


Article

Validation of a Commercial Loop-Mediated Isothermal Amplification (LAMP) Assay for the Rapid Detection of *Anisakis* spp. DNA in Processed Fish Products

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Abstract: Parasites belonging to the *Anisakis* genera are organisms of interest for human health because they are responsible for the Anisakiasis zoonosis, caused by the ingestion of raw or undercooked fish. Furthermore, several authors have reported this parasite to be a relevant inducer of acute or chronic allergic diseases. In this work, a rapid commercial system based on Loop-Mediated Isothermal Amplification (LAMP) was optimised and validated for the sensitive and rapid detection of *Anisakis* spp. DNA in processed fish products. The specificity and sensitivity of the LAMP assay for processed fish samples experimentally infected with *Anisakis* spp. larvae and DNA were determined. The LAMP system proposed in this study was able to give positive amplification for all the processed fish samples artificially contaminated with *Anisakis* spp., giving sensitivity values equal to 100%. Specificity tests provided no amplification for the *Contracaecum*, *Pseudoterranova*, or *Hysterothylacium* genera and uninfected samples. The limit of detection (LOD) of the LAMP assay proposed was 10² times lower than the real-time PCR method compared. To the best of our knowledge, this is the first report regarding the application of the LAMP assay for the detection of *Anisakis* spp. in processed fish products. The results obtained indicate that the LAMP assay validated in this work could be a reliable, easy-to-use, and convenient tool for the rapid detection of *Anisakis* DNA in fish product inspection.

Keywords: *Anisakis* spp.; molecular methods; LAMP; validation; anisakidae family

1. Introduction

The Anisakidae family includes a vast number of parasites with a worldwide distribution. The life-cycle of Anisakidae nematodes involves invertebrates, fish, cephalopods, and marine mammals so these parasites can be found in the muscles and viscera of numerous fish and cephalopod species [1–5]. In the Mediterranean, the *Anisakis* spp. parasites can be found in different teleosts belonging to different ecological distributions [6]. The fish species mainly involved in the life cycle of *Anisakis* belong to the pelagic, benthopelagic, and benthodemersal domains. Indeed, the three *Anisakis* species are widely

occurring in pelagic, benthopelagic, and demersal species of the Gadidae, Merlucciidae, Scombridae, Carangidae, and Trichiuridae families [2,7,8]. *Anisakis* spp. parasites are marine nematodes of health interest because of their high zoonotic potential, being responsible for a human disease called Anisakiasis. Anisakiasis is a zoonotic disease caused by the ingestion of raw or undercooked fish infected with Anisakidae larvae [9–13]. Anisakiasis has become an increasing human health concern, particularly in Asian countries, representing more than 50% of all cases, where the consumption of raw or undercooked fish is frequent and/or has become increasingly popular. The remaining cases are from European countries, especially Italy (57 cases from 1996 until now), Spain (124 cases from 1996 until now), and France (15 cases from 1975 until now), whereas there are much fewer cases in Scandinavian countries, despite comparatively high fish consumption rates per capita in these countries [7,14]. Moreover, several authors have reported this parasite to be a relevant inducer of acute or chronic allergic diseases [15–18]. *Anisakis* is implicated in allergic IgE-mediated reactions, such as urticaria, angioedema, asthma, and anaphylaxis, in highly sensitized people [19,20]. The European Food Safety Authority (EFSA 2010) confirmed that all wild saltwater fish must be considered at risk of containing viable parasites of human health concern and no sea fishing grounds can be considered free of *A. simplex complex* larvae [21].

EFSA also recommends further studies and methods to improve the surveillance and diagnostic awareness of pathologies to parasites in fishery products. Even the Italian Ministry of Health encourages fish sector operators to carry out correct evisceration protocols of fish products in order to prevent *Anisakis*-related pathologies by reducing the possibility of migration of L3 larvae in the musculature [22]. At present, European authorities perform laborious and unreliable inspection methods, such as visual control and transillumination with UV [23], which cannot be applied for processed fish products such as anchovy paste, marinated anchovies, infant formulas, etc. [24]. Furthermore, the current immunological methods for *Anisakis* allergy diagnosis give a high number of false positives due to the cross-reactivity with numerous panallergens [25,26].

In this case, molecular biology methods are valuable tools in the detection of *Anisakis* spp. nematodes in processed seafood [24,27–30]. Several studies have shown that immunological and molecular methodologies yield comparable results concerning the detection of allergens in processed foods as sensitive and specific tools [31–33]. According to literature, Real-Time PCR is recognized to be the only molecular method capable of detecting the presence of *Anisakis* spp. in processed fish products, showing satisfactory sensitivity and specificity values [24,27,30,33,34]. Nonetheless, this sophisticated and expensive molecular method is currently indispensable and requires operational skills limiting their broad applicability. Loop-mediated isothermal amplification (LAMP) is considered a highly sensitive and rapid method for DNA amplification at constant temperature (60–65 °C) [35,36].

The food sector operators must have systems and procedures that allow competent authorities to access information on the products in order to guarantee their hygiene. In this work, a commercial LAMP assay for *Anisakis* spp. DNA detection was optimised and validated in order to obtain a simple, fast, and cheap tool, which can identify possible risks to consumer health due to the presence of these organisms in processed fish products.

2. Materials and Methods

2.1. Fish Samples and *Anisakis* Larvae Collection

All the processed fish samples used for the method optimisation and validation came from a large-scale distribution, in order to reduce any bias from local food specialities and extend the range of validation. Homogenised farmed trout (*Oncorhynchus mykiss*; $n = 40$), homogenised farmed sea bream (*Saprus aurata*; $n = 40$), and homogenised farmed salmon (*Salmo salar*; $n = 40$) were chosen as naturally negative (noncontaminated by *Anisakis*) samples [4,21,37–39]; whereas anchovy (*Engraulis encrasicolus*) paste ($n = 40$), anchovy in oil ($n = 40$), and salted sardines (*Sardina pilchardus*; $n = 40$) samples were chosen as positive samples for the validation of the method and for matrix effects evaluation. All the

processed fish samples came from Italian supermarkets. The Anisakidae larvae used for the artificial contamination of the samples were collected from *Lepidopus caudatus*, *Clupea harengus*, and *Merluccius merluccius* samples after visual inspection and modified chloro-peptic digestion [40]. The larvae isolated were washed in physiological saline serum (pH 7) and morphologically identified by B-800 light microscopy (Optika, Ponteranica, Italy) according to taxonomic keys [41]. Furthermore, *Anisakis* morphotype II, *Hysterothylacium* sp., *Contracaecum* sp., and *Pseudoterranova* sp. larvae were taken from the reference materials of the Centro di Referenza Nazionale per le Anisakiasi (Palermo, Italy). The number of *Anisakis* spp. larvae used for artificial contamination has been defined according to the prevalence of infestation of the fish species examined and reported in the literature [8,42–44]. The larvae were cut into small pieces and then were carefully mixed with the processed fish samples.

2.2. DNA Extraction

Genomic DNA was extracted from positive and negative fish samples, contaminated or not with *Anisakis* spp., respectively. The extraction was also carried out for the samples artificially infested with *Contracaecum* sp., *Pseudoterranova* sp., and *Hysterothylacium* sp. The DNA extraction was performed using a ready-to-use buffer contained in the Anisakis Screen Glow kit (Enbitech S.r.l., Palermo, Italy). Then, 250 ± 50 mg of sample was directly placed into 15 mL tubes containing 4 mL of the ready-to-use extraction buffer (Enbitech S.r.l., Palermo, Italy) and then incubated for 40 ± 5 min at room temperature.

2.3. Primers Design and LAMP Assays

To design the primer set targeting *Anisakis* spp. gene, the genomic sequences of the internal transcribed spacer 2 gene from various species were collected from GenBank™ (EU624342.1, AY826720.1, AB277823.1, AB196671.1, AB277821.1, AB551660.1, HF911524.1, AY826723.1, EU718479.1, JQ912690.1, KF512840.1, JX535521.1, KF032062.1, EU327691.1). A set of six primers, two outer (F3 and B3), two inner (FIP and BIP), and two loop (LF and LB), which recognised eight distinct regions of the target gene, was designed.

The analytical and diagnostic assays to recognise *Anisakis* spp. DNA was performed using Anisakis Screen Glow commercial kit (Enbitech Group S.r.l., Palermo, Italy) with ICGENE mini portable instrument (Figure 1) (Enbitech Group s.r.l., Palermo, Italy), consisting of a real-time fluorimeter, monitored and regulated by the the ICGENE application (Enbitech Group s.r.l., Palermo, Italy), downloadable on various smart devices. The Anisakis Screen Glow commercial kit includes ready-to-use reaction tubes (containing primers, fluorescent dye, etc.) to achieve a rapid amplification of DNA template. The protocol to obtain the specific amplification of the target *Anisakis* spp. DNA was carried out in a mixture of a final volume of 55 μ L, including 22 μ L of Anisakis Screen Glow LAMP mix (Enbitech Group s.r.l., Palermo, Italy), 30 μ L of mineral oil, and 3 μ L of the extracted DNA samples. The mineral oil was added to the top of the reaction mixture to prevent evaporation. The amplification was optimised and performed at 65 °C for 35 min. Real-time monitoring of the fluorescence associated with the amplification was possible using the fluorimeter of the ICGENE portable instrument and the ICGENE application interface.

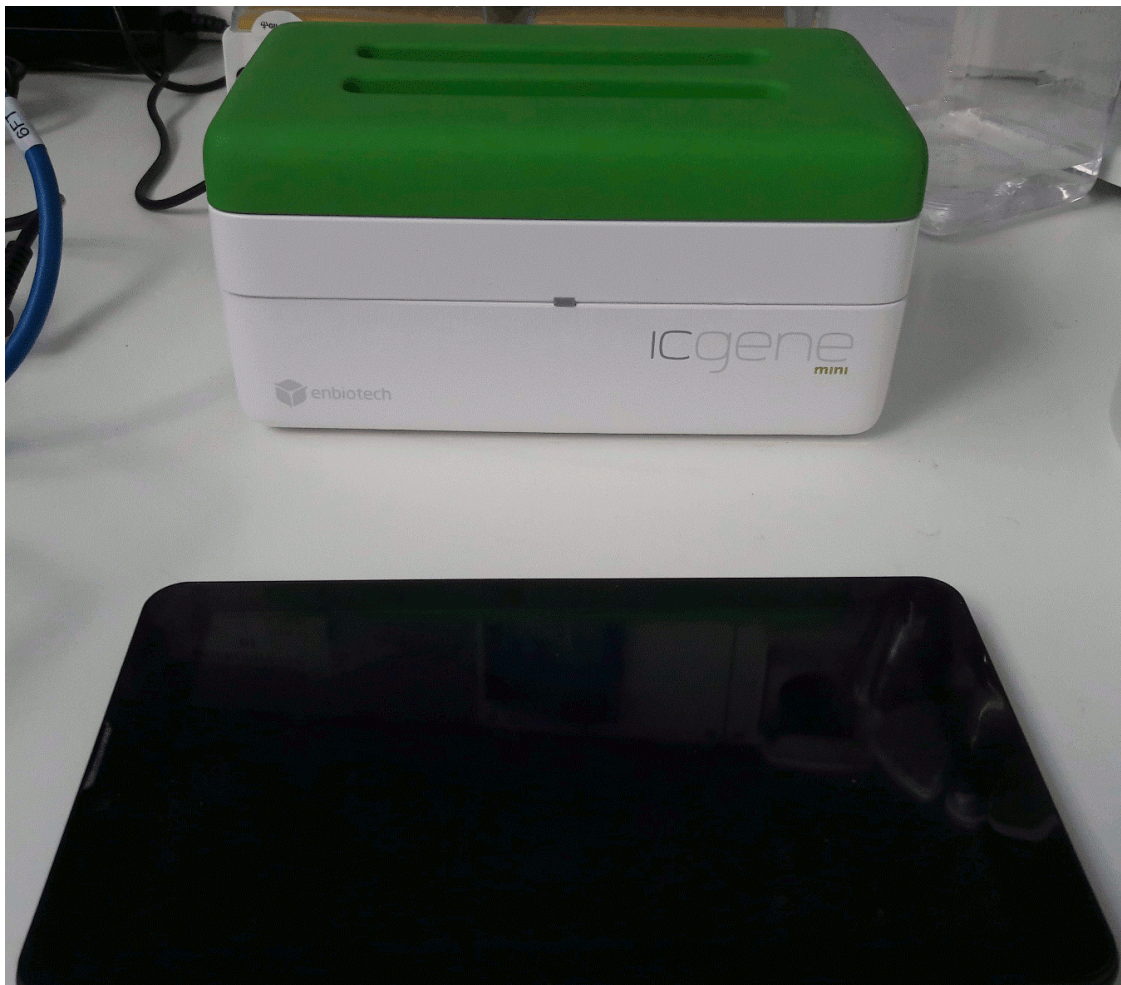


Figure 1. ICGENE mini portable instrument.

2.4. Specificity of the LAMP Assay

Based on the evolutionary relationships and their feasible genetic similarity, parasitic material belonging to the Anisakidae and Raphidascaridadae family was screened by the method proposed to have evidence on the diagnostic specificity of the LAMP assay. The parasitic material was previously characterised as belonging to *Hysterothylacium*, *Contraecum*, and *Pseudoterranova* genera using specific molecular diagnostic keys and methods reported in the literature [45–47]. Two different experiments were carried out for the specificity evaluation: (i) In the first experiment performed using genomic DNA, *P. decipiens* sensu stricto, *P. krabbei*, *P. cattani*, *P. azarasi*, *C. rudolphii* A, and *Hysterothylacium aduncum* were tested in duplicate, also including internal positive and negative controls of the Anisakis Screen Glow commercial kit (Enbiotech S.r.l., Palermo, Italy); (ii) in the second experiment, processed fish samples artificially contaminated with *Pseudoterranova* sp., *Contraecum* sp., and *Hysterothylacium* sp. larvae were subjected to DNA extraction and analysed in duplicate, together with positive and negative internal controls. Twenty samples of each processed fish product type, of which ten were artificially infested (5 with genomic DNA and 5 with larvae), were tested for the specificity assessment.

2.5. Sensitivity of the LAMP Assay

The sensitivity or inclusivity [48] was established as the ability of the LAMP method to detect DNA of *Anisakis* spp. (expressed as a percentage). Two different experiments were carried out for the inclusivity evaluation: (i) In the first experiment performed using genomic DNA, *A. pegreffii*, *A. simplex* sensu stricto, *A. typica*, *A. ziphidarum*, and *A. physeteris* were tested in duplicate, also including

internal positive and negative controls; (ii) in the second experiment, processed fish samples artificially contaminated with *Anisakis* spp. type I and type II larvae were subjected to DNA extraction and analysed in duplicate, together with positive and negative internal controls. Twenty samples of each processed fish product type were tested for the inclusivity assessment.

2.6. Limit of Detection

The limit of detection (LOD) was established as the lowest concentration of DNA of *Anisakis* species, which provided a significantly different signal to the negative control. The determination of the LOD of the LAMP method was assessed by serial 10-fold dilution of the DNA extracted from *Anisakis* spp. larvae with nuclease-free water. All measurements were performed in ten replicates from each sample type independently. The range of the DNA extracted varied between $2.22 \text{ ng } \mu\text{L}^{-1}$ and $8.40 \text{ ng } \mu\text{L}^{-1}$ with good 260/280 and 260/230 ratios (1.8 to 2.1). A test was considered acceptable when it ensured the detection of positive samples successfully with DNA content equal to or greater than LOD.

2.7. Real-Time PCR Assay

Real-Time PCR (RT-PCR) assays of the same samples analysed by LAMP were also carried out for a comparative purpose and as a confirmation method for the results of the LAMP assays. The RT-PCR amplification was carried out using the materials and following the validated protocols described by Cavallero et al. (2017) [27]. The extraction of DNA from artificially or noncontaminated processed fish samples was carried out using the Ion Force Fast kit (Generon, Modena, Italy), following the manufacturer's instructions. The PCR reactions were carried out by the commercial kit PATHfinder *Anisakis*/Pseudoterranova DNA detection assay (Generon, Modena, Italy). Five microliters of DNA extracted were mixed with 15 μL of PATHfinder *Anisakis*/Pseudoterranova kit for a total volume of 20 μL . The RT-PCR was carried out in a BIOER 9600 series Thermocycler (BIOER, Hangzhou, China) following the present thermal cycling conditions: A Taq activation at 95°C for 3 min, and 45 cycles of amplification (95°C for 10 s and 57°C for 16 s).

3. Results

3.1. Optimization of the LAMP Assay

The method was optimised for DNA extraction phase by testing in triplicate the initial weight of the samples at 50, 100, 250, and 350 mg with 0.5, 2, 4, and 8 mL of extraction buffer. The extract was tested with undiluted and diluted 1:5 and 1:10 for matrix effect assessment. An initial weight of 250 mg with 4 mL of extraction buffer and a 1:5 dilution after the extraction were found to be the best conditions for effective real-time detection of DNA amplification by the LAMP method proposed, giving the fluorescence intensity required for detection. The method was optimised by real-time monitoring of the time and temperature of reaction. The optimal reaction temperature and time for the LAMP assay was proved to be 65°C and 35 min, respectively.

3.2. Sensitivity and Specificity

The LAMP method proposed was able to amplify *Anisakis* spp. DNA from artificially infested fish samples, giving a sensitivity of 100% for each sample type analysed (Figure 2). Moreover, the method was also able to detect each sample contaminated with *A. simplex* s.s., *A. pegreffii*, *A. physeteris*, *A. ziphidarum*, and *A. typica* DNA. The assay detected *Anisakis* spp. DNA to a dilution of 10^{-4} ($0.00022 \text{ ng } \mu\text{L}^{-1}$), giving an amplification for all the replicates, with fluorescence values necessary for detection.

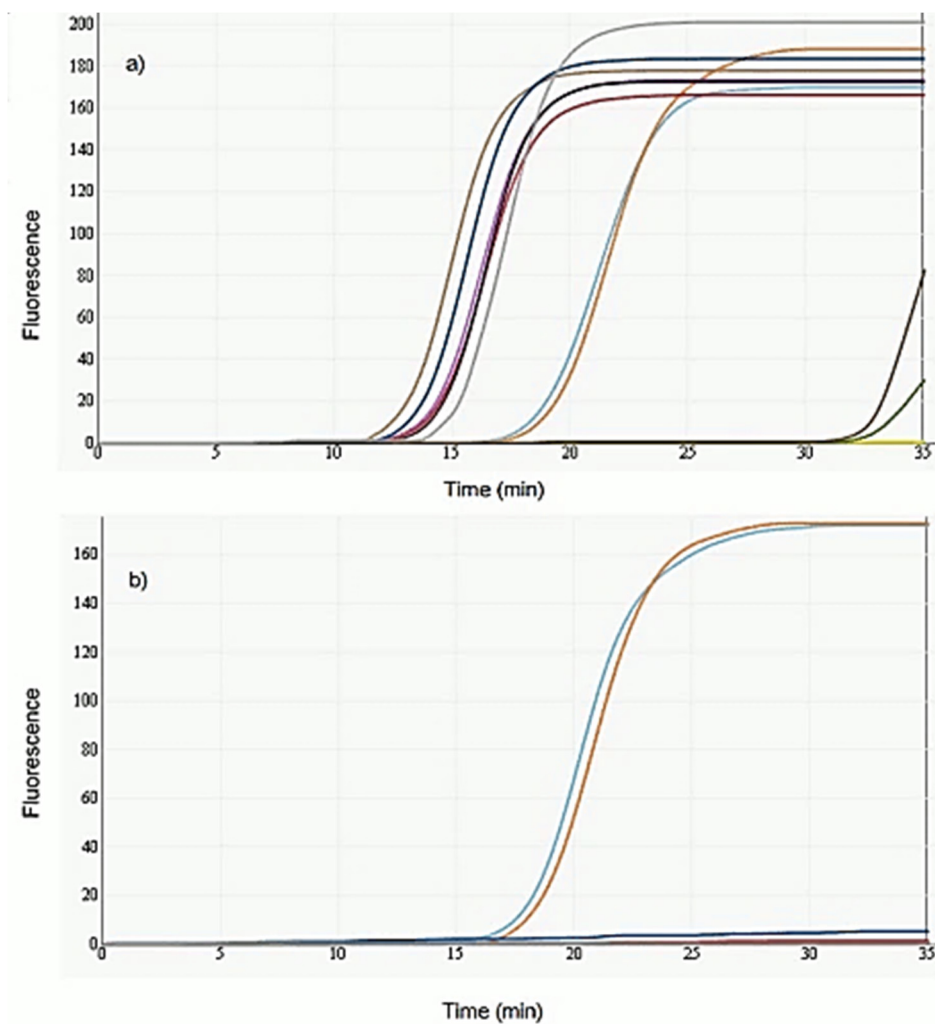


Figure 2. Monitoring of Loop-mediated isothermal amplification (LAMP) amplification for sensitivity (a) and specificity (b) tests from homogenized farmed salmon samples. The analysis shows the DNA amplification detection in real-time (colored curves shown in (a) and (b)) by measuring the increasing fluorescence of DNA binding to the dye. The amplification plots displayed in the specificity correspond to the positive control (duplicate).

All the LAMP analysis were carried out using positive and negative controls contained in the *Anisakis* Screen Glow kit. No amplification products were detected in uninfected samples, giving a very high specificity rate. Furthermore, no amplification was obtained on processed fish samples contaminated with *Contracaecum* sp., *Pseudoterranova* sp., and *Hysterothylacium* sp. larvae and DNA. The RT-PCR assay showed a sensitivity rate of 100% for the fish samples artificially infested with *Anisakis* sp. larvae and DNA. However, the analysis of uninfected samples and samples contaminated with *Pseudoterranova* sp., *Hysterothylacium* sp., and *Contracaecum* sp. larvae and DNA for the specificity test did not obtain any amplification.

In agarose gel analysis, the LAMP amplicons revealed a ladder-like pattern, according to what was reported in the literature, with many bands of different molecular weights, indicating the production of stem-loop DNA with inverted repeats of the target sequence (Parida et al., 2008; Li et al., 2012; Figure 3).

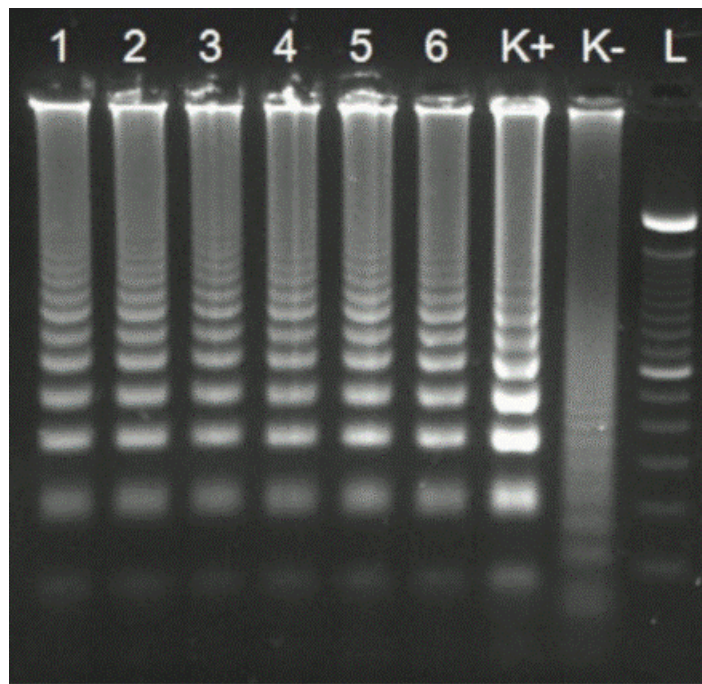


Figure 3. Amplification of LAMP for DNA extracted from anchovy paste samples experimentally infected with *Anisakis* spp. larvae (Lanes 1–6). Lane K+: Positive control, Lane K–: Negative control, Lane L: 100 bp DNA ladder.

4. Discussion

The method optimised with the set of primers employed was able to amplify *Anisakis* spp. DNA in 35 min at 65 °C from a considerable initial weight of samples, giving a satisfactory sensibility and specificity. The time needed for the LAMP assay was much lower than the RT-PCR method (about 94 min). The use of two loop primers and the ability of the two sets of primers (F3–B3, FIP–BIP) to recognise two distinct regions on the target DNA allowed acceleration of the reaction time and provided extremely high specificity. The specificity test has demonstrated that the LAMP assay proposed in this work can discriminate the *Anisakis* genera with respect to species belonging to other related genera that are not present in the Mediterranean Sea, such as *Pseudoterranova* spp. Furthermore, the method was able to discriminate the *Anisakis* genera with respect to nonallergic parasites such as *Hysterothylacium* spp, which proved that the LAMP primers are highly specific for the detection of *Anisakis* spp. The LAMP reaction primers recognise specifically eight independent regions in comparison with PCR primers, which can only recognise two independent regions, enhancing the sensitivity and specificity and decreasing the probability of false-positive results [49]. Generally, the fluorescence-based real-time monitoring of LAMP reaction is considerably faster than that performed by a real-time turbidimeter [50]. Moreover, compared to the real-time turbidity method, the real-time fluorescence method possesses two further advantages: The first is higher sensitivity; the second is that the sensitivity is less affected by the presence of opaque substances in the mixture, such as proteins [51]. The LAMP assay optimised was able to amplify *Anisakis* spp. DNA from different sample types, suggesting that the type of fish processing does not affect the quality of the assay by matrix interferences. The specificity and sensitivity of the LAMP method do not seem impaired by sample type, as confirmed by other studies [52]. However, there is a high risk of aerosol contamination due to the large amount of LAMP products. To reduce this risk, we adopted the use of mineral oil inside the reaction tubes.

The LAMP system we described here can detect the concentration of *Anisakis* spp. DNA up to 10^2 times lower than the RT-PCR method. The high sensitivity of such a method is undoubtedly an advantage, but needs special care to avoid false positives. The utilisation of ready-to-use reagents allowed us to minimise any operator error and the occurrence of possible false positives. Moreover,

the presence of non-target DNA and inhibitors in the LAMP reaction was shown to not affect the amplification results. The present method in combination with the ICGENE mini portable instrument proved to be accurate as of the Real-Time PCR method, but more rapid, easy to use, and with a lower limit of detection. Furthermore, the LAMP system proposed in this work is inexpensive, requires little equipment and technical support, is not space-consuming, and is easy to use.

The IC GENE instrument allows one to analyze up to ten samples per test, excluding positive and negative controls, with very low costs. Given the satisfactory results obtained during the validation and the ease of use, the samples do not need to be analyzed in duplicate. *Anisakis* spp. nematodes are considered one of the most critical biological hazards present in seafood products [53].

These food-borne parasites are also a hidden food allergen [54]. Concerning the *A. simplex* complex, there is no information about the minimum allergen concentration that causes allergic diseases. In this respect, [34] concluded that a single larva contains sufficient allergens to induce an antibody response in sensitive individuals. Exposure to this parasite seems to increase due to the increasing consumption of seafood worldwide and the increase of new gastronomic trends based on the consumption of raw and undercooked fish. Because all the *Anisakis* spp. studied originate the same disease and allergic reactions, the species identification is less relevant [24]. To prevent the occurrence of this pathogen in fish sectors, effective and efficient control procedures must be adopted throughout the fish industry and processing continuum. Therefore, it is necessary to use more efficient, simple, and cost- and time-effective methods. Rapid or alternative methods are important, as they reduce the detection time for *Anisakis* spp. considerably in comparison to other inspective methods and prevent the dissemination of these nematodes along the food chain.

The assay validated in this work has the advantages of being more straightforward and more sensitive than PCR and real-time PCR [55]. The LAMP method also has the characteristics of not requiring special reagents and sophisticated temperature control devices so the detection of LAMP products is also suitable for on-site conditions. Furthermore, the preservation of the reagents used for the LAMP assay only requires storage at >4 °C. Therefore, the LAMP assay proposed in this work should be considered a new and reliable tool for food quality and security control for prevention of *Anisakis* allergy in fish sectors.

The present system could be suitable for use by operators of the fish product processing industry for self-monitoring purposes, given its extreme ease and speed, in order to verify the correct evisceration practices of fish sector operators, following the suggestion of the Italian Ministry of Health [22]. Furthermore, due to its capability of discriminating *Anisakis* spp. DNA with respect to species belonging to the *Pseudoterranova* genus, the LAMP system proposed in this work can be a valuable tool to trace Mediterranean fish products.

5. Conclusions

The results obtained confirmed the reliability of the method analysed, which is faster and has more sensibility (in terms of limit of detection) than the real-time PCR method compared. In conclusion, the great rapidity, sensitivity, and ease of use suggest that LAMP assay can be a valid alternative for routine examination in the fishery sector to help manufacturers establish concepts for hazard analysis and critical control points (HACCPs) evaluation, where sophisticated and expensive equipment is not sustainable.

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References

- Abollo, E.; Gestal, C.; Pascual, S. *Anisakis* infestation in marine fish and cephalopods from Galician waters: An updated perspective. *Parasitol. Res.* **2001**, *87*, 492–499. [PubMed]
- Ferrantelli, V.; Costa, A.; Graci, S.; Buscemi, M.D.; Giangrosso, G.; Porcarello, C.; Palumbo, S.; Cammilleri, G. *Anisakid* Nematodes as Possible Markers to Trace Fish Products. *Ital. J. Food Saf.* **2015**, *4*, 4090. [CrossRef] [PubMed]
- Mattiucci, S.; Nascetti, G. Chapter 2 Advances and Trends in the Molecular Systematics of *Anisakid* Nematodes, with Implications for Their Evolutionary Ecology and Host—Parasite Co-evolutionary Processes. In *Advances in Parasitology*; Academic Press: Cambridge, MA, USA, 2008; Volume 66, pp. 47–148. Available online: <https://www.sciencedirect.com/science/article/abs/pii/S0065308X08002029> (accessed on 21 January 2017).
- Cammilleri, G.; Costa, A.; Graci, S.; Buscemi, M.D.; Collura, R.; Vella, A.; Pulvirenti, A.; Cicero, A.; Giangrosso, G.; Schembri, P.; et al. Presence of *Anisakis pegreffii* in farmed sea bass (*Dicentrarchus labrax* L.) commercialized in Southern Italy: A first report. *Vet. Parasitol.* **2018**, *259*, 13–16. [CrossRef] [PubMed]
- Chen, H.-X.; Zhang, L.-P.; Gibson, D.I.; Lü, L.; Xu, Z.; Li, H.-T.; Ju, H.-D.; Li, L. Detection of ascaridoid nematode parasites in the important marine food-fish *Conger myriaster* (Brevoort) (Anguilliformes: Congridae) from the Zhoushan Fishery, China. *Parasites Vectors* **2018**, *11*, 274. [CrossRef]
- Gaglio, G.; Battaglia, P.; Costa, A.; Cavallaro, M.; Cammilleri, G.; Graci, S.; Buscemi, M.D.; Ferrantelli, V.; Andaloro, F.; Marino, F. *Anisakis* spp. larvae in three mesopelagic and bathypelagic fish species of the central Mediterranean Sea. *Parasitol. Int.* **2018**, *67*, 23–28. [CrossRef]
- Mattiucci, S.; Cipriani, P.; Levsen, A.; Paoletti, M.; Nascetti, G. Chapter Four—Molecular Epidemiology of *Anisakis* and *Anisakiasis*: An Ecological and Evolutionary Road Map. In *Advances in Parasitology*; Rollinson, D., Stothard, J.R., Eds.; Academic Press: Cambridge, MA, USA, 2018; Volume 99, pp. 93–263.
- Cammilleri, G.; Pulvirenti, A.; Costa, A.; Graci, S.; Collura, R.; Buscemi, M.D.; Sciortino, S.; Badaco, V.V.; Vazzana, M.; Brunone, M.; et al. Seasonal trend of *Anisakidae* infestation in South Mediterranean bluefish. *Nat. Prod. Res.* **2020**, *34*, 158–161. [CrossRef]
- Arizono, N.; Yamada, M.; Tegoshi, T.; Yoshikawa, M. *Anisakis simplex* sensu stricto and *Anisakis pegreffii*: Biological Characteristics and Pathogenetic Potential in Human *Anisakiasis*. *Foodborne Pathog. Dis.* **2012**, *9*, 517–521. [CrossRef]
- Bao, M.; Pierce, G.J.; Pascual, S.; González-Muñoz, M.; Mattiucci, S.; Mladineo, I.; Cipriani, P.; Bušelić, I.; Strachan, N.J.C. Assessing the risk of an emerging zoonosis of worldwide concern: *Anisakiasis*. *Sci. Rep.* **2017**, *7*, 43699. [CrossRef]
- Fumarola, L.; Monno, R.; Ierardi, E.; Rizzo, G.; Giannelli, G.; Lalle, M.; Pozio, E. *Anisakis pegreffii* Etiological Agent of Gastric Infections in Two Italian Women. *Foodborne Pathog. Dis.* **2009**, *6*, 1157–1159. [CrossRef]
- Polimeno, L.; Loiacono, M.; Pesetti, B.; Mallamaci, R.; Mastrodonato, M.; Azzarone, A.; Annoscia, E.; Gatti, F.; Amoroso, A.; Ventura, M.T. *Anisakiasis*, an Underestimated Infection: Effect on Intestinal Permeability of *Anisakis simplex*—Sensitized Patients. *Foodborne Pathog. Dis.* **2010**, *7*, 809–814. [CrossRef]
- Sakanari, J.A.; McKerrow, J.H. *Anisakiasis*. *Clin. Microbiol. Rev.* **1989**, *2*, 278–284. [CrossRef]
- Audicana, M.T.; Ansotegui, I.J.; de Corres, L.F.; Kennedy, M.W. *Anisakis simplex*: Dangerous—Dead and alive? *Trends Parasitol.* **2002**, *18*, 20–25. [CrossRef]
- Alonso, A.; Daschner, A.; Moreno-Ancillo, A. Anaphylaxis with *Anisakis simplex* in the Gastric Mucosa. *N. Engl. J. Med.* **1997**, *337*, 350–351. [CrossRef] [PubMed]
- Alonso-Gómez, A.; Moreno-Ancillo, A.; López-Serrano, M.C.; Suarez-de-Parga, J.M.; Daschner, A.; Caballero, M.T.; Barranco, P.; Cabañas, R. *Anisakis simplex* only provokes allergic symptoms when the worm parasitises the gastrointestinal tract. *Parasitol. Res.* **2004**, *93*, 378–384. [CrossRef] [PubMed]
- Anadón, A.M.; Romarís, F.; Escalante, M.; Rodríguez, E.; Gárate, T.; Cuéllar, C.; Ubeira, F.M. The *Anisakis simplex* Ani s 7 major allergen as an indicator of true *Anisakis* infections. *Clin. Exp. Immunol.* **2009**, *156*, 471–478. [CrossRef] [PubMed]
- Rodríguez-Mahillo, A.I.; González-Muñoz, M.; De las Heras, C.; Tejada, M.; Moneo, I. Quantification of *Anisakis simplex* Allergens in Fresh, Long-Term Frozen, and Cooked Fish Muscle. *Foodborne Pathog. Dis.* **2010**, *7*, 967–973. [CrossRef]
- Choi, S.-J.; Lee, J.-C.; Kim, M.-J.; Hur, G.-Y.; Shin, S.-Y.; Park, H.-S. The Clinical Characteristics of *Anisakis* Allergy in Korea. *Korean J. Intern. Med.* **2009**, *24*, 160–163. [CrossRef]

20. Mazzucco, W.; Raia, D.D.; Marotta, C.; Costa, A.; Ferrantelli, V.; Vitale, F.; Casuccio, A. *Anisakis* sensitization in different population groups and public health impact: A systematic review. *PLoS ONE* **2018**, *13*, e0203671. [[CrossRef](#)]
21. European Food Safety Authority. Scientific Opinion on risk assessment of parasites in fishery products. *EFSA J.* **2010**, *8*, 1543. [[CrossRef](#)]
22. Salute, M. *Sicurezza Alimentare, Linee Guida in Materia di Igiene dei Prodotti della Pesca*; Ministero della Salute: Rome, Italy, 2015.
23. Seo, J.-S.; Jun, E.-J.; Jung, S.-H.; Kim, M.-S.; Park, M.-A.; Lee, C.-H.; Han, M.-C.; Kim, J.-W.; Jee, B.-Y. Prevalence of Anisakid larvae in chum salmon *Oncorhynchus keta* in Korea. *J. Fish Pathol.* **2010**, *23*, 123–129.
24. Herrero, B.; Vieites, J.M.; Espiñeira, M. Detection of anisakids in fish and seafood products by real-time PCR. *Food Control* **2011**, *22*, 933–939. [[CrossRef](#)]
25. Lluch-Bernal, M.; Sastre, J.; Fernández-Caldas, E.; Marañón, F.; Cuesta-Herranz, J.; las Heras De, M.; Quirce, S.; Novalbos, A. Conjunctival provocation tests in the diagnosis of *Anisakis simplex* hypersensitivity. *J. Investig. Allergol. Clin. Immunol.* **2002**, *12*, 21–24. [[PubMed](#)]
26. García-Palacios, L.; González, M.L.; Esteban, M.I.; Mirabent, E.; Perteguer, M.J.; Cuéllar, C. Enzyme-linked immunosorbent assay, immunoblot analysis and RAST fluoroimmunoassay analysis of serum responses against crude larval antigens of *Anisakis simplex* in a Spanish random population. *J. Helminthol.* **1996**, *70*, 281–289. [[CrossRef](#)] [[PubMed](#)]
27. Cavallero, S.; Bruno, A.; Arletti, E.; Caffara, M.; Fioravanti, M.L.; Costa, A.; Cammilleri, G.; Graci, S.; Ferrantelli, V.; D’Amelio, S. Validation of a commercial kit aimed to the detection of pathogenic anisakid nematodes in fish products. *Int. J. Food Microbiol.* **2017**, *257*, 75–79. [[CrossRef](#)]
28. Godínez-González, C.; Roca-Geronès, X.; Cancino-Faure, B.; Montoliu, I.; Fisa, R. Quantitative SYBR Green qPCR technique for the detection of the nematode parasite *Anisakis* in commercial fish-derived food. *Int. J. Food Microbiol.* **2017**, *261*, 89–94. [[CrossRef](#)] [[PubMed](#)]
29. Mossali, C.; Palermo, S.; Capra, E.; Piccolo, G.; Botti, S.; Bandi, C.; D’Amelio, S.; Giuffra, E. Sensitive Detection and Quantification of Anisakid Parasite Residues in Food Products. *Foodborne Pathog. Dis.* **2009**, *7*, 391–397. [[CrossRef](#)]
30. Zhang, S.L.; Wang, H.L.; Liu, J.; Ni, F.; Xu, S.S.; Luo, D.M. Detection of anisakid nematodes by an SYBR green I real-time PCR. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* **2010**, *28*, 194–199.
31. Costa, J.; Ansari, P.; Mafra, I.; Oliveira, M.B.P.P.; Baumgartner, S. Assessing hazelnut allergens by protein- and DNA-based approaches: LC-MS/MS, ELISA and real-time PCR. *Anal. Bioanal. Chem.* **2014**, *406*, 2581–2590. [[CrossRef](#)] [[PubMed](#)]
32. Stephan, O.; Weisz, N.; Vieths, S.; Weiser, T.; Rabe, B.; Vatterott, W. Protein Quantification, Sandwich ELISA, and Real-Time PCR Used to Monitor Industrial Cleaning Procedures for Contamination with Peanut and Celery Allergens. *J. AOAC Int.* **2004**, *87*, 1448–1457. Available online: <https://www.ingentaconnect.com/content/aoac/jaoac/2004/00000087/00000006/art00022> (accessed on 20 May 2019).
33. Stephan, O.; Vieths, S. Development of a Real-Time PCR and a Sandwich ELISA for Detection of Potentially Allergenic Trace Amounts of Peanut (*Arachis hypogaea*) in Processed Foods. *J. Agric. Food Chem.* **2004**, *52*, 3754–3760. [[CrossRef](#)]
34. Lopez, I.; Pardo, M.A. Evaluation of a Real-Time Polymerase Chain Reaction (PCR) Assay for Detection of *Anisakis simplex* Parasite as a Food-Borne Allergen Source in Seafood Products. *J. Agric. Food Chem.* **2010**, *58*, 1469–1477. [[CrossRef](#)]
35. Notomi, T.; Okayama, H.; Masubuchi, H.; Yonekawa, T.; Watanabe, K.; Amino, N.; Hase, T. Loop-mediated isothermal amplification of DNA. *Nucleic Acids Res.* **2000**, *28*, e63. [[CrossRef](#)] [[PubMed](#)]
36. Nagamine, K.; Hase, T.; Notomi, T. Accelerated reaction by loop-mediated isothermal amplification using loop primers. *Mol. Cell. Probes* **2002**, *16*, 223–229. [[CrossRef](#)] [[PubMed](#)]
37. Peñalver, J.; Dolores, E.M.; Muñoz, P. Absence of Anisakid Larvae in Farmed European Sea Bass (*Dicentrarchus labrax* L.) and Gilthead Sea Bream (*Sparus aurata* L.) in Southeast Spain. *J. Food Prot.* **2010**, *73*, 1332–1334.
38. Lunestad, B.T. Absence of Nematodes in Farmed Atlantic Salmon (*Salmo salar* L.) in Norway. *J. Food Prot.* **2003**, *66*, 122–124. [[CrossRef](#)]
39. Angot, V.; Brasseur, P. European farmed Atlantic salmon (*Salmo salar* L.) are safe from anisakid larvae. *Aquaculture* **1993**, *118*, 339–344. [[CrossRef](#)]

40. Cammilleri, G.; Chetta, M.; Costa, A.; Graci, S.; Collura, R.; Buscemi, M.D.; Cusimano, M.; Alongi, A.; Principato, D.; Giangrosso, G.; et al. Validation of the TrichinEasy[®] digestion system for the detection of *Anisakidae* larvae in fish products. *Acta Parasitol.* **2016**, *61*, 369–375. [CrossRef]
41. Berland, B. Nematodes from some Norwegian marine fishes. *Sarsia* **1961**, *2*, 1–50. [CrossRef]
42. Cavallero, S.; Magnabosco, C.; Civettini, M.; Boffo, L.; Mingarelli, G.; Buratti, P.; Giovanardi, O.; Fortuna, C.M.; Arcangeli, G. Survey of *Anisakis* sp. and *Hysterothylacium* sp. in sardines and anchovies from the North Adriatic Sea. *Int. J. Food Microbiol.* **2015**, *200*, 18–21. [CrossRef]
43. Ciccarelli, C.; Aliventi, A.; Trani, V.D.; Semeraro, A.M. Assessment of the prevalence of *Anisakidae* larvae prevalence in anchovies in the central Adriatic Sea. *Ital. J. Food Saf. 1(1zero)* **2011**, *1*, 15–19. [CrossRef]
44. Cipriani, P.; Acerra, V.; Bellisario, B.; Sbaraglia, G.L.; Cheleschi, R.; Nascetti, G.; Mattiucci, S. Larval migration of the zoonotic parasite *Anisakis pegreffii* (Nematoda: Anisakidae) in European anchovy, *Engraulis encrasicolus*: Implications to seafood safety. *Food Control* **2016**, *59*, 148–157. [CrossRef]
45. Cavallero, S.; Nadler, S.A.; Paggi, L.; Barros, N.B.; D'Amelio, S. Molecular characterization and phylogeny of anisakid nematodes from cetaceans from southeastern Atlantic coasts of USA, Gulf of Mexico, and Caribbean Sea. *Parasitol. Res.* **2011**, *108*, 781–792. [CrossRef] [PubMed]
46. Costa, A.; Cammilleri, G.; Graci, S.; Buscemi, M.D.; Vazzana, M.; Principato, D.; Giangrosso, G.; Ferrantelli, V. Survey on the presence of *A. simplex* s.s. and *A. pegreffii* hybrid forms in Central-Western Mediterranean Sea. *Parasitol. Int.* **2016**, *65*, 696–701. [CrossRef] [PubMed]
47. D'Amelio, S.; Mathiopoulou, K.D.; Santos, C.P.; Pugachev, O.N.; Webb, S.C.; Picanço, M.; Paggi, L. Genetic markers in ribosomal DNA for the identification of members of the genus *Anisakis* (Nematoda: Ascaridoidea) defined by polymerase-chain-reaction-based restriction fragment length polymorphism. *Int. J. Parasitol.* **2000**, *30*, 223–226. [CrossRef]
48. Feldsine, P.; Abeyta, C.; Andrews, W.H.; AOAC International Methods Committee. AOAC International methods committee guidelines for validation of qualitative and quantitative food microbiological official methods of analysis. *J. AOAC Int.* **2002**, *85*, 1187–1200. [PubMed]
49. Li, X.; Liu, W.; Wang, J.; Zou, D.; Wang, X.; Yang, Z.; Yin, Z.; Cui, Q.; Shang, W.; Li, H.; et al. Rapid detection of *Trichinella spiralis* larvae in muscles by loop-mediated isothermal amplification. *Int. J. Parasitol.* **2012**, *42*, 1119–1126. [CrossRef]
50. Zhang, X.; Lowe, S.B.; Gooding, J.J. Brief review of monitoring methods for loop-mediated isothermal amplification (LAMP). *Biosens. Bioelectron.* **2014**, *61*, 491–499. [CrossRef]
51. Francois, P.; Tangomo, M.; Hibbs, J.; Bonetti, E.-J.; Boehme, C.C.; Notomi, T.; Perkins, M.D.; Schrenzel, J. Robustness of a loop-mediated isothermal amplification reaction for diagnostic applications. *FEMS Immunol. Med. Microbiol.* **2011**, *62*, 41–48. [CrossRef]
52. Kong, Q.-M.; Lu, S.-H.; Tong, Q.-B.; Lou, D.; Chen, R.; Zheng, B.; Kumagai, T.; Wen, L.-Y.; Ohta, N.; Zhou, X.-N. Loop-mediated isothermal amplification (LAMP): Early detection of *Toxoplasma gondii* infection in mice. *Parasites Vectors* **2012**, *5*, 2. [CrossRef]
53. Scientific Opinion on Risk Assessment of Parasites in Fishery Products|Autorità Europea per la Sicurezza Alimentare. Available online: <https://www.efsa.europa.eu/it/efsajournal/pub/1543> (accessed on 30 June 2017).
54. Nieuwenhuizen, N.E.; Lopata, A.L. Allergic Reactions to *Anisakis* Found in Fish. *Curr. Allergy Asthma Rep.* **2014**, *14*, 455. [CrossRef]
55. Parida, M.; Sannarangaiah, S.; Dash, P.K.; Rao, P.V.L.; Morita, K. Loop mediated isothermal amplification (LAMP): A new generation of innovative gene amplification technique; perspectives in clinical diagnosis of infectious diseases. *Rev. Med. Virol.* **2008**, *18*, 407–421. [CrossRef] [PubMed]

