

# **HHS Public Access**

Author manuscript *Curr Opin Cell Biol.* Author manuscript; available in PMC 2021 February 01.

Published in final edited form as:

Curr Opin Cell Biol. 2020 February ; 62: 37-45. doi:10.1016/j.ceb.2019.08.001.

# Retinal Pigment Epithelium Polarity in Health and Blinding Diseases

## Paulo S. Caceres, Enrique Rodriguez-Boulan

Weill Cornell Medical College, Department of Ophthalmology, Margaret Dyson Vision Research Institute, New York, NY, USA, 10065.

# Abstract

The polarized phenotype of the retinal pigment epithelium is critical for the outer retina-blood barrier and support of photoreceptors and underlying choroid, and its disruption plays a central role in degenerative retinopathies. Although the mechanisms of polarization remain mostly unknown, they are fundamental for homeostasis of the outer retina. Recent research is revealing a growing picture of interconnected tissues in the outer retina, with the retinal pigment epithelium at the center. This review discusses how elements of epithelial polarity relate to emerging apical interactions with the neural retina, basolateral cross-talk with the underlying Bruch's membrane and choriocapillaris, and tight junction biology. An integrated view of outer retina physiology is likely to provide insights into the pathogenesis of blinding diseases.

# Keywords

apical- basolateral polarity; tight junction; outer retina- blood barrier; tissue cross- talk; choroid

# Introduction

The retinal pigment epithelium (RPE) is located strategically between the photoreceptors and the choroidal vasculature (Figure 1A), the only blood supply for the outer retina [1]. As many epithelia, the RPE is polarized into apical and basolateral plasma membrane domains separated by tight junctions. However, many plasma membrane proteins display a reversed polarity compared with extraocular epithelia, e.g. Na/K-ATPase [2], Integrin  $\alpha\nu\beta5$  [3], NCAM [4], CD147 [5], CAR [6] and Monocarboxylate transporters [7]. The cellular mechanisms that determine RPE polarity are poorly understood, but they involve sorting and transit of proteins through endosomal compartments (Reviewed in refs [8,9]). Characterizing these mechanisms is a critical need in retina physiology since the polarized organization of the RPE mediates important support functions to the neural retina and maintains access to

Conflicts of interest: None

**Corresponding authors:** E. Rodriguez-Boulan and P.S. Caceres are co-corresponding authors. Rodriguez-Boulan, Enrique (boulan@med.cornell.edu), Caceres, Paulo S. (psc2004@med.cornell.edu), Address: 1300 York Avenue, LC-305, New York, NY 10065, Phone: (212) 746-2277.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

the choroidal blood supply [1,9]. Basolateral RPE interactions with a laminar extracellular matrix called Bruch's membrane and the choriocapillaris constitutes the outer-retina blood barrier (ORBB), which regulates exchange of substances between the neural retina and the circulation [8,10]. Here we will discuss an emerging picture of cross-talk mechanisms between the neural retina and the ORBB, and also within the components of the ORBB itself (Figure 1B). We will consider how the three defining characteristics of the epithelial phenotype: tight junctions, apical plasma membrane, and basolateral domain including the Bruch's membrane, relate to maintenance of the outer retina as a functional unit.

# **Tight Junctions**

The RPE tight junctions constitute a key component of the ORBB since they restrict the passage of substances between the choroid and the outer retina. Recent work highlighted a crucial role of the tight junction protein Claudin-19 in RPE trans-epithelial permeability, retinal differentiation and disease [11]. Disruption of RPE cell-cell contacts was recently found to stimulate production of Vascular endothelial growth factor (VEGF) in cell culture [12], a secreted pro-angiogenic factor that we will discuss later. This is part of an emerging concept that associates intact tight junctions with VEGF secretion, as several groups have proposed strategies that target RPE tight junctions to minimize the detrimental effects of VEGF during diabetic retinopathy and choroidal neovascularization [13–15].

Given the importance of tight junctions for maintenance of the epithelial phenotype, recent interest has been directed towards novel factors that increase tight junction permeability like C-reactive protein [16] and tissue factor [17]; or stimulate tight junction formation like somatostatin [13], nicotinamide [18], lysophosphatidic acid [19] and HIWI2-mediated activation of Akt [20]. Most of these recent studies were performed in RPE cell lines, therefore their translational relevance in native RPE still remains to be determined. A special mechanism of tight junction formation is mediated by interactions with the basement membrane [21]. Loss of RPE tight junctions could lead to epithelial-mesenchymal transition and recent reports have addressed the possible contributions of this process to proliferative retinopathy [22,23], diabetic retinopathy [24] and age-related macular degeneration (AMD) [25].

# Apical Domain

RPE barrier functions rely on directional solute and fluid transport through polarized epithelial transporters and channels. This activity is the foundation of many RPE support functions to the photoreceptors, since otherwise they would be isolated from the circulation. These support functions are summarized in Figure 2 and reviewed elsewhere [1]. They explain the close relationship between the RPE apical membrane and photoreceptors.

#### Polarized RPE solute transport

Many RPE support functions rely on the plasma membrane distribution of transporters. Recent research has advanced our understanding of key membrane transporters in the RPE. For instance, glucose is transported from the choroid to the photoreceptors via GLUT1 [26– 28]. The photoreceptors utilize this glucose to produce lactate, which is eliminated and

absorbed by the RPE via apical MCT1 transporters [7] (Figure 2). Recently, Kanow et al. [29] showed that RPE cells favor lactate as a nutrient over glucose, which is instead preferred by the photoreceptors. In addition, the RPE also utilizes proline to produce citrate and other metabolites, which are secreted apically for photoreceptor utilization [30]. Importantly, dietary proline protected the RPE from oxidative damage [31]. This emerging retinal ecosystem of coordinated nutrient transport and utilization provides the framework for membrane distribution of RPE transporters (Figure 2).

Fluid transport by the RPE maintains the low volume of the apical subretinal space and it has been proposed to depend on KCl transport [1,8]. Bestrophin is a basolateral Cl<sup>-</sup> channel affected in retinal degenerative diseases (recently reviewed [32]). Recently, Bestrophin has been shown to be regulated by ATP binding to a site mutated in patients [33], and also possibly by trafficking to the plasma membrane [34,35], although the latter remains to be confirmed in RPE. K<sup>+</sup> is transported via cation channels, like the newly recognized Anoctamin 4 in the RPE [36]. Further characterization of polarized ion transport will undoubtedly contribute to our full understanding of directional fluid transport by the RPE.

#### Apical receptor-mediated RPE interactions with photoreceptors

A major support function of the RPE is circadian clearing of shed photoreceptor outer segments by phagocytosis. This requires binding of milk fat globule–EGF 8 (MFG-E8) to RPE apical  $\alpha\nu\beta5$  Integrin receptors [3,37] (Figure 2), and internalization via activity of focal adhesion kinase (FAK) and Mer tyrosine kinase (MerTK) [38–40]. Recently, it was found that apical Plexin-B1 serves as a receptor for Semaphorin-4D, inhibiting outer segment internalization, therefore contributing to the circadian pattern [41]. The Plexin-B1/ Semaphorin-4D pair may be important for retina homeostasis as knockout mice have higher phagosome content [41].

Phagocytosed outer segments are a source of cholesterol that needs to be eliminated from the RPE (Figure 2). Recent work has shown that cholesterol efflux occurs via apical and basolateral ABCA1 transporters [42]. This bidirectional efflux is relevant because cholesterol accumulates in basolateral (drusen) and apical (subretinal drusenoid) deposits, which have been independently associated with macular degeneration [43–45]. However, the bidirectional efflux could be subject to differential regulation since cholesterol loading stimulates efflux preferentially towards the apical side [46]. Defects in RPE phagocytosis have been linked to photoreceptor degeneration [47], and further characterization of the RPE endocytic pathway may shed light into the mechanisms of retinal disease [48,49]. An important step is to identify the physiological roles of additional endocytic receptors, for instance the apical receptor Megalin is necessary for retina structure and visual function as described in recently generated RPE-specific Megalin knockout mice [50].

#### **RPE primary cilium**

Early studies suggested that the primary cilium disappeared in mature RPE [51]. However, this observation may have been due to artefacts in the histological preparation [9]. The functional role of the RPE primary cilium has recently been confirmed by two groups. Buskin et al. [52] found polarity and functionality defects in RPE cells obtained from

retinitis pigmentosa patients carrying mutations in mRNA splicing factors (PRPF) that control cilia genes. May-Simera et al. [53] showed that disruption of ciliary proteins in cultured RPE cells causes abnormal RPE phenotype, possibly by interfering with PKC8, a mediator of apico-basal polarity. The authors also observed abnormal morphology in RPE cells obtained from patients harboring a mutation in CEP290 and mice knockout for Bbs8, two ciliary proteins illustrated in Figure 2.

#### Neural retina-RPE interaction regulates choroid development

An underdeveloped area of research is how the neural retina influences the choroidal vasculature, despite this being potentially critical to maintain a blood supply to the outer retina. A remarkable study by Goto et al. [54] described a pathway involving retinoic acids produced via Aldehyde Dehydrogenase 1 A1 (Aldh1a1) in the neural retina, from where they reach the RPE and activate the transcription factor Sox9, which controls VEGF expression and choroid development (Figure 2). In agreement with this, another study found that the transcriptional relationship Pax6-Sox9 controls coordinated RPE and choroid vasculature development [55]. It was also found that Pax6-Sox9 control expression of other factors in addition to VEGF that could be secreted by the RPE towards the choroid. These recent reports reveal a growing picture of cell-cell communication between tissues at the outer retina. An important question is the nature of these pathways in adult tissue and their contributions to retinopathies.

#### **Basolateral Domain**

An important aspect of RPE polarity is the ability to basolaterally secrete components of the Bruch's membrane and choroidal trophic factors (Figure 3). The Bruch's membrane is a five-layer sheet rich in elastin and collagen [56]. The outermost layer is the RPE basement membrane, composed of collagen-IV, laminin, fibronectin and glycosaminoglycans [57]. Each of these components has a distinct effect in RPE adhesion [58] and barrier properties [59]. The RPE secretes VEGF basolaterally [60], a growth factor for the choriocapillaris supplying the outer retina. Diffusion of substances to and from the choriocapillaris and across the Bruch's membrane is central to ORBB function.

#### Bruch's membrane

The composition of the Bruch's membrane depends on synthesis and degradation of ECM components [57], which change with age [61] and during AMD [62]. This impairs RPE adhesion [63,64] and is relevant for RPE-based therapies [65]. A Bruch's membrane-modifying factor of recent interest is Fibulin-3 (*EFEMP1*), an ECM-remodeling protein secreted by the RPE (Figure 3) and associated with macular degeneration [66]. In AMD eyes, Fibulin-3 accumulates between the RPE and Bruch's membrane [66,67]. The role of Fibulin-3 in the Bruch's membrane is not well understood. However, it is likely to regulate matrix-metalloproteases (MMP), its regulators (TIMP) and proteoglycans as shown recently in mice carrying a Fibulin-3 mutation that causes macular degeneration and impaired diffusion across the Bruch's membrane [68]. Polarized secretion of MMPs and TIMPs may be of great relevance in ECM-remodeling occurring during RPE wound-healing, which is associated with PVR [69].

Caceres and Rodriguez-Boulan

Several recent studies have focused on Bruch's membrane-mediated RPE signaling. For instance, the complement pathway, essential for RPE defense functions, can be activated by modifications in the ECM [70–72]. Such alterations could be post-translational modifications induced by oxidative stress, e.g. nitration and advanced glycation end products [72–74], which accumulate during retinal disease. The recent emphasis in these areas can benefit stem cell and RPE transplantation therapies, which require correct differentiation of RPE cells on the adequate substrate [75,76]. It has also been shown that aged Bruch's membrane impairs RPE cell adhesion and function [63].

The close association between the RPE and the choroid vasculature can also modify the Bruch's membrane. It was recently shown that when co-cultured with endothelial cells, an RPE cell line increases ECM deposition and expression of ECM genes [77]. It is unknown how this compare to native Bruch's membrane since this study used cell lines and the endothelial cells were not of choroidal origin. There is also evidence that the choroid endothelium can directly modify the Bruch's membrane. We recently observed a switch in expression profile of ECM genes in mice choroid during early postnatal development [21]. This was associated with secretion of collagen cross-linker lysyl oxidases by endothelial cells, which increased the assembly of ECM collagen bundles in co-cultures with primary RPE cells (Figure 3). Importantly, this EC-mediated modification of the ECM enhanced the barrier properties of the RPE [21]. These findings illustrate the ability of endothelial cells to influence the RPE phenotype and the ORBB.

#### **RPE/choroid cross-talk**

Our findings that endothelial cells promote RPE barrier properties [21], together with a recent similar report [77], are part of an emerging notion of endothelia as instructive niches for regeneration and homeostasis of surrounding tissues [78,79]. The signals originating in the choroid can modify the Bruch's membrane as discussed, and we found that this activates basolateral  $\beta$ 1-integrin receptors in RPE cells [21] (Figure 3). Lysyl oxidases are secreted factors that we found modified the Bruch's membrane and preserved vision [21]. However, additional choroidal factors could also target the RPE directly. Future work will need to identify novel secreted factors in choroid-RPE cross talk.

One of the best characterized RPE secreted factors is VEGF, which is released basolaterally (Figure 3) and maintains the choriocapillaris [60,80,81]. RPE-specific knockout of VEGF in adult mice resulted in dramatic ablation of the choriocapillaris and deficient vision [80]. Recent work has shown that VEGF inhibition results in decreased complement regulators, including complement factor H through a mechanism that involves VEGF receptor 2, PKC-alpha and CREB signaling [82]. These results suggest that VEGF protects the choroid vasculature through both trophic and complement regulatory functions. The ramifications of VEGF signaling in the choroid are expected to continue to dominate the field since it has important implications in therapy of neovascular (wet) AMD. However, additional regulators of angiogenesis secreted by the RPE may represent alternatives to anti-VEGF treatment. It was recently reported that Galectin-1 (Figure 3) is increased in RPE in a mouse model of choroidal neovascularization and it promotes choroid angiogenesis and fibrosis [83]. It was recently proposed that exosomes secreted by RPE cells in culture (Figure 3) could promote

angiogenesis via a VEGF-independent pathway [84]. However, in vivo confirmation is still pending. A word of caution, a separate study using the same RPE and endothelial cell lines established a negative relationship in angiogenesis [77], while it is known that RPE promotes angiogenesis in native choroid [85].

# **Concluding remarks**

Despite the importance of RPE polarization to retina physiology, how this polarity is achieved is poorly understood. Part of this phenotype may be explained by the absence of the clathrin adaptor protein AP-1B, which is characteristic of epithelial cells [6]. However, this small aspect of RPE polarity still doesn't explain the complex phenotype of this epithelium. Recently, we used quantitative surface proteomics to uncover extended roles of clathrin adaptors in epithelial polarity [86]. Similar approaches applied to RPE cells could shed light on the mechanisms of RPE polarity [87], and the growing body of interactions between the RPE and neighboring tissues. A holistic picture of the connections among the cell types in the outer retina will be fundamental in our understanding of retinopathies like macular degeneration, proliferative and diabetic retinopathies.

# Acknowledgments:

We would like to thank Dr. Silvia Finnemann for critical reading of the manuscript. E.R.B is funded by grant EY008538 and P.S.C. by award EY008538–28S1 from the National Institutes of Health (National Eye Institute).

## References

\* of special interest

\*\* of outstanding interest

- Strauss O: The retinal pigment epithelium in visual function. Physiol Rev 2005, 85:845–881. [PubMed: 15987797]
- Caldwell RB, McLaughlin BJ: Redistribution of Na-K-ATPase in the dystrophic rat retinal pigment epithelium. J Neurocytol 1984, 13:895–910. [PubMed: 6100117]
- 3. Finnemann SC, Bonilha VL, Marmorstein AD, Rodriguez-Boulan E: Phagocytosis of rod outer segments by retinal pigment epithelial cells requires alpha(v)beta5 integrin for binding but not for internalization. Proc Natl Acad Sci U S A 1997, 94:12932–12937. [PubMed: 9371778]
- 4. Gundersen D, Powell SK, Rodriguez-Boulan E: Apical polarization of N-CAM in retinal pigment epithelium is dependent on contact with the neural retina. J Cell Biol 1993, 121:335–343. [PubMed: 8468350]
- Deora AA, Gravotta D, Kreitzer G, Hu J, Bok D, Rodriguez-Boulan E: The basolateral targeting signal of CD147 (EMMPRIN) consists of a single leucine and is not recognized by retinal pigment epithelium. Mol Biol Cell 2004, 15:4148–4165. [PubMed: 15215314]
- Diaz F, Gravotta D, Deora A, Schreiner R, Schoggins J, Falck-Pedersen E, Rodriguez-Boulan E: Clathrin adaptor AP1B controls adenovirus infectivity of epithelial cells. Proc Natl Acad Sci U S A 2009, 106:11143–11148. [PubMed: 19549835]
- Philp NJ, Yoon H, Grollman EF: Monocarboxylate transporter MCT1 is located in the apical membrane and MCT3 in the basal membrane of rat RPE. Am J Physiol 1998, 274:R1824–1828. [PubMed: 9841555]
- 8. Caceres PS, Benedicto I, Lehmann GL, Rodriguez-Boulan EJ: Directional Fluid Transport across Organ-Blood Barriers: Physiology and Cell Biology. Cold Spring Harb Perspect Biol 2017, 9.

- Lehmann GL, Benedicto I, Philp NJ, Rodriguez-Boulan E: Plasma membrane protein polarity and trafficking in RPE cells: past, present and future. Exp Eye Res 2014, 126:5–15. [PubMed: 25152359]
- Bhutto I, Lutty G: Understanding age-related macular degeneration (AMD): relationships between the photoreceptor/retinal pigment epithelium/Bruch's membrane/choriocapillaris complex. Mol Aspects Med 2012, 33:295–317. [PubMed: 22542780]
- Wang SB, Xu T, Peng S, Singh D, Ghiassi-Nejad M, Adelman RA, Rizzolo LJ: Disease-associated mutations of claudin-19 disrupt retinal neurogenesis and visual function. Commun Biol 2019, 2:113. [PubMed: 30937396] \* This article shows that disease-causing mutations in the RPE tight junction protein claudin-19 impair RPE maturation and function, retina neurogenesis and vision.
- Farjood F, Vargis E: Physical disruption of cell-cell contact induces VEGF expression in RPE cells. Mol Vis 2017, 23:431–446. [PubMed: 28761317]
- Fonollosa A, Valcarcel M, Salado C, Pereiro X, Vecino E: Effect of somatostatin on human retinal pigment epithelial cells permeability. Exp Eye Res 2019, 184:15–23. [PubMed: 30978347]
- Obert E, Strauss R, Brandon C, Grek C, Ghatnekar G, Gourdie R, Rohrer B: Targeting the tight junction protein, zonula occludens-1, with the connexin43 mimetic peptide, alphaCT1, reduces VEGF-dependent RPE pathophysiology. J Mol Med 2017, 95:535–552. [PubMed: 28132078]
- 15. Ved N, Hulse RP, Bestall SM, Donaldson LF, Bainbridge JW, Bates DO: Vascular endothelial growth factor-A165b ameliorates outer-retinal barrier and vascular dysfunction in the diabetic retina. Clin Sci 2017, 131:1225–1243. [PubMed: 28341661] \* This article describes how the VEGF splice variant VEGFA165b protects RPE tight junctions and barrier functions, minimizing the development of diabetic retinopathy in rats.
- Molins B, Pascual A, Mendez, Llorenc V, Zarranz-Ventura J, Mesquida M, Adan A, Martorell J: Creactive protein isoforms differentially affect outer blood-retinal barrier integrity and function. Am J Physiol Cell Physiol 2017, 312:C244–c253. [PubMed: 28003224]
- Zou XL, Wang GF, Li DD, Chen JX, Zhang CL, Yu YZ, Zhou WJ, Zou YP, Rao BQ: Protection of tight junction between RPE cells with tissue factor targeting peptide. Int J Ophthalmol 2018, 11:1594–1599. [PubMed: 30364251]
- Hazim RA, Volland S, Yen A, Burgess BL, Williams DS: Rapid differentiation of the human RPE cell line, ARPE-19, induced by nicotinamide. Exp Eye Res 2019, 179:18–24. [PubMed: 30336127]
- Lidgerwood GE, Morris AJ, Conquest A, Daniszewski M, Rooney LA, Lim SY, Hernandez D, Liang HH, Allen P, Connell PP, et al.: Role of lysophosphatidic acid in the retinal pigment epithelium and photoreceptors. Biochim Biophys Acta Mol Cell Biol Lipids 2018, 1863:750–761. [PubMed: 29660533]
- Sivagurunathan S, Palanisamy K, Arunachalam JP, Chidambaram S: Possible role of HIWI2 in modulating tight junction proteins in retinal pigment epithelial cells through Akt signaling pathway. Mol Cell Biochem 2017, 427:145–156. [PubMed: 28025795]
- 21. Benedicto I, Lehmann GL, Ginsberg M, Nolan DJ, Bareja R, Elemento O, Salfati Z, Alam NM, Prusky GT, Llanos P, et al.: Concerted regulation of retinal pigment epithelium basement membrane and barrier function by angiocrine factors. Nat Commun 2017, 8:15374. [PubMed: 28524846] \*\* This study describes an angiocrine mechanism by which extracellular matrix-remodeling lysyl oxidases secreted by endothelial cells increase the stiffness of the Bruch's membrane, which is sensed by RPE β1-integrin receptors, triggering Rho GTPase signals that strengthen tight junctions and barrier function.
- Feng H, Zhao X, Guo Q, Feng Y, Ma M, Guo W, Dong X, Deng C, Li C, Song X, et al.: Autophagy resists EMT process to maintain retinal pigment epithelium homeostasis. Int J Biol Sci 2019, 15:507–521. [PubMed: 30745838]
- 23. Huang L, Zhang C, Su L, Song Z: GSK3beta attenuates TGF-beta1 induced epithelialmesenchymal transition and metabolic alterations in ARPE-19 cells. Biochem Biophys Res Commun 2017, 486:744–751. [PubMed: 28342867]
- Ding Y, Xu H, Li L, Yuan Y, Xu Y: Megakaryocytic leukemia 1 (MKL1) mediates high glucose induced epithelial-mesenchymal transition by activating LOX transcription. Biochem Biophys Res Commun 2019, 509:633–640. [PubMed: 30553442]

- 25. Ghosh S, Shang P, Terasaki H, Stepicheva N, Hose S, Yazdankhah M, Weiss J, Sakamoto T, Bhutto IA, Xia S, et al.: A Role for betaA3/A1-Crystallin in Type 2 EMT of RPE Cells Occurring in Dry Age-Related Macular Degeneration. Invest Ophthalmol Vis Sci 2018, 59:Amd104–amd113. [PubMed: 30098172]
- 26. Sugasawa K, Deguchi J, Okami T, Yamamoto A, Omori K, Uyama M, Tashiro Y: Immunocytochemical analyses of distributions of Na, K-ATPase and GLUT1, insulin and transferrin receptors in the developing retinal pigment epithelial cells. Cell Struct Funct 1994, 19:21–28. [PubMed: 8069944]
- Takata K, Kasahara T, Kasahara M, Ezaki O, Hirano H: Ultracytochemical localization of the erythrocyte/HepG2-type glucose transporter (GLUT1) in cells of the blood-retinal barrier in the rat. Invest Ophthalmol Vis Sci 1992, 33:377–383. [PubMed: 1740368]
- Tserentsoodol N, Shin BC, Suzuki T, Takata K: Colocalization of tight junction proteins, occludin and ZO-1, and glucose transporter GLUT1 in cells of the blood-ocular barrier in the mouse eye. Histochem Cell Biol 1998, 110:543–551. [PubMed: 9860252]
- 29. Kanow MA, Giarmarco MM, Jankowski CS, Tsantilas K, Engel AL, Du J, Linton JD, Farnsworth CC, Sloat SR, Rountree A, et al.: Biochemical adaptations of the retina and retinal pigment epithelium support a metabolic ecosystem in the vertebrate eye. Elife 2017, 6.\*\* This study describes a metabolic environment in the outer retina achieved by preferential utilization and transport of nutrients by the RPE and photoreceptors. The authors propose a model where glucose is transported from the choroidal blood vessels through the RPE to the photoreceptors, where it is utilized to produce lactate. Lactate is exported to the RPE where it is used as a nutrient.
- 30. Chao JR, Knight K, Engel AL, Jankowski C, Wang Y, Manson MA, Gu H, Djukovic D, Raftery D, Hurley JB, et al.: Human retinal pigment epithelial cells prefer proline as a nutrient and transport metabolic intermediates to the retinal side. J Biol Chem 2017, 292:12895–12905. [PubMed: 28615447] \* This study systematically analyzed nutrient utilization in RPE cells by mass spectrometry. The authors found that RPE cells utilize proline at a higher rate than other nutrients and transport the resulting citrate to the apical side, where it can be imported to the neural retina.
- 31. Yam M, Engel AL, Wang Y, Zhu S, Hauer A, Zhang R, Lohner D, Huang J, Dinterman M, Zhao C, et al.: Proline mediates metabolic communication between retinal pigment epithelial cells and the retina. J Biol Chem 2019.
- 32. Johnson AA, Guziewicz KE, Lee CJ, Kalathur RC, Pulido JS, Marmorstein LY, Marmorstein AD: Bestrophin 1 and retinal disease. Prog Retin Eye Res 2017, 58:45–69. [PubMed: 28153808]
- 33. Zhang Y, Kittredge A, Ward N, Ji C, Chen S, Yang T: ATP activates bestrophin ion channels through direct interaction. Nat Commun 2018, 9:3126. [PubMed: 30087350] \*\* This study shows that ATP activates the Cl– channel Bestrophin in RPE cells by direct binding to the intracellular loop 2, next to the activation gate, without the need of ATP hydrolysis. A disease-causing mutation in Bestrophin involved the ATP-binding site and impaired ATP-activation of channel conductance.
- 34. Park M, Jung HG, Kweon HJ, Kim YE, Park JY, Hwang EM: The E3 ubiquitin ligase, NEDD4L (NEDD4–2) regulates bestrophin-1 (BEST1) by ubiquitin-dependent proteolysis. Biochem Biophys Res Commun 2019, 514:344–350. [PubMed: 31036321]
- 35. Oh SJ, Woo J, Lee YS, Cho M, Kim E, Cho NC, Park JY, Pae AN, Justin Lee C, Hwang EM: Direct interaction with 14–3–3gamma promotes surface expression of Best1 channel in astrocyte. Mol Brain 2017, 10:51. [PubMed: 29121962]
- 36. Reichhart N, Schoberl S, Keckeis S, Alfaar AS, Roubeix C, Cordes M, Crespo-Garcia S, Haeckel A, Kociok N, Fockler R, et al.: Anoctamin-4 is a bona fide Ca(2+)-dependent non-selective cation channel. Sci Rep 2019, 9:2257. [PubMed: 30783137] \*\* This report identifies Anoctamin-4 as a novel cation channel in the RPE with weak selectivity for K+ > Na+ > Li+. The cause of the cation selectivity was mapped to a single negative amino acid within the putative pore region that, when mutated, turned Anocatamin-4 into a Cl– channel.
- Nandrot EF, Anand M, Almeida D, Atabai K, Sheppard D, Finnemann SC: Essential role for MFG-E8 as ligand for alphavbeta5 integrin in diurnal retinal phagocytosis. Proc Natl Acad Sci U S A 2007, 104:12005–12010. [PubMed: 17620600]
- Finnemann SC: Focal adhesion kinase signaling promotes phagocytosis of integrin-bound photoreceptors. EMBO J 2003, 22:4143–4154. [PubMed: 12912913]

- Feng W, Yasumura D, Matthes MT, LaVail MM, Vollrath D: Mertk triggers uptake of photoreceptor outer segments during phagocytosis by cultured retinal pigment epithelial cells. J Biol Chem 2002, 277:17016–17022. [PubMed: 11861639]
- Nandrot EF, Silva KE, Scelfo C, Finnemann SC: Retinal pigment epithelial cells use a MerTKdependent mechanism to limit the phagocytic particle binding activity of alphavbeta5 integrin. Biol Cell 2012, 104:326–341. [PubMed: 22289110]
- 41. Bulloj A, Maminishkis A, Mizui M, Finnemann SC: Semaphorin4D-PlexinB1 Signaling Attenuates Photoreceptor Outer Segment Phagocytosis by Reducing Rac1 Activity of RPE Cells. Mol Neurobiol 2018, 55:4320–4332. [PubMed: 28624895] \*\* This study describes a novel role for the Semaphorin4D-PlexinB1 pair in the circadian internalization of shed photoreceptor outer segments by the RPE. Semaphorin4D was found at the photoreceptors and, when shed, it reached the apical membrane of the RPE where it bound the receptor PlexinB1, inhibiting Rac1 signaling and preventing outer segment internalization but not binding.
- 42. Storti F, Raphael G, Griesser V, Klee K, Drawnel F, Willburger C, Scholz R, Langmann T, von Eckardstein A, Fingerle J, et al.: Regulated efflux of photoreceptor outer segment-derived cholesterol by human RPE cells. Exp Eye Res 2017, 165:65–77. [PubMed: 28943268] \* This study shows that RPE cells export cholesterol via ABCA1 plasma membrane transporters bidirectionally towards the apical and basolateral sides.
- 43. Klein R, Meuer SM, Knudtson MD, Iyengar SK, Klein BE: The epidemiology of retinal reticular drusen. Am J Ophthalmol 2008, 145:317–326. [PubMed: 18045568]
- 44. Sarks SH: Council Lecture. Drusen and their relationship to senile macular degeneration. Aust J Ophthalmol 1980, 8:117–130. [PubMed: 6160841]
- Zweifel SA, Imamura Y, Spaide TC, Fujiwara T, Spaide RF: Prevalence and significance of subretinal drusenoid deposits (reticular pseudodrusen) in age-related macular degeneration. Ophthalmology 2010, 117:1775–1781. [PubMed: 20472293]
- 46. Lyssenko NN, Haider N, Picataggi A, Cipollari E, Jiao W, Phillips MC, Rader DJ, Chavali VRM: Directional ABCA1-mediated cholesterol efflux and apoB-lipoprotein secretion in the retinal pigment epithelium. J Lipid Res 2018, 59:1927–1939. [PubMed: 30076206] \* This study shows that RPE primary culture cells secreted apoB-lipoprotein apically, and exported cholesterol via ABCA1 bidirectionally. However, cholesterol loading increased efflux at the apical side.
- Nandrot EF, Kim Y, Brodie SE, Huang X, Sheppard D, Finnemann SC: Loss of synchronized retinal phagocytosis and age-related blindness in mice lacking alphavbeta5 integrin. J Exp Med 2004, 200:1539–1545. [PubMed: 15596525]
- 48. Anderson DMG, Ablonczy Z, Koutalos Y, Hanneken AM, Spraggins JM, Calcutt MW, Crouch RK, Caprioli RM, Schey KL: Bis(monoacylglycero)phosphate lipids in the retinal pigment epithelium implicate lysosomal/endosomal dysfunction in a model of Stargardt disease and human retinas. Sci Rep 2017, 7:17352. [PubMed: 29229934]
- 49. Kaur G, Tan LX, Rathnasamy G, La Cunza N, Germer CJ, Toops KA, Fernandes M, Blenkinsop TA, Lakkaraju A: Aberrant early endosome biogenesis mediates complement activation in the retinal pigment epithelium in models of macular degeneration. Proc Natl Acad Sci U S A 2018, 115:9014–9019. [PubMed: 30126999]
- 50. Storm T, Burgoyne T, Dunaief JL, Christensen EI, Futter C, Nielsen R: Selective Ablation of Megalin in the Retinal Pigment Epithelium Results in Megaophthalmos, Macromelanosome Formation and Severe Retina Degeneration. Invest Ophthalmol Vis Sci 2019, 60:322–330. [PubMed: 30665232] \*\* This report used conditional RPE-specific megalin knockout mice to study the postnatal role of megalin. Loss of megalin in the RPE resulted in abnormal fusion of melanosomes, megaophthalmos and severe retinal thinning by postnatal day 14.
- 51. Nishiyama K, Sakaguchi H, Hu JG, Bok D, Hollyfield JG: Claudin localization in cilia of the retinal pigment epithelium. Anat Rec 2002, 267:196–203. [PubMed: 12115268]
- 52. Buskin A, Zhu L, Chichagova V, Basu B, Mozaffari-Jovin S, Dolan D, Droop A, Collin J, Bronstein R, Mehrotra S, et al.: Disrupted alternative splicing for genes implicated in splicing and ciliogenesis causes PRPF31 retinitis pigmentosa. Nat Commun 2018, 9:4234. [PubMed: 30315276] \*\* This article describes mis-splicing of genes controlling ciliogenesis and cell adhesion in RPE cells from patients harboring a mutation in the pre-mRNA processing factor PRPF31 that causes retinitis pigmentosa.

- 53. May-Simera HL, Wan Q, Jha BS, Hartford J, Khristov V, Dejene R, Chang J, Patnaik S, Lu Q, Banerjee P, et al.: Primary Cilium-Mediated Retinal Pigment Epithelium Maturation Is Disrupted in Ciliopathy Patient Cells. Cell Rep 2018, 22:189–205. [PubMed: 29298421] \*\* This article shows that ciliary proteins in the RPE are important for RPE maturation and physiology. RPE cells derived from ciliopathy patients or with knockdown of a ciliary protein failed to properly mature and had a defective epithelial phenotype with reduced functionality.
- 54. Goto S, Onishi A, Misaki K, Yonemura S, Sugita S, Ito H, Ohigashi Y, Ema M, Sakaguchi H, Nishida K, et al.: Neural retina-specific Aldh1a1 controls dorsal choroidal vascular development via Sox9 expression in retinal pigment epithelial cells. Elife 2018, 7.\*\* This study describes a novel pathway of communication between the neural retina and the RPE, which results in maintenance of the choroid vasculature. The authors propose a model where retinoic acids produced in the neural retina via Aldh1a1 reach the RPE, stimulating the transcription factor Sox9, which increases VEGF expression and in turn stimulates choroidal angiogenesis.
- 55. Cohen-Tayar Y, Cohen H, Mitiagin Y, Abravanel Z, Levy C, Idelson M, Reubinoff B, Itzkovitz S, Raviv S, Kaestner KH, et al.: Pax6 regulation of Sox9 in the mouse retinal pigmented epithelium controls its timely differentiation and choroid vasculature development. Development 2018, 145.
- 56. Takei Y, Ozanics V: Origin and development of Bruch's membrane in monkey fetuses: an electron microscopic study. Invest Ophthalmol 1975, 14:903–916. [PubMed: 811582]
- Booij JC, Baas DC, Beisekeeva J, Gorgels TG, Bergen AA: The dynamic nature of Bruch's membrane. Prog Retin Eye Res 2010, 29:1–18. [PubMed: 19747980]
- 58. Li R, Maminishkis A, Zahn G, Vossmeyer D, Miller SS: Integrin alpha5beta1 mediates attachment, migration, and proliferation in human retinal pigment epithelium: relevance for proliferative retinal disease. Invest Ophthalmol Vis Sci 2009, 50:5988–5996. [PubMed: 19608542]
- 59. Sorkio A, Hongisto H, Kaarniranta K, Uusitalo H, Juuti-Uusitalo K, Skottman H: Structure and barrier properties of human embryonic stem cell-derived retinal pigment epithelial cells are affected by extracellular matrix protein coating. Tissue Eng Part A 2014, 20:622–634. [PubMed: 24044751]
- 60. Blaauwgeers HG, Holtkamp GM, Rutten H, Witmer AN, Koolwijk P, Partanen TA, Alitalo K, Kroon ME, Kijlstra A, van Hinsbergh VW, et al.: Polarized vascular endothelial growth factor secretion by human retinal pigment epithelium and localization of vascular endothelial growth factor receptors on the inner choriocapillaris. Evidence for a trophic paracrine relation. Am J Pathol 1999, 155:421–428. [PubMed: 10433935]
- 61. Pauleikhoff D, Harper CA, Marshall J, Bird AC: Aging changes in Bruch's membrane. A histochemical and morphologic study. Ophthalmology 1990, 97:171–178. [PubMed: 1691475]
- 62. Spraul CW, Lang GE, Grossniklaus HE, Lang GK: Histologic and morphometric analysis of the choroid, Bruch's membrane, and retinal pigment epithelium in postmortem eyes with age-related macular degeneration and histologic examination of surgically excised choroidal neovascular membranes. Surv Ophthalmol 1999, 44 Suppl 1:S10–32. [PubMed: 10548114]
- 63. Cai H, Gong J, Del Priore LV, Tezel TH, Fields MA: Culturing of Retinal Pigment Epithelial Cells on an Ex Vivo Model of Aged Human Bruch's Membrane. J Vis Exp 2018.
- Ho TC, Del Priore LV: Reattachment of cultured human retinal pigment epithelium to extracellular matrix and human Bruch's membrane. Invest Ophthalmol Vis Sci 1997, 38:1110–1118. [PubMed: 9152230]
- 65. Heller JP, Martin KR: Enhancing RPE Cell-Based Therapy Outcomes for AMD: The Role of Bruch's Membrane. Transl Vis Sci Technol 2014, 3:11.
- 66. Marmorstein LY, Munier FL, Arsenijevic Y, Schorderet DF, McLaughlin PJ, Chung D, Traboulsi E, Marmorstein AD: Aberrant accumulation of EFEMP1 underlies drusen formation in Malattia Leventinese and age-related macular degeneration. Proc Natl Acad Sci U S A 2002, 99:13067–13072. [PubMed: 12242346]
- Rajapakse D, Peterson K, Mishra S, Wistow G: Serum starvation of ARPE-19 changes the cellular distribution of cholesterol and Fibulin3 in patterns reminiscent of age-related macular degeneration. Exp Cell Res 2017, 361:333–341. [PubMed: 29097185]
- 68. Zayas-Santiago A, Cross SD, Stanton JB, Marmorstein AD, Marmorstein LY: Mutant Fibulin-3 Causes Proteoglycan Accumulation and Impaired Diffusion Across Bruch's Membrane. Invest Ophthalmol Vis Sci 2017, 58:3046–3054. [PubMed: 28622396] \*\* This study describes the role of

Fibulin-3 in RPE-mediated remodeling of the Bruch's membrane and its diffusion properties. The authors studied macular degeneration-causing mutation in Fibulin-3 and Bruch's membrane from Fibulin-3 knockout mice. They found alterations in proteoglycan composition, activity of metalloproteases and diffusion across the Bruch's membrane.

- 69. Greene WA, Burke TA, Kaini RR, Por ED, Wang HC: Polarized Secretion of Matrix Metalloproteinases and Their Inhibitors by Retinal Pigment Epithelium Derived from Induced Pluripotent Stem Cells During Wound Healing. J Ocul Pharmacol Ther 2017, 33:132–140. [PubMed: 28384031]
- Fernandez-Godino R: Alterations in Extracellular Matrix/Bruch's Membrane Can Cause the Activation of the Alternative Complement Pathway via Tick-Over. Adv Exp Med Biol 2018, 1074:29–35. [PubMed: 29721924]
- Fernandez-Godino R, Bujakowska KM, Pierce EA: Changes in extracellular matrix cause RPE cells to make basal deposits and activate the alternative complement pathway. Hum Mol Genet 2018, 27:147–159. [PubMed: 29095988]
- 72. Fields MA, Bowrey HE, Gong J, Moreira EF, Cai H, Del Priore LV: Extracellular matrix nitration alters growth factor release and activates bioactive complement in human retinal pigment epithelial cells. PLoS One 2017, 12:e0177763. [PubMed: 28505174]
- 73. Sharif U, Mahmud NM, Kay P, Yang YC, Harding SP, Grierson I, Kamalden TA, Jackson MJ, Paraoan L: Advanced glycation end products-related modulation of cathepsin L and NF-kappaB signalling effectors in retinal pigment epithelium lead to augmented response to TNFalpha. J Cell Mol Med 2019, 23:405–416. [PubMed: 30338926]
- 74. Thao MT, Karumanchi DK, Yacout SM, Gaillard ER: Nitrite ion modifies tyrosine and lysine residues of extracellular matrix proteins. Nitric Oxide 2018, 79:51–56. [PubMed: 30055286]
- 75. Gandhi JK, Knudsen T, Hill M, Roy B, Bachman L, Pfannkoch-Andrews C, Schmidt KN, Metko MM, Ackerman MJ, Resch Z, et al.: Human Fibrinogen for Maintenance and Differentiation of Induced Pluripotent Stem Cells in Two Dimensions and Three Dimensions. Stem Cells Transl Med 2019.
- McLenachan S, Hao E, Zhang D, Zhang L, Edel M, Chen F: Bioengineered Bruch's-like extracellular matrix promotes retinal pigment epithelial differentiation. Biochem Biophys Rep 2017, 10:178–185. [PubMed: 28955745]
- 77. Spencer C, Abend S, McHugh KJ, Saint-Geniez M: Identification of a synergistic interaction between endothelial cells and retinal pigment epithelium. J Cell Mol Med 2017, 21:2542–2552. [PubMed: 28402065]
- 78. Nolan DJ, Ginsberg M, Israely E, Palikuqi B, Poulos MG, James D, Ding BS, Schachterle W, Liu Y, Rosenwaks Z, et al.: Molecular signatures of tissue-specific microvascular endothelial cell heterogeneity in organ maintenance and regeneration. Dev Cell 2013, 26:204–219. [PubMed: 23871589]
- Rafii S, Butler JM, Ding BS: Angiocrine functions of organ-specific endothelial cells. Nature 2016, 529:316–325. [PubMed: 26791722]
- Kurihara T, Westenskow PD, Bravo S, Aguilar E, Friedlander M: Targeted deletion of Vegfa in adult mice induces vision loss. J Clin Invest 2012, 122:4213–4217. [PubMed: 23093773]
- Sonoda S, Sreekumar PG, Kase S, Spee C, Ryan SJ, Kannan R, Hinton DR: Attainment of polarity promotes growth factor secretion by retinal pigment epithelial cells: relevance to age-related macular degeneration. Aging 2009, 2:28–42. [PubMed: 20228934]
- Keir LS, Firth R, Aponik L, Feitelberg D, Sakimoto S, Aguilar E, Welsh GI, Richards A, Usui Y, Satchell SC, et al.: VEGF regulates local inhibitory complement proteins in the eye and kidney. J Clin Invest 2017, 127:199–214. [PubMed: 27918307]
- 83. Wu D, Kanda A, Liu Y, Kase S, Noda K, Ishida S: Galectin-1 promotes choroidal neovascularization and subretinal fibrosis mediated via epithelial-mesenchymal transition. FASEB J 2019, 33:2498–2513. [PubMed: 30277820] \*\* This study addresses the role of Galectin-1 in choroidal neovascularization. Galectin-1 was found in RPE cells and associated with choroidal VEGF receptor type 2. Genetic deletion of Galectin-1 protected mice from laser-induced choroidal neovascularization, associated with a decrease in markers of epithelial-mesenchymal transition.

- 84. Atienzar-Aroca S, Serrano-Heras G, Freire Valls A, Ruiz de Almodovar C, Muriach M, Barcia JM, Garcia-Verdugo JM, Romero FJ, Sancho-Pelluz J: Role of retinal pigment epithelium-derived exosomes and autophagy in new blood vessel formation. J Cell Mol Med 2018, 22:5244–5256. [PubMed: 30133118] \* This study establishes a pro-angiogenic role for exosomes secreted by ARPE-19 cells in culture subject to oxidative stress and with a functional autophagy pathway. The exosome-mediated angiogenic response, measured in HUVEC endothelial cell line, required expression of VEGF receptor but it was not affected by a VEGF-blocking agent.
- Shao Z, Friedlander M, Hurst CG, Cui Z, Pei DT, Evans LP, Juan AM, Tahiri H, Duhamel F, Chen J, et al.: Choroid sprouting assay: an ex vivo model of microvascular angiogenesis. PLoS One 2013, 8:e69552. [PubMed: 23922736]
- 86. Caceres PS, Gravotta D, Zager PJ, Dephoure N, Rodriguez-Boulan E: Quantitative proteomics of MDCK cells identify unrecognized roles of clathrin adaptor AP-1 in polarized distribution of surface proteins. Proceedings of the National Academy of Sciences 2019, 116:11796–11805.\* This work introduces a surface proteomics approach to study polarity of epithelial cells in culture at a large scale. The approach was used to describe novel roles of a clathrin adaptor in polarized distribution of surface proteins.
- Khristov V, Wan Q, Sharma R, Lotfi M, Maminishkis A, Bharti K: Polarized Human Retinal Pigment Epithelium Exhibits Distinct Surface Proteome on Apical and Basal Plasma Membranes. Methods Mol Biol 2018, 1722:223–247. [PubMed: 29264809]

Caceres and Rodriguez-Boulan



#### Figure 1: Retinal pigment epithelium in relation to photoreceptors and choroid.

A) Localization of the retinal pigment epithelium in the eye at the interface between the neural retina and choroid. **B**) Polarized trafficking of secreted factors, intracellular proteins, organelles and membrane receptors and transporters dictates the identity of the apical (green), basolateral (red) plasma membrane domains and tight junctions. Apical trafficking determines interactions with photoreceptors at the neural retina. Basolateral trafficking mediates interactions with the Bruch's membrane and choroidal vasculature. Green and red arrows represent pathways acting at the apical and basolateral domains respectively. Blue arrows represent cross-talk pathways originating at the neural retina or choroid. The polarized organization of the retinal pigment epithelium permits the formation of the outer retina-blood barrier, also composed by the Bruch's membrane and the choroidal vasculature.



# **Barrier function**

Figure 2: Key interactions at the apical domain of the retinal pigment epithelium.

The retinal pigment epithelium provides many photoreceptor-support functions (indicated in red font) which require interactions through the apical domain. Only the processes discussed in this review that contribute to these functions are represented here in detail. Dotted lines represent pathways that may contain intermediate steps. Abbreviations used: RPE, retinal pigment epithelium; MFG-E8, milk fat globule–EGF 8; ABCA1, ATP binding cassette subfamily A member 1; GLUT1, Glucose Transporter Type-1; MCT1, Monocarboxylate Transporter-1; AQP1, aquaporin-1; VEGF, Vascular Endothelial Growth Factor; Pax6, Paired Box 6; Sox9, SRY (Sex-Determining Region Y)-Box 9, CEP290, Centrosomal Protein 290; Bbs8, Bardet-Biedl Syndrome 8.

Caceres and Rodriguez-Boulan



# Figure 3: Basolateral retinal pigment epithelium interactions with the Bruch's membrane and choriocapillaris.

Basolateral targeting of membrane receptors, secreted factors and extracellular matrix components dictates the key interactions at the outer retina-blood barrier discussed in this review and represented here. The pathway represented by the blue arrows indicates a hypothetical mechanism of direct communication between the choroid endothelium and the retinal pigment epithelium involving unidentified (?) secreted endothelial factors and epithelial receptors. Dotted lines represent pathways that may involve intermediate steps. Abbreviations used: RPE, retinal pigment epithelium; VEGF, Vascular Endothelial Growth Factor; Gal1, Galectin-1; MMP, matrix metalloproteinase; TIMP-3, Tissue Inhibitor of Metalloproteinases 3; RhoA, Ras Homolog Family Member A; ROCK, Rho Associated Coiled-Coil Containing Protein Kinase; Rac1, Rac family small GTPase 1; LOX, Lysyl oxidases;  $\alpha_{(n)}\beta1$  represents integrin receptors formed by association of a  $\beta1$  subunit and an  $\alpha$  subunit.