World Journal of WÍ Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2020 February 21; 26(7): 696-705

DOI: 10.3748/wjg.v26.i7.696

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

Proteomic insights on the metabolism in inflammatory bowel disease

Laura Francesca Pisani, Manuela Moriggi, Cecilia Gelfi, Maurizio Vecchi, Luca Pastorelli

ORCID number: Laura Francesca Pisani (0000-0002-9490-3723); Manuela Moriggi (0000-0002-4718-0307); Cecilia Gelfi (0000-0002-2996-6912); Maurizio Vecchi (0000-0003-1558-8604); Luca Pastorelli (0000-0002-2810-9951).

Author contributions: Pisani LF performed the majority of the writing; Moriggi M prepared the figure and wrote the technical proteomic paragraphs; Vecchi M provided the input in writing the review; Gelfi C revised the review and gave her support as proteomics expert; and Pastorelli L revised the review and gave his support as clinical expert.

Supported by Italy's Ministero Italiano della Salute (Italian Ministry of Health Grant), No. GR-2016-02364736.

Conflict-of-interest statement:

There is no conflict of interest associated with any of the senior author or other coauthors contributed their efforts in this manuscript.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative **Commons Attribution** NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licen Laura Francesca Pisani, Manuela Moriggi, Luca Pastorelli, Gastroenterology and Digestive Endoscopy Unit, IRCCS Policlinico San Donato, San Donato Milanese 20097, Italy

Cecilia Gelfi, Department of Biomedical Science for Health, University of the Study of Milan, IRCCS Istituto Ortopedico Galeazzi, Milan 20122, Italy

Maurizio Vecchi, Gastroenterology and Endoscopy Unit, IRCCS Ca' Granda Foundation, Policlinico Hospital, University of the Study of Milan, Milan 20122, Italy

Luca Pastorelli, Department of Biomedical Science for Health, University of the Study of Milan, Milan 20122, Italy

Corresponding author: Luca Pastorelli, DPhil, MD, Assistant Professor, Doctor, Gastroenterology and Digestive Endoscopy Unit, IRCCS Policlinico San Donato, Piazza Malan, San Donato Milanese 20097, Italy. luca.pastorelli@unimi.it

Abstract

Inflammatory bowel diseases (IBD) are chronic and relapsing inflammatory conditions of the gut that include Crohn's disease and ulcerative colitis. The pathogenesis of IBD is not completely unraveled, IBD are multi-factorial diseases with reported alterations in the gut microbiota, activation of different immune cell types, changes in the vascular endothelium, and alterations in the tight junctions' structure of the colonic epithelial cells. Proteomics represents a useful tool to enhance our biological understanding and to discover biomarkers in blood and intestinal specimens. It is expected to provide reproducible and quantitative data that can support clinical assessments and help clinicians in the diagnosis and treatment of IBD. Sometimes a differential diagnosis of Crohn's disease and ulcerative colitis and the prediction of treatment response can be deducted by finding meaningful biomarkers. Although some non-invasive biomarkers have been described, none can be considered as the "gold standard" for IBD diagnosis, disease activity and therapy outcome. For these reason new studies have proposed an "IBD signature", which consists in a panel of biomarkers used to assess IBD. The above described approach characterizes "omics" and in this review we will focus on proteomics.

Key words: Proteomics; Inflammatory bowel disease; Cronh's disease; Ulcerative colitis; Proteins; Biomarkers discovery

©The Author(s) 2020. Published by Baishideng Publishing Group Inc. All rights reserved.



Baiahidena® WJG | https://www.wjgnet.com

ses/by-nc/4.0/

Manuscript source: Invited manuscript

Received: November 25, 2019 Peer-review started: November 25, 2019

First decision: December 23, 2019 Revised: January 2, 2020 Accepted: February 10, 2020 Article in press: February 10, 2020 Published online: February 21, 2020

P-Reviewer: Poullis A, Can G S-Editor: Dou Y L-Editor: A E-Editor: Ma YJ



Core tip: Patients' heterogeneity is a hallmark for inflammatory bowel diseases (IBD). Some patients present limited bowel involvement and a mild course of the disease, others develop very extensive, aggressive disease and variable response to therapy. In IBD, there is a great need of patient stratification and of new biomarkers as part of a personalized medicine approach to patient care. Biological therapies are more and more widely used for IBD patients, because of their efficacy in patient's refractory to other drugs; still, biological treatments fail in 20%-40% of patients and, to date, no reliable clinical or molecular predictor of response to biological therapeutic strategy has been described. This review aims to collect the "omics" approach for research of serological biomarkers of diagnosis, response to specific biological therapies in the IBD field.

Citation: Pisani LF, Moriggi M, Gelfi C, Vecchi M, Pastorelli L. Proteomic insights on the metabolism in inflammatory bowel disease. *World J Gastroenterol* 2020; 26(7): 696-705 **URL**: https://www.wjgnet.com/1007-9327/full/v26/i7/696.htm **DOI**: https://dx.doi.org/10.3748/wjg.v26.i7.696

INFLAMMATORY BOWEL DISEASE

Ulcerative colitis (UC) and Crohn's disease (CD) are the two main inflammatory bowel diseases (IBD)^[1-4]. Despite some shared characteristics, they can be distinguished by differences in genetic predisposition, risk factors, and clinical, endoscopic and histological features. CD is characterized by diffuse chronic inflammation throughout the gastrointestinal tract, in a non-continuous manner^[5]; UC presents with inflammation limited to the colon, spreading continuously from the rectum^[6]. The pathogenesis of IBD is at present not completely unraveled; however, genetically susceptible individuals seem to have a dysregulated mucosal immune response to the commensal gut flora, which results in bowel inflammation^[7]. IBD are multi-factorial diseases^[8] with reported alterations in the gut microbiota^[9-12], activation of different immune cell types^[13-15], changes in the vascular endothelium^[16,17], and alterations in the tight junctions structure of colon epithelial cells^[18-20].

Nowadays, the diagnostic and prognostic tools for IBD and the outcome of therapy are largely based on evaluation of clinical symptoms in combination with endoscopy, histology, radiology and non-specific biomarkers from serum or stools^[21].

BIOMARKERS IN INFLAMMATORY BOWEL DISEASE

Inflammation in IBD is characterized by the increased levels of some molecules extensively validated but not all included in the laboratory routine. Some of them are related to the inflammatory acute-phase response, coagulation and fibrinolysis (fibrinogen, plasminogen, complement components), proteinase inhibitors (a1antitrypsin and a1-anti-chymotrypsin), transport proteins (haptoglobin and ceruplasmin) and other serum proteins^[22] and cytokines^[23]. Elevated platelet and white blood cell counts may also indicate inflammation but they cannot be considered strictly related to bowel inflammation^[23]. C-reactive protein (CRP), anti-Saccaromyces cerevisiae (ASCA) and anti-neutrophil cytoplasmic antibody are the most widely used indicators. CRP has a short reaction time (6-10 h) and it is useful for the identification of inflammatory disease activity especially in CD, but not in UC^[24]. CRP has low specificity enabling to differentiate between CD, UC and infectious colitis^[21], and also the 25% of IBD patients with demonstrable disease activity have CRP levels above the normal threshold^[22]. ASCA is an antibody used for the identification of CD patients who are often positive (39%-79% of CD patients, 5%-15% UC patients)^[25,26], however a large part of healthy controls is also positive (14%-18%) to this antibody, limiting the diagnostic value of its detection^[27]. anti-neutrophil cytoplasmic antibodies are antibodies found in immune-mediated pathologies, such as rheumatoid arthritis and Wegener's granulomatosis^[28], and have shown a different staining pattern in UC and CD patients^[29-31], but as for ASCA 32% of healthy population is also positive to them^[32].

Another explored field in the search for IBD biomarkers is the analysis of stool proteins, which can be dysregulated or abnormally present in patients. Stool markers have the advantage of increased specificity for bowel inflammation and reflect any mucosal barrier disruption. Fecal markers can be useful to diagnose CD, where

¹⁵ WJG https://www.wjgnet.com

inflammation is patchy and is possibly missed at endoscopy^[33]. Fecal calprotectin (FC) accounts for up to 5% of the neutrophil granulocytes' protein content with chemotactic and antimicrobial activities. It is stable in stool for more than a week and can resists to bacterial degradation^[34]. FC is not a specific marker for IBD, but it correlates with increased disease activity at least in adults^[35], but not in pediatric patients where was found with high sensitivity (98%), but only modest specificity (68%)^[36]. Disease location should also be taken into account when interpreting FC levels. Patients with ileal CD may have ulcers even in the absence of markedly elevated FC levels. Consequently, the cut-off values for ileal CD may differ from those with ileocolic disease^[37,38]. A study conducted by De Vos et al^[39] has demonstrated that Calprotectin decreased 2 wk after Infliximab administration predicts remission in anti-TNF-naïve patients with UC. The increase of FC can also be a suitable marker for the identification of relapse, given the fact that the levels are increased as early as 6 mo before clinical and endoscopic relapse^[40]. Lactoferrin is an iron-binding protein expressed by neutrophils during inflammation and represents a defense against infection as part of the innate immune system^[41,42]. As a biomarker, Lactoferrin can distinguish IBD from Irritable Bowel Syndrome, but not between CD and UC^[27].

Although many non-invasive biomarkers have been described, none can be considered as the "gold standard" for IBD diagnosis, disease activity and therapy outcome. A single ideal biomarker is very unlikely to be found. As for other pathologies as pancreatic cancer^[43-46], non-small cell lung cancer^[47] and colorectal cancer^[48] new studies have proposed the idea of a "Biomarker Signature", which consists in a panel of biomarkers used to assess various pathological conditions and response to therapy^[49], and which is applicable also to IBD diagnosis and prognosis. Table 1 summarizes the biomarkers commonly used for IBD.

PROTEOMIC APPROACH TO INFLAMMATORY BOWEL DISEASE RESEARCH

Proteomics comprehensively studies the protein composition and abundance in a given cell population and its changes under biological perturbations^[50,51]. The proteome may be considered the signature of a disease, in fact it is the result of the interactions between the genetic background and environmental factors^[40]. The novel proteomic technologies now facilitate the analysis of transcriptome variations also in the IBD context and have already provided with new candidate biomarkers^[52]. They help to investigate the inflammatory response, epithelial barrier function and gut microbiome from different biological samples, *i.e.*, serum/blood, colon samples and feces. The proteomic strategies can be bottom-up and top-down (Figure 1). In the bottom-up approach, purified proteins or complex protein mixtures are subjected to proteolytic cleavage and the peptide products are analyzed by mass spectrometry (MS). Conversely, the top-down approach is based either on the analysis of intact proteins followed by the direct measurement of fragment ions by MS or on the isolation of the protein by gel-based separative methods, protein gel elution and MS analysis.

Proteomics in the study of IBD pathogenesis

By LC-MS analysis of colon mucosal biopsies from 10 patients with UC, Bennike *et* $al^{[53]}$ identified 5711 quantifiable proteins classified by biological function, sub-cellular location and molecular function. Forty-six proteins demonstrated statistically significant changes in mean abundance between UC biopsies and control biopsies; among those proteins, the one with the largest mean fold abundance change was lactotransferrin, which was 219 times more abundant in the UC group. The relative abundance of lactotransferrin also correlated to the severity of tissue inflammation in the patients with UC, as determined by the colon inflammation grade score based on histology. Good correlation was found between the colon inflammation grade score and the relative abundance of lactotransferrin in the tissue (0.82)^[53]. Eleven of the 46 proteins identified in the UC biopsies are present in neutrophils and are associated with the formation of neutrophil extra-cellular traps which are released from neutrophils in response to inflammatory stimuli^[54,56,57].

Proteomics has also investigated IBD-related immune-cell responses. Riaz *et al*^[58] compared Th1 and Th17 clones isolated from the intestinal mucosa of CD patients by means of label-free quantitative mass-spectrometry analysis, which led to the identification of a total number of 7401 unique protein groups and demonstrated that 334 proteins were differentially expressed. The largest differences between the two phenotypes were observed in such proteins with cytotoxic function as Granzyme B

[®] WJG https://www.wjgnet.com

Table 1 Biomarkers in inflammatory bowel disease						
Marker	Setting	Diagnostic accuracy	Ref.			
C-Reactive Protein (CRP)	Serum	Higher in CD vs UC	Henriksen <i>et al</i> ^[24] , 2008			
		25% IBD patients have levels above normal	Vermeire et al ^[22] , 2004			
Anti-Saccharomyces cerevisiae Antibodies (ASCA)	Serum	39%-79% CD positive	Peyrin-Biroulet et al ^[25] , 2015;			
		5%-15% UC positive	Reumaux <i>et al</i> ^[26] , 2004			
		14%-18% HC positive	Bennike <i>et al</i> ^[27] , 2014			
Anti-neutrophil cytoplasmic antibodies (ANCA)	Serum	Different pattern in CD and UC	Peeters <i>et al</i> ^[31] , 2001;			
			Peyrin-Biroulet <i>et al</i> ^[30] , 2007;			
			Reumaux <i>et al</i> ^[29] , 2003			
		32% HC positive	Bernstein <i>et al</i> ^[32] , 2011			
Calprotectin	Colorectal mucus	Higher in IBD vs HC	Loktionov <i>et al</i> ^[79] , 2016			
		Higher in UC vs CD				
Calgranulin C (S100A12)		Higher in UC vs CD				
Eosinophil-derived neurotoxin (EDN)		Higher in IBD vs HC				
		Higher in UC vs CD				
Fecal calprotectin (FC)	Stool	It correlates with disease activity in adults	Gisbert <i>et al</i> ^[35] , 2009			
Lactoferrin	Stool	It distinguishes IBD from IBS	Bennike <i>et al</i> ^[27] , 2014			

CD: Crohn's disease; UC: Ulcerative colitis; HC: Healthy controls; IBS: Irritable bowel syndrome; IBD: Inflammatory bowel disease.

and perforin, which are lower in Th17 cells than in Th1 cells. Other differentially expressed proteins with higher expression in the Th1 clones included several transcription factors with both known and unknown functions in CD4⁺ T-cells. The most striking differences at quantitative analysis are about CD4⁺ T cells with Th1 phenotype having a much higher degree of cytotoxic features as compared with Th1/Th17 phenotype^[58].

As discussed above, the disruption of the intestinal barrier is a typical event in IBD pathogenesis. The intestinal epithelium is the largest surface exposed and coming into contact with the external environment. The intestinal epithelial cells (IECs) are the main component of the physical barrier between the luminal micro-environment and the host and act as the host's first line-of-defense against potential harmful stimulants. They also represent the innate immunity within the gut mucosa^[59]. Normally, the intestinal epithelium is covered by a single layer of IECs, which are characterized by a fast renewal rate, and act as a protective barrier against luminal antigens, but this barrier can be damaged, thus promoting a state of chronic inflammation due to mucosal immune cell infiltration, as is typically observed in IBD patients^[59]. The molecular changes in the epithelial layer, extra-cellular matrix and junction proteins in inflamed and non-inflamed intestinal tissue have been only partially addressed to date. In 2012 Poulsen et al[60] analyzed the proteomic profiles of whole colonic biopsies from UC patients using 2D-gel electrophoresis and MALDI-TOF MS for the identification of differently expressed protein spots. Forty-three proteins were identified differentially expressed between UC inflamed and non-inflamed tissue, including proteins involved in the energy metabolism and in oxidative stress^[60]. Proteomic studies on isolated IECs obtained from surgical specimens of full-thickness colonic tissues from UC-, CD-affected patients and non-inflamed controls were analyzed by gel-based stable-isotope label technologies (2D-DIGE and ICPL LC-MS/MS) and immunoblot assay to evaluate any proteome changes. Moreover, the results were verified on a group of patients not participating in the discovery phase^[61]. The differential proteomic approaches have revealed changes in several molecules involved in extracellular matrix, mechano-transduction, metabolic rewiring and autophagy that characterize quiescent UC and quiescent CD epithelial cells and they may help understanding the complex mechanisms associated to IBD. UC patients are characterized by cytoskeletal rearrangement and increased level of specific enzymes that contribute to cell homeostasis, enabling cells to cope with energy requirements and macro-autophagy. CD patients are characterized by metabolic rewiring to sustain the cell metabolism, whereas autophagy and cell renewal are blunted^[61-63]. Table 2 provides a summary of the proteins and pathways identified by the proteomic approach as involved in IBD pathogenesis.

WJG https://www.wjgnet.com





Figure 1 Schematic illustration of the difference between protein-based top-down and peptide-based bottom-up proteomics.

Proteomics for the identification of novel biomarkers

Another approach is the identification of biomarkers useful for the diagnosis, treatment selection and response monitoring. A recent study focused on diagnosis has identified a serological panel which demonstrates transmural intestinal injury and is able to indicate complications in CD patients with 70% sensitivity and 72.5% specificity^[64]. The increase of circulating epithelial component proteins may be a sign of transmural intestinal injury and stricturing or fistulizing intestinal complications. The serum biomarkers for the stratification of IBD patients are unable to distinguish between CD and UC^[65], while the proteomic profiles of colon biopsies can identify a more precise signature of these diseases^[61,66,67]. In 2016 Starr *et al*^[68] established two candidate biomarker panels: A 5-protein panel to discriminate IBD from control patients and a 12-protein panel to distinguish CD from UC patients in children with a new IBD diagnosis.

Proteomics has been applied to the identification of treatment-response biomarkers. The anti-TNF drug called Infliximab is one of the most used drugs in IBD, but the factors predicting the response and the molecular mechanisms that are related to the loss of response or non-responsiveness are not completely known. Meuwis *et al*^[69] have analyzed sera from responder and non-responder CD patients at baseline and then comparing sera throughout the induction period (week 4 for non-fistulizing and week 10 for fistulizing patients) and have shown that the platelet aggregation Factor 4 (PF4) was higher in non-responders than responders to Infliximab therapy (both before and after treatment). PF4 is considered as an acute-phase reactant because its level increases with general inflammation, as already observed in the plasma of CD

[®] WJG https://www.wjgnet.com

Table 2 Proteomics in inflammatory bowel disease pathogenesis							
Protein	Setting	Diagnostic accuracy	Ref.				
Lactotransferrin	UC vs HC biopsies	It correlates to the colon inflammation grade score	Bennike <i>et al</i> ^[53] , 2015				
Neutrophil extracellular traps (NETs)		Sign of chronic inflammation					
Granzyme B and Perforin	CD Th1 and Th17 clones from	Higher in Th1 vs Th17	Riaz <i>et al</i> ^[58] , 2016				
RORC and FOXP3	intestinal mucosa						
Glycerol-3- phosphatedehydrogenase	UC biopsies inflamed vs non- inflamed	Higher in inflamed <i>vs</i> non-inflamed tissue	Poulsen <i>et al</i> ^[60] , 2012				
Alphaenolase		Lower in inflamed <i>vs</i> non- inflamed tissue					
Keratins 10, 14, 19	UC intestinal epithelial cells	Higher in QUC vs HC	Moriggi <i>et al</i> ^[61] , 2017				
Keratin 8		Lower in QUC vs HC					
Tricarboxylic acid cycle enzymes							
Oxidative phosphorylation enzymes	3						
Vinculin and α-tubulin							
Keratin 8, 18	CD intestinal epithelial cells	Lower in QCD vs HC					
Heat shock cognate-70 (HSC70)							
Vinculin and α-tubulin		Higher in QCD vs HC					
Fibrinopeptide A (FPA)	CD serum	Higher in CD vs HC	Nanni <i>et al</i> ^[62] , 2009				
Complement 3 protein (C3)							
Apolipoprotein A-IV							
Apolipoprotein E		Lower in CD vs HC					
L-lactate dehydrogenase	IBD and HC intestinal epithelial cells	Higher in IBD vs HC; Higher in CD	Shkoda <i>et al</i> ^[63] , 2007				
Carbonyl reductase		vs UC					
Keratin 19							
Rho-GDI dissociation inhibitor $\boldsymbol{\alpha}$							
Annexin 2	UC intestinal epithelial cells	Higher in UC vs HC					
Programmed cell death protein 8 (PDCD8)							

IBD: Inflammatory bowel disease; CD: Crohn's disease; UC: Ulcerative colitis; QCD: Quiescent Crohn's disease; QUC: Quiescent ulcerative colitis; HC: Healthy controls; CRC: Colorectal carcinoma.

patients^[70-72]. Gazouli et al^[73] have compared sera before treatment and after IFX induction (week 12) and successfully identified 15 proteins that were differentially accumulated in the sera, most of them modifying the activation of monocytes /macrophages and directly and indirectly regulating the differentiation and activation of CD4+ T-lymphocytes. Also, a recent study by Magnusson et al^[74] reported on the proteomic analysis on biopsies obtained from 6 UC patients (3 responders and 3 non-responders) treated in vitro with or without Infliximab and also from 43 UC patients' sera at different time points: Baseline, week 2 and week 14. Those authors have shown that the response in UC patients is associated with reduced monocyte activation 2 wk after therapy initiation, suggesting that the monocytes of these patients are less responsive to inflammatory stimuli when reaching the intestinal mucosa. In therapy responders Infliximab has had influence on Tenascin C, which might be a down-regulator of the two chemokines CCL2 (mcp-1) and CXCL10 (IP-10)^[74], which are produced by inflammatory cells and stromal cells, recruit leucocytes, and are induced in inflamed UC mucosa^[75-77]. Table 3 summarized the potential biomarkers identified by proteomics in IBD.

CONCLUSION

In the IBD micro-environment a multitude of components interact. No information about a single gene, a single molecule or microbe can exhaustively explain the events that result from such a complex signaling. Also, the wide range of variability between patients' disease features and medical histories makes it difficult to understand how every component of IBD acts and influences other components. On the other hand,

Zaishideng® WJG | https://www.wjgnet.com

Table 3 Proteomics in inflammato	ole 3 Proteomics in inflammatory bowel disease diagnosis and response to therapy						
Proteins	Setting	Diagnostic accuracy	Ref.				
Platelet aggregation factor 4 (PF4)	Responder vs non-responder's CD serum	Higher in non-responders	Mewuis <i>et al</i> ^[69] , 2008				
Proteins that regulate CD4 ⁺ T-cell activation	Serum before IFX treatment <i>vs</i> serum after IFX induction period	Higher before treatment	Gazouli <i>et a</i> [^{73]} , 2013				
Proteins that regulate monocytes/macrophages activation							
Tenascin C	Responder <i>vs</i> non-responder's UC serum	Higher in non-responders	Magnusson <i>et al</i> ^[74] , 2015				

CD: Crohn's disease; UC: Ulcerative colitis.

even if the diagnostic gold standard is endoscopy, the introduction of novel molecular biomarkers in clinical practice has always nurtured hopes for new tools that can lead to improvements in diagnostic accuracy. However, the low diagnostic performance of the available markers strongly limits their use in clinical practice. Still, it is reasonable to hypothesize that combining the modification of several biomarkers may identify a sort of fingerprint for IBD with specific disease features.

Indeed, techniques and methodologies that can deal with a very large volume of data and describe a wide picture, rather than focus on single alteration, are likely to represent the necessary step forward in describing and comprehending IBD^[78]. For all these reasons omics can support the discovery of novel molecular interactions through a better definition of relevant biological pathways and interactions, rather than the analysis of the role of the perturbation of a single element. Omics can lead to the identification of representative patterns of disease which may replace simple biomarkers in clinical practice for the diagnosis, monitoring of IBD and for the personalization of therapies and treatments. Exploiting omic techniques and mastering big data analysis will help researchers to embrace the complexity and overcome the limitations of deciphering inflammatory disorders away from any restricted point of view. Table 3 provides a summary of the potential biomarkers identified by proteomics in IBD.

REFERENCES

- Odze R. Diagnostic problems and advances in inflammatory bowel disease. *Mod Pathol* 2003; 16: 347-358 [PMID: 12692200 DOI: 10.1097/01.MP.0000064746.82024.D1]
- 2 Bosani M, Ardizzone S, Porro GB. Biologic targeting in the treatment of inflammatory bowel diseases. Biologics 2009; 3: 77-97 [PMID: 19707398]
- 3 Ullman T, Lazarev M. Scope early and often in ulcerative colitis and Crohn's colitis? *Gastroenterology* 2009; **136**: 718-9; discussion 719-20 [PMID: 19105962 DOI: 10.1053/j.gastro.2008.12.032]
- 4 Neurath MF. Cytokines in inflammatory bowel disease. Nat Rev Immunol 2014; 14: 329-342 [PMID: 24751956 DOI: 10.1038/nri3661]
- 5 Baumgart DC, Sandborn WJ. Crohn's disease. Lancet 2012; 380: 1590-1605 [PMID: 22914295 DOI: 10.1016/S0140-6736(12)60026-9]
- 6 Ordás I, Eckmann L, Talamini M, Baumgart DC, Sandborn WJ. Ulcerative colitis. *Lancet* 2012; 380: 1606-1619 [PMID: 22914296 DOI: 10.1016/S0140-6736(12)60150-0]
- 7 Abraham C, Cho JH. Inflammatory bowel disease. N Engl J Med 2009; 361: 2066-2078 [PMID: 19923578 DOI: 10.1056/NEJMra0804647]
- 8 Fiocchi C. Genes and 'in-vironment': how will our concepts on the pathophysiology of inflammatory bowel disease develop in the future? *Dig Dis* 2012; **30** Suppl 3: 2-11 [PMID: 23295686 DOI: 10.1159/000342585]
- 9 Yu CG, Huang Q. Recent progress on the role of gut microbiota in the pathogenesis of inflammatory bowel disease. J Dig Dis 2013; 14: 513-517 [PMID: 23848393 DOI: 10.1111/1751-2980.12087]
- Vetrano S, Danese S. Colitis, microbiota, and colon cancer: an infernal triangle. *Gastroenterology* 2013; 144: 461-463 [PMID: 23260490 DOI: 10.1053/j.gastro.2012.12.016]
- 11 Morgan XC, Tickle TL, Sokol H, Gevers D, Devaney KL, Ward DV, Reyes JA, Shah SA, LeLeiko N, Snapper SB, Bousvaros A, Korzenik J, Sands BE, Xavier RJ, Huttenhower C. Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment. *Genome Biol* 2012; 13: R79 [PMID: 23013615 DOI: 10.1186/gb-2012-13-9-r79]
- 12 Jostins L, Ripke S, Weersma RK, Duerr RH, McGovern DP, Hui KY, Lee JC, Schumm LP, Sharma Y, Anderson CA, Essers J, Mitrovic M, Ning K, Cleynen I, Theatre E, Spain SL, Raychaudhuri S, Goyette P, Wei Z, Abraham C, Achkar JP, Ahmad T, Amininejad L, Ananthakrishnan AN, Andersen V, Andrews JM, Baidoo L, Balschun T, Bampton PA, Bitton A, Boucher G, Brand S, Büning C, Cohain A, Cichon S, D'Amato M, De Jong D, Devaney KL, Dubinsky M, Edwards C, Ellinghaus D, Ferguson LR, Franchimont D, Fransen K, Gearry R, Georges M, Gieger C, Glas J, Haritunians T, Hart A, Hawkey C, Hedl M, Hu X, Karlsen TH, Kupcinskas L, Kugathasan S, Latiano A, Laukens D, Lawrance IC, Lees CW, Louis E, Mahy G, Mansfield J, Morgan AR, Mowat C, Newman W, Palmieri O, Ponsioen CY, Potocnik U, Prescott NJ,



Regueiro M, Rotter JI, Russell RK, Sanderson JD, Sans M, Satsangi J, Schreiber S, Simms LA, Sventoraityte J, Targan SR, Taylor KD, Tremelling M, Verspaget HW, De Vos M, Wijmenga C, Wilson DC, Winkelmann J, Xavier RJ, Zeissig S, Zhang B, Zhang CK, Zhao H. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* 2012; **491**: 119-124 [PMID: 23128233 DOI: 10.1038/nature11582]

- 13 Blumberg R. What are innate and acquired immunity, and why are they important in IBD? Inflamm Bowel Dis 2008; 14 Suppl 2: S93-S94 [PMID: 18816702 DOI: 10.1002/ibd.20689]
- 14 Wallace KL, Zheng LB, Kanazawa Y, Shih DQ. Immunopathology of inflammatory bowel disease. World J Gastroenterol 2014; 20: 6-21 [PMID: 24415853 DOI: 10.3748/wjg.v20.i1.6]
- 15 Cader MZ, Kaser A. Recent advances in inflammatory bowel disease: mucosal immune cells in intestinal inflammation. *Gut* 2013; 62: 1653-1664 [PMID: 24104886 DOI: 10.1136/gutjnl-2012-303955]
- 16 D'Alessio S, Tacconi C, Fiocchi C, Danese S. Advances in therapeutic interventions targeting the vascular and lymphatic endothelium in inflammatory bowel disease. *Curr Opin Gastroenterol* 2013; 29: 608-613 [PMID: 24100721 DOI: 10.1097/MOG.0b013e328365d37c]
- 17 Rieder F, Kessler SP, West GA, Bhilocha S, de la Motte C, Sadler TM, Gopalan B, Stylianou E, Fiocchi C. Inflammation-induced endothelial-to-mesenchymal transition: a novel mechanism of intestinal fibrosis. *Am J Pathol* 2011; 179: 2660-2673 [PMID: 21945322 DOI: 10.1016/j.ajpath.2011.07.042]
- 18 Raleigh DR, Boe DM, Yu D, Weber CR, Marchiando AM, Bradford EM, Wang Y, Wu L, Schneeberger EE, Shen L, Turner JR. Occludin S408 phosphorylation regulates tight junction protein interactions and barrier function. *J Cell Biol* 2011; 193: 565-582 [PMID: 21536752 DOI: 10.1083/jcb.201010065]
- 19 Al-Sadi R, Guo S, Dokladny K, Smith MA, Ye D, Kaza A, Watterson DM, Ma TY. Mechanism of interleukin-1β induced-increase in mouse intestinal permeability in vivo. *J Interferon Cytokine Res* 2012; 32: 474-484 [PMID: 22817402 DOI: 10.1089/jir.2012.0031]
- 20 Ye D, Guo S, Al-Sadi R, Ma TY. MicroRNA regulation of intestinal epithelial tight junction permeability. Gastroenterology 2011; 141: 1323-1333 [PMID: 21763238 DOI: 10.1053/j.gastro.2011.07.005]
- 21 Stein J, Dignass AU. Laboratory diagnostics in IBD What the gastroenterologist should know. European Gastroenterology Journal 2015; 32-47
- 22 Vermeire S, Van Assche G, Rutgeerts P. C-reactive protein as a marker for inflammatory bowel disease. Inflamm Bowel Dis 2004; 10: 661-665 [PMID: 15472532]
- 23 Cioffi M, Rosa AD, Serao R, Picone I, Vietri MT. Laboratory markers in ulcerative colitis: Current insights and future advances. *World J Gastrointest Pathophysiol* 2015; 6: 13-22 [PMID: 25685607 DOI: 10.4291/wjgp.v6.i1.13]
- 24 Henriksen M, Jahnsen J, Lygren I, Stray N, Sauar J, Vatn MH, Moum B; IBSEN Study Group. C-reactive protein: a predictive factor and marker of inflammation in inflammatory bowel disease. Results from a prospective population-based study. *Gut* 2008; 57: 1518-1523 [PMID: 18566104 DOI: 10.1136/gut.2007.146357]
- 25 Peyrin-Biroulet L, Sandborn W, Sands BE, Reinisch W, Bemelman W, Bryant RV, D'Haens G, Dotan I, Dubinsky M, Feagan B, Fiorino G, Gearry R, Krishnareddy S, Lakatos PL, Loftus EV, Marteau P, Munkholm P, Murdoch TB, Ordás I, Panaccione R, Riddell RH, Ruel J, Rubin DT, Samaan M, Siegel CA, Silverberg MS, Stoker J, Schreiber S, Travis S, Van Assche G, Danese S, Panes J, Bouguen G, O'Donnell S, Pariente B, Winer S, Hanauer S, Colombel JF. Selecting Therapeutic Targets in Inflammatory Bowel Disease (STRIDE): Determining Therapeutic Goals for Treat-to-Target. *Am J Gastroenterol* 2015; 110: 1324-1338 [PMID: 26303131 DOI: 10.1038/ajg.2015.233]
- 26 Reumaux D, Duthilleul P, Roos D. Pathogenesis of diseases associated with antineutrophil cytoplasm autoantibodies. *Hum Immunol* 2004; 65: 1-12 [PMID: 14700590]
- 27 Bennike T, Birkelund S, Stensballe A, Andersen V. Biomarkers in inflammatory bowel diseases: current status and proteomics identification strategies. *World J Gastroenterol* 2014; 20: 3231-3244 [PMID: 24696607 DOI: 10.3748/wjg.v20.i12.3231]
- 28 Iskandar HN, Ciorba MA. Biomarkers in inflammatory bowel disease: current practices and recent advances. *Transl Res* 2012; 159: 313-325 [PMID: 22424434 DOI: 10.1016/j.trsl.2012.01.001]
- 29 Reumaux D, de Boer M, Meijer AB, Duthilleul P, Roos D. Expression of myeloperoxidase (MPO) by neutrophils is necessary for their activation by anti-neutrophil cytoplasm autoantibodies (ANCA) against MPO. J Leukoc Biol 2003; 73: 841-849 [PMID: 12773517]
- 30 Peyrin-Biroulet L, Standaert-Vitse A, Branche J, Chamaillard M. IBD serological panels: facts and perspectives. *Inflamm Bowel Dis* 2007; 13: 1561-1566 [PMID: 17636565 DOI: 10.1002/ibd.20226]
- 31 Peeters M, Joossens S, Vermeire S, Vlietinck R, Bossuyt X, Rutgeerts P. Diagnostic value of anti-Saccharomyces cerevisiae and antineutrophil cytoplasmic autoantibodies in inflammatory bowel disease. *Am J Gastroenterol* 2001; 96: 730-734 [PMID: 11280542 DOI: 10.1111/j.1572-0241.2001.03613.x]
- 32 Bernstein CN, El-Gabalawy H, Sargent M, Landers C, Rawsthorne P, Elias B, Targan SR. Assessing inflammatory bowel disease-associated antibodies in Caucasian and First Nations cohorts. *Can J Gastroenterol* 2011; 25: 269-273 [PMID: 21647462]
- 33 Tibble J, Teahon K, Thjodleifsson B, Roseth A, Sigthorsson G, Bridger S, Foster R, Sherwood R, Fagerhol M, Bjarnason I. A simple method for assessing intestinal inflammation in Crohn's disease. *Gut* 2000; 47: 506-513 [PMID: 10986210]
- 34 Røseth AG, Fagerhol MK, Aadland E, Schjønsby H. Assessment of the neutrophil dominating protein calprotectin in feces. A methodologic study. *Scand J Gastroenterol* 1992; 27: 793-798 [PMID: 1411288]
- 35 Gisbert JP, Bermejo F, Pérez-Calle JL, Taxonera C, Vera I, McNicholl AG, Algaba A, López P, López-Palacios N, Calvo M, González-Lama Y, Carneros JA, Velasco M, Maté J. Fecal calprotectin and lactoferrin for the prediction of inflammatory bowel disease relapse. *Inflamm Bowel Dis* 2009; 15: 1190-1198 [PMID: 19291780 DOI: 10.1002/ibd.20933]
- 36 Henderson P, Anderson NH, Wilson DC. The diagnostic accuracy of fecal calprotectin during the investigation of suspected pediatric inflammatory bowel disease: a systematic review and meta-analysis. *Am J Gastroenterol* 2014; 109: 637-645 [PMID: 23670113 DOI: 10.1038/ajg.2013.131]
- 37 Gecse KB, Brandse JF, van Wilpe S, Löwenberg M, Ponsioen C, van den Brink G, D'Haens G. Impact of disease location on fecal calprotectin levels in Crohn's disease. Scand J Gastroenterol 2015; 50: 841-847 [PMID: 25636819 DOI: 10.3109/00365521.2015.1008035]
- 38 Manceau H, Chicha-Cattoir V, Puy H, Peoc'h K. Fecal calprotectin in inflammatory bowel diseases: update and perspectives. *Clin Chem Lab Med* 2017; 55: 474-483 [PMID: 27658156 DOI: 10.1515/cclm-2016-0522]
- 39 De Vos M, Louis EJ, Jahnsen J, Vandervoort JG, Noman M, Dewit O, D'haens GR, Franchimont D, Baert FJ, Torp RA, Henriksen M, Potvin PM, Van Hootegem PP, Hindryckx PM, Moreels TG, Collard A,

Karlsen LN, Kittang E, Lambrecht G, Grimstad T, Koch J, Lygren I, Coche JC, Mana F, Van Gossum A, Belaiche J, Cool MR, Fontaine F, Maisin JM, Muls V, Neuville B, Staessen DA, Van Assche GA, de Lange T, Solberg IC, Vander Cruyssen BJ, Vermeire SA. Consecutive fecal calprotectin measurements to predict relapse in patients with ulcerative colitis receiving infliximab maintenance therapy. Inflamm Bowel Dis 2013; 19: 2111-2117 [PMID: 23883959 DOI: 10.1097/MIB.0b013e31829b2a37]

- 40 Molander P, Färkkilä M, Ristimäki A, Salminen K, Kemppainen H, Blomster T, Koskela R, Jussila A, Rautiainen H, Nissinen M, Haapamäki J, Arkkila P, Nieminen U, Kuisma J, Punkkinen J, Kolho KL, Mustonen H, Sipponen T. Does fecal calprotectin predict short-term relapse after stopping TNFa-blocking agents in inflammatory bowel disease patients in deep remission? J Crohns Colitis 2015; 9: 33-40 [PMID: 25052347 DOI: 10.1016/j.crohns.2014.06.012]
- Kane SV, Sandborn WJ, Rufo PA, Zholudev A, Boone J, Lyerly D, Camilleri M, Hanauer SB. Fecal 41 lactoferrin is a sensitive and specific marker in identifying intestinal inflammation. Am J Gastroenterol 2003; 98: 1309-1314 [PMID: 12818275 DOI: 10.1111/j.1572-0241.2003.07458.x]
- 42 Angriman I, Scarpa M, D'Incà R, Basso D, Ruffolo C, Polese L, Sturniolo GC, D'Amico DF, Plebani M. Enzymes in feces: useful markers of chronic inflammatory bowel disease. Clin Chim Acta 2007; 381: 63-68 [PMID: 17368600 DOI: 10.1016/j.cca.2007.02.025]
- Nixon AB, Pang H, Starr MD, Friedman PN, Bertagnolli MM, Kindler HL, Goldberg RM, Venook AP, 43 Hurwitz HI; Alliance for Clinical Trials In Oncology. Prognostic and predictive blood-based biomarkers in patients with advanced pancreatic cancer: results from CALGB80303 (Alliance). Clin Cancer Res 2013; 19: 6957-6966 [PMID: 24097873 DOI: 10.1158/1078-0432.CCR-13-0926]
- Ingvarsson J, Wingren C, Carlsson A, Ellmark P, Wahren B, Engström G, Harmenberg U, Krogh M, 44 Peterson C, Borrebaeck CA. Detection of pancreatic cancer using antibody microarray-based serum protein profiling. Proteomics 2008; 8: 2211-2219 [PMID: 18528842 DOI: 10.1002/pmic.200701167]
- 45 Vigren E, Hamberg M, Zhaunerchyk V, Kaminska M, Thomas RD, Trippel S, Zhang M, Kashperka I, Ugglas MA, Walsh C, Wester R, Semaniak J, Larsson M, Geppert WD. Dissociative recombination of the acetaldehyde cation, CH(3)CHO(+). Phys Chem Chem Phys 2010; 12: 11670-11673 [PMID: 20714489 DOI: 10.1039/c003857a]
- Wingren C, Sandström A, Segersvärd R, Carlsson A, Andersson R, Löhr M, Borrebaeck CA 46 Identification of serum biomarker signatures associated with pancreatic cancer. Cancer Res 2012; 72: 2481-2490 [PMID: 22589272 DOI: 10.1158/0008-5472.CAN-11-2883]
- 47 Mehan MR, Williams SA, Siegfried JM, Bigbee WL, Weissfeld JL, Wilson DO, Pass HI, Rom WN, Muley T, Meister M, Franklin W, Miller YE, Brody EN, Ostroff RM. Validation of a blood protein signature for non-small cell lung cancer. Clin Proteomics 2014; 11: 32 [PMID: 25114662 DOI: 10.1186/1559-0275-11-32
- Pommier AJ, Shaw R, Spencer SK, Morgan SR, Hoff PM, Robertson JD, Barry ST, Jürgensmeier JM. 48 Serum protein profiling reveals baseline and pharmacodynamic biomarker signatures associated with clinical outcome in mCRC patients treated with chemotherapy ± cediranib. Br J Cancer 2014; 111: 1590-1604 [PMID: 25121956 DOI: 10.1038/bjc.2014.436]
- Viennois E, Zhao Y, Merlin D. Biomarkers of Inflammatory Bowel Disease: From Classical Laboratory 49 Tools to Personalized Medicine. Inflamm Bowel Dis 2015; 21: 2467-2474 [PMID: 25985250 DOI: 10.1097/MIB.000000000000444
- Anderson NL, Anderson NG. Proteome and proteomics: new technologies, new concepts, and new words. 50 Electrophoresis 1998; 19: 1853-1861 [PMID: 9740045 DOI: 10.1002/elps.1150191103]
- Barnett M, Young W, Cooney J, Roy N. Metabolomics and Proteomics, and What to Do with All These 51 'Omes': Insights from Nutrigenomic Investigations in New Zealand. J Nutrigenet Nutrigenomics 2014; 7: 274-282 [PMID: 25997469 DOI: 10.1159/000381349]
- Hong SN, Joung JG, Bae JS, Lee CS, Koo JS, Park SJ, Im JP, Kim YS, Kim JW, Park WY, Kim YH. 52 RNA-seq Reveals Transcriptomic Differences in Inflamed and Noninflamed Intestinal Mucosa of Crohn's Disease Patients Compared with Normal Mucosa of Healthy Controls. Inflamm Bowel Dis 2017; 23: 1098-1108 [PMID: 28613228 DOI: 10.1097/MIB.0000000000001066]
- Bennike TB, Carlsen TG, Ellingsen T, Bonderup OK, Glerup H, Bøgsted M, Christiansen G, Birkelund S, 53 Stensballe A, Andersen V. Neutrophil Extracellular Traps in Ulcerative Colitis: A Proteome Analysis of Intestinal Biopsies. Inflamm Bowel Dis 2015; 21: 2052-2067 [PMID: 25993694 DOI: 10.1097/MIB.000000000000460
- Mantovani A, Cassatella MA, Costantini C, Jaillon S. Neutrophils in the activation and regulation of 54 innate and adaptive immunity. Nat Rev Immunol 2011; 11: 519-531 [PMID: 21785456 DOI: 10.1038/nri3024
- O'Donoghue AJ, Jin Y, Knudsen GM, Perera NC, Jenne DE, Murphy JE, Craik CS, Hermiston TW. 55 Global substrate profiling of proteases in human neutrophil extracellular traps reveals consensus motif predominantly contributed by elastase. PLoS One 2013; 8: e75141 [PMID: 24073241 DOI: 10.1371/journal.pone.0075141
- Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, Weinrauch Y, Zychlinsky A. 56 Neutrophil extracellular traps kill bacteria. Science 2004; 303: 1532-1535 [PMID: 15001782 DOI: 10.1126/science.1092385]
- Delgado-Rizo V, Martínez-Guzmán MA, Iñiguez-Gutierrez L, García-Orozco A, Alvarado-Navarro A, 57 Fafutis-Morris M. Neutrophil Extracellular Traps and Its Implications in Inflammation: An Overview. Front Immunol 2017; 8: 81 [PMID: 28220120 DOI: 10.3389/fimmu.2017.00081]
- Riaz T, Sollid LM, Olsen I, de Souza GA. Quantitative Proteomics of Gut-Derived Th1 and Th1/Th17 58 Clones Reveal the Presence of CD28+ NKG2D- Th1 Cytotoxic CD4+ T cells. Mol Cell Proteomics 2016; 15: 1007-1016 [PMID: 26637539 DOI: 10.1074/mcp.M115.050138]
- 59 Pastorelli L, De Salvo C, Mercado JR, Vecchi M, Pizarro TT. Central role of the gut epithelial barrier in the pathogenesis of chronic intestinal inflammation: lessons learned from animal models and human genetics. Front Immunol 2013; 4: 280 [PMID: 24062746 DOI: 10.3389/fimmu.2013.00280]
- Poulsen NA, Andersen V, Møller JC, Møller HS, Jessen F, Purup S, Larsen LB. Comparative analysis of 60 inflamed and non-inflamed colon biopsies reveals strong proteomic inflammation profile in patients with ulcerative colitis. BMC Gastroenterol 2012; 12: 76 [PMID: 22726388 DOI: 10.1186/1471-230X-12-76
- Moriggi M, Pastorelli L, Torretta E, Tontini GE, Capitanio D, Bogetto SF, Vecchi M, Gelfi C 61 Contribution of Extracellular Matrix and Signal Mechanotransduction to Epithelial Cell Damage in Inflammatory Bowel Disease Patients: A Proteomic Study. Proteomics 2017; 17 [PMID: 29027377 DOI: 10.1002/pmic.201700164]
- 62 Nanni P, Mezzanotte L, Roda G, Caponi A, Levander F, James P, Roda A. Differential proteomic analysis



of HT29 Cl.16E and intestinal epithelial cells by LC ESI/QTOF mass spectrometry. *J Proteomics* 2009; **72**: 865-873 [PMID: 19168159 DOI: 10.1016/j.jprot.2008.12.010]

- 63 Shkoda A, Werner T, Daniel H, Gunckel M, Rogler G, Haller D. Differential protein expression profile in the intestinal epithelium from patients with inflammatory bowel disease. *J Proteome Res* 2007; 6: 1114-1125 [PMID: 17330946 DOI: 10.1021/pr060433m]
- 64 Yau YY, Leong RWL, Pudipeddi A, Redmond D, Wasinger VC. Serological Epithelial Component Proteins Identify Intestinal Complications in Crohn's Disease. *Mol Cell Proteomics* 2017; 16: 1244-1257 [PMID: 28490445 DOI: 10.1074/mcp.M116.066506]
- 65 Korolkova OY, Myers JN, Pellom ST, Wang L, M'Koma AE. Characterization of Serum Cytokine Profile in Predominantly Colonic Inflammatory Bowel Disease to Delineate Ulcerative and Crohn's Colitides. *Clin Med Insights Gastroenterol* 2015; 8: 29-44 [PMID: 26078592 DOI: 10.4137/CGast.S20612]
- 66 M'Koma AE, Seeley EH, Washington MK, Schwartz DA, Muldoon RL, Herline AJ, Wise PE, Caprioli RM. Proteomic profiling of mucosal and submucosal colonic tissues yields protein signatures that differentiate the inflammatory colitides. *Inflamm Bowel Dis* 2011; 17: 875-883 [PMID: 20806340 DOI: 10.1002/ibd.21442]
- 67 Seeley EH, Washington MK, Caprioli RM, M'Koma AE. Proteomic patterns of colonic mucosal tissues delineate Crohn's colitis and ulcerative colitis. *Proteomics Clin Appl* 2013; 7: 541-549 [PMID: 23382084 DOI: 10.1002/prca.201200107]
- 68 Starr AE, Deeke SA, Ning Z, Chiang CK, Zhang X, Mottawea W, Singleton R, Benchimol EI, Wen M, Mack DR, Stintzi A, Figeys D. Proteomic analysis of ascending colon biopsies from a paediatric inflammatory bowel disease inception cohort identifies protein biomarkers that differentiate Crohn's disease from UC. *Gut* 2017; 66: 1573-1583 [PMID: 27216938 DOI: 10.1136/gutjnl-2015-310705]
- 69 Meuwis MA, Fillet M, Lutteri L, Marée R, Geurts P, de Seny D, Malaise M, Chapelle JP, Wehenkel L, Belaiche J, Merville MP, Louis E. Proteomics for prediction and characterization of response to infliximab in Crohn's disease: a pilot study. *Clin Biochem* 2008; 41: 960-967 [PMID: 18489908 DOI: 10.1016/j.clin-biochem.2008.04.021]
- 70 Slungaard A. Platelet factor 4 modulation of the thrombomodulin-protein C system. Crit Care Med 2004; 32: S331-S335 [PMID: 15118540]
- 71 Bikfalvi A. Platelet factor 4: an inhibitor of angiogenesis. Semin Thromb Hemost 2004; 30: 379-385 [PMID: 15282661 DOI: 10.1055/s-2004-831051]
- 72 Simi M, Leardi S, Tebano MT, Castelli M, Costantini FM, Speranza V. Raised plasma concentrations of platelet factor 4 (PF4) in Crohn's disease. *Gut* 1987; 28: 336-338 [PMID: 3570037]
- 73 Gazouli M, Anagnostopoulos AK, Papadopoulou A, Vaiopoulou A, Papamichael K, Mantzaris G, Theodoropoulos GE, Anagnou NP, Tsangaris GT. Serum protein profile of Crohn's disease treated with infliximab. J Crohns Colitis 2013; 7: e461-e470 [PMID: 23562004 DOI: 10.1016/j.crohns.2013.02.021]
- 74 Magnusson MK, Strid H, Isaksson S, Bajor A, Lasson A, Ung KA, Öhman L. Response to infliximab therapy in ulcerative colitis is associated with decreased monocyte activation, reduced CCL2 expression and downregulation of Tenascin C. J Crohns Colitis 2015; 9: 56-65 [PMID: 25518051 DOI: 10.1093/ecco-jcc/jju008]
- 75 Banks C, Bateman A, Payne R, Johnson P, Sheron N. Chemokine expression in IBD. Mucosal chemokine expression is unselectively increased in both ulcerative colitis and Crohn's disease. *J Pathol* 2003; 199: 28-35 [PMID: 12474223 DOI: 10.1002/path.1245]
- 76 Uguccioni M, Gionchetti P, Robbiani DF, Rizzello F, Peruzzo S, Campieri M, Baggiolini M. Increased expression of IP-10, IL-8, MCP-1, and MCP-3 in ulcerative colitis. *Am J Pathol* 1999; 155: 331-336 [PMID: 10433925 DOI: 10.1016/S0002-9440(10)65128-0]
- 77 Leal RF, Planell N, Kajekar R, Lozano JJ, Ordás I, Dotti I, Esteller M, Masamunt MC, Parmar H, Ricart E, Panés J, Salas A. Identification of inflammatory mediators in patients with Crohn's disease unresponsive to anti-TNFα therapy. *Gut* 2015; 64: 233-242 [PMID: 24700437 DOI: 10.1136/gutjnl-2013-306518]
- 78 Fiocchi C. Inflammatory Bowel Disease: Complexity and Variability Need Integration. Front Med (Lausanne) 2018; 5: 75 [PMID: 29619371 DOI: 10.3389/fmed.2018.00075]
- 79 Loktionov A, Chhaya V, Bandaletova T, Poullis A. Inflammatory bowel disease detection and monitoring by measuring biomarkers in non-invasively collected colorectal mucus. J Gastroenterol Hepatol 2017; 32: 992-1002 [PMID: 27787913 DOI: 10.1111/jgh.13627]

Gaishideng® WJG | https://www.wjgnet.com



Published By Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk:http://www.f6publishing.com/helpdesk http://www.wjgnet.com



© 2020 Baishideng Publishing Group Inc. All rights reserved.