. Alexa Fluor 647 fluorescence was detected with an FL4 detector (675/25 BP).

CRISPR screen: Library preparation and sequencing

To prepare samples for sequencing, genomic DNA from the different cell populations was extracted from frozen cell pellets using Gentra Puregene Cell kit and concentration of DNA was measured on the Nanodrop. PCR was carried out in quadruplicate to amplify an amplicon containing the guide, with primers used to attach barcodes, stagger regions and sequencing

adaptors for use in sequencing, as previously described by Doench *et al.*¹⁸. Briefly, each well was set up to contain 10 µg genomic DNA, 0.5 µM uniquely barcoded P7 primer, 0.5 µM P5 stagger primer mix, 200 µM each dNTP, 7.5 units ExTaq and 1x ExTaq buffer in a total volume of 100 µl. PCR cycles were: initial denaturation step at 95°C for 1 min; 28 cycles of 95°C for 30 s, 53°C for 30 s, 72°C for 30s; final extension at 72°C for 10 min. One replicate from each sample was analysed on a 2% agarose gel to confirm amplification was successful. PCR products were pooled, adding 30 µl from each PCR reaction, then purified using AMPure XP magnetic beads to retain only DNA fragments larger than 100 bp¹⁸. An equal volume of the pooled product and AMPure XP magnetic beads were mixed at room temperature for 5 min, then placed on a magnet to retain beads. Beads were washed three times with 70% ethanol and purified

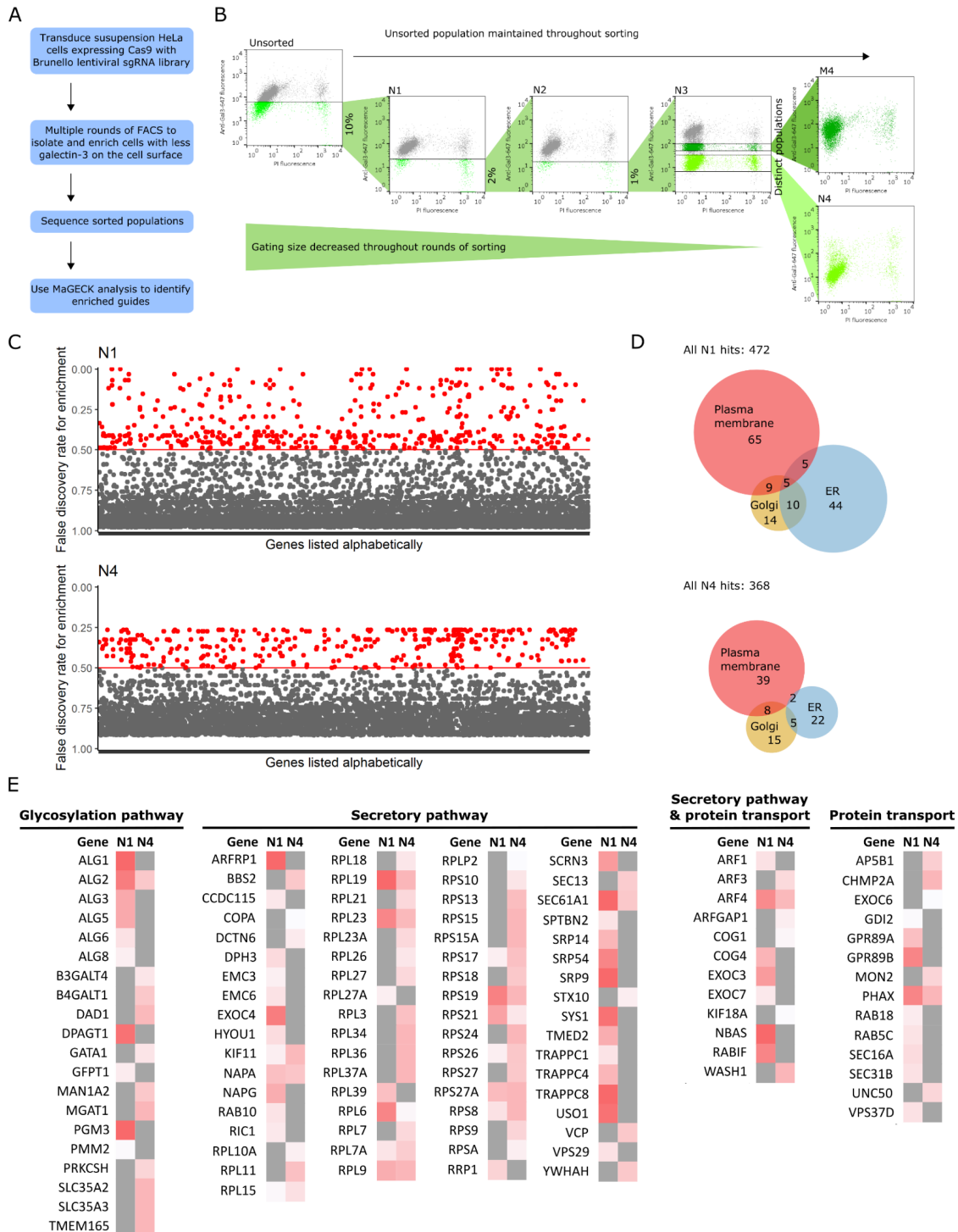


Figure 1. CRISPR screen. (A) Workflow of genome-wide CRISPR screen. (B) Multiple rounds of cell sorting by flow cytometry, decreasing gating percentage with progressive rounds, resulted in final populations of cells with a low level of galectin-3 on the cell surface. FACS plots are of samples taken after cells had been expanded after cell sorting, and show cell surface galectin-3 levels as measured by anti-galectin-3-647 fluorescence, and viability as measured by PI. Labels of FACS plots refer to population names: N1–4 = populations after negative sorts 1–4; M4, cells expressing medium level of galectin-3 after sort 4. (C) Hits from populations N1 and N4; genes with a false discovery rate (FDR) < 0.5 were defined as hit and shown in red. Genes with had guides that were enriched, but not significantly enriched (i.e. FDR ≥ 0.5), are shown in grey. (D) Euler diagram showing the number of hits known to be associated with organelles in the secretory pathway. Many ER-resident hits are lost when progressing from sorts N1 to N4. (E) Heat map of genes known to be in the secretory pathway, glycosylation pathway or involved in protein transport, as annotated by GO_Slim lists. Grey indicates an FDR greater than 0.5; hits are shown on a white-red scale, where darker red indicates a more significant FDR, closer to 0. The identity of hits changes significantly from populations N1 to N4.

PCR product was eluted with 500 μ l EB buffer. Sequencing was carried out by the CRUK genomics facility using SE50 sequencing on the Illumina HiSeq 4000.

CRISPR screen: Bioinformatics analysis using MaGECK

Data were analysed using both MaGECK-MLE²⁰ and MaGECK-RRA²¹ (version 0.5.7), using an enrichment (positive) sort type. In all analyses, median normalisation was used. PCR replicates were treated as technical replicates and sort replicates were treated as biological replicates. Each population was given an identification code; descriptions are available in data file S2 (see *Underlying data*)²². Genes were annotated with their function using UniProt²², and with the organelle they reside in using Gene Ontology subsets^{23,24}.

Secondary screen: HRP assay

Based on having an unknown or unclear function in secretion, hits from the MaGECK analyses were selected to create a shortlist of 368 genes to be used in a secondary screen. This shortlist was screened in an arrayed siRNA screen, using siGenome smart pools (Dharmacon) in which each plate contained two replicates of non-targeting siRNA as a negative control. To screen for conventional secretion, HeLa cells expressing ss-HRP were reverse transfected in with the secondary screen library, as previously described²⁵. Briefly, 80 μ l of 31 000 cells ml⁻¹ were plated into wells of a 96 well plate containing a mixture of 20 μ l 250 nM siRNA and 0.5% (v/v) lipofectamine 2000 in OptiMEM and incubated at 37°C, 5% CO₂. Medium was changed to fresh culture medium 48 h after transfection. 72 h after transfection both supernatants and cell lysates were assessed for HRP levels in white CulturPlate 96 well plates by chemiluminescence using a Tecan M1000 Pro. Luminescence signal from the cell lysate (CL), supernatant (SN) and the CL:SN ratio were each normalised to signal obtained from siRNA control cells according to the formula:

$$\frac{x - \text{mean background}}{\text{mean siRNA control} - \text{mean background}}$$

To identify genes required for HRP secretion, a line with a gradient of 1 was fit to the normalised CL vs normalised SN data. Outliers of this line, as identified by the ROUT method²⁶, are genes that result in an up or down regulation of HRP secretion. The same method was also used to identify outliers from the line of $y = 1$ for the normalised CL:SN ratio line. The [Gene Ontology \(GO\) Term Mapper](#) was used to classify hits identified into broad categories, using the ontology aspect “process” and ontology category “Generic slim” (GO_Slim) for Homo sapiens (GOA @EBI + Ensembl) proteins²⁷. A gene was defined as related to protein transport if it was annotated with any of the GO_Slim annotations: transport (GO:0006810), transmembrane transport (GO:0055085) or vesicle-mediated transport (GO:0016192).

Secondary screen: Golgi morphology

HeLa cells were reverse-transfected as described above, although here transfected in 96-well ViewPlates. At 72 h after transfection, cells were fixed in ice cold methanol for 5 min, washed in PBS, stained with appropriate primary and secondary

antibodies and ProLong with DAPI, and observed using an Opera Phenix High-Content Screening System to obtain unbiased confocal pictures. Analysis was performed on CellProfiler²⁸ (version 3.1.5) to define cells and Golgi, and to measure Golgi intensity, size, shape and granularity. Briefly, cells were defined as secondary objects by propagation from nuclei, using a global “minimum cross entropy” thresholding method. Golgi were defined within a broad pixel diameter range of 2-60 px, and using an adaptive, two-class, Otsu thresholding method. The classifier of CellProfiler Analyst (CPA)²⁹ (version 2.2.1) was then trained using cells randomly chosen from the whole experiment. These were classified as having aberrant or intact Golgi until the sensitivity reached an acceptable level: 82% and 77% for the intact Golgi and aberrant Golgi classes, respectively; here, 65 cells were used. The CPA classifier was then used to define all cells as having intact or aberrant Golgi.

Transfection with cDNA

HeLa cells at ~50% confluency were transfected with a mix of cDNA at 1 μ g μ l⁻¹ and 0.2% (v/v) TransIT in OptiMEM. Cells were incubated at 37°C, 5% CO₂ for 4 h, then medium was changed to fresh culture medium; cells were then incubated for a further 20 h.

Immunofluorescence microscopy

For immunofluorescence microscopy, cells were cultured on coverslips, fixed with either 4% paraformaldehyde in PBS for 5 min and permeabilised with 0.1% Triton X100 in PBS for 5 min, or fixed and permeabilised with ice cold methanol for 5 min. Cells were then blocked in 10% (v/v) FBS / 1x PBS for 30 min, incubated with primary antibodies for 2 h, washed three times with PBS, and incubated with secondary antibodies for 30 min. Primary and secondary antibodies, and the concentration they were used for immunofluorescence, are listed in [Table 1](#). Samples were mounted using ProLong Gold antifade reagent with DAPI and observed using a Leica SP8 laser confocal microscope.

Analysis of hits

To find interacting partners of proteins of interest, we used the [BioPlex network](#), a resource of proteins shown to interact by affinity purification-mass spectrometry^{30,31}. Default parameters were used to search the network.

Co-localisation analysis

To measure co-localisation between golgin A5 and TGN46, 17 cells were selected from each treatment from confocal micrographs and analysed using the coloc 2 plugin in Fiji (downloaded 25th May 2018)³². For cells with TMEM220 or GPR161 knocked down, only cells with aberrant Golgi were selected from confocal micrographs to be confident that siRNA knockdown was effective. Pearson’s coefficient with no threshold was recorded for each cell analysed. Data were tested for normality and equality of variance by the Shapiro-Wilk test and Bartlett’s K squared test, respectively. After passing normality tests, data were analysed by one-way ANOVA followed by Tukey HSD post-hoc test between groups. All statistical analysis was performed using R (version 3.5.1).

An earlier version of this article can be found on bioRxiv, DOI: <https://doi.org/10.1101/522334>.

Results

A genome-wide CRISPR screen revealed novel regulators of glycoprotein secretion

We carried out a genome-wide CRISPR screen with the aim of identifying novel genes involved in glycoprotein secretion, using the level of galectin-3 on the surface of live cells to look at glycoprotein secretion via the ER-Golgi pathway. The workflow for the screen is shown in [Figure 1A](#). Suspension HeLa cells stably expressing Cas9 were transduced with the Brunello lentiviral guide (sgRNA) library, an optimised library for the human genome¹⁸, at a low multiplicity of infection such that the majority of the transduced cells received exactly one sgRNA. After transduction, we performed three rounds of cell sorting by fluorescence activated with increasing selection stringency for low levels of cell surface galectin-3. This method initially allows more cells through cell sorting, then becomes gradually more stringent by decreasing the gating percentage, increasing the number of true positives in later sorted populations. This resulted in the enrichment of two distinct populations with low levels of cell-surface galectin-3 after the third sort, in population N3 ([Figure 1B](#)). As there are lots of cells with very little or no galectin-3 on the cell surface in population N3, the other populations may have more galectin-3, transferred from the extracellular pool, or a higher effective concentration of anti-galectin-3 antibody. A fourth round of cell sorting was used to separate these distinct populations, resulting in populations M4 and N4 ([Figure 1B](#)). We extracted genomic DNA from all populations and carried out deep sequencing to determine the raw sgRNA counts in each population, available in data file S1 (see *Underlying data*)²². MaGECK analysis allowed the identification of hits from population N1 onwards; full MaGECK results are available in data file S2 (see *Underlying data*)²². Hits enriched in populations N1 and N4 are shown in [Figure 1C](#).

The identity of the hits changed as cell sorting progressed. This was particularly true of hits which localise to the ER: in total, 64 hits localise to the ER in population N1, compared to 29 in population N4 ([Figure 1D](#)). As many genes in the ER are involved in critical biological processes, it is likely that these genes were lost in later sorts as cells lacking these genes have a growth or survival defect. For example, many of the enzymes involved in N-linked glycosylation, such as the asparagine-linked glycosylation (ALG) enzymes ALG1, ALG3, ALG5, ALG6 and ALG8, are identified as hits in population N1 but are lost in N4. Conversely, some hits are enriched after multiple cell sorting rounds in N4 but were not identified in N1 ([Figure 1E](#)). As such, we took a selection hits from across all populations forward into the secondary screen. MaGECK-MLE and MaGECK-RRA analysis revealed that there is little difference between the later populations: N3, M4 and N4 (data file S2, see *Underlying data*)²². As we intended to validate hits with further screening, we used a false discovery rate (FDR) of 0.5 to classify hits. This lenient FDR is validated by the presence of hits previously shown to play a role in secretory and glycosylation pathways, such as ribosomal proteins involved

in protein translocation into the ER machinery, enzymes involved in N-linked glycosylation, RAB proteins and trafficking protein particle complex (TRAPPC) proteins ([Figure 1E](#)). This shows that our screen was very efficient in identifying previously described regulators of glycoprotein secretion. Additionally, there were many hits identified that did not have a clear link to functions in protein secretion. Therefore, we hypothesised that many genes enriched in our screen with either no characterised function, or with no described function in the secretory pathway, may be new regulators of protein secretion.

An RNAi secondary screen validated hits from the CRISPR screen

Due to the lenient FDR used for MaGECK, it was important to validate the hits. Around 400 hits, identified from populations N1 - N4/M4 of the CRISPR screen, were selected for validation by secondary screening. Of these 400 hits, 113 had no annotated function in UniProt; 32 also had no GO annotations. As many of these hits were uncharacterised, they had the potential to represent novel regulators of protein secretion. Twelve positive controls annotated with GO terms related to protein secretion were also included in the screen to validate the statistical analysis. For secondary screening, we used siRNA knockdown of hits in HeLa cells that stably express ss-HRP, which is a glycosylated protein secreted into the extracellular space^{33,34}. HeLa-ss-HRP were transfected with arrayed siRNA in a 96 well plate. At 72 h after transfection, we collected the supernatant, lysed the cells and assessed HRP by chemiluminescence. Knockdown that led to a decrease in the ratio of supernatant luminescence to cell lysate luminescence indicated genes that were important for glycoprotein secretion. Knockdown of 92 of the genes screened resulted in less secretion of HRP ([Figure 2A](#)). Positive controls also resulted in less HRP secretion, as expected ([Figure 2A, B](#)). The 92 genes validated by the siRNA screen are listed in [Figure 2B and 2C](#). Of these hits, 55 were not annotated with GO_Slim terms related to secretion or protein transport; 22 of these had no GO_Slim annotations and 6 had no GO annotations at all ([Figure 2C](#)).

Further screening revealed many genes with altered glycoprotein secretion also had a fragmented aberrant Golgi apparatus

To further probe how the identified genes affect glycoprotein secretion, the 92 genes identified in the HRP screen were further analysed for changes in Golgi apparatus morphology using the same siRNA knockdown. At 72 h after siRNA transfection, cells were fixed and analysed by immunofluorescence, using an antibody against GM130, a marker of *cis*-Golgi membranes. Images were collected and classified as having aberrant or intact *cis*-Golgi using the machine learning platform within CellProfiler, as shown in [Figure 3A and 3B](#) (raw results are provided in S6, *Underlying data*)²². Many of the silenced genes resulted in an increase in aberrant *cis*-Golgi compared to the control (~20%). Genes with more than the median percentage of cells with aberrant Golgi are shown in [Figure 3C](#), and all genes are shown in [Figure E1](#) (see *Extended data*)²². Based on having unknown functions in secretion, five genes from the top half of these hits were chosen to further

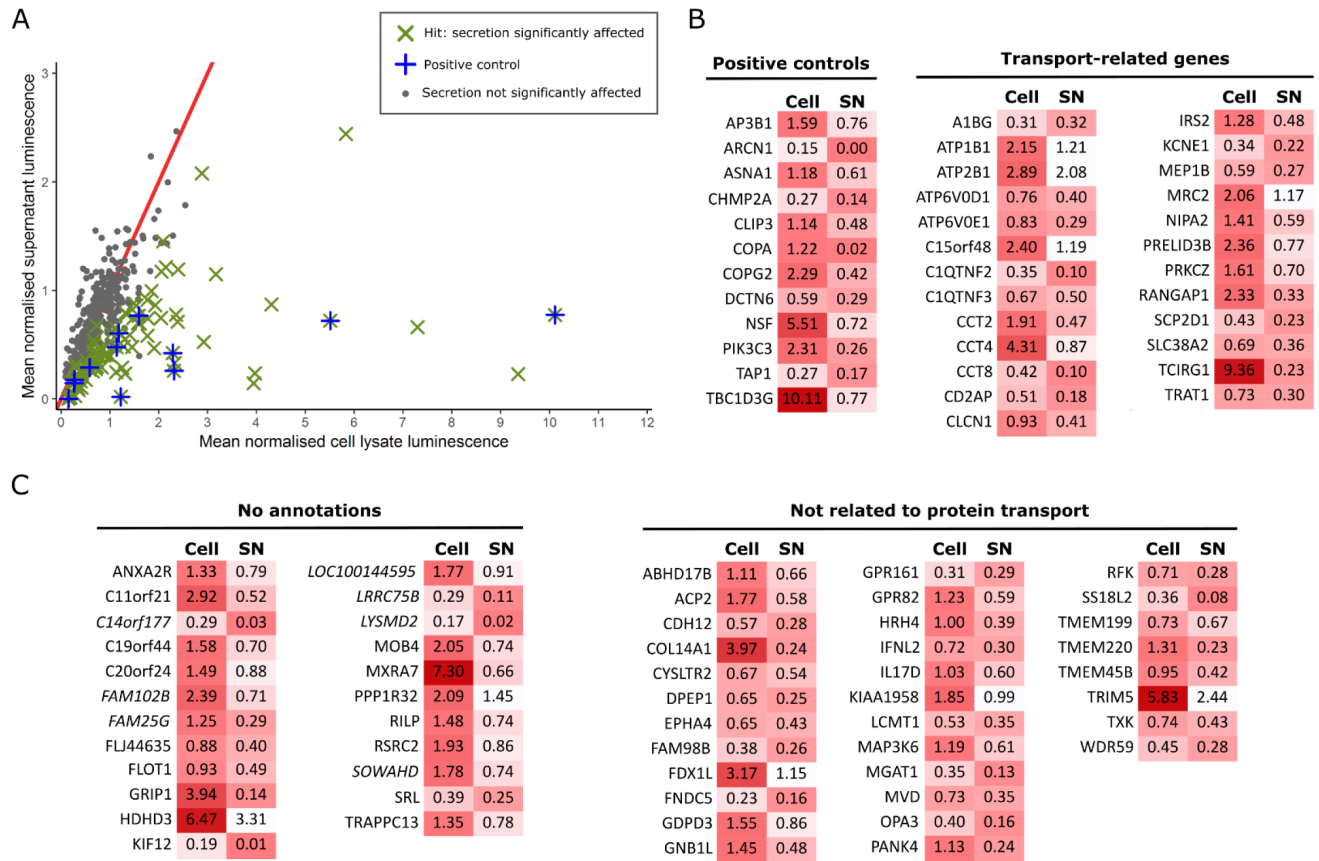


Figure 2. Horseradish peroxidase (HRP) screen to identify genes important for secretion. Approximately 400 genes were screened in an arrayed siRNA knockdown format. **(A)** Mean of the normalised luminescence for both cell lysate and supernatant signals from two independent replicates. ~400 genes were screened; those with significant divergence from control siRNAs, as identified by the ROUT method, are indicated by a green cross (x). Twelve genes known to be involved in secretion or protein trafficking were included in the screen as positive controls, indicated by a blue plus (+); these known controls were also identified as hits, validating the statistical method used here. **(B, C)** Mean values for normalised luminescence for cell lysate and supernatant (SN) signals of the 92 hits identified. Here these hits are shown on a white-red scale, where darker red indicates a greater secretion defect. The generic GO_Slim was used to categorise hits as either related to protein transport, unrelated to protein transport or not annotated. **(B)** A gene was defined as related to protein transport if it was annotated with any of the GO_Slim annotations transport (GO:0006810), transmembrane transport (GO:0055085) or vesicle-mediated transport (GO:0016192). **(C)** Genes unrelated to protein transport or not annotated by GO_Slim terms represent novel hits involved in protein secretion. Of the genes with no GO_Slim annotations, genes with no GO annotations at all are indicated in italics. Genes defined as not related to protein transport have other annotations in the generic GO_Slim.

investigate the mechanism by which these hits lead to a secretion defect: *GPR161*, *TMEM220*, *FAM98B*, *FAM102B* and *MXRA7*. We confirmed that the knockdown of these five genes led to a perturbation in the architecture of the Golgi and affected protein secretion. This can be seen by the more diffuse localisation of two Golgi-resident proteins, TGN46 and GM130; by the more diffuse localisation of SEC31, a marker of ER exit sites; and by the defect in the transport of the glycoprotein MHC-I to the cell surface (Figure 3D).

Subcellular localisation of new regulators of glycoprotein secretion

Given the poor characterisation of *GPR161*, *TMEM220*, *FAM98B*, *FAM102B* and *MXRA7* in the literature and the limited resources available to study their localisation, we obtained

cDNA constructs of each of these gene fused to a FLAG-tag, to study the localisation of the protein inside the cells (Figure 4). HeLa cells were transiently transfected with cDNA, then fixed 24 h after transfection. Fixed cells were immunostained for FLAG and either calnexin or TGN46, markers of the ER and the Golgi, respectively. *GPR161* localised to the Golgi, as seen by its co-localisation with TGN46 (Figure 4A), whereas *TMEM220* and *MXRA7* both partially colocalised with calnexin, indicating that they are found at the ER (Figure 4B, C). *FAM98B* was primarily found in the cytosol, with a portion of *FAM98B* also localising to the nucleus (Figure 4D). Finally, *FAM102B* was found both at the plasma membrane and in the cytosol (Figure 4E). However, it is important to note that the over-expression of these FLAG-tagged constructs may not reflect the endogenous protein localisation, as some dominant

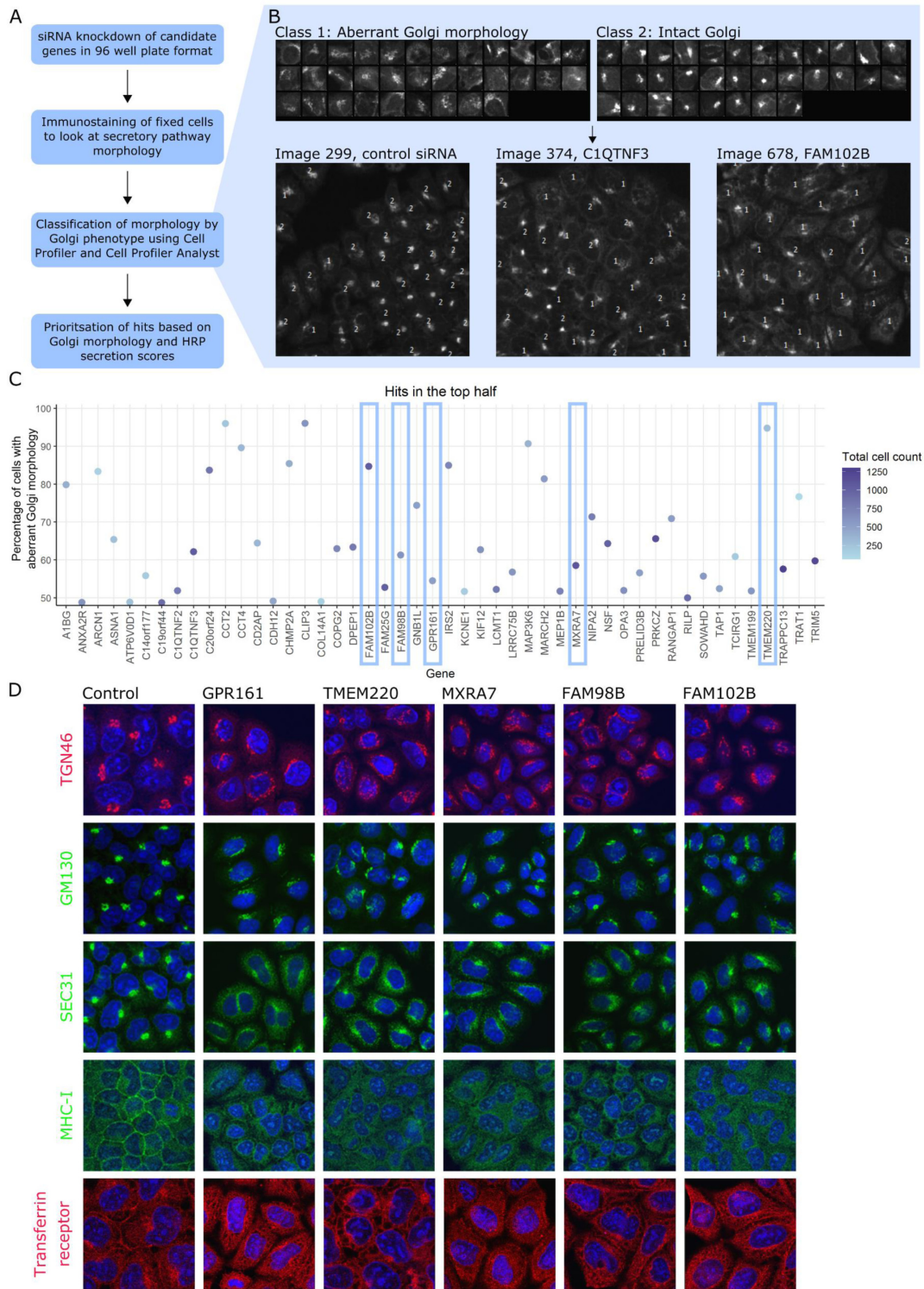


Figure 3. Further secondary screen for altered Golgi morphology. (A) Workflow of Golgi morphology screen. (B) Training set used to train the CellProfiler Analyst (CPA) classifier to distinguish between fragmented and intact Golgi. Using this training set, sensitivity was 77% for fragmented Golgi class and 82% for the intact Golgi class. Examples of control and test images classified using CPA are also shown. Cells with fragmented Golgi are labelled with 1; cells with intact Golgi are labelled with 2. (C) Percentage of cells with fragmented Golgi for the top 48 hits from tertiary screen. Hits are arranged alphabetically and coloured by cell count, with darker blue spots representing more confluent wells. Five of these hits were further characterised, indicated with blue rectangles. (D) Immunostaining of HeLa cells (original magnification 20x) treated with siRNA to knockdown each of the five genes indicated above the images. Cells were stained with the antibodies against the proteins indicated in green or red on the left. For all images, DAPI staining is shown in blue.

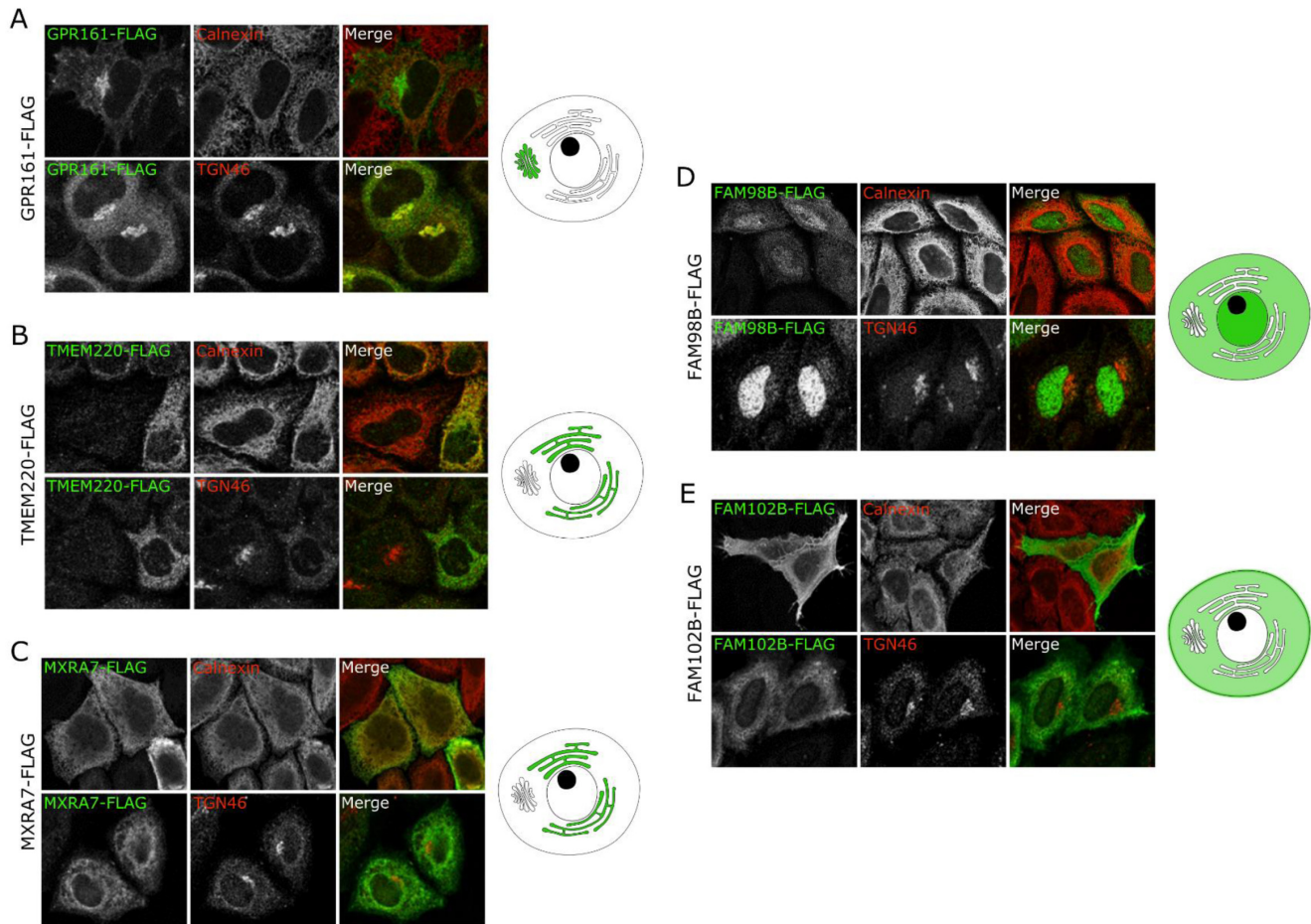


Figure 4. Localisation study for new regulators of glycoprotein secretion. Fluorescence microscopy images of HeLa cells expressing a FLAG-tagged cDNA construct show each protein's localisation by co-staining with either an ER marker, calnexin, or a Golgi marker, TGN46. In the merged images, FLAG staining is shown in green and calnexin or TGN46 staining is shown in red. The subcellular localisation of each protein construct is indicated in green on the cell diagrams. Darker shades of green are used to indicate the most prominent location if the construct localises to more than one compartment. **A–E** show expression of FLAG-tagged GPR161 (**A**), TMEM220 (**B**), MXRA7 (**C**), FAM98B (**D**) and FAM102B (**E**). Original magnification 20x.

negative effect might happen when using the cDNA construct. We believed this was the case for TMEM220, as the TGN46 immunostaining seems compromised in the transfected cells (Figure 4B). It was therefore important to validate the localisation of the protein using specific antibodies. We studied the localisation of the endogenous proteins GPR161 and TMEM220 and confirmed that GPR161 localised mainly in the Golgi (Figure 5Ai). This was further demonstrated using siRNA against GPR161; here the Golgi localisation of GPR161 disappeared (Figure 5Bi). However, endogenous TMEM220 localised to the Golgi (Figure 5Aii), in contrast to the cDNA overexpression showing an ER localisation. This Golgi residence was similarly confirmed using siRNA (Figure 5Bii). It is therefore likely that the overexpression of TMEM220 causes a perturbation in the trafficking of proteins from the ER to the Golgi.

GPR161 is a novel Golgi resident protein involved in protein secretion

Our study shows that GPR161 is a new Golgi-resident protein. To understand its role in secretion, we used the [BioPlex network](#),

a resource of proteins shown to interact by affinity purification-mass spectrometry, to look at its interacting partners^{30,31}. Interestingly, the BioPlex network shows that GPR161 interacts with golgin A5 (also known as golgin 84). Golgin A5 is a coiled-coil membrane protein that plays a role in intra-Golgi vesicle capture³⁵ and contributes to maintaining Golgi morphology, likely by its interaction with Rab1A^{36–38}. We showed that golgin A5 and GPR161 colocalised (Figure 5C), and the knockdown of GPR161 decreased the colocalisation between golgin A5 and TGN46 (Figure 5D). This appears to be specific to GPR161, since the knockdown of TMEM220, which we also found localises to the Golgi, resulted in similar perturbation of Golgi, but did not affect golgin A5-TGN46 colocalisation. Therefore, an aberrant Golgi structure alone does not change golgin A5 localisation to the Golgi (Figure 5D). These data suggest that GPR161 may serve to recruit golgin A5 to the Golgi membrane.

Discussion

Here we identified novel genes involved in glycoprotein secretion using a combination of pooled CRISPR screening and

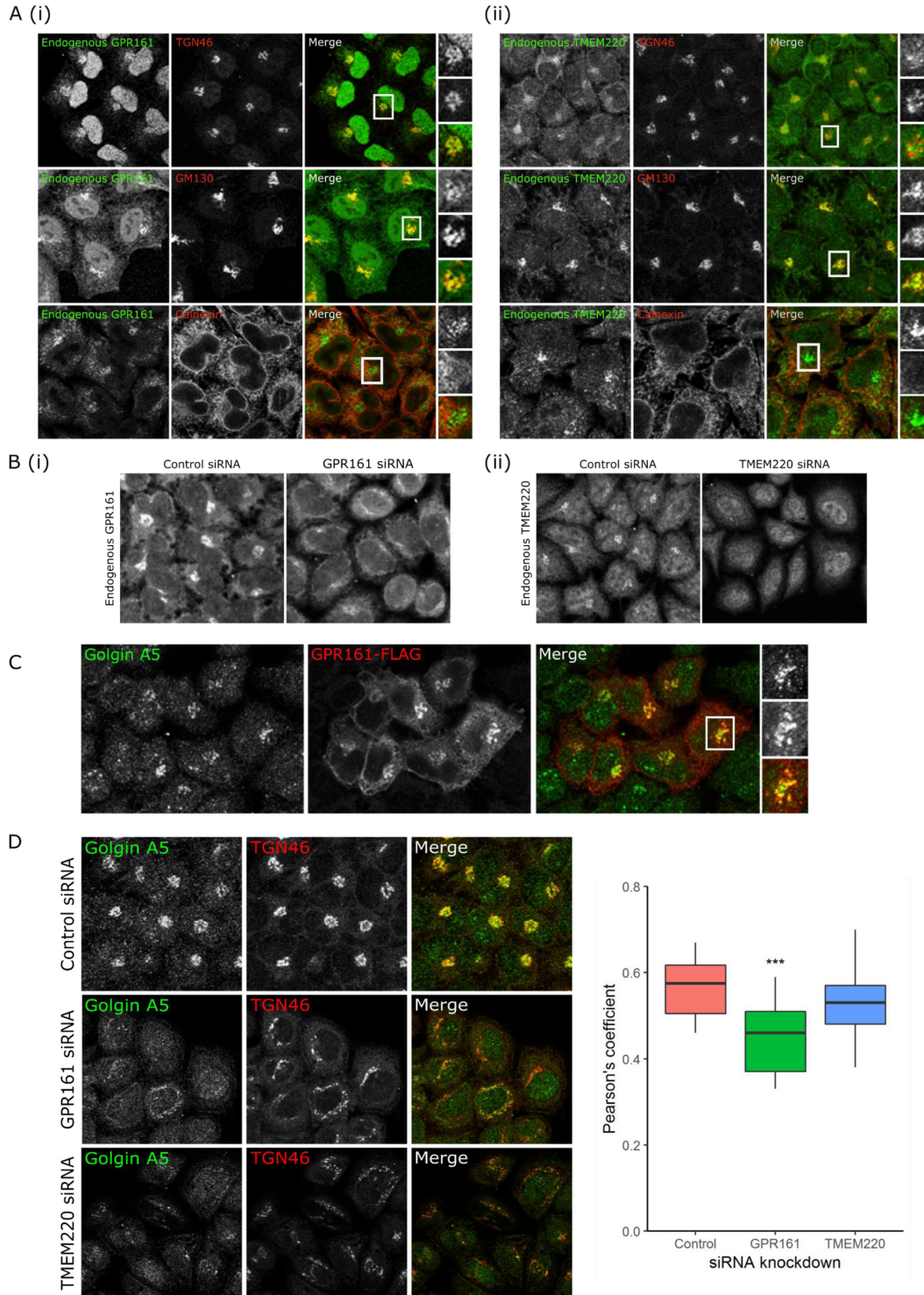


Figure 5. GPR161 and TMEM220, two novel Golgi-resident proteins involved in protein secretion, regulate glycoprotein secretion by different mechanisms. (A) Colocalisation microscopy shows that both (i) GPR161 and (ii) TMEM220 colocalise with TGN46 and GM130, two Golgi markers, and not with calnexin, a marker of the ER. (B) siRNA knockdown of (i) GPR161 or (ii) TMEM220 demonstrates that the antibodies used to detect these endogenous proteins only specifically detect protein at the Golgi. (C) GPR161-FLAG (red) colocalises with golgin A5 by microscopy. (D) Microscopy shows that knockdown of both GPR161 and TMEM220 lead to Golgi fragmentation seen by the TGN46 staining, but only GPR161 knockdown results in less colocalisation of golgin A5 and TGN46. Quantification of this change in colocalisation, measured by Pearson's coefficient, shows that GPR161 knockdown cells have significantly less colocalisation than control cells (** $p < 0.001$), but there is no significant difference in colocalisation between control cells and TMEM220 knockdown cells ($p = 0.302$). Original magnification 20x.

siRNA screening. This work expands on our results from a previous CRISPR screen measuring cell-surface galectin-3, which had identified a number of regulators of glycosylation and protein trafficking; however, most of those hits had had well-known functions¹⁴. Although galectin-3 itself is unconventionally secreted, this type of screen should primarily identify genes responsible for the transport of glycoproteins to the cell surface, as secreted galectin-3 can be transferred between cells^{14,39}. Here, we sought to build on the findings of our previous screen by using galectin-3 to identify novel proteins with roles in glycoprotein secretion. We used the improved Brunello sgRNA library¹⁸ rather than GeCKOv2 library, and an altered cell sorting strategy which would allow more hits through cell sorting initially, in order to increase the number of true positives in later populations. The Brunello library is an optimised library for the human genome, which was designed to increase the percentage of active guides while reducing off target effects compared to other libraries¹⁸. Furthermore, here we analysed populations at all stages of cell sorting rounds performed, rather than only the final sorted population.

Overall, our data suggest that the enhanced cell sorting strategy we employed here does improve results. While there is little enrichment visible by anti-galectin-3 staining in the earlier sorted populations, deep sequencing showed that enrichment had taken place. Many of the hits enriched in the first sorted population, N1, localise to the ER and the plasma membrane; later populations had fewer hits localised to the ER. This is likely due to problems with cell survival; over the extended time period and stress derived from the four sorts, cells with ER defects may be less able to survive. Enrichment of two distinct galectin-3 negative populations was visible by flow cytometry after the third round of cell sorting, in population N3. The most negative population within N3 consisted of cells that had little or no galectin-3 bound to the cell surface but were still able to secrete galectin-3 into the supernatant, so the excess of extracellular galectin-3 could bind to both the mid-negative population and the unaffected population of cells. Combined with the higher effective concentration of anti-galectin-3 antibody during staining, this meant that the unaffected population appeared to have more galectin-3 staining after earlier sorts. Across all sorted populations, the enrichment of many hits that are known to be in secretory and glycosylation pathways provides an initial validation of the success of the CRISPR screen. In comparison to our previous screen, we identified more hits known to have roles in the maturation or secretion of glycoproteins; moreover, we found a much higher number of genes not known to have such roles. These hits may have roles in wider protein secretion; further work is required to investigate this possibility.

Due to the large number of hits identified by the MaGECK algorithms, we selected hits to be taken forward for secondary screening for protein secretion. Via this screening method, we identified 92 hits that result in a secretion defect, of which 55 are not annotated with GO terms related to secretion. The 92 hits validated for secretion represent approximately one quarter of the hits screened, highlighting the importance of

secondary screening after a CRISPR screen due to the high chance of false positives. However, the fact that 55 of the validated hits have previously unknown or unclear functions in secretion demonstrates the power of this screening strategy. In comparison to other siRNA screens investigating protein secretion, there is very little overlap with our validated list of 92 hits⁴⁰⁻⁴²; however, this is not surprising as many of our hits taken forward for secondary screening were selected based on having an unclear function in protein secretion. Additionally, these screens looked specifically at the early secretory pathway⁴⁰, or started with smaller-scale libraries^{41,42}, so an overlap of hits after secondary screening in the different pipelines may not be expected. To further validate these hits, we investigated Golgi morphology. There was a clear increase in the percentage of cells with aberrant Golgi morphology for many of the hits. As siRNA knockdown efficiency was not quantified for each gene, the percentage of cells with aberrant Golgi does not give a definitive ranking of the scale of the effect each gene has on protein secretion. Additionally, while the presence of aberrant Golgi morphology gives a reason for the altered secretion and therefore validates both the CRISPR screen and HRP assay results, it does not explain how each individual gene product contributes to protein secretion. As such, we selected five hits with little known about their function in protein secretion to investigate further. Most of these had very strong phenotypes in both assays, such as MXRA7, but some had a weaker, yet still significant, phenotype, including GPR161; we studied this selection in order to validate the full list of genes as a resource for further investigation.

Among the five hits that we studied for further mechanistic insight, GPR161 and TMEM220 localised to the Golgi, whereas MXRA7 localised to the ER. FAM98B and FAM102B localised to the cytosol or the plasma membrane. Further studies will be required to fully understand how each gene regulates protein secretion. Interestingly, a recent study has shown the importance of the extracellular matrix (ECM) in regulating Golgi organisation and function via the activation of ARF1⁴³. MXRA7, a component of the ECM, might play a similar role. Little is known about FAM98B and FAM102B, but FAM102B shows a clear localisation at the plasma membrane, so it could regulate protein secretion through an identified signalling pathway. FAM98B shows cytosolic and nuclear localisation, and interacts with other such proteins to form a complex involved in shuttling RNA between the nucleus and cytoplasm⁴⁴, suggesting that FAM98B may affect protein secretion by a transcriptional route.

Our further investigation of the two proteins that localised to the Golgi, TMEM220 and GPR161, suggests that they regulate Golgi morphology by different mechanisms. TMEM220 is a transmembrane protein that has been shown to interact with both actin and testis-specific glyceraldehyde-3-phosphate dehydrogenase (GAPDH) in the BioPlex network^{30,31}. Actin, and the cytoskeleton in general, is well known to contribute to protein trafficking along the secretory pathway; for example, it is recruited to the Golgi to by a complex of SPCA1 and active cofilin, where it is suggested to form a membrane domain required

to initiate sorting of secretory cargo^{7,33,45}. It is possible that TMEM220 acts as an anchor for actin on the Golgi membrane. This possibility needs to be further investigated using *in vitro* experiments. Alternatively, TMEM220 could affect protein secretion via an interaction with GAPDH, which has recently been implicated in protein secretion via the inhibition of COPI vesicle biogenesis⁴⁶.

GPR161 is a G-protein-coupled receptor (GPCR) that is involved in neural tube development and acts as a regulator of cell signalling pathways, including Shh signalling, protein kinase A (PKA) signalling, retinoic acid signalling and Wnt signalling^{47,48}. This was a particularly interesting hit as it has recently become clear that GPCRs function at membranes other than the plasma membrane⁴⁹, and previous work has suggested that a Golgi-resident GPCR regulates transport from the Golgi, although the specific GPCR involved remains unknown⁵⁰. Our results showing that GPR161 is both localised to the Golgi and involved in regulation of Golgi morphology and glycoprotein traffic fit with this emerging idea that GPCRs can regulate protein trafficking. The BioPlex network for GPR161 shows an interaction with golgin A5, as well as with five PKA subunits^{30,31}; other work has also demonstrated an interaction between GPR161 and PKA⁵¹. The interaction with PKA subunits is interesting, as a PKA signalling pathway has previously been shown to regulate retrograde transport from the Golgi to the ER, which indirectly also affects anterograde traffic⁵². However, previous work found that PKA regulatory subunits binding to GPR161 results in its transport to the plasma membrane to signal through PKA⁵¹. Here, we did not observe any localisation of either overexpressed or endogenous GPR161 at the plasma membrane, suggesting that a different mechanism may be important here. Furthermore, our data suggest that the interaction between GPR161 and golgin A5 is important for maintenance of the Golgi architecture and function. One mechanism for this may be that GPR161 acts to recruit golgin A5 to the Golgi. To confirm this mechanism, further experiments involving mutated forms of GPR161 and golgin A5 will have to be performed, along with higher resolution imaging.

In summary, here we describe an optimised CRISPR screening strategy that successfully identifies new regulators of glycoprotein secretion. Our secondary screening validated 55 hits not previously known to be directly involved in protein secretion; many of these hits also regulate Golgi morphology. We hope that these validated hits can serve as a resource for other researchers investigating protein secretion and Golgi morphology. We also highlight GPR161, a particularly interesting protein given that Golgi-localised GPCRs have recently been implicated in protein trafficking. We find that it is a novel Golgi-localised protein that appears to interact with golgin A5 in order to maintain Golgi structure.

Data availability

Underlying data

Raw sequencing files from deep sequencing of the screened populations on Gene Expression Omnibus, Accession number GSE133692: <https://identifiers.org/geo/GSE133692>.

Apollo: Research data supporting 'Genome-wide CRISPR screening identifies new regulators of glycoprotein secretion'. <https://doi.org/10.17863/CAM.4040822>

This project contains the following underlying data:

- S1.xlsx (sgRNA counts of unsorted and sorted population from the CRISPR screen)
- S2.xlsx (MaGECK [version 0.5.7] analysis of sgRNA counts from the CRISPR screen. Both MaGECK-MLE and MaGECK-RRA results are shown. False discovery rates [FDR] from this table were used to create [Figure 1C–E](#))
- S3.zip (raw FCS files from flow cytometry staining of sorted samples, used to create [Figure 1B](#)).
- S4.xlsx (raw chemiluminescence data, used to create [Figure 2](#))
- S5.zip (unedited image files used in [Figure 3D](#), [Figure 4](#) and [Figure 5](#))
- S6.xlsx (data output from Cell Profiler, showing total number of cells identified per image and number of cells classified as having fragmented or intact Golgi)

Data are available under the terms of the [Creative Commons Attribution 4.0 International license](#) (CC-BY 4.0).

Extended data

Apollo: Research data supporting 'Genome-wide CRISPR screening identifies new regulators of glycoprotein secretion'. <https://doi.org/10.17863/CAM.40408>

This project contains the following extended data:

- E1.pdf (all hits from the secondary screen for Golgi morphology, top hits of which are shown in [Figure 3C](#))
- E2.xlsx (sequence of P5 primers used for PCR amplification of sgRNA)
- E3.xlsx (sequence of P7 primers used for PCR amplification of sgRNA)
- E4.xlsx (details of the siGenome pooled siRNA library used in secondary screening)

Data are available under the terms of the [Creative Commons Attribution 4.0 International license](#) (CC-BY 4.0).

Acknowledgments

We thank Paul Lehner for sHeLa cells, Vivek Malhotra for HeLa-ss-HRP cells, David Root and John Doench for the Brunello library, Marcella Ma, Brian Lam and Giles Yeo for valuable help and advice with the gRNA sequencing, Matt Castle for advice on statistical analysis, and James Smith and Greg Strachan for technical assistance with the Opera Phenix.

References

1. Benham AM: **Protein secretion and the endoplasmic reticulum.** *Cold Spring Harb Perspect Biol.* 2012; 4(8): a012872.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
2. Dancourt J, Barlowe C: **Protein sorting receptors in the early secretory pathway.** *Annu Rev Biochem.* 2010; 79: 777–802.
[PubMed Abstract](#) | [Publisher Full Text](#)
3. Han J, Pluhackova K, Böckmann RA: **The Multifaceted Role of SNARE Proteins in Membrane Fusion.** *Front Physiol.* 2017; 8: 5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
4. Beznoussenko GV, Parashuraman S, Rizzo R, et al.: **Transport of soluble proteins through the Golgi occurs by diffusion via continuities across cisternae.** *eLife.* 2014; 3: e02009.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
5. Kienzle C, von Blume J: **Secretory cargo sorting at the trans-Golgi network.** *Trends Cell Biol.* 2014; 24(10): 584–593.
[PubMed Abstract](#) | [Publisher Full Text](#)
6. Spang A: **The Road not Taken: Less Traveled Roads from the TGN to the Plasma Membrane.** *Membranes (Basel).* 2015; 5(1): 84–98.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
7. Pakdel M, von Blume J: **Exploring new routes for secretory protein export from the trans-Golgi network.** *Mol Biol Cell.* 2018; 29(3): 235–240.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
8. Kurata M, Yamamoto K, Moriarity BS, et al.: **CRISPR/Cas9 library screening for drug target discovery.** *J Hum Genet.* 2018; 63(2): 179–186.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. le Sage C, Lawo S, Panicker P, et al.: **Dual direction CRISPR transcriptional regulation screening uncovers gene networks driving drug resistance.** *Sci Rep.* 2017; 7(1): 17693.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
10. Li M, Yu JSL, Tilgner K, et al.: **Genome-wide CRISPR-KO Screen Uncovers mTORC1-Mediated Gsk3 Regulation in Naive Pluripotency Maintenance and Dissolution.** *Cell Rep.* 2018; 24(2): 489–502.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
11. Morita K, Hama Y, Izume T, et al.: **Genome-wide CRISPR screen identifies TMEM41B as a gene required for autophagosome formation.** *J Cell Biol.* 2018; 217(11): 3817–3828.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. Moretti F, Bergman P, Dodgson S, et al.: **TMEM41B is a novel regulator of autophagy and lipid mobilization.** *EMBO Rep.* 2018; 19(9): pii: e45889.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
13. Zhang R, Kim AS, Fox JM, et al.: **Mxra8 is a receptor for multiple arthritogenic alphaviruses.** *Nature.* 2018; 557(7706): 570–574.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
14. Stewart SE, Menzies SA, Popa SJ, et al.: **A genome-wide CRISPR screen reconciles the role of N-linked glycosylation in galectin-3 transport to the cell surface.** *J Cell Sci.* 2017; 130(19): 3234–3247.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Popa SJ, Stewart SE, Moreau K: **Unconventional secretion of annexins and galectins.** *Semin Cell Dev Biol.* 2018; 83: 42–50.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. Nabi IR, Shankar J, Dennis JW: **The galectin lattice at a glance.** *J Cell Sci.* 2015; 128(13): 2213–2219.
[PubMed Abstract](#) | [Publisher Full Text](#)
17. Agrotis A, Ketteler R: **A new age in functional genomics using CRISPR/Cas9 in arrayed library screening.** *Front Genet.* 2015; 6: 300.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
18. Doench JG, Fusi N, Sullender M, et al.: **Optimized sgRNA design to maximize activity and minimize off-target effects of CRISPR-Cas9.** *Nat Biotechnol.* 2016; 34(2): 184–191.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
19. Fehse B, Kustikova OS, Bubenheim M, et al.: **Pois(s)on--It's a question of dose...** *Gene Ther.* 2004; 11(11): 879–881.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. Li W, Köster J, Xu H, et al.: **Quality control, modeling, and visualization of CRISPR screens with MAGeCK-VISPR.** *Genome Biol.* 2015; 16: 281.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Li W, Xu H, Xiao T, et al.: **MAGeCK enables robust identification of essential genes from genome-scale CRISPR/Cas9 knockout screens.** *Genome Biol.* 2014; 15(12): 554.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. The UniProt Consortium: **UniProt: the universal protein knowledgebase.** *Nucleic Acids Res.* 2017; 45(D1): D158–D169.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
23. Ashburner M, Ball CA, Blake JA, et al.: **Gene ontology: tool for the unification of biology.** *The Gene Ontology Consortium. Nat Genet.* 2000; 25(1): 25–29.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
24. The Gene Ontology Consortium: **Expansion of the Gene Ontology knowledgebase and resources.** *Nucleic Acids Res.* 2017; 45(D1): D331–D338.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. **Reverse siRNA transfection | Pelkmans Lab.**
[Reference Source](#)
26. Motulsky HJ, Brown RE: **Detecting outliers when fitting data with nonlinear regression - a new method based on robust nonlinear regression and the false discovery rate.** *BMC Bioinformatics.* 2006; 7: 123.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
27. Boyle EI, Weng S, Gollub J, et al.: **GO::TermFinder--open source software for accessing Gene Ontology information and finding significantly enriched Gene Ontology terms associated with a list of genes.** *Bioinformatics.* 2004; 20(18): 3710–3715.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
28. Carpenter AE, Jones TR, Lamprecht MR, et al.: **CellProfiler: image analysis software for identifying and quantifying cell phenotypes.** *Genome Biol.* 2006; 7(10): R100.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
29. Jones TR, Carpenter AE, Lamprecht MR, et al.: **Scoring diverse cellular morphologies in image-based screens with iterative feedback and machine learning.** *Proc Natl Acad Sci U S A.* 2009; 106(6): 1826–1831.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
30. Huttlin EL, Ting L, Bruckner RJ, et al.: **The BioPlex Network: A Systematic Exploration of the Human Interactome.** *Cell.* 2015; 162(2): 425–440.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
31. Huttlin EL, Bruckner RJ, Paulo JA, et al.: **Architecture of the human interactome defines protein communities and disease networks.** *Nature.* 2017; 545(7655): 505–509.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
32. Schindelin J, Arganda-Carreras I, Frise E, et al.: **Fiji: an open-source platform for biological-image analysis.** *Nat Methods.* 2012; 9(7): 676–682.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
33. von Blume J, Alleaume AM, Cantero-Recasens G, et al.: **ADF/cofilin regulates secretory cargo sorting at the TGN via the Ca²⁺ ATPase SPCA1.** *Dev Cell.* 2011; 20(5): 652–662.
[PubMed Abstract](#) | [Publisher Full Text](#)
34. Bard F, Casano L, Mallabiarrena A, et al.: **Functional genomics reveals genes involved in protein secretion and Golgi organization.** *Nature.* 2006; 439(7076): 604–607.
[PubMed Abstract](#) | [Publisher Full Text](#)
35. Wong M, Gillingham AK, Munro S: **The golgin coiled-coil proteins capture different types of transport carriers via distinct N-terminal motifs.** *BMC Biol.* 2017; 15(1): 3.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
36. Satoh A, Wang Y, Malsam J, et al.: **Golgin-84 is a rab1 binding partner involved in Golgi structure.** *Traffic.* 2003; 4(3): 153–161.
[PubMed Abstract](#) | [Publisher Full Text](#)
37. Diao A, Rahman D, Pappin DJ, et al.: **The coiled-coil membrane protein golgin-84 is a novel rab effector required for Golgi ribbon formation.** *J Cell Biol.* 2003; 160(2): 201–212.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
38. Liu X, Wang Z, Yang Y, et al.: **Rab1A mediates proinsulin to insulin conversion in β -cells by maintaining Golgi stability through interactions with golgin-84.** *Protein Cell.* 2016; 7(9): 692–696.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
39. Patnaik SK, Potvin B, Carlsson S, et al.: **Complex N-glycans are the major ligands for galectin-1, -3, and -8 on Chinese hamster ovary cells.** *Glycobiology.* 2006; 16(4): 305–317.
[PubMed Abstract](#) | [Publisher Full Text](#)
40. Simpson JC, Joggerst B, Laketa V, et al.: **Genome-wide RNAi screening identifies human proteins with a regulatory function in the early secretory pathway.** *Nat Cell Biol.* 2012; 14(7): 764–774.
[PubMed Abstract](#) | [Publisher Full Text](#)
41. Farhan H, Wendeler MW, Mitrovic S, et al.: **MAPK signaling to the early secretory pathway revealed by kinase/phosphatase functional screening.** *J Cell Biol.* 2010; 189(6): 997–1011.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
42. Chia J, Goh G, Racine V, et al.: **RNAi screening reveals a large signaling network controlling the Golgi apparatus in human cells.** *Mol Syst Biol.* 2012; 8: 629.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
43. Singh V, Erady C, Balasubramanian N: **Cell-matrix adhesion controls Golgi organization and function through Arf1 activation in anchorage-dependent cells.** *J Cell Sci.* 2018; 131(16): pii: jcs215855.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
44. Pérez-González A, Pazo A, Navajas R, et al.: **hCLE/C14orf166 associates with DDX1-HSPC117-FAM98B in a novel transcription-dependent shuttling RNA-transporting complex.** *PLoS One.* 2014; 9(3): e90957.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

45. Gurel PS, Hatch AL, Higgs HN: **Connecting the cytoskeleton to the endoplasmic reticulum and Golgi.** *Curr Biol.* 2014; **24**(14): R660–R672.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
46. Yang JS, Hsu JW, Park SY, *et al.*: **GAPDH inhibits intracellular pathways during starvation for cellular energy homeostasis.** *Nature.* 2018; **561**(7722): 263–267.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
47. Mukhopadhyay S, Wen X, Ratti N, *et al.*: **The ciliary G-protein-coupled receptor Gpr161 negatively regulates the Sonic hedgehog pathway via cAMP signaling.** *Cell.* 2013; **152**(1–2): 210–223.
[PubMed Abstract](#) | [Publisher Full Text](#)
48. Li BI, Matteson PG, Ababon MF, *et al.*: **The orphan GPCR, Gpr161, regulates the retinoic acid and canonical Wnt pathways during neurulation.** *Dev Biol.* 2015; **402**(1): 17–31.
[PubMed Abstract](#) | [Publisher Full Text](#)
49. Eichel K, von Zastrow M: **Subcellular Organization of GPCR Signaling.** *Trends Pharmacol Sci.* 2018; **39**(2): 200–208.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
50. Díaz Añel AM, Malhotra V: **PKCeta is required for beta1gamma2/beta3gamma2- and PKD-mediated transport to the cell surface and the organization of the Golgi apparatus.** *J Cell Biol.* 2005; **169**(1): 83–91.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
51. Bachmann VA, Mayrhofer JE, Ilouz R, *et al.*: **Gpr161 anchoring of PKA consolidates GPCR and cAMP signaling.** *Proc Natl Acad Sci U S A.* 2016; **113**(28): 7786–7791.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
52. Cancino J, Capalbo A, DiCampli A, *et al.*: **Control systems of membrane transport at the interface between the endoplasmic reticulum and the Golgi.** *Dev Cell.* 2014; **30**(3): 280–294.
[PubMed Abstract](#) | [Publisher Full Text](#)

Open Peer Review

Current Peer Review Status:  

Version 2

Reviewer Report 23 January 2020

<https://doi.org/10.21956/wellcomeopenres.17182.r37605>

© 2020 Farhan H. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Hesso Farhan 

Institute of Basic Medical Sciences, Department of Molecular Medicine, University of Oslo, Oslo, Norway

I have no further comments or suggestion.

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Membrane trafficking, Signal transduction, COPII, Golgi, Cell migration, Rho GTPases, Kinases

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 21 January 2020

<https://doi.org/10.21956/wellcomeopenres.17182.r37604>

© 2020 Stephens D. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



David Stephens 

School of Biochemistry, Faculty of Life Sciences, University of Bristol, Bristol, UK

Some minor text changes have been made but further experiments don't address key concerns e.g. around the effects on glycoprotein versus general protein secretion.

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Cell biology, membrane traffic and cytoskeletal dynamics.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 1

Reviewer Report 28 August 2019

<https://doi.org/10.21956/wellcomeopenres.16623.r36252>

© 2019 Stephens D. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



David Stephens 

School of Biochemistry, Faculty of Life Sciences, University of Bristol, Bristol, UK

This is an interesting manuscript that applies CRISPR gene editing approaches to screen for components of the glycoprotein secretion machinery. It appears to be carefully conducted with appropriate analysis applied to the screen. I am not an expert in that area and so cannot comment on the design, execution, or analysis of the screen further.

The results are clearly presented and some key "hits" defined. I would have liked to have read more about the overlap between the current work and previous efforts e.g. whole genome RNAi screens in mammalian cells and flies^{1,2,3}. Particularly given the previous use of ssHRP as a reporter.

Fundamentally the screen is really testing those genes required for localization of galectin-3 to the cell surface, this is not quite the same as a global analysis of glycoprotein secretion. Perhaps validation with other lectins, or of specific glycoproteins, or even an unbiased glycoproteomics approach might have been appropriate - at least for the "hits" that were worked up. Really, I would have liked to see some analysis of other endogenous glycoproteins as well as non-glycosylated secretory proteins. In particular, it is not clear to me whether the effects seen are indeed general or specific to galectin-3.

The analysis of five of the key "hits" is quite preliminary but the work is carefully conducted and nicely presented. This provides some framework for others to build on these data. The link between defects in glycoprotein secretion and Golgi organization are not really explored and this could have been built on to provide more insight.

A fundamental limitation for me is that none of the validation work has been done in any system other than HeLa cells. Given the literature on the role of GPR161 in cilia⁴, one might have hoped to see some work-up done in ciliated cells.

I expect that this will stand alone as a useful data resource and can certainly be built on by others.

Specific comments:

- RNAi efficacy (important for Figure 5) is not validated anywhere that I could see.

- The colocalization analysis is limited. GolginA5 and TGN46 should not overlap significantly as they localize to distinct parts of the Golgi/TGN (5D), therefore I have doubts over the robustness of this readout. In Figure 5C colocalization of GPR161 and GolginA5 does not appear robust to me and is not quantified. The images shown are low resolution and more detailed analysis is warranted. Nocodazole treatment of cells to scatter the stacked Golgi would be useful here.
- The integrity of the microtubule network should be checked as disruption of this could result in altered Golgi morphology (but not likely to secretion per se). The two phenotypes might be unconnected.
- The conclusion that GPR161 might recruit GolginA5 is premature. The data do not provide sufficient support for this.

References

1. Bard F, Casano L, Mallabiabarrena A, Wallace E, Saito K, Kitayama H, Guizzunti G, Hu Y, Wendler F, Dasgupta R, Perrimon N, Malhotra V: Functional genomics reveals genes involved in protein secretion and Golgi organization. *Nature*. 2006; **439** (7076): 604-7 [PubMed Abstract](#) | [Publisher Full Text](#)
2. Simpson JC, Joggerst B, Laketa V, Verissimo F, Cetin C, Erfle H, Bexiga MG, Singan VR, Hériché JK, Neumann B, Mateos A, Blake J, Bechtel S, Benes V, Wiemann S, Ellenberg J, Pepperkok R: Genome-wide RNAi screening identifies human proteins with a regulatory function in the early secretory pathway. *Nat Cell Biol*. 2012; **14** (7): 764-74 [PubMed Abstract](#) | [Publisher Full Text](#)
3. Wendler F, Gillingham AK, Sinka R, Rosa-Ferreira C, Gordon DE, Franch-Marro X, Peden AA, Vincent JP, Munro S: A genome-wide RNA interference screen identifies two novel components of the metazoan secretory pathway. *EMBO J*. 2010; **29** (2): 304-14 [PubMed Abstract](#) | [Publisher Full Text](#)
4. Mukhopadhyay S, Wen X, Ratti N, Loktev A, Rangell L, Scales SJ, Jackson PK: The ciliary G-protein-coupled receptor Gpr161 negatively regulates the Sonic hedgehog pathway via cAMP signaling. *Cell*. 2013; **152** (1-2): 210-23 [PubMed Abstract](#) | [Publisher Full Text](#)

Is the work clearly and accurately presented and does it cite the current literature?

Partly

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Partly

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Cell biology, membrane traffic and cytoskeletal dynamics.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Reviewer Report 27 August 2019

<https://doi.org/10.21956/wellcomeopenres.16623.r36185>

© 2019 Farhan H. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Hesso Farhan 

Institute of Basic Medical Sciences, Department of Molecular Medicine, University of Oslo, Oslo, Norway

The work by Popa and colleagues from the Moreau group reports on the identification of new regulators of glycoprotein secretion using a CRISPR screen.

The manuscript is written well and is easy to follow. Most of the data are of strong quality and therefore, I am supportive of the indexing of this work. I have a few concerns that could be easily addressed by a revision.

1. The title implies that the paper identifies regulators of glycoprotein secretion. How sure are the authors that these new proteins regulate glycoprotein trafficking, and not general trafficking? If they want to maintain the prefix glyco-, then they should show that non-glycosylated proteins such as albumin are not affected.
2. The authors use the term “Golgi fragmentation”, but this is not what I see. In many cases, the Golgi is simply aberrant in shape (stretched, or extended). Is it possible to revisit the data to show quantification of Golgi “fragments”? I think it is safer to use the term “aberrant Golgi structure”. The authors could use the Golgi compactness index developed originally by Fred Bard (2003).
3. It is hard to appreciate the effects on ERES, with the magnification and image quality the authors use. In my experience, a 20X magnification does not permit reliable judging of ERES. The authors should do better ERES imaging and quantification for the 2-3 most important hits. Especially for TMEM220, which they consider a regulator of ER-Golgi trafficking. This needs to be experimentally supported.
4. The authors state/conclude that GRP161 serves to recruit GolginA5 to the Golgi. I don't think that the data support this claim. A similar perturbation of GolgiA5 staining is seen with TMEM220. Could these not be simply a reflection of a distorted Golgi structure?
5. General comment: the work does not compare the results to previous screens by Rainer Pepperkok¹, Hesso Farhan² and Fred Bard³. I am not sure why the authors chose to ignore this previous work, despite its obvious relevance to the current paper. Not only should these three papers be cited, but a comparison of the hits should be made.

References

1. Simpson JC, Joggerst B, Laketa V, Verissimo F, Cetin C, Erfle H, Bexiga MG, Singan VR, Hériché JK, Neumann B, Mateos A, Blake J, Bechtel S, Benes V, Wiemann S, Ellenberg J, Pepperkok R: Genome-wide RNAi screening identifies human proteins with a regulatory function in the early secretory pathway. *Nat Cell Biol.* 2012; **14** (7): 764-74 [PubMed Abstract](#) | [Publisher Full Text](#)
2. Farhan H, Wendeler MW, Mitrovic S, Fava E, Silberberg Y, Sharan R, Zerial M, Hauri HP: MAPK signaling to the early secretory pathway revealed by kinase/phosphatase functional screening. *J Cell Biol.* 2010; **189** (6): 997-1011 [PubMed Abstract](#) | [Publisher Full Text](#)
3. Chia J, Goh G, Racine V, Ng S, Kumar P, Bard F: RNAi screening reveals a large signaling network controlling the Golgi apparatus in human cells. *Mol Syst Biol.* 2012; **8**: 629 [PubMed Abstract](#) | [Publisher Full Text](#)

Is the work clearly and accurately presented and does it cite the current literature?

No

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

I cannot comment. A qualified statistician is required.

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Partly

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Membrane trafficking, Signal transduction, COPII, Golgi, Cell migration, Rho GTPases, Kinases

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.
