To Be or Not To Be Pathogenic: Transcriptional Reprogramming Dictates a Fungal Pathogen's Response to Different Hosts^[OPEN]

The fungal pathogen Fusarium virguliforme invades a wide spectrum of host plants, ranging from eudicots to monocots. Interestingly, fungal root colonization results in disease-producing phenotypes only in certain eudicot hosts (Kolander et al., 2012). For instance, F. virguliforme causes sudden death syndrome, one of the most destructive diseases in soybean (Glycine max). Diseased soybean plants exhibit an array of symptoms such as root rot, leaf chlorosis/necrosis, and loss of plant biomass. While several F. virguliforme virulence factors responsible for the disease phenotype have been identified, little is known about how this pathogen evokes symptomatic or asymptomatic responses in different hosts.

A transcriptome-based study by Baetsen-Young and colleagues (2020) provides a detailed insight into this phenomenon. Instead of looking at a single host-pathogen interaction, the authors compared fungal root colonization and gene expression dynamics in infected soybean (symptomatic) and maize (Zea mays; asymptomatic) hosts. In both of these hosts, germinating F. virguliforme spores can be detected soon after inoculation. Examination of fungal mycelia structures indicated that maize roots undergo a similar, albeit slower, infection process to that of soybean roots. However, fungal infection does not trigger host cell death or other negative consequences in maize (see figure), whereas in soybean such symptoms appear as early as 10 d postinoculation.

To detect fungal transcripts expressed at the pathogen-host interface, Baetsen-Young et al. (2020) first generated a long read-based high-quality *F. virguliforme* genome assembly with improved gene annotations. Next, the authors sequenced RNAs from fungus-infected maize or soybean roots (harvested at six different time

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F. virguliforme Utilizes Dynamic Transcriptional Reprogramming to Switch to Alternative Lifestyles in Asymptomatic and Symptomatic Hosts.

Coexpressed *F. virguliforme* gene modules in symptomatic (soybean) or asymptomatic (maize) roots show distinct temporal patterns (groups G1 to G4). Modules from different hosts belonging to the same temporal group show little overlap in their gene content, indicating host-dependent transcriptional reprogramming. (*Adapted from Baetsen-Young et al. [2020], Figure 5.*)

points during infection), uninfected maize or soybean roots, and in vitrogerminated fungal spores without any host. Transcriptome analysis based on a total of 39 RNA-seq libraries revealed that F. virguliforme exhibits both hostspecific and inoculation stage-specific gene expression patterns. Compared with the in vitro-germinated spores, thousands of fungal genes are differentially expressed in each of the infected hosts. Interestingly, a majority of these fungal genes are highly upregulated (more than fourfold) in both hosts around 7 to 14 d postinoculation. Infected maize and soybean roots share little overlap in terms of which fungal genes are upregulated at any given time point. However, in general, fungal genes important for growth and host colonization are induced in both hosts. In soybean roots,

fungal genes involved in protein dephosphorylation and small molecule biosynthesis are also upregulated, suggesting that these may be important for symptomatic fungal infection.

Global coexpression network analyses identified multiple fungal gene modules associated with F. virguliforme-maize or F. virguliforme-soybean interactions. In both hosts, these modules can be assigned to one of four major groups based on distinct temporal patterns. However, modules belonging to the same temporal group vary greatly in their gene content between the two hosts (see figure). In the case of soybean, Gene Ontology terms enriched in the temporal groups 1 and 2 correlate to the faster fungal colonization phenotype of this host. Additionally, the temporal expression patterns of these groups reflect fungal transcriptome

reprogramming to facilitate the transition from initial manipulation of soybean host defense to subsequent necrotrophic mode for fungal proliferation. At the same time, fungal gene modules upregulated in maize were enriched for processes involved in host-nutrient acquisition. To assess whether host physiology shapes the F. virguliforme transcriptome, Baetsen-Young et al. (2020) directly compared fungal gene expression between maize and soybean for specific time points. This further confirmed distinct changes in the fungal transcriptome that are linked to the manifestation of symptomatic or asymptomatic phenotypes. For example, F. virguliforme expresses a greater diversity of necrosis-inducing effectors and pectin-degrading enzymes at a much higher level in infected soybean roots than in maize roots. Soybean has a higher pectin content than maize, so the fungi might produce relatively more pectin lyase enzymes to facilitate host cell breakdown in soybean.

Overall, the findings of Baetsen-Young et al. (2020) show that rather than expressing unique transcripts in different hosts, F. virguliforme extensively reprograms its transcriptome to adopt a necrotrophic or biotrophic lifestyle in soybean or maize, respectively. This transcriptional plasticity might be a common strategy for generalist pathogens to ensure survival on a wide range of morphologically and biochemically diverse hosts in the absence of a compatible host nearby. Questions that remain to be answered include how F. virguliforme accomplishes this temporal regulation and whether host gene expression is also differentially manipulated by F. virguliforme

during asymptomatic versus symptomatic colonization.

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