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## Mechanisms of Interaction of Ultrasound with Cancellous Bone: A Review

**Keith A. Wear [Senior Member, IEEE]**

U.S. Food and Drug Administration, Silver Spring, MD, 20993

### Abstract

Ultrasound is now a clinically-accepted modality in the management of osteoporosis. The most common commercial clinical devices assess fracture risk from measurements of attenuation and sound speed in cancellous bone. This review discusses fundamental mechanisms underlying the interaction between ultrasound and cancellous bone. Because of its two-phase structure (mineralized trabecular network embedded in soft tissue—marrow), its anisotropy, and its inhomogeneity, cancellous bone is more difficult to characterize than most soft tissues. Experimental data for the dependences of attenuation, sound speed, dispersion, and scattering on ultrasound frequency, bone mineral density, composition, microstructure, and mechanical properties are presented. The relative roles of absorption, scattering, and phase cancellation in determining attenuation measurements *in vitro* and *in vivo* are delineated. Common speed of sound metrics, which entail measurements of transit times of pulse leading edges (to avoid multipath interference), are greatly influenced by attenuation, dispersion, and system properties including center frequency and bandwidth. However, a theoretical model has been shown to be effective for correction for these confounding factors *in vitro* and *in vivo*. Theoretical and phantom models are presented to elucidate why cancellous bone exhibits negative dispersion, unlike soft tissue, which exhibits positive dispersion. Signal processing methods are presented for separating “fast” and “slow” waves (predicted by poro-elasticity theory and supported in cancellous bone) even when the two waves overlap in time and frequency domains. Models to explain dependences of scattering on frequency and mean trabecular thickness are presented and compared with measurements. Anisotropy, the effect of the fluid filler medium (marrow *in vivo* or water *in vitro*), phantoms, computational modeling of ultrasound propagation, acoustic microscopy, and nonlinear properties in cancellous bone are also discussed.

### Keywords

cancellous bone; ultrasound; bone sonometry; osteoporosis

### I. INTRODUCTION

IN 1984, Langton *et al.* published the finding that ultrasonic attenuation in calcaneus (heel bone) *in vivo* has a strong correlation with bone mineral content, an important determinant of fracture risk [1]. This seminal finding launched a worldwide effort to investigate the

characterization of cancellous bone with ultrasound. Cancellous bone (a.k.a. trabecular bone or spongy bone) consists of a mineralized, porous trabecular network embedded in bone marrow, with filament-like trabeculae tending to align roughly along loading directions. It is mostly found in calcaneus, vertebrae, and in medullary cavities near the ends of tubular bones. Cancellous bone is surrounded by a denser, less-porous shell of cortical bone. Fig. 1 shows a human calcaneus with lateral cortical endplates removed. Fig. 2 shows human cancellous femur.

Early ultrasound studies in bone focused on the calcaneus rather than other skeletal sites such as hip and spine, where the most serious osteoporotic fractures occur. There are many reasons to support this approach.

First, the calcaneus is accessible to interrogation from two opposite sides, has relatively simple shape, and has minimal surrounding soft tissue. These features make it a more practical target for through-transmission measurements than other skeletal sites. (Through-transmission measurements use one transducer as a transmitter and a second, opposing, co-axially-aligned transducer on the other side of the bone as a receiver.)

Second, the calcaneus contains approximately 90% cancellous bone and 10% cortical bone [2] (see Fig. 1). Because of high surface area to bone matrix volume ratio, cancellous bone exhibits faster loss than cortical bone at the onset of unbalanced bone remodeling associated with osteoporosis [3]. Therefore, skeletal sites rich in cancellous bone would be expected to reflect changes due to osteoporosis earlier than other skeletal sites.

Third, the calcaneus is a weight-bearing bone that is thought to experience a mechanical environment similar to clinically important sites such as hip and spine [2].

Finally, an important prospective x-ray study involving 8134 women established that bone density at the calcaneus is a strong indicator of risk of hip fracture (the most debilitating kind of osteoporotic fracture), superior to bone density at the radius and spine (but of course inferior to bone density at the proximal femur) [4].

Clinical utility for through-transmission ultrasound to assess hip fracture risk was established by studies in the mid-1990s [5, 6]. This led to the development of commercial, clinical bone sonometers based on the through-transmission geometry. Now, calcaneal bone sonometry is recognized by major professional organizations as effective in the management of osteoporosis [7, 8].

Future development of medical ultrasound technology involving bone (including but not limited to devices for measuring fracture risk) would benefit from a comprehensive understanding of the mechanisms of the interaction between ultrasound and cancellous bone. With this motivation, the present paper attempts to summarize the current state of knowledge on physical interpretation of ultrasound measurements in cancellous bone. The emphasis is on human cancellous bone, but examples in other species are considered. This topic is more than vast enough to fill a review paper of this size. Therefore, the interaction of ultrasound with cortical bone [9–11] and skull [12–14] is left to others. In addition, comprehensive

discussions of clinical diagnostic findings [15] and therapeutic applications [16, 17] in bone may be found elsewhere.

## II. ATTENUATION

### A. Metrics for Attenuation

Attenuation coefficient is a frequency-dependent material property that describes losses due to absorption and scattering. Attenuation coefficient and speed of sound (SOS) are usually measured in through-transmission. One transducer transmits a broadband pulse into the bone. A second, opposing, co-axially-aligned transducer receives the attenuated signal that passed through the bone. In addition, a calibration measurement is performed with only water between the two transducers. Complex amplitude spectra are usually obtained by applying the Fast Fourier Transform (FFT) to digitized radio-frequency (RF) signals. If  $f$  is frequency,  $X(f)$  is the calibration amplitude spectrum,  $Y(f)$  is the amplitude spectrum of the signal propagated through bone, and attenuation in water is neglected, then [18]

$$20\log\left|\frac{X(f)}{Y(f)}\right| = \alpha(f)d - 10\log\left|T_{ws}^I(f)\right| \left|T_{sw}^I(f)\right| \quad (1)$$

where  $T_{ws}(f)$  and  $T_{sw}(f)$  are intensity transmission coefficients at water-sample and sample-water interfaces,  $\alpha_s(f)$  is the attenuation coefficient in dB per unit length, and  $d$  is the sample thickness.

This substitution technique assumes that the effects of beam diffraction are identical for the two measurements and therefore cancel out when the spectral ratio is taken. This assumption gets weaker as the difference in sound speed between the bone sample and water increases. A diffraction correction method to account for sound speed mismatch has been developed [19, 20]. This effect has been shown to be significant for attenuation and velocity metrics in samples with velocity exceeding 2000 m/s [19–21] but small for attenuation ( $\pm 0.05$  dB/cm) and phase velocity ( $\pm 1$  m/s) in typical cancellous bone samples [21].

The slope of a linear fit of  $20\log[X(f)/Y(f)]$  vs.  $f$  over the usable frequency band of the ultrasound measurement system (*e.g.*, 300 kHz – 700 kHz) is known as broadband ultrasound attenuation (BUA) [22]. Normalized BUA or nBUA is BUA divided by sample thickness  $d$  [22]. Typical units for BUA and nBUA are dB/MHz and dB/cmMHz respectively.

Often the frequency dependences of transmission coefficients may be neglected over the experimental frequency band, especially for *in vitro* experiments in which the surfaces of pure cancellous bone samples are machined to be approximately planar. With this assumption, nBUA gives the slope of the attenuation coefficient, which is an intrinsic material property. BUA is not an intrinsic material property because it is the product of nBUA and bone thickness  $d$ . However, since fracture risk is negatively correlated with both nBUA and bone thickness, the product makes an effective clinical fracture risk predictor.

The range of frequencies for the BUA measurement may be enhanced using coded excitation [23]. BUA images of calcaneus *in vivo* have been shown to be feasible [24–27].

Cortical endplates have been reported to increase nBUA by an average of 13% [22] and BUA by an average of 15% [28] in human calcaneus. Algorithms have been proposed to compensate for the effects of cortical endplates [28–30].

While calcaneus is the most straightforward site for measurement of BUA, BUA can also be measured at the femur [31–33]. Many commercial clinical devices measure BUA in calcaneus [2, 9, 34].

An alternative attenuation metric, frequency-modulated attenuation (FMA) is also measured in through-transmission mode. Instead of using broadband ultrasound pulses, FMA uses linear-frequency-swept signals.  $FMA = 10 \log (E_r / E_b)$  where  $E_r$  and  $E_b$  are time-integrals for the squared through-transmission signal envelopes for reference (*i.e.*, water-only) and bone measurements [35, 36].

For *in vivo* measurements, there is uncertainty in  $d$  because bones (*e.g.*, calcaneus) are surrounded by soft tissue. However,  $d$  can be estimated by performing pulse-echo measurements with both transducers in addition to through-transmission measurements [37–40].

## B. Attenuation vs. Frequency

Many publications have reported that attenuation coefficient in human calcaneus varies approximately linearly with frequency in a clinical range of about 300–600 kHz [1, 41–44]. Over broader frequency ranges, attenuation coefficient can show deviations from linear frequency dependence in human cancellous calcaneus *in vitro* [45–48], as shown in Fig. 3. For example, attenuation coefficient from 0.2–1.7 MHz has been reported to be proportional to  $f^{1.09 \pm 0.3}$  in 14 human cancellous calcaneus specimens *in vitro* [45]. Even over a clinical range (300–700 kHz), polynomial fits of attenuation coefficients as functions of frequency showed quadratic coefficients statistically different from zero in 12 of 30 human cancellous calcaneus samples *in vitro* [48]. A higher degree of nonlinearity has been reported in human cancellous femur and tibia, with rate of change of attenuation coefficient (dB/cmMHz) higher below 1 MHz and lower above 1 MHz [49].

## C. Attenuation vs. BMD and Composition

In studies on human cancellous calcaneus [50–54], vertebrae [55], femur [49, 56, 57] and tibia [49, 58] *in vitro*, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between nBUA and site-matched, volumetric bone mineral density (BMD) near the clinical range of frequencies are  $r^2 = 0.71$ ,  $0.67 \pm 0.10$ . A similar value,  $r^2 = 0.69$ , has been reported for intact human femur *in vitro* [59, 60].

In studies in human calcaneus *in vivo* [43, 44, 61–64], the median and mean  $\pm$  standard deviation of the squared correlation coefficient between BUA and site-matched, areal BMD (volumetric BMD multiplied by thickness) near the clinical range of frequencies are  $r^2 = 0.53$ ,  $0.56 \pm 0.08$ .

In another study in human calcaneus *in vivo*, BUA correlated positively and linearly with areal BMD only in calcanei of low or moderate density ( $r^2 = 0.72$ ). At high levels of BMD,

BUA plateaued or possibly declined with increasing BMD, as shown in Fig. 4. When calcaneal areas of low and high areal BMD were included, the relationship between BMD and BUA was best described by a second-order polynomial ( $r^2 = 0.62$ ) [65]. This behavior is consistent with earlier measurements of nBUA in human and bovine cancellous bone *in vitro*, which exhibited a parabolic relationship with porosity with a maximum value near 75% [66–68].

Two studies found that demineralization (by treating bone with nitric acid [42] or EDTA [58]) significantly reduced bovine cancellous femur BUA (300–800 kHz) [42] and human cancellous tibia nBUA (0.5–2 MHz) [58] *in vitro*. However, another study, in which no active demineralization was applied, found no significant correlation between nBUA (1–2.8 MHz) and bone mineral content (normalized to the calcified matrix volume) in human cancellous tibia and femur *in vitro* [69]. The lack of significant correlation may have been partially due to the relatively low natural variation of mineral content in the samples (8.8%) [69] and also to the complex parabolic relationship between BUA/nBUA and BMD mentioned above [65].

One study found that decollagenization (by treating bone with sodium hypochlorite) significantly increased nBUA (0.5–2 MHz) in human cancellous tibia *in vitro* [58]. However, another study, in which no active decollagenization was applied, found no significant correlation between nBUA (1–2.8 MHz) and calcified matrix collagen (normalized to the calcified matrix volume) in human cancellous tibia and femur *in vitro* [69]. The lack of significant correlation may have been partially due to the relatively low natural variation of collagen content in the samples (11.2%) [69].

One study found a significant negative correlation ( $r = -0.47$ ) between nBUA (1–2.8 MHz) and fat content in human cancellous tibia and femur *in vitro* but no significant correlations between nBUA and water content or proteoglycan content [69].

The studies discussed in this section and throughout this review paper include measurements on human and animal cancellous bone. Animal cancellous bone studies provide a useful complement to human cancellous bone studies because they extend ranges of BMD and bone volume fraction to higher values than found in humans. However, because animal bone (particularly bovine bone) can be considerably denser than human bone, trends observed in animal bone do not always exactly match trends observed in human bone.

#### D. Attenuation vs. Microstructure

Histomorphometric analysis of 3D micro-computed tomography ( $\mu$ CT) scans of cancellous bone samples can provide descriptive parameters such as the ratio of bone volume to total volume (BV/TV), mean trabecular thickness (Tb.Th), mean trabecular separation (Tb.Sp), trabecular number (Tb.N), degree of anisotropy (DA), and structural model index (SMI) [70, 71]. While BV/TV is mostly an indication of bone quantity, the other parameters describe trabecular microstructure. Fig. 5 shows steps involved in analyzing human cancellous calcaneus with ultrasound and  $\mu$ CT. Fig. 6 shows a  $\mu$ CT reconstruction of a rectangular volume from a human calcaneus sample. High resolution peripheral CT (HR-pQCT) may be used to analyze microstructure in the distal radius and tibia *in vivo* [72–74].

Investigators have found many significant correlations between QUS parameters and histomorphometric parameters in human cancellous bone. In addition, many have compared univariate regression models of QUS vs. a measure of bone quantity (*e.g.*, apparent density, BMD, or BV/TV) with multiple regression models of QUS vs. bone quantity and histomorphometric parameters. This analysis offers insight into the added variance of QUS parameters explained by microstructure beyond that explained by bone quantity alone. This has implications regarding additional diagnostic information that QUS could provide beyond BMD, the standard clinical measurement.

In multiple regression studies on human cancellous vertebrae [55, 75], calcaneus [52, 53, 75, 76], tibia [77] and femur [57, 77], the median for the squared correlation coefficient between nBUA and site-matched, bone quantity (*e.g.*, BV/TV) near the clinical range of frequencies is  $r^2 = 0.75$ . In these studies, the median value for the added variance of nBUA,  $r^2$ , explained by microstructure is 0.07.

FMA has been shown to be useful for investigating microstructure of ovine [35] and bovine [36] cancellous femur.

### E. Attenuation vs. Mechanical Properties

Mechanical properties of cancellous bone are important because they are closely related to fracture risk, the primary clinical endpoint [78–82]. Cancellous bone contains hydroxyapatite mineral and fibrous collagen components that team up to create an adaptive and flexible structure to withstand a variety of loading conditions. Correlations between QUS parameters (which may be measured *in vivo*) and mechanical properties in cancellous bone (which are usually measured *ex vivo*) are of interest for diagnostic applications.

Mechanical testing is the most widely-accepted method for measuring mechanical properties on specimens but is impractical *in vivo*. Mechanical testing of cancellous bone specimens often entails preconditioning with low-amplitude cyclic compression followed by compression to approximately 5% strain in order to generate a stress-strain curve, from which mechanical parameters such as Young's modulus, yield stress, yield strain and ultimate strength, may be derived [49, 83].

Finite element analysis (FEA) based on high-resolution 3D image data is also effective for estimating mechanical properties on bone specimens [80, 84]. (Although  $\mu$ CT is the most common imaging modality to provide input data for FEA, ultrasound tomography has also been shown to be effective for estimating stiffness in cancellous bone replica models [85].)

In studies on human cancellous calcaneus [22, 86, 87], femur [49, 88] and tibia [49, 88] specimens, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between nBUA and mechanical parameters (*e.g.*, Young's modulus, ultimate strength) are  $r^2 = 0.60, 0.57 \pm 0.14$ . Fig. 7 shows a scatter plot of log strength vs. log nBUA in human calcaneus *in vitro*. Lower correlations have been reported for bovine cancellous femur [83].

In order to investigate the sensitivity of BUA to elastic properties, BUA was measured with 1 MHz transducers in human cancellous calcaneus cores before and after destructive testing that reduced elastic modulus without significant changes to porosity or microstructure [89].

BUA decreased by less than 2% despite decreases in elastic modulus greater than 75%. The investigators concluded that BUA is not directly sensitive to elastic properties of human calcaneus and therefore empirical correlations between BUA and elastic properties must be due to correlations among microstructure, porosity, elastic properties and BUA in undamaged cancellous bone [89, 90].

## F. Mechanisms of Attenuation

Sources of measured attenuation (*e.g.*, BUA or nBUA) include absorption, longitudinal-longitudinal (LL) scattering, longitudinal-shear (LS) scattering, and phase cancellation. A discussion of the role of scattering in determining attenuation is deferred to the section on scattering (Sec. V). However, phase cancellation can be treated here.

The importance of phase cancellation can be assessed by comparing measurements of attenuation performed using phase sensitive (PS) and phase insensitive (PI) methods [18, 44, 91–95]. Fig. 8 shows measurements of BUA assessed by PS and PI methods in 73 women [44]. For  $BUA < 75$  dB/MHz, the two measures are approximately equal, suggesting that phase cancellation at the receiving transducer is negligible. However, for  $BUA > 75$  dB/MHz, the PS values are much greater than PI values, suggesting the phase cancellation at the receiving transducer is substantial. A similar trend has been reported for nBUA in human calcaneus *in vitro*, with a break point near 15 dB/cmMHz [94]. As BUA (or nBUA) increases, scattering also increases and creates more opportunity for phase cancellation and therefore greater discrepancy between PS and PI BUA (or nBUA). Phase cancellation has also been demonstrated in sheep femoral trabecular bone by comparing PS and PI BUA [95] with difference increasing with frequency.

One model for phase cancellation represents the ultrasound beam as a set of parallel “sonic rays,” propagating through a medium with two components with contrasting sound speeds (trabecular bone and marrow) [96]. The transit time of each ray is determined by the proportion of bone and marrow in the path of the ray. The transit time spectrum describes the proportion of sonic rays at a particular transit time [97]. Phase cancellation in this model results from sonic rays with different transit times and therefore different phase shifts. A deconvolution algorithm to recover transit time spectra has been validated in cancellous bone replica samples [98, 99]. The transit time spectrum has been shown to be very effective for predicting solid volume fraction of simplified bone / marrow replica models consisting of acrylic and water [100]. In addition, bone volume fraction obtained from the transit time spectrum has been shown to be effective for estimation of mechanical stiffness and failure load in human cancellous femur *in vitro* [101].

## III. SPEED OF SOUND

### A. Metrics for Speed of Sound

In general, velocity may be described by phase velocity (velocity of a single-frequency component as a function of frequency), group velocity (velocity of the center of a pulse), and signal velocity (velocity of the leading edge of a pulse) [102]. “Speed of sound” (SOS) is measured in through-transmission. One transducer transmits a broadband pulse into the

bone. A second, opposing, co-axially-aligned transducer receives the attenuated signal that passed through the bone. In addition, a calibration measurement is performed with only water between the two transducers. SOS is often computed from

$$SOS = \frac{c_w}{1 + \frac{c_w \Delta t}{d}} \quad (2)$$

where  $c_w$  is the acoustic velocity in water.  $t$  is the difference in transit times of the two pulses, and  $d$  is the thickness of the sample. As mentioned in Sec. II, diffraction-related errors due to speed-of-sound mismatch [20] in the substitution experiment are small for typical cancellous bone samples ( $\pm 1$  m/s) [21].

A marker on the pulse (*e.g.*, a zero crossing) is chosen to measure transit times. The same marker is chosen for bone measurement and calibration (*i.e.*, water only) measurement. Markers are often chosen near the leading edge in order to avoid multipath interference (*e.g.*, forward scattering, refraction, reverberations in cortical plates, multiple scattering). Several investigators have reported that the SOS measurement in cancellous bone depends on marker location with higher values near the leading edge and lower values near the trailing edge *in vitro* [103–107], *in vivo* [108], and in simulation [109]. The main reason for this is that frequency-dependent attenuation (which is a low-pass filter) stretches the attenuated signal in time and causes  $t$  to depend on marker choice and the extent of pulse spreading [106, 110], as shown in Fig. 9. This is a small effect in soft tissues but a much bigger effect in bone because the attenuation coefficient is much larger, leading to greater pulse spreading [106]. This phenomenon has been studied theoretically in soft tissues that were assumed to be non-dispersive [110] and more generally in dispersive media including bone [106]. In Fig. 9, the black bar on the time axis represents the mean  $\pm$  one standard deviation of marker locations from 43 studies that used the following markers: first detectable deviation from zero (L3) [53, 58, 88, 104, 105, 111–119], thresholding at 3 times the noise standard deviation [120], thresholding at 10–20% of the first rising half cycle [49, 69, 77, 105, 119, 121–123], thresholding at 10% of maximum amplitude [124], first maximum [40, 124–126], “first” zero crossing (L2) [31, 32, 55, 75, 76, 89, 104, 105, 119, 122, 124, 127–131], “second” zero crossing (L1) [39, 51, 83, 104, 132], and envelope maximum [51, 83, 124, 130, 133–135]. See Table II.

It can be shown that the dependence of SOS on marker choice and other experimental parameters is approximately given by [107, 108]

$$SOS_n - c_g \approx - \frac{\tau_n c_g^2 \sigma_f^2 \beta}{f_0^2} \frac{1}{1 - (\sigma_f^2 \beta d / f_0)} \quad (3)$$

where the subscript  $n$  denotes a transit-time marker choice (*e.g.*, a zero crossing such as L3 in Fig. 9),  $c_g$  is the group velocity,  $\beta$  is the slope of attenuation coefficient with respect to frequency (analogous to *nBUA*),  $f_0$  is the center frequency, and the calibration signal is assumed to have a Gaussian amplitude spectrum proportional to  $\exp[-(f - f_0)^2 / 2\sigma_f^2]$ . The time variable  $\tau_n$  is the time lag between the transit-time marker ( $n$ ) and the envelope



maximum measured in units of the calibration waveform period ( $T_0 = 1/f_0$ ) (see Table I and Fig. 9).

Eq. (3) suggests that SOS depends not only on a true velocity property ( $c_g$ ) but also on confounding variables such as  $\tau_n$ ,  $f_0$ ,  $\sigma_f$ ,  $\beta$ , and  $d$ . These confounding variables complicate the physical interpretation of SOS measurements and cause difficulties in comparisons of data from different investigations. This likely contributes to the disparity in SOS measurements seen in clinical bone sonometers [136, 137], suggesting the potential benefit of standardization [34, 108, 137].

Fig. 10 shows how (3) applied to data from 73 women *in vivo* compensates for confounding variables to provide more consistent SOS measurements [108]. Since  $\beta$  increases with BMD (see Section II), (3) shows that differences between SOS measurements and group velocity increase with BMD. Therefore, Eq. (3) explains why the correlation between SOS and BMD has been found to increase as marker location moves from the center of the pulse toward the leading edge in human femur [124, 130]. A more general formula than (3) that includes the effects of dispersion is available [106, 138].

Cortical endplates have been reported to introduce a 2% increase in SOS measurements in human calcaneus [112].

## B. Model for Velocity

At 50 kHz, measurements of SOS in bovine cancellous femur have been shown to be predicted well (correlation coefficient squared  $r^2 = 0.94$ ) by bar wave theory [139–141], which is valid at low frequencies such that the wavelength is much greater than both the pore size and the lateral dimensions of the specimen [90, 111]

$$SOS_{bar} = \sqrt{\frac{E}{\rho}} \quad (4)$$

where  $SOS_{bar}$  is the bar wave velocity,  $E$  is Young's modulus, and  $\rho$  is mass density. At a more clinically relevant frequency of 1 MHz, SOS in 18 bovine cancellous femur samples (if longitudinal waves are measured instead of bar waves) has been fit to a linear function of the form  $SOS = A + B(E/\rho)^{1/2}$  with a squared correlation coefficient of  $r^2 = 0.85$  but with a nonzero intercept  $A$  near 1400 m/s [111]. Similarly, at 1.25 MHz, SOS in equine cancellous vertebrae,  $r^2 = 0.94$  and an intercept  $A$  also near 1400 m/s have been reported [126].

## C. Phase Velocity vs. Frequency (Dispersion)

Like SOS, phase velocity,  $c(f)$ , is often measured in through-transmission experiments. Unlike SOS, phase velocity measurement requires Fourier transformation of the digitized through-transmission signals. Phase velocity is computed from [142, 143]

$$c(f) = \frac{c_w}{1 - \frac{c_w \Delta\phi(f)}{2\pi f d}} \quad (5)$$

where  $f$  = frequency, and  $\phi(f)$  is the phase shift between the bone measurement and the calibration measurement. The equation for phase velocity (5) is often reported with a plus sign instead of a minus sign in the denominator. The ambiguity arises from ambiguity in  $\phi(f)$ , which may be computed as  $\phi(f) - \phi_w(f)$  or  $\phi_w(f) - \phi(f)$ . Phase velocity has also been measured in cancellous bone using a phase tracking method [144].

The frequency dependence of phase velocity is called dispersion. Dispersion is generally positive in soft tissues. (That is, phase velocity increases with frequency). For media with attenuation coefficients that vary linearly with frequency, the Kramers-Kronig relations (which are required for causal systems) imply that [145–147]

$$c(f) = c(f_0) + [c(f_0)]^2 \frac{\beta}{\pi^2} \ln\left(\frac{f}{f_0}\right) \quad (6)$$

Eq. (6) predicts that linearly-attenuating media will exhibit positive dispersion with a logarithmic frequency dependence, consistent with empirical evidence for soft tissues [145].

Unlike soft tissues, cancellous bone exhibits negative dispersion [21, 105, 148–153]. Two explanations for this have been proposed. The first explanation is that apparent negative dispersion arises from the interference of two waves, each of which is positively dispersive [154–157]. The two waves could be the fast and slow waves predicted for poro-elastic media by Biot theory [158, 159] (see Section IV). The second explanation is multiple scattering, which also results in interference of waves. This has been demonstrated analytically [160] and experimentally [161, 162] in phantoms with cylindrical scatterers representing trabeculae. It has also been demonstrated analytically and experimentally using a one-dimensional multiple scattering model called the stratified model in bovine tibia and femur [163], ovine femur [164], and human calcaneus *in vitro* [165]. The “restricted-bandwidth form” of the Kramers-Kronig dispersion relations may be used to improve agreement between measured and theoretical phase velocities in cancellous bone [143].

#### D. SOS vs. BMD and Composition

In studies on human cancellous calcaneus [50–53], vertebrae [6, 55], femur [49, 56, 57] and tibia [49, 58] *in vitro*, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between SOS and site-matched, volumetric BMD near the clinical range of frequencies are  $r^2 = 0.77$ ,  $0.72 \pm 0.12$ . A similar value,  $r^2=0.71$ , has been reported for intact human femur *in vitro* [59, 60]. Fig. 11 shows SOS vs apparent density in bovine cancellous tibia *in vitro* in three orientations.

Three studies found that demineralization (by treating bone with nitric acid [42] or EDTA [58, 126]) significantly reduced bovine cancellous femur SOS (600 kHz) [42], human cancellous tibia SOS (2.25 MHz) [58], and equine cancellous vertebrae (1.25 MHz) [126] *in vitro*. However, another study, in which no active demineralization was applied, found no significant correlation between SOS (2.25 MHz) and bone mineral content (normalized to the calcified matrix volume) in human cancellous tibia and femur *in vitro* [69]. The lack of significant correlation may have been partially due to the relatively low natural variation of mineral content in the samples (8.8%) [69].

One study found that decollagenization (by treating bone with sodium hypochlorite) significantly decreased SOS (2.25 MHz) in human cancellous tibia *in vitro* [58]. However, another study, in which no active decollagenization was applied, found no significant correlation between SOS (2.25 MHz) and calcified matrix collagen (normalized to the calcified matrix volume) in human cancellous tibia and femur *in vitro* [69]. The lack of significant correlation may have been partially due to the relatively low natural variation of collagen content in the samples (11.2%) [69].

One study found a significant negative correlation ( $r = -0.43$ ) between SOS (2.25 MHz) and fat content in human cancellous tibia and femur *in vitro* but no significant correlations between SOS and water content or proteoglycan content [69].

Experiments comparing phase velocity measurements in human cancellous femur *in vitro* obtained using marrow, water, and alcohol as fluid fillers suggest that the phase velocity of the saturating fluid is a primary determinant of phase velocity of the composite structure [166].

#### E. SOS vs. Microstructure

In multiple regression studies on human cancellous vertebrae [55, 75], calcaneus [52, 53, 75, 76], tibia [77] and femur [57, 77], the median for the squared correlation coefficient between SOS and site-matched bone quantity (*e.g.*, BV/TV) near the clinical range of frequencies is  $r^2 = 0.78$ . In these studies, the median for the added variance of SOS,  $r^2$ , explained by microstructure is 0.04.

#### F. SOS vs. Mechanical Properties

In studies on human cancellous vertebra [167], calcaneus [22, 86, 87], femur [49, 88] and tibia [49, 88] *in vitro*, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between SOS and mechanical parameters (*e.g.*, Young's modulus, ultimate strength) are  $r^2 = 0.46$ ,  $0.48 \pm 0.10$ . In studies in bovine [83, 111, 116] and ovine [132] femur, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between SOS and mechanical parameters are  $r^2 = 0.79$ ,  $0.71 \pm 0.20$ .

In order to investigate the sensitivity of SOS to elastic properties, SOS (1 MHz) was measured in human cancellous calcaneus cores before and after destructive testing that reduced elastic modulus without significant changes to porosity or microstructure [89]. SOS decreased by less than 0.25% despite decreases in elastic modulus greater than 75%. The investigators concluded that SOS is not directly sensitive to elastic properties of human calcaneus and therefore empirical correlations between SOS and elastic properties must be due to correlations among microstructure, porosity, elastic properties and SOS in undamaged cancellous bone [89, 90]. These experiments illustrate limitations in applying the bar equation to cancellous bone at clinical frequencies near 1 MHz [89].

In order to assess mechanical integrity of vertebral cancellous bone *in vivo*, investigators have developed a method to measure SOS from reflections from metallic pins inserted during surgery [168].

## IV. TWO-WAVE PHENOMENON

### A. Biot Theory and Evidence for Fast and Slow Waves

Biot theory predicts that fluid-saturated porous solids can support two longitudinal waves that travel with different velocities [158, 159]. The fast wave is associated with the fluid (blood and marrow) moving in phase with the solid (mineralized trabeculae), and the slow wave is associated with the fluid moving out of phase with the solid [169–172].

Many authors have demonstrated the existence of two longitudinal waves that may correspond to Biot's predictions in bovine cancellous bone in through-transmission experiments *in vitro*, as shown in Fig. 12. Bovine cancellous femur [173] and tibia [114] specimens *in vitro* can support a low-amplitude fast wave and a higher amplitude slow wave when ultrasound propagates approximately parallel to the predominant trabecular orientation. Similarly, human cancellous vertebrae can support fast and slow waves *in vitro* when propagation is along the craniocaudal axis [55]. Human cancellous femur can support fast and slow waves *in vitro* in the main load direction [122]. Temporal separation between fast and slow waves can be increased if alcohol is substituted for water as the filling fluid *in vitro* [174].

Properties of fast and slow waves are anisotropic [175, 176], with fast wave velocity achieving its maximum value when propagation is parallel to the predominant trabecular orientation [119, 123, 177], as shown in Fig. 12. Fast and slow waves can be identified even when cortical plates are attached to equine cancellous bone specimens *in vitro* [178, 179]. When ultrasound is transmitted through the radius *in vitro*, three waves can propagate and be distinguished: circumferential wave through radial cortex and surrounding soft tissue, fast and slow waves due to cancellous bone in radial interior [180, 181]. The evolution of fast and slow waves as they propagate through cancellous bone has been investigated *in vitro* by alternately performing through-transmission measurements and cutting thin slices from bone samples [182, 183], as shown in Fig. 13. Fast and slow waves have been observed in 3D printed cancellous bone phantoms based on  $\mu$ CT reconstructions of equine femur [184]. Detection of fast and slow waves can be improved through the use of coded excitation based on Golay code modulation [185].

Fast and slow waves have also been identified *in vivo*, in the human distal radius. It has been reported that fast and slow wave properties *in vivo* yield information regarding cancellous bone microstructure and elasticity [186–188].

There have many studies to test variants of Biot theory and related theories for prediction of fast and slow wave properties in cancellous bone. A complete accounting of these investigations [135, 170, 171, 173–175, 189–220] is beyond the scope of this paper, but a thorough review is available elsewhere [169].

Separation of fast and slow waves can be more difficult in human cancellous calcaneus than other species / skeletal sites mentioned above because it is less anisotropic (and separation is maximum when ultrasound propagates along a clearly-defined trabecular orientation). In addition, human calcaneus has a lower bone volume fraction than found in many animal

skeletal sites (and separation increases with BV/TV as shown in Fig. 8 in [122]). However, Biot theory accurately predicts the dependence of velocity on porosity in human calcaneus [221].

## B. Signal Processing for Separation of Fast and Slow Waves

Separate reconstruction of individual fast and slow waves is challenging when the two waves overlap in time and frequency domains. Several methods proposed for this decomposition, including Bayesian [146, 222–224], modified least-squares Prony's (MLSP) [225], space alternating generalized expectation maximization (SAGE) [226], MLSP plus curve fitting (MLSP+CF) [227, 228], and adaptive beamforming [229, 230] algorithms, are predicated on a model for the transfer function of the bone specimen that contains terms for fast and slow waves [156, 222]

$$Y(f) = X(f)[H_{fast}(f) + H_{slow}(f)] \quad (7)$$

where  $Y(f)$  and  $X(f)$  are complex amplitude spectra of the signals passing through bone and water-path-only respectively. For the Bayesian and MLSP+CF algorithms, the fast and slow wave transfer functions are

$$H_k(f) = A_k \exp[-\beta_k f d] \exp\left[\frac{i2\pi f d}{c_k(f)}\right] \quad (8)$$

where  $k$  can be “fast” or “slow.”  $A_k$  are frequency-independent wave amplitudes,  $\beta_k$  are slopes of attenuation coefficients (with respect to frequency),  $c_k(f)$  are phase velocities, and  $d$  is the sample thickness [154]. Causality implies that frequency-dependent phase velocities  $c_k(f)$  obey (6). The adaptive beamforming method assumes similar transfer functions  $H_k(f)$  that are augmented with phase rotation parameters that compensate for wave propagation through inhomogeneous media [229].

The Bayesian method maximizes the joint posterior probability for all the wave parameters (magnitudes, attenuation slopes, velocities) given the measured waveform using Markov chain Monte Carlo with simulated annealing [222]. The Bayesian method provides estimates not just of wave parameters but also of probability density functions for those parameters.

The MLSP method employs Prony's method, which fits a signal (in this case, the sum of fast- and slow-wave transfer functions) to the sum of complex exponentially-modulated sinusoids [225]. The MLSP method assumes nondispersive waves. However, if the outputs of the MLSP method are used as inputs to a curve-fitting routine (MLSP+CF method), then the solution can be constrained to obey the dispersion relation (6) given above [227]. Consequently, the MLSP+CF method has superseded the MLSP method.

The SAGE algorithm begins with “dictionaries” for amplitude and phase that are constructed from the Fourier transform of the reference signal and used to form a good initial guess for the solution. Then, an iterative nonlinear optimization scheme based on the Levenberg-Marquardt algorithm is used to obtain a final solution [226].

Generalized harmonic analysis begins by finding the frequency, amplitude, and phase of the sine wave that best matches the measured signal. It repeats this process on the residual and on subsequent residuals to generate a set of complex sine waves to approximate the measured signal [231].

The multi-channel instantaneous frequency method applies a filter bank to the measured signal and then computes the time-dependent phase of each filter output using a Hilbert transform. The instantaneous frequency is the time derivative of the phase. The multi-channel approach is more stable than the single channel approach when the waveform contains multiple components and/or background noise [232].

The adaptive beam forming method is a two-step process. First, initial estimates of fast and slow waves are performed using frequency-domain interferometry. Second, final estimates are obtained by performing least-squares fitting in the time domain [229, 230].

Although bandlimited deconvolution is not restricted to attenuation coefficients that vary linearly with frequency, it has recovered highly linear attenuation coefficients in bovine cancellous femur as reflected by average correlation coefficients between attenuation coefficients and frequency:  $0.997 \pm 0.002$  (fast wave) and  $0.986 \pm 0.013$  (slow wave) [233]. This finding provides substantial support for the two-wave model (8), which assumes attenuation coefficients that vary linearly with frequency.

There have not been many side-by-side comparisons of the various algorithms applied to the same experimental data. However, bandlimited deconvolution, MLSP+CF, and Bayesian methods gave similar results in bovine cancellous femur samples [228]. In addition, the Bayesian and MLSP+CF algorithms gave similar results in equine cancellous radius [146]. Simulations suggest that the relative performances of MLSP and SAGE depend on parameters of the experiment modeled, with MLSP having a comparative advantage in high signal-to-noise ratio (SNR) conditions and small temporal separation of waves but SAGE having a comparative advantage in low SNR conditions [226].

Most methods require less than 5 s on an ordinary laptop computer [142, 225, 226, 228, 229, 233]. The Bayesian method is more computationally intensive, requiring 100 min on a Sun Enterprise 250 dual 400-MHz workstation or 3 min using 32 processors of an SGI Altix 3000 with Itanium2 processors running at 900 MHz [222]. However, computation times would be expected to decrease as computer technology evolves. In addition, as mentioned previously, the Bayesian method provides estimates not just of wave parameters but also of probability density functions for those parameters.

## V. SCATTERING

### A. Metrics for Scattering

Comprehensive discussions of scattering from cancellous bone have been published previously [234, 235]. Here a more concise presentation (with some recent updates and added emphasis on mechanisms underlying scattering) is given. Fig. 14 shows acquisition of ultrasonic backscatter data from human calcaneus *in vivo* [236].

Scattering from cancellous bone arises from interfaces between the solid mineralized trabecular network and the fluid filler, which is marrow *in vivo* or water *in vitro*. Backscattering may be described quantitatively by the backscatter coefficient, which is defined as follows. If a plane wave  $P_{inc}(f)$  is incident upon a scatterer with backscattering amplitude  $\Phi_b(f)$ , then the scattered wave  $P_{scat}(f)$  measured a distance  $r$  away from the scatterer may be described by [237–241]

$$P_{scat}(r, f) = P_{inc}(f)\Phi_b(f)e^{ikr}/r \quad (9)$$

if  $kr \gg 1$ ,  $ka_s \ll 1$ , and  $a_s$  is the scatterer radius. The backscatter coefficient  $\eta(f)$  from a volume of unresolved scatterers is given by [241]

$$\eta(f) = n_0 |\Phi_b(f)|^2 \quad (10)$$

where  $n_0$  is the number of scatterers per unit volume. It has been assumed that the scatterers are positioned sufficiently randomly in space that phase differences between scattered signals from pairs of scatterers are uniformly distributed between 0 and  $2\pi$  radians. Backscatter measurements are inherently noisier than attenuation and sound speed measurements because they are impacted by varying degrees of constructive and destructive interference from signals scattered by randomly-positioned scatterers [242–244], as is the case with speckle noise in ultrasound B-mode scans [245, 246]. When the number of scatterers (*i.e.* trabeculae) per resolution cell is large enough (so that the central limit theorem applies), it can be shown that the envelope of the backscattered signal obeys a Rayleigh distribution with a characteristic mean to standard deviation ratio of 1.91 [245, 246], which is consistent with measurements in human calcaneus *in vivo* [247] and a model for scattering from thin cylindrical scatterers with randomly varying diameters [248].

Like attenuation coefficient and phase velocity, backscatter coefficient is an intrinsic material property and, if measured properly, exhibits minimal dependence on the measurement system. There are many methods for measurement of ultrasonic backscatter coefficient [237–241, 249, 250]. The average backscatter coefficient over a band of frequencies (usually centered about the transducer resonance frequency) is sometimes called broadband ultrasonic backscatter (BUB).

The backscatter coefficient provides insight into the size and distribution of scatterers. For spherical scatterers that are much smaller than the wavelength, the backscatter coefficient is proportional to frequency to the fourth power [102]. For cylindrical scatterers that are much thinner than a wavelength, the backscatter coefficient is proportional to frequency to the third power [251].

Although the backscatter coefficient is useful for elucidating physical mechanisms underlying the scattering process, it is difficult to measure *in vivo*. Other metrics may not be measurement-system-independent descriptions of scattering properties but still may provide useful diagnostic information. Scattering indexes may be obtained from the apparent backscatter transfer function (ABTF), which is the ratio of the spectrum of backscatter from a gated volume of cancellous bone to the spectrum of an echo from a planar reflector

(usually expressed in dB) [252]. The adjective “apparent” refers to the fact that the signal is not corrected for attenuation or diffraction [253]. Apparent integrated backscatter (AIB) is the average of the ABTF over a band of frequencies that usually corresponds to the usable band of the transducer. The frequency slope of apparent backscatter (FSAB) is the slope of a linear regression of ABTF vs. frequency [253]. AIB and FSAB depend on two time-gate parameters that affect the volume within the tissue that is selected for analysis: gate delay (from the beginning of the bone signal) and gate width, both of which may be optimized [88, 252, 254]. Autoregressive spectral estimation (instead of the Fast Fourier Transform) may be used to improve estimation of ABTF [255]. ABTF has been measured in human cancellous femur at frequencies from 0.6 – 15 MHz [256]. Usage of backscatter difference measurements (that is, differences in parameters derived from near and far windows in the gated backscatter signal) can reduce dependence on transducer properties, beam properties, and the effects of intervening tissues (soft tissue and cortical plates) [257–260]. The integrated reflection coefficient (IRC) is the mean of the frequency-dependent energy reflection coefficient over the range of frequencies corresponding to the usable frequency band of the transducer [49, 69, 83, 88, 261–263]. The backscattered spectral centroid shift (BSCS) describes the frequency downshift in backscattered signals due to the low-pass filter effect of attenuation in cancellous bone [264, 265].

The effects of overlying tissues on scattering metrics can be suppressed using dual-frequency ultrasound measurements [261, 262, 266–268]. The effects of the cortical shell on scattering metrics can be suppressed by estimation of and compensation for the integrated transmission coefficient through the cortex [269]. The presence of cortex has been reported to have a small effect on correlations between backscatter parameters and BMD in human femoral heads [270].

## B. Models for Scattering

While several models for scattering from cancellous bone have been proposed [271–278], two models have received far more experimental validation than the others: the incoherent scattering cylinder model [274] and the weak scattering model [275].

Two variants of cylinder scattering models, based on theory for scattering from a solid cylinder [279], were developed independently. One variant assumes coherent scattering from a two-dimensional array of regularly-spaced, parallel cylinders. Cylinder spacing and diameter could be estimated from measurements of scattering as a function of angle and frequency. Because only two or three bone samples were interrogated (one bovine and one or two human), it is difficult to assess the robustness of the model and method.

Another variant of the cylinder scattering model assumes that trabeculae are positioned sufficiently randomly that the incoherent contribution to scattering dominates the coherent contribution (*i.e.*, the phase differences between scattered signals from pairs of trabeculae are uniformly distributed between 0 and  $2\pi$  radians) [251, 274]. In the incoherent limit, the dependence of scattering on frequency and cylinder diameter is the same for a single scatterer or an ensemble of unresolved scatterers [251, 274]. When the cylinder diameter (mean value of 127  $\mu\text{m}$  in human calcaneal trabeculae [70]) is much smaller than a wavelength (about 3 mm at 500 kHz), the cylinder model predicts that backscatter should be



proportional to frequency cubed, which is consistent with measurements in the clinical frequency range ( $< 1$  MHz) as shown in Fig. 15. The cylinder model has been extended to include cylinders with finite lengths [280], quasi-periodic positions [281, 282], and randomly-varying diameters [248]. As mentioned in Section III. E, the cylinder scattering model also explains the dependence of phase velocity and dispersion on trabecular thickness and trabecular spacing in cancellous bone mimicking phantoms.

The incoherent assumption explains measurements of backscattered envelope mean-to-standard deviation ratio in human calcaneus *in vivo* of  $1.81 \pm 0.08$  (2.25 MHz) [247],  $1.92 \pm 0.12$  (580 kHz) [283] and  $1.73 \pm 0.12$  (1.3 MHz) [283], which are close to the theoretical Rayleigh distribution value of 1.91 for purely incoherent scattering [245, 246]. The fact that experimental values tend to be a little lower than 1.91 can be explained by a mixed model composed of long, thick trabeculae and short, thin trabeculae [283].

The other scattering model that has received extensive experimental validation is the weak scattering model [275, 284]. This model predicts the backscatter coefficient based on the structural autocorrelation function, which may be measured from cancellous bone samples using  $\mu$ CT. Like the incoherent cylinder model, the weak scattering model considers only the incoherent component of scattering [284]. This model has been extended to a 2-component form [285]. Substantial agreement between the incoherent scattering cylinder model and the weak scattering model in human cancellous femur *in vitro* has been reported [234, 235, 286].

The binary (marrow fat and bone matrix) mixture model [273, 276], which has received less experimental validation than the models discussed above, predicts a scattering coefficient that is proportional to mean fluctuations in velocity, neglecting contributions due to fluctuations in density as has been done with soft tissue scattering models [287, 288].

### C. Backscatter vs. Frequency

Fig. 15 shows means and standard errors for backscatter coefficient as a function of frequency from 16 human cancellous calcaneus samples *in vitro* [274]. Fig. 15 also shows the prediction of the incoherent scattering cylinder model. At low frequencies ( $< 1$  MHz), backscatter is approximately proportional to frequency cubed, as predicted by the incoherent scattering cylinder model [251, 274]. Several studies support this approximate cubic frequency dependence in human cancellous calcaneus *in vitro* [152, 274, 275, 289, 290] and bovine cancellous femur *in vitro* [291] at low frequencies. Values of the exponent of frequency dependence of slightly greater than 3 in human cancellous calcaneus may be explained by finite effective cylinder lengths [280], combined contributions from cylinders (trabeculae) and point-like scatterers (plates), multiple scattering [274], or some combination. The dependence of backscatter coefficient on frequency deviates from the cubic behavior for frequencies above 1 MHz [49, 274, 281], with some studies suggesting that backscatter coefficient from human cancellous calcaneus *in vitro* hits a plateau between 2 and 2.5 MHz [274, 281]. Studies in bovine cancellous bone *in vitro* suggest backscatter coefficient increases approximately monotonically with frequency below 1 MHz [281, 285, 291] and can be shown to be consistent with a binary mixture model [273, 276, 291].

Theoretically, the far field scattering response of a thin cylinder to an incident plane wave varies as frequency cubed when the wavelength is much bigger than the cylinder diameter [279]. An experimental study indicates that the theoretical far field scattering response from a single cylinder has the same dependences on frequency and diameter as the backscatter coefficient measured with a focused transducer from an ensemble of cylindrical scatterers [251].

The incoherent cylinder scattering model and the weak scattering model have been shown to exhibit very similar predictions of backscatter coefficient in 26 human cancellous femur samples *in vitro* [286].

The ABTF (in dB) usually decreases with frequency over the range from 0.6 MHz to 9.1 MHz in human cancellous femur *in vitro* [253], except when gate delay is very short (< about 1 or 2  $\mu$ s) [252].

#### D. Backscatter vs. BMD and Composition

In studies on human cancellous calcaneus [50–53, 247, 292, 293], femur [49, 56, 57] and tibia [49] *in vitro*, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between BUB and site-matched, volumetric BMD near the clinical range of frequencies are  $r^2 = 0.66$ ,  $0.65 \pm 0.13$ .

In human spine *in vivo*, the square of the correlation coefficient between the BSCS at 2.5 MHz and volumetric BMD has been reported to be  $r^2 = 0.37$  [294]. BSCS and AIB at 3.5 MHz and 5.0 MHz measured in human calcaneus *in vivo* exhibit squared correlation coefficients with areal BMD measured at the hip and spine in the range of  $0.41 < r^2 < 0.61$  [295, 296]. AIB measured in intact human proximal femur *ex vivo* exhibits squared correlation coefficient with areal BMD in the femoral neck of  $r^2 = 0.44$  [297].

Low correlations have been reported ( $r^2 = 0.01$  and  $r^2 = 0.23$ ) in transverse and longitudinal orientations between AIB and volumetric BMD in bovine cancellous tibiae *in vitro* near 1 MHz, but higher correlations ( $r^2 = 0.82$  and  $r^2 = 0.49$ ) have been reported in transverse and longitudinal orientations near 5 MHz [298]. In cases of significant correlation, AIB usually decreases with BMD, except when gate delay is very short (< 1  $\mu$ s) [252]. The opposite trends of backscatter coefficient and AIB as BMD increases may be attributable to the fact that AIB includes the effects of attenuation (which increases with BMD) while backscatter coefficient does not.

The correlation between BUB (200–600 kHz) and volumetric BMD in bovine cancellous femur *in vitro* has been reported to be  $r^2 = 0.37$  [83]. Correlations in the range  $0.48 < r^2 < 0.66$  between BUB and volumetric BMD in human cancellous femur and tibia *in vitro* have been reported for measurements with five transducers with center frequencies ranging from 0.5–5 MHz [49]. The correlation between BUB (1.5–3.8 MHz) and bone volume fraction (which, like BMD, is a good indicator of bone quantity) in human cancellous femur and tibia *in vitro* has been reported to be  $r^2 = 0.76$  [69].

One study found that demineralization (by treating bone with EDTA [58]) significantly reduced AIB in human cancellous tibia *in vitro* (1–3 MHz) [58] in the superoinferior (SI)

direction but not in mediolateral (ML) or anteroposterior (AP) directions *in vitro*. However, another study, in which no active demineralization was applied, found no significant correlation between BUB (2.25 MHz) and mineral content (normalized to the calcified matrix volume) in human cancellous tibia and femur *in vitro* [69]. The lack of significant correlation may have been partially due to the relatively low natural variation of mineral content in the samples (8.8%) [69].

One study found that decollagenization (by treating bone with sodium hypochlorite) significantly increased AIB (1–3 MHz) in all three directions in human cancellous tibia *in vitro* [58]. Another study, in which no active decollagenization was applied, found a significant negative correlation ( $r = -0.50$ ) between BUB (1–2.8 MHz) and calcified matrix collagen (normalized to the calcified matrix volume) in human cancellous tibia and femur *in vitro* [69], as shown in Fig. 16. A follow-up study found similar significant negative correlations ( $-0.46 > r > -0.75$ ) between BUB/AIB and calcified matrix collagen in human cancellous tibia and femur *in vitro* for transducers with center frequencies of 1, 2.25, 3.5, and 5 MHz [263].

One study found a significant negative correlation ( $r = -0.55$ ) between BUB (1–2.8 MHz) and fat content in human cancellous tibia and femur *in vitro* but no significant correlations between BUB and water content or proteoglycan content [69].

#### E. Backscatter vs. Microstructure

In multiple regression studies on human cancellous calcaneus [52, 53], tibia [77] and femur [57, 77] specimens *in vitro*, the median for the squared correlation coefficient between BUB and bone quantity (*e.g.*, BV/TV) near the clinical range of frequencies is  $r^2 = 0.63$ . In these studies, the median for the added variance of BUB,  $r^2$ , explained by microstructure is 0.07. Similar results have been reported for backscatter difference parameters in human cancellous femur *in vitro* at 3.5 MHz (although  $r^2$  values were sometimes higher when measurements were averaged over 6 orthogonal directions) [260] and multiple backscatter parameters in bovine cancellous femur at 2.25 MHz [299].

Fig. 17 shows a scatter plot of backscatter coefficient vs. mean trabecular thickness (Tb.Th) in 43 human calcaneus samples *in vitro*. Fig. 17 also shows a power law fit in which backscatter coefficient at 500 kHz is proportional to trabecular thickness to the 2.8 power. (95% confidence interval: 1.7 – 3.9). This dependence is very close to the prediction of the incoherent scattering cylinder model (2.9) [300]. The weak scattering model has been shown to be effective for estimating trabecular thickness from ultrasonic backscatter measurements in cancellous bone specimens [243, 244, 284].

#### F. Backscatter vs. Mechanical Properties

In studies on human cancellous calcaneus [22, 87], femur [49, 88, 263] and tibia [49, 88, 263] *in vitro*, the median and mean  $\pm$  standard deviation of the squared correlation coefficient between BUB and mechanical parameters (*e.g.*, Young's modulus, ultimate strength) are  $r^2 = 0.51, 0.46 \pm 0.09$ .

## G. Multiple Scattering

Scattering models that ignore multiple scattering are consistent with measurements of frequency dependence of backscatter in the clinical frequency range [274, 275]. Therefore, multiple scattering is usually assumed to be small compared to single scattering in this range.

However, indirect evidence for multiple scattering exists in the clinical frequency range. A theoretical model for multiple scattering from cylinders [160] has been shown to be accurate for predicting negative dispersion observed in the clinical frequency range in cancellous bone *in vitro* [21, 105, 148, 149], *in vivo* [150] and cancellous bone-mimicking phantoms [161, 162]. Another model provides additional insight into multiple scattering processes in cancellous bone [277, 278].

A theoretical model based on cylindrical trabeculae predicts that multiple scattering in cancellous bone is relatively small below 1.5 MHz but is substantial above 1.5 MHz [301]. Direct evidence for multiple scattering from human cancellous femur has been observed *in vitro* at 3 MHz [302]. Multiple scattering may be inferred from the angular dependence of backscattered intensity, which may be measured using a linear array. A single element is used for transmission while all elements are used for reception. The process is repeated using a different transmit element each time to acquire the complete dataset. At short times, the angular dependence of backscattered intensity is nearly flat, but at later times there is an enhancement in the backscattering direction that is a signature of multiple scattering [302]. Such measurements from human cancellous femur *in vitro* indicate scattering mean-free path between 2.3 and 8 mm [302]. The method may be used to measure diffusion constant, which characterizes the rate of growth of the diffusive halo due to multiple scattering. Measurements at 3 MHz in human cancellous femur *in vitro* suggest that the diffusion constant may have diagnostic value [303]. Simulations at 5 MHz in equine cancellous femur suggest that the diffusion constant is effective for quantifying anisotropy [117].

## H. Scattering as a Component of Attenuation

Potential sources of measured attenuation (*e.g.*, BUA or nBUA) include absorption, longitudinal-longitudinal (LL) scattering, longitudinal-shear (LS) scattering, and phase cancellation. These sources are not mutually exclusive. For example, some ultrasound energy can be initially scattered (LL or LS) at a trabecular interface and then subsequently absorbed as it propagates away from the trabecula. Phase cancellation was considered in Section II.F.

In the clinical frequency range ( $< 1$  MHz), LL scattering would not seem to be a significant component of attenuation [274, 304] because it is highly nonlinear with frequency [251, 274, 275, 284, 289, 304] while attenuation is quasi-linear with frequency [1, 41–43].

One finite difference time domain (FDTD) simulation solved the 2D viscoelastic wave equation with and without viscous loss terms set to zero and found little difference (4.4%) in attenuation (300-900 kHz) in the two cases for a human cancellous calcaneus sample subjected to computational erosions and dilations [305]. This result suggested that direct absorption is a small component of attenuation. Another FDTD simulation solved the 3D

linear elastic wave propagation equation without taking absorption into account and was able predict magnitude and frequency dependence of attenuation (0.4-1.2 MHz) for 31 human cancellous femur samples consistent with measurements on cancellous bone (*e.g.*, quasi-linear frequency dependence) [306]. This finding led the investigators to suggest that LS scattering could be a significant contribution to attenuation [306]. Subsequent paired comparison of this simulation approach with measurements on 28 human cancellous femur samples *in vitro* showed that the simulation correctly predicted experimental attenuation (0.4-1.2 MHz) values for low BV/TV, but tended to underestimate experimental attenuation magnitude and frequency dependence for higher BV/TV [307]. The authors concluded that the relative contribution of scattering to attenuation increases with frequency, becoming predominant (>50%) over absorption for frequencies above 600 kHz [307]. This simulation and another simulation (3D FDTD simulation based on 11  $\mu$ CT images of cancellous bone from human cadaver knees) suggest that the importance of absorption relative to scattering increases as BV/TV increases [307, 308].

If, in the clinical range of frequencies, 1) attenuation varies quasi-linearly with frequency, 2) scattering is a significant component of attenuation, and 3) LL scattering varies highly nonlinearly with frequency, then LS scattering must vary quasi-linearly with frequency and dominate LL scattering. In order to investigate these mechanisms experimentally, attenuation and backscattering were measured over a broad range of frequencies on cancellous-bone-mimicking phantoms containing nylon wires (simulating trabeculae) in a soft tissue-mimicking medium (simulating marrow) [309]. Frequency-dependent attenuation coefficients,  $\alpha(f)$ , were decomposed into three components as illustrated in Fig. 18:

$$\alpha(f) = \alpha_{FL}(f) + \alpha_{L2}(f) + \alpha_{NL}(f) \quad (11)$$

The first component,  $\alpha_{FL}(f)$ , corresponds to absorption in the fluid medium (*e.g.*, marrow). It varies approximately linearly with frequency and may be measured in a phantom containing only soft tissue-mimicking medium without nylon wires. The second linear component,  $\alpha_{L2}(f)$ , contains absorption in the wires (trabeculae) and LS scattering by the wires, which was hypothesized [309] to vary quasi-linearly with frequency based on previous studies of suspensions of particles in fluids [310–312] (see Fig. 8 in [313]). The second linear component,  $\alpha_{L2}(f)$ , may be measured by performing a linear fit to low-frequency attenuation measurements and then subtracting  $\alpha_{FL}(f)$  as shown in Fig. 18. The nonlinear component,  $\alpha_{NL}(f)$ , was hypothesized to be due to LL scattering [309].

Fig. 18 shows measurements of total attenuation ( $\alpha$ , left column), nonlinear attenuation ( $\alpha_{NL}$ , middle column), and LL backscatter coefficient ( $\eta$ , right column) in five phantoms with different nylon wire thicknesses [309]. For each row (*i.e.*, each phantom) in Fig. 18, the frequencies at which nonlinear attenuation (middle column) and LL backscatter coefficient (right column) become non-negligible are approximately equal (increasing from 1 MHz in the top row to 2.5 MHz in the bottom row), supporting the association of  $\alpha_{NL}(f)$  with LL scattering. The two functions are similar in shape but not identical because  $\alpha_{NL}(f)$  corresponds to the integral of LL scattering over all solid angles while LL backscatter corresponds only to LL scattering in the reverse direction. Linear regression analysis of low-frequency (clinical range) attenuation coefficient slope (due to  $\alpha_{FL}$  and  $\alpha_{L2}$ ) vs. volume

fraction occupied by nylon filaments yielded a correlation coefficient of  $r = 0.96$  (95% confidence interval: 0.82–0.99), supporting the relevance of the phantom model to cancellous bone [309].

Another experimental investigation into sources of attenuation compared attenuation measurements performed on 26 human cancellous femur samples *in vitro* with three different filling fluids: marrow, water, and alcohol [166]. No significant influence of the fluid choice on attenuation was observed despite a wide variety of fluid viscosity and acoustic impedance mismatch between fluid and trabeculae. This led the investigators to conclude that LS scattering and absorption in the trabeculae were candidates as main sources for attenuation [166].

Shear waves due to mode conversion at scatterer interfaces are likely to be transient. For example, shear waves generated from graphite particles suspended in gelatin are described as “evanescent” because they are quickly absorbed [313]. Similar rapid absorption is likely in cancellous bone. Shear attenuation coefficients in bovine cancellous bone are approximately 17 dB/inm at 1 MHz [314], which means that shear wave power decreases by 98% for each mm of propagation. Therefore, characterization of LS-scattering from trabeculae as an absorption mechanism is not unreasonable, although it overlooks a brief, highly-confined, transitional phase of energy in the form of a transient shear wave that propagates on the order of 1 mm prior to nearly-complete absorption [304].

## VI. ADDITIONAL TOPICS

### A. Anisotropy

QUS parameters depend on the orientation of ultrasound propagation relative to the predominant trabecular orientation. Regarding BUA and nBUA, results are mixed with one study in bovine radius [315] indicating higher BUA in the parallel orientation and studies in human vertebrae [55] and bovine tibia [114] indicating lower nBUA in the parallel orientation. Studies in bovine femur [111], human vertebrae [55, 167] and bovine tibia [114] consistently indicate that SOS is faster in the parallel orientation, as shown in Fig. 11. One study in human calcaneus [304] indicates that backscatter coefficient is higher in the perpendicular (ML) orientation. This is plausible because echoes tend to be stronger when ultrasound strikes a target from the perpendicular direction. In bovine tibia, the difference in AIB between parallel and perpendicular orientations has been found to be small, perhaps due to increases in both nBUA and backscatter coefficient in the perpendicular orientation cancelling each other out to produce little change in AIB [114]. In human distal femur, for short gate delays ( $< 2 \mu\text{s}$ ) AIB is significantly larger in the perpendicular orientation while for longer delays AIB is similar in perpendicular and parallel orientations [252].

In bovine femur, the fast wave speed has been found to be maximum along the main trabecular orientation [119, 216] while the slow wave speed was relatively isotropic [119]. The fast wave can exhibit significant refraction when the propagation direction is not coincident with the main trabecular alignment [176, 316]. The attenuation coefficient of the fast wave has also been found to be maximum along the main trabecular orientation [216].

## B. The Effect of Fluid Filler

For convenience, many *in vitro* experiments are conducted in defatted (marrow removed) cancellous bone samples immersed in water. This raises the question of the effects of substituting water for marrow on QUS parameters. Measurements of nBUA and SOS in human cancellous calcaneus / femur at 1 MHz suggest that replacing marrow with water has anywhere from no statistically significant effect on either parameter [22, 112] to a mean decrease of nBUA of 5.6 dB/cmMHz [317] and a mean increase of SOS of 20-43 m/s [166, 317]. Measurements in bovine cancellous femur at 500 kHz and 1 MHz suggest a mean decrease of nBUA of 2 - 10 dB/cmMHz and mean increases of SOS of 35 – 48 m/s [120, 133, 318]. Measurements in bovine tibia at 2.25 MHz suggest no significant change in either parameter [115]. A simulation study suggests that these effects depend on BV/TV [308]. Replacing marrow with water was found not to have a big effect on dispersion or frequency dependence of backscatter in bovine cancellous femur [152]. The ultrasonic properties of human [319] and bovine [320] marrow samples have been studied extensively.

## C. Cancellous Bone-mimicking Phantoms

Experiments on bone-mimicking phantoms are useful for modeling the interaction between ultrasound and cancellous bone. An early phantom design, based on epoxy mixed with cubic granules of gelatin, exhibits nBUA values consistent with the clinical range and negative dispersion similar to cancellous bone [321, 322].

Phantoms consisting of light-cured resin manufactured using stereo lithography, with silicone rubber as a marrow mimic, exhibit values of BUA and SOS consistent with cancellous bone [194, 323]. Water-saturated stereo-lithographical bone replicas of cancellous bone have been used to perform measurements of through-transmission ultrasound with values similar to predictions based on modified anisotropic Biot-Allard theory for porous media [204, 205]. Three-dimensional printed phantoms generated from synchrotron x-ray  $\mu$ CT images of equine cancellous femur have been interrogated at 1 MHz and exhibit both fast and slow longitudinal waves [184].

Phantoms consisting of alternating parallel layers of water and polystyrene (simulating marrow and trabecular material) exhibit negative dispersion similar to cancellous bone and consistent with the analytic stratified model [165].

Nylon wires exhibit frequency-dependent scattering similar to cancellous bone [251]. Phantoms consisting of parallel nylon wires in two-dimensional arrays immersed in water exhibit dependences of phase velocity and dispersion on bone volume fraction similar to cancellous bone [161]. Phantoms that use nylon wire segments in random orientations immersed in tissue-mimicking gel also exhibit dependences of phase velocity and dispersion on bone volume fraction similar to cancellous bone [162]. Phantoms that use nylon wire segments in random orientation immersed in tissue-mimicking gel are useful for elucidating the relative roles of absorption, LS scattering, and LL scattering in cancellous bone [309].

Polyacetal cuboid bone-mimicking phantoms exhibit dependences of phase velocity and nBUA on porosity and dependence of attenuation coefficient on frequency consistent with cancellous bone and cancellous-bone-mimicking phantoms [324].

Water-saturated metal foams have been shown to be useful for mimicking some properties of cancellous bone. Aluminum foams (see Fig. 19) can exhibit slow wave phase velocity consistent with Biot theory [325–327]. A study on one set of aluminum foams indicates that SOS increases with porosity, but nBUA decreases with porosity [328]. Studies on nickel foams [329] and copper foams [330] indicate several properties that are consistent with cancellous bone: attenuation coefficient varies approximately as frequency to the first power [1], phase velocity exhibits negative dispersion [21, 105, 148–150], and backscatter coefficient varies approximately as frequency cubed [274, 275]. Water-saturated polymer foams exhibit values of nBUA, SOS, and backscatter parameters consistent with cancellous bone [118].

Phantoms consisting of a custom composite material using an epoxy resin, alumina powder, and inclusions (poppy seeds or hemp seeds) to control porosity can exhibit BUA and SOS in the range of values reported for cancellous bone [331].

#### D. Simulations

Simulations are helpful for elucidating mechanisms underlying the interaction of ultrasound with cancellous bone. A previous review discusses simulation examples from cortical bone, cancellous bone, whole bones, skull, and therapeutic applications [332]. The present section provides developments since the previous review and places greater emphasis on cancellous bone.

A pioneering FDTD simulation based on the 2D elastic wave equation predicted dependences of ultrasound velocity and mean frequency in cancellous human calcaneus (derived from  $\mu$ CT) on bone volume fraction consistent with experimental results [134]. The same simulation was used to investigate the relative roles of absorption and scattering in cancellous human calcaneus [305]. A 3D version of this simulation was used to validate net time delay (difference between transit time through cancellous bone and transit time through hypothetical object of equal thickness containing soft tissue only) as an effective index of BMD [333]. The 3D version has also been used to investigate the effect of bone marrow on attenuation and speed in cancellous bone [308], Fig. 20 shows snapshots of waves propagating through cancellous bone, showing ballistic and scattered components.

FDTD simulation based on 3D linear elastic wave propagation predicts several phenomena in cancellous human femur samples (derived from synchrotron  $\mu$ CT) that are consistent with experimental evidence: attenuation varies approximately linearly with frequency, nBUA and SOS increase with bone volume fraction, and most samples exhibit negative dispersion [306, 334]. It can reveal fast and slow waves when the ultrasound propagation direction is aligned with the predominant trabecular orientation. It predicts that mode conversion of incident longitudinal waves to shear waves is a significant contributor to attenuation [306] and that the dependence of nBUA on BV/TV can be mostly explained by scattering [307]. Using image processing to induce “virtual osteoporosis” in  $\mu$ CT data, it predicts that nBUA and SOS are mostly determined by volume fraction but that material properties and microstructure also play roles [131, 335]. 3D FDTD simulations indicate that except when BV/TV is high, variations of BUA induced purely by changes in BV/TV exceed technique imprecision and therefore can be detected [336]. However, variations of QUS properties



induced by changes in compressive or shear stiffness are more difficult to model due to sparse description of elastic properties at the tissue level [336].

A one-dimensional finite difference approach has been applied to model velocity in cancellous bone [337], but this approach has some limitations [338].

FDTD simulation based on the elastic wave equation has been studied extensively on bovine cancellous bone models (usually measured with  $\mu$ CT), which are more anisotropic than human cancellous bone and therefore elucidate the effects of trabecular orientation. In two-dimensions, FDTD simulation based on Biot theory is better for identifying fast and slow waves than FDTD simulation based on the elastic wave equation because of the limitations of the 2D elastic model [314]. However, viscoelastic FDTD simulation has closer agreement to experiment than Biot's FDTD for single wave propagation perpendicular to the trabeculae [339]. The effects of porosity on amplitude and speed of fast and slow waves have been elucidated by performing 3D FDTD solution of the viscoelastic wave equation on 3D  $\mu$ CT images of cancellous bone subjected to varying degrees of erosion [340, 341] or, alternatively, with a simplified model for cancellous bone consisting of spherical pores in otherwise solid bone [342]. Similarly, the effects of orientation of the ultrasound beam relative to the trabecular direction has been investigated by digitally rotating the  $\mu$ CT cancellous bone image [316]. This FDTD method suggests that reflection coefficients of fast and slow waves at the boundary between cancellous bone and cortical bone increase with porosity [343]. Finally, this approach has been used to characterize reflected and backscattered waves from cancellous bone [344, 345].

FDTD simulation based on the 3D elastic wave equation shows that the attenuation of the fast wave is higher in the early state of propagation and gradually decreases as the wave propagates in bovine cancellous bone [183]. It also predicts dependences of fast wave speed and fast and slow wave amplitudes on bone volume fraction similar to measurements in bovine cancellous bone [346, 347]. It has been used to investigate the effect of surrounding cortical bone on fast and slow waves propagating through equine cancellous radius [178, 179].

A 3D mixture model, in which each point in the cancellous bone has both fluid and solid phases coexisting has been used to predict attenuation monotonically increasing with frequency as has been observed experimentally [348].

3D FDTD simulation has been used to model and compensate for the effects of the cortical layer on cancellous bone backscatter metrics [269].

3D FDTD simulation using randomly distributed clusters of ellipsoidal scatterers in a fluid can predict the propagation of fast and slow waves, as in Biot theory [349]. Under the independent scattering approximation (ISA), this approach successfully predicts the attenuation coefficient (unlike Biot's theory) and the existence of negative dispersion [215]. However, the ISA does not model wave speeds in two-wave propagation as well as Biot's theory [215].

3D Finite element modeling (FEM), in which only the solid part of the bone was considered (*i.e.*, vacuum filler), suggests that the bar equation successfully predicts *SOS* near 50 kHz but does not perform as well at a more clinical frequency of 1 MHz [125]. 3D Finite element analysis of models based on a weak variational formulation of the equations of motions in solid and neighboring fluid media that simulate elastic scattering, refraction, and mode conversion have been shown to predict the linear dependence of attenuation with frequency and increases in nBUA and *SOS* with BV/TV consistent with reported experimental measurements for cancellous bone [350]. FEM that considered reflection, refraction, elastic scattering and mode conversion has been shown to be accurate for predicting frequency-dependent attenuation and phase velocity of water-saturated aluminum foam cancellous-bone-mimicking phantoms [351]. FEM based on Biot's model has been used to investigate anisotropy of reflection and transmission of plane waves in a human cancellous bone specimen [203].

A 2D pseudo-spectral time domain numerical model has been developed to investigate effects of microcracks in cancellous bone [352].

Nonlinear propagation through cancellous bone has been modeled using the Khokhlov–Zabolotskaya–Kuznetsov (KZK) equation and suggests challenges associated with detecting harmonics of the fundamental frequency [353].

## E. Acoustic Microscopy

Discussions of acoustic microscopy for both cortical and cancellous bone may be found elsewhere [11, 354], but the present discussion focuses on applications in cancellous bone. Scanning acoustic microscopy (SAM) is possible at frequencies of 0.1-1 GHz. Due to high absorption at such high frequencies, only surface or subsurface images are obtained [355]. For a flat sample, the reflected signal depends on the reflection coefficient at the sample surface  $r = (Z_2 - Z_1) / (Z_2 + Z_1)$  where  $Z = \rho v$ ,  $\rho$  is the material density,  $v$  is the longitudinal velocity, and the indexes 1 and 2 refer to the sample medium and coupling medium. A SAM system may be calibrated by scanning a set of materials with known physical properties so that both density and elasticity may be obtained [355,356].

SAM has been used to measure Young's moduli of human trabeculae of 17.5 GPa (400 MHz, femur) [355,357] and 19.9 GPa (100 MHz, pelvis) [356]. Both of these values are close to the measurement obtained with nanoindentation, 18.1 GPa [355]. SAM at 200 MHz has been used to measure Young's moduli of murine trabeculae of  $12.9 \pm 2.0$  GPa and  $17.7 \pm 1.4$  GPa in B6 and C3H mice respectively [358].

SAM at 200 MHz was used to study the effects of dynamic compressive strain loading in bovine cancellous sternum and ulna [359], Fig. 21 shows a close relationship between acoustic impedance measured with acoustic microscopy and degree of bone mineralization measured with synchrotron radiation  $\mu$ CT.

Ultrasound longitudinal wave velocity in an individual trabecula may be inferred from measurements of Brillouin scattering, which is the interaction between light and thermally excited acoustic phonons [360]. Micro-Brillouin scattering has been used to measure

longitudinal wave velocities in bovine femoral trabecula with a spatial resolution of 10  $\mu\text{m}$  and to study the dependence of elastic properties on trabecular type (rod-like vs. plate-like) and direction of trabecular alignment [360, 361]. The squared correlation coefficient between acoustic impedance measured with 200 MHz SAM and longitudinal wave velocity measured with micro-Brillouin scattering has been reported to be  $R^2 = 0.63 - 0.67$  [362].

The effects of sample preparation for acoustic microscopy have been characterized [363].

## F. Nonlinear Properties of Cancellous Bone

Linear acoustics is valid at low pressures and based on an equation of state for which variations of pressure are directly proportional to variations in density. At higher pressures, the quadratic term in a Taylor series for the equation of state cannot be neglected. The ratio of quadratic to linear terms in the equation of state is denoted by B/A, which may be used to describe nonlinear behavior of media [364].

The ratio of the amplitudes of second harmonic relative to the fundamental (236 kHz) has been shown to be sensitive to BMD in through-transmission measurements in human calcaneus *in vivo* in seven volunteers [365]. Correlation coefficients of B/A with apparent bone density ( $r = 0.95$ ), BMD ( $r = 0.77$ ), BV/TV ( $r = 0.80$ ), and Tb.Th ( $r = 0.44$ ) have been reported in bovine cancellous femur (0.5 MHz transmitter, 0.5 or 1 MHz receiver) [366–368]. Similar findings have been reported in aluminum-foam cancellous-bone-mimicking phantoms [328].

Vibroacoustography is a nonlinear imaging method that has been demonstrated to be sensitive to porosity of phosphocalcic ceramic samples (that simulate bone) and also to be capable of generating images of human calcaneus *in vitro* that reveal spatial distribution of mineralization [369]. Nonlinear measurement methods that use a mechanical vibrator to shake samples during acoustic measurement have been shown to be sensitive to bone density in human calcaneus samples *in vitro* [370] and to distinguish between healthy and osteoporotic human cancellous femur samples *in vitro* [371].

## VII. CONCLUSION

Experimental, computational, and theoretical analysis elucidates mechanisms underlying the interaction between ultrasound and cancellous bone.

Attenuation at clinical frequencies in cancellous bone is primarily determined by bone quantity but is also influenced by composition, microstructure, and mechanical properties. Attenuation usually varies quasi-linearly with frequency in the clinical range. Phase cancellation (especially for denser bones) and longitudinal-shear scattering make important contributions to attenuation at clinical frequencies.

Speed of sound at clinical frequencies in cancellous bone is primarily determined by bone quantity but is also influenced by composition, microstructure, and mechanical properties. Common speed of sound metrics, which entail measurements of transit times of pulse leading edges (to avoid multipath interference), are greatly influenced by attenuation, dispersion, and system properties including center frequency and bandwidth. A formula has

been derived to accurately account for these distortions. Unlike soft tissue, which exhibits positive dispersion, cancellous bone tends to exhibit negative dispersion, which may be understood using theoretical and phantom models.

Cancellous bone supports two longitudinal waves (“fast” and “slow” waves), as predicted from poro-elasticity theory. Fast wave velocity is highest when ultrasound propagates parallel to the predominant trabecular orientation. Signal processing methods have been developed to separate fast and slow waves in cancellous bone even when they overlap in time and frequency domains.

Backscatter at clinical frequencies in cancellous bone is primarily determined by bone quantity but is also influenced by composition, microstructure, and mechanical properties. Backscatter coefficient from cancellous bone at clinical frequencies varies approximately as frequency to the third power and mean trabecular thickness to the third power, which may be explained by cylinder or weak-scattering models.

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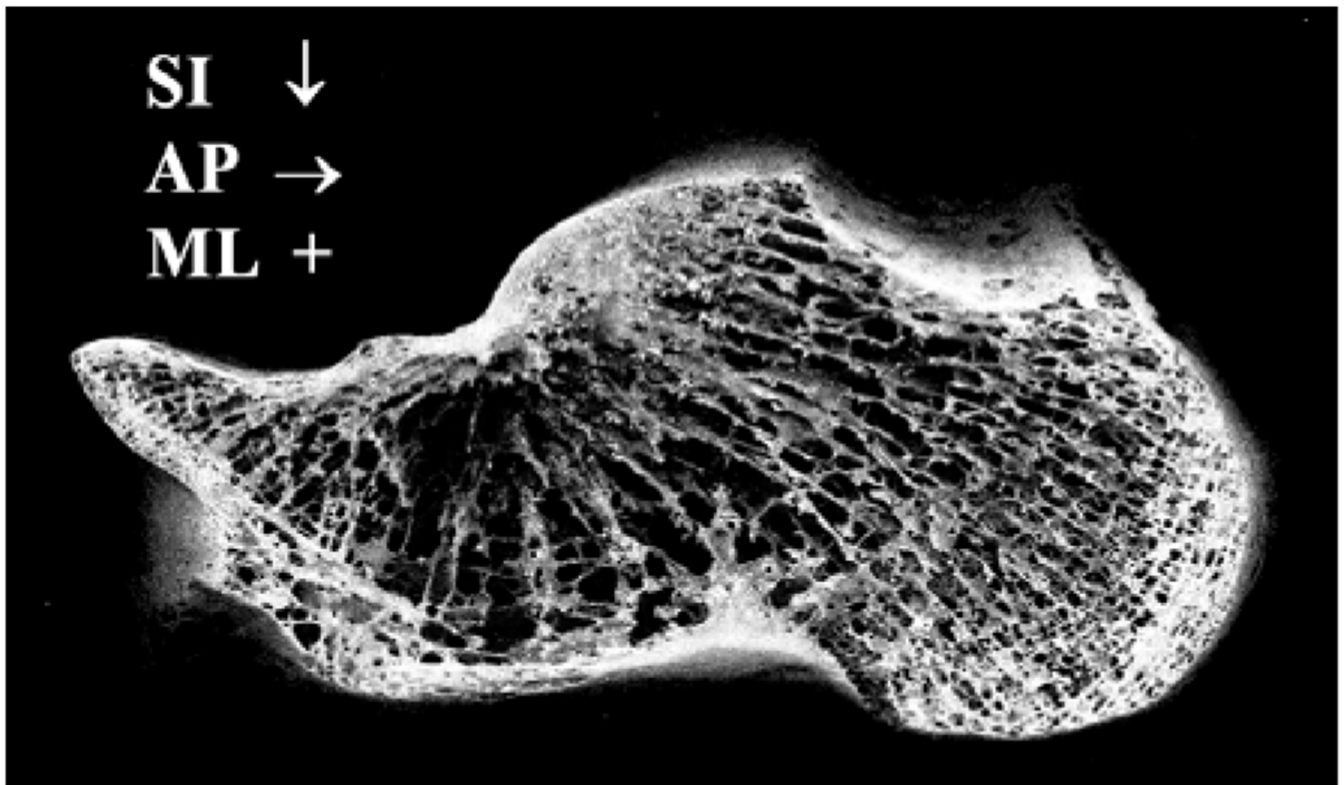
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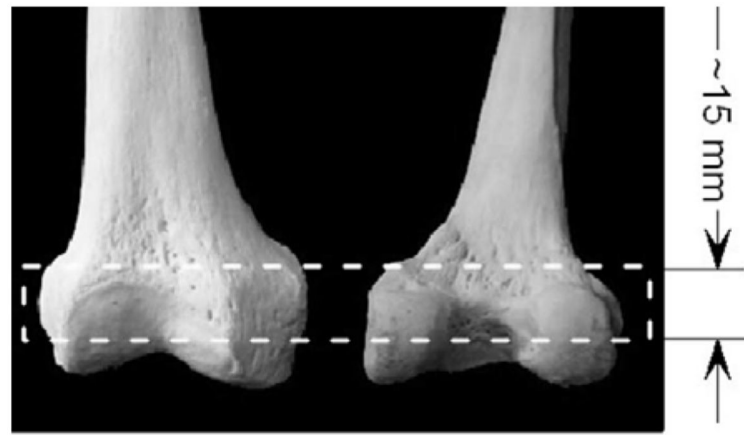
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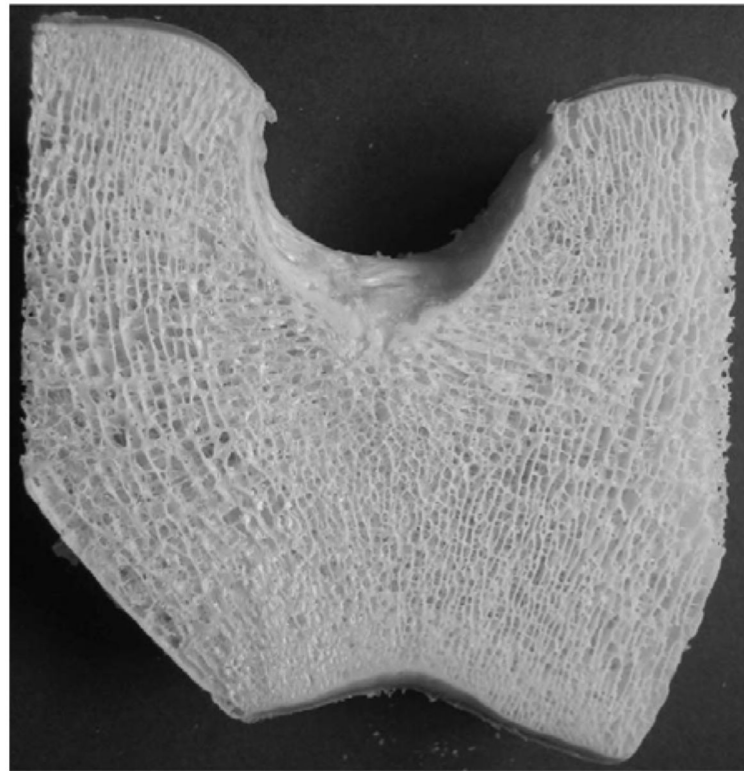


**Fig. 1.**  
Human calcaneus with lateral cortical endplates removed. Three orientations for ultrasound propagation are shown: superoinferior (SI), anteroposterior (AP), and mediolateral (ML). The most common orientation for ultrasound measurements *in vitro* and *in vivo* is ML.





(a)



(b)

**Fig. 2.** (a) human femurs. (b) cut slice showing cancellous bone corresponding to dashed box in (a). Reprinted with permission from M. Pakula *et al.*, Influence of the filling fluid on frequency-dependent velocity and attenuation in cancellous bones between 0.35 and 2.5 MHz, *J. Acoust. Soc. Am.*, 126, 3301-3310, 2009. Copyright 2009, Acoustical Society of America.

Power fit Attenuation= $-0.078+4.64 f^{1.49}$   $R^2 = 0.995$

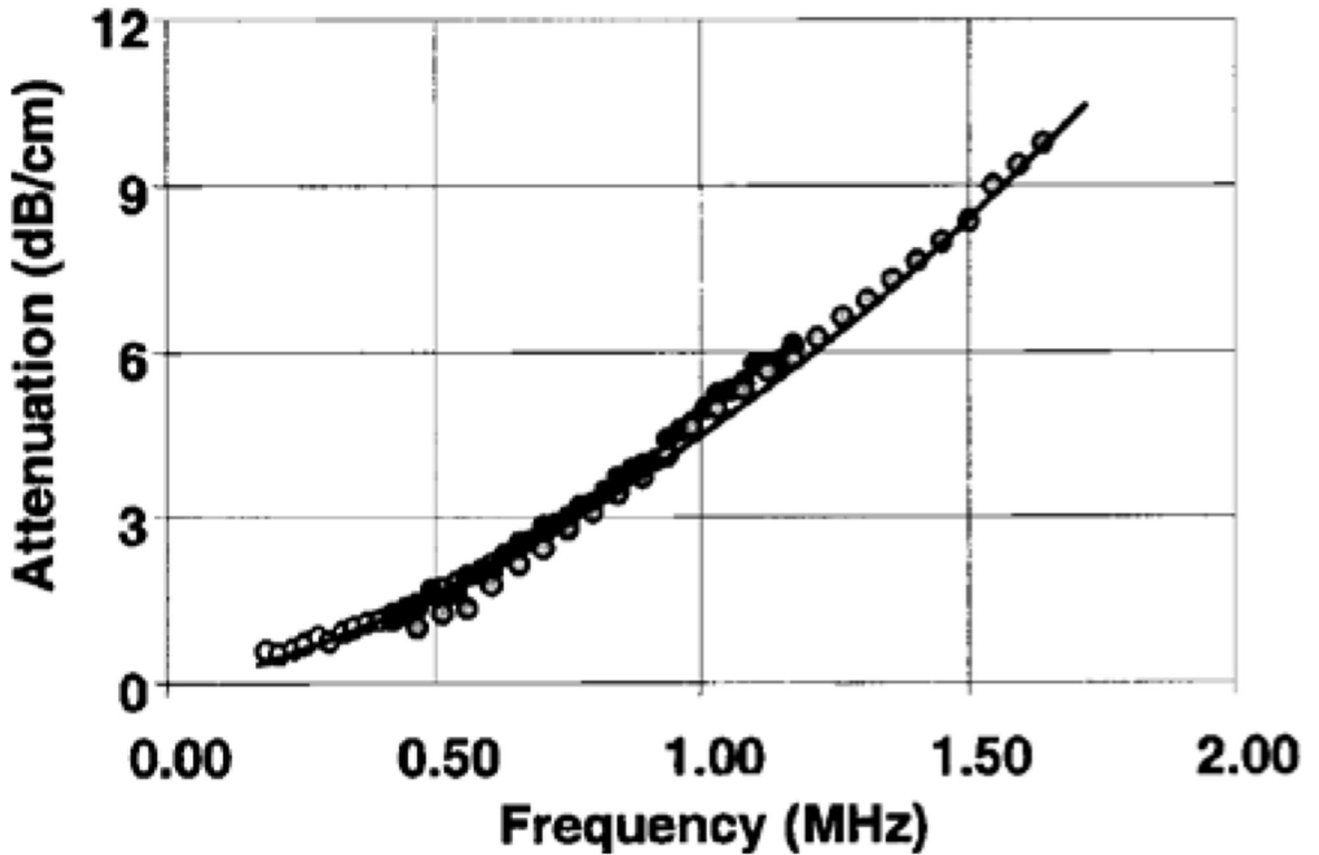
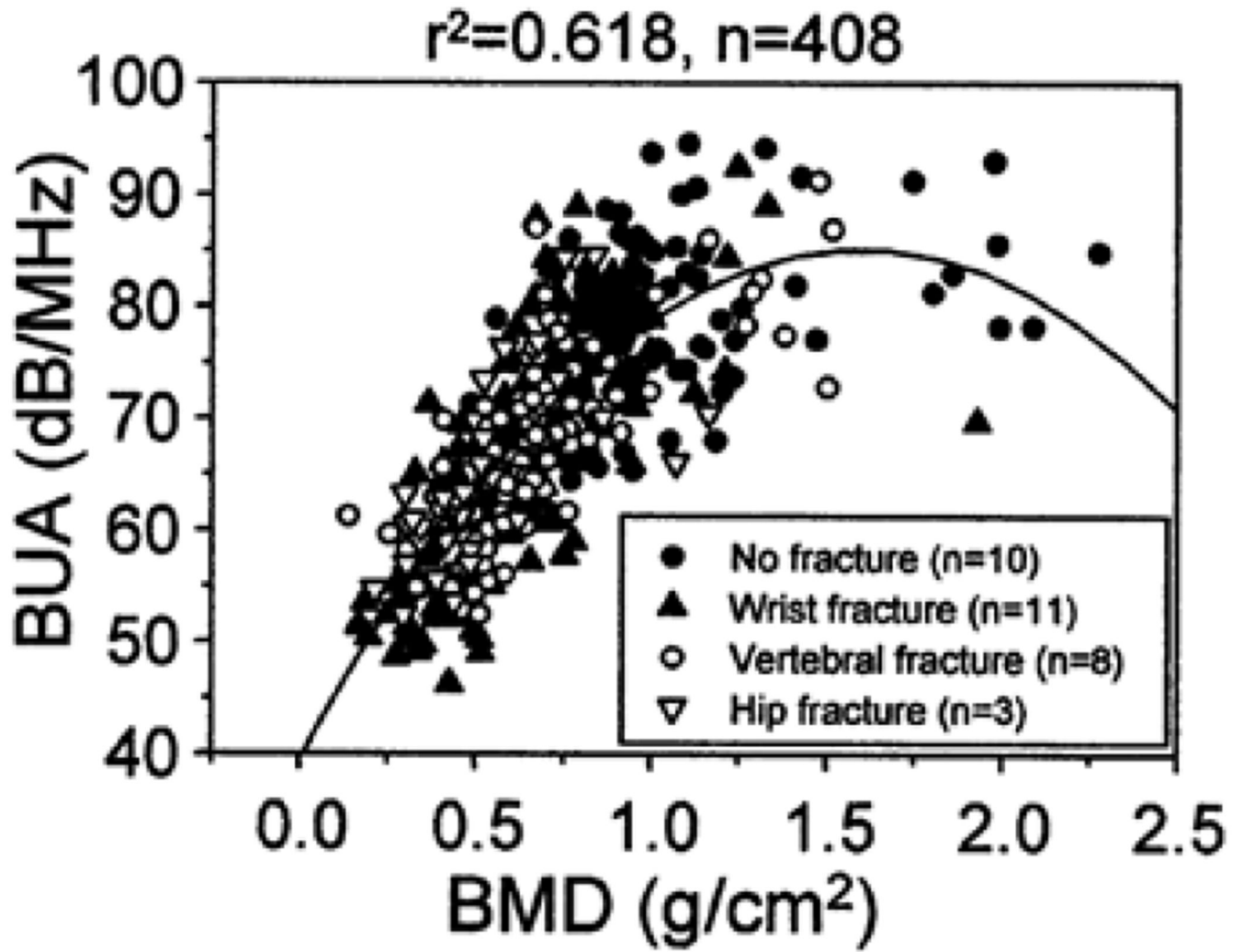


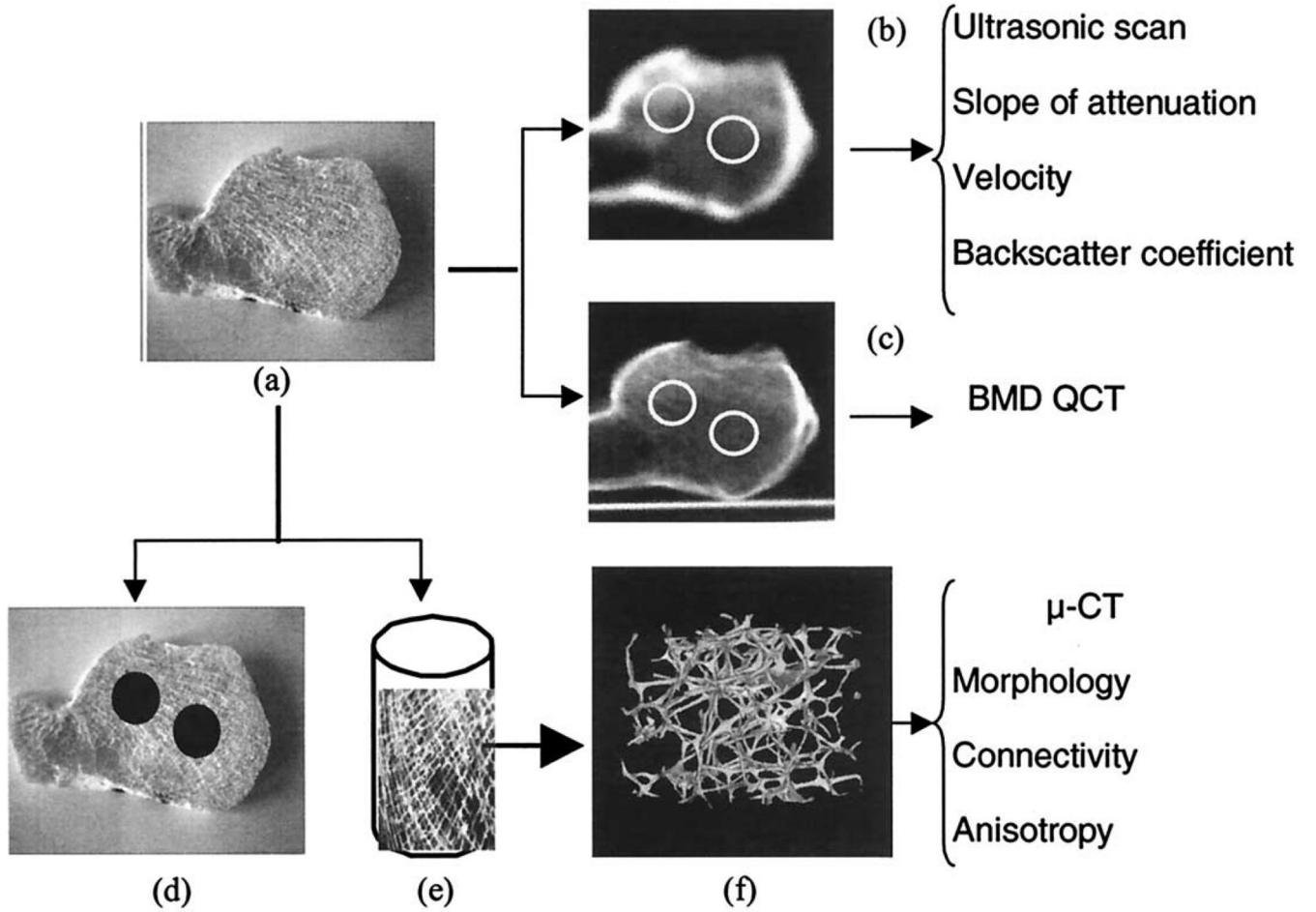
Fig. 3.

Attenuation coefficient of human cancellous calcaneus *in vitro* from 0.2-1.7 MHz. Although attenuation seems linear with frequency over a clinical bandwidth (*e.g.*, 300-700 kHz), it may be nonlinear over a broader bandwidth. Reprinted with permission from S. Chaffai *et al.*, In vitro measurement of the frequency-dependent attenuation in cancellous bone between 0.2 and 2 MHz, *J. Acoust. Soc. Am.*, 108, 1281-1289, 2000. Copyright 2000, Acoustical Society of America.



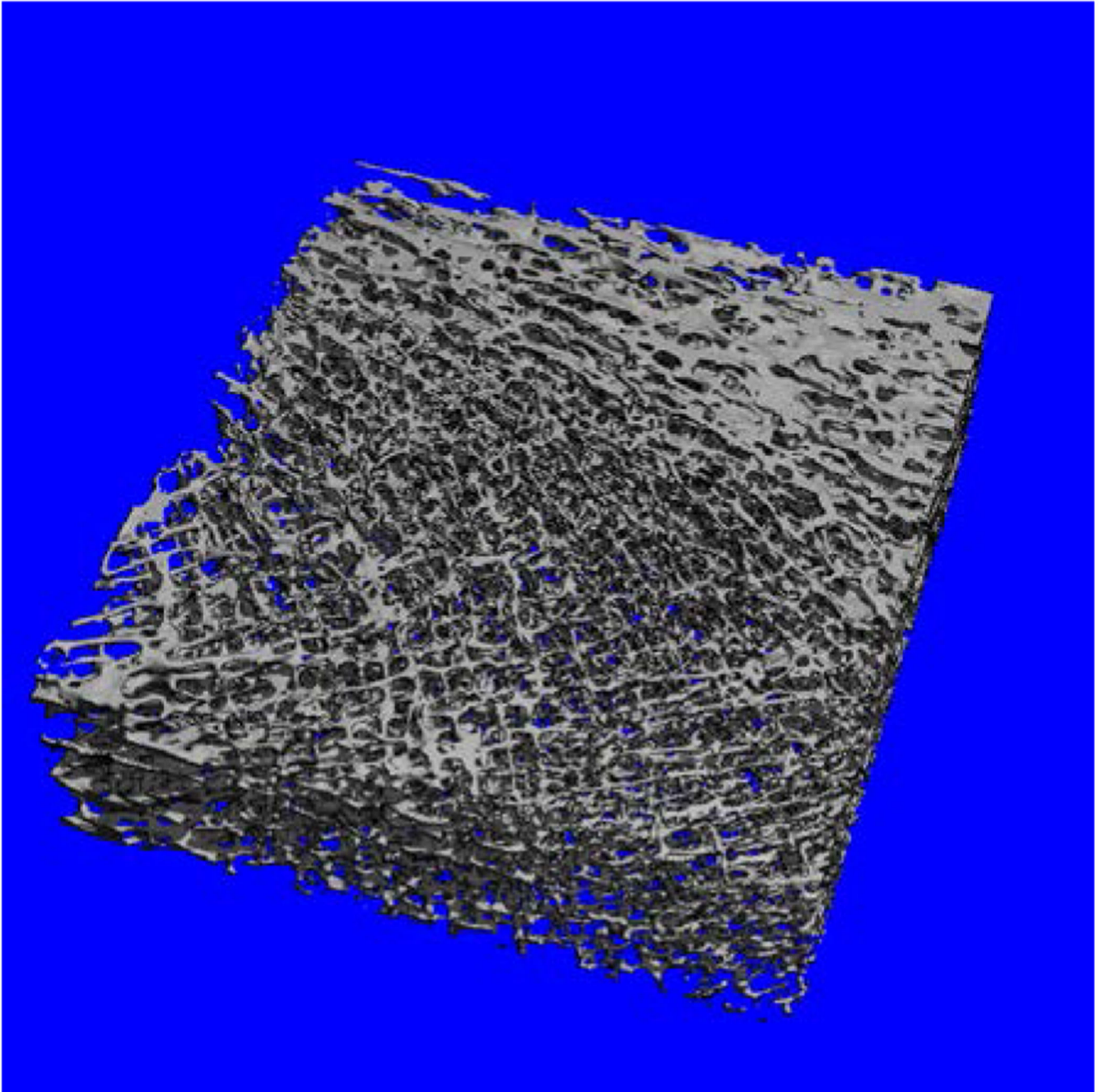
**Fig. 4.**

BUA vs. areal BMD in human calcaneus *in vivo*. Reprinted, with permission from *Bone*, 31, Toyras *et al.*, Bone mineral density, ultrasound velocity, and broadband attenuation predict mechanical properties of trabecular bone differently, 503-5-7, Copyright (2002), with permission from Elsevier.

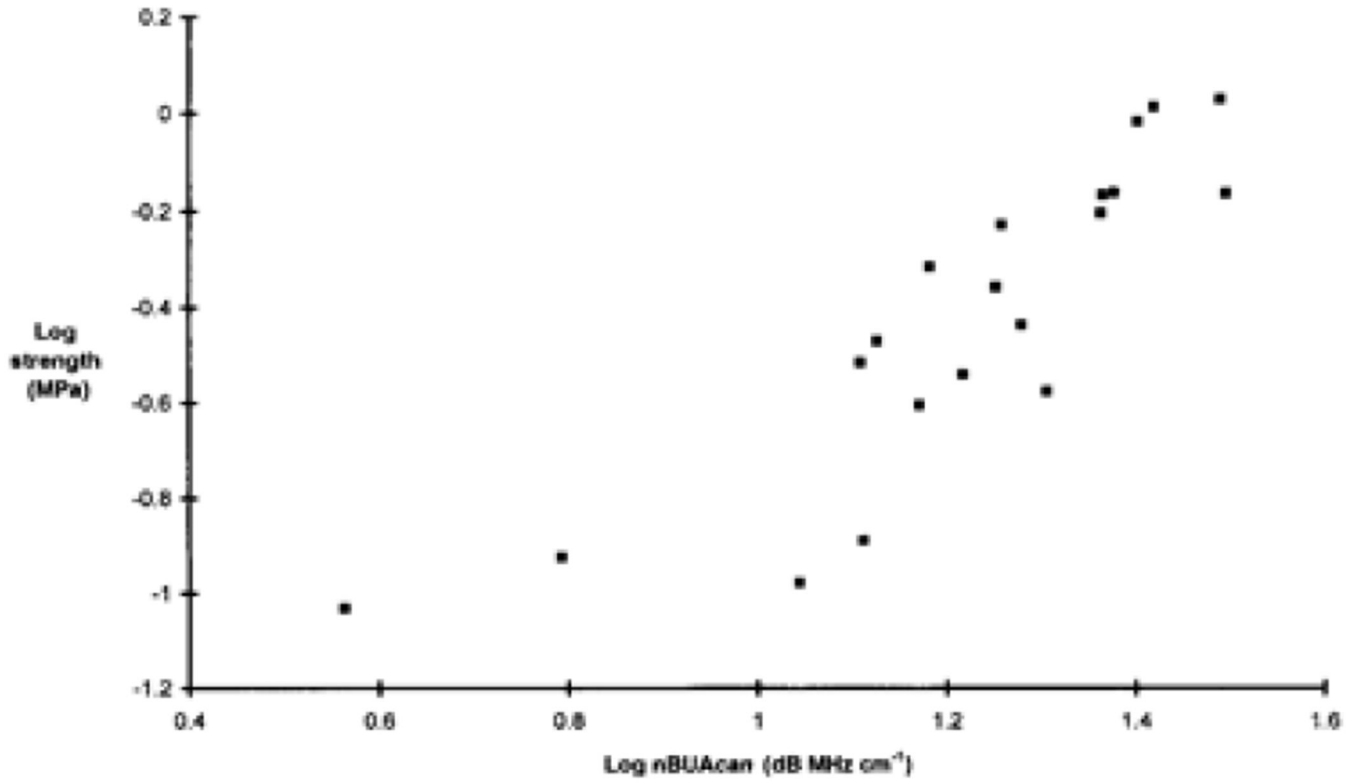


**Fig. 5.**

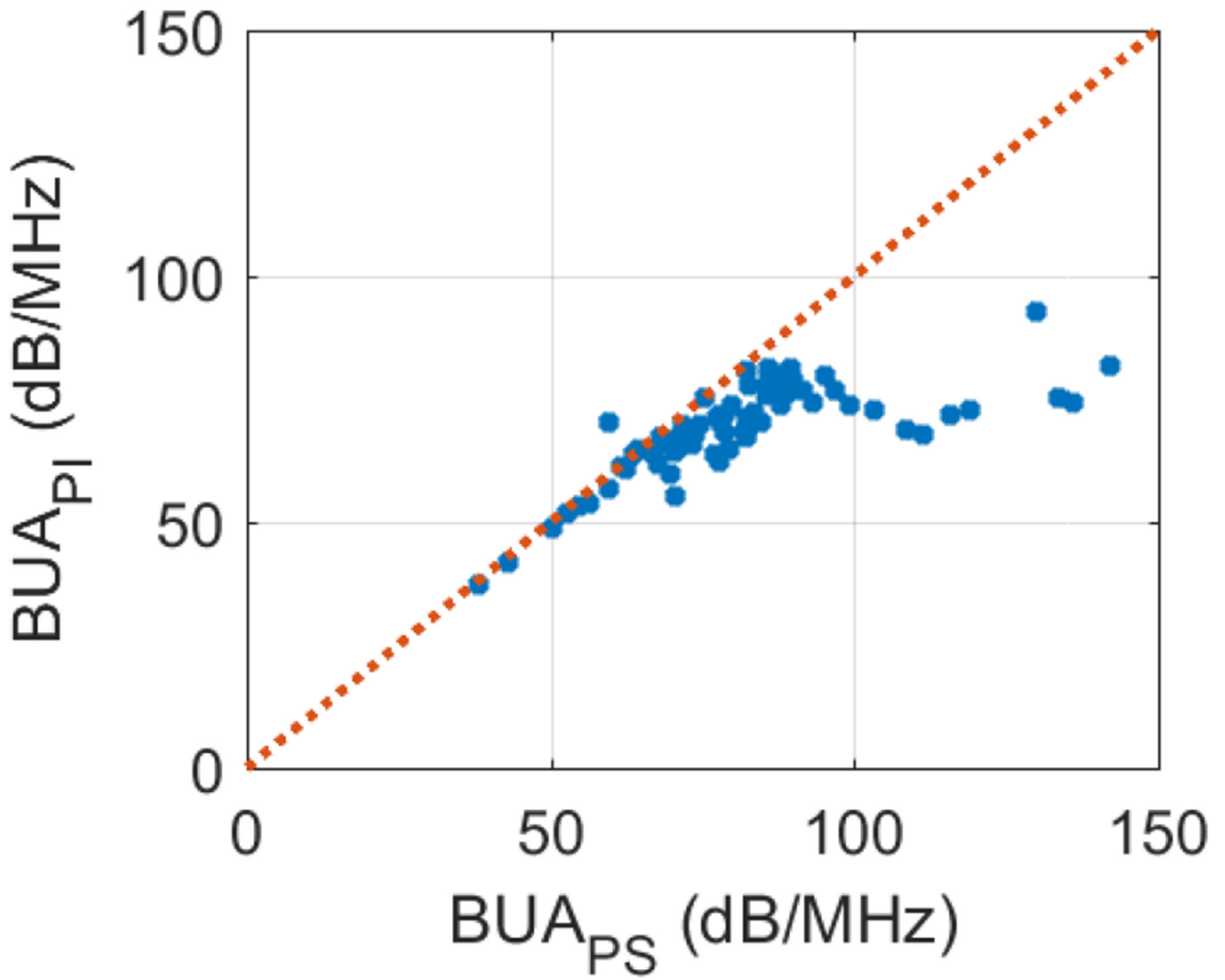
Steps for ultrasound and  $\mu$ CT analysis of cancellous calcaneus. Reprinted from *Bone*, 30, S. Chaffai *et al.*, Ultrasonic characterization of human cancellous bone using transmission and backscatter measurements: relationships to density and microstructure, 229-237, Copyright (2002), with permission from Elsevier.



**Fig. 6.**  
 $\mu$ CT reconstruction of rectangular volume from human calcaneus with cortical endplates removed. © 2003 IEEE. Reprinted, with permission, from K. A. Wear and A. Laib, The dependence of ultrasonic backscatter on trabecular thickness in human calcaneus, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 50(8). 979-986. 2003.

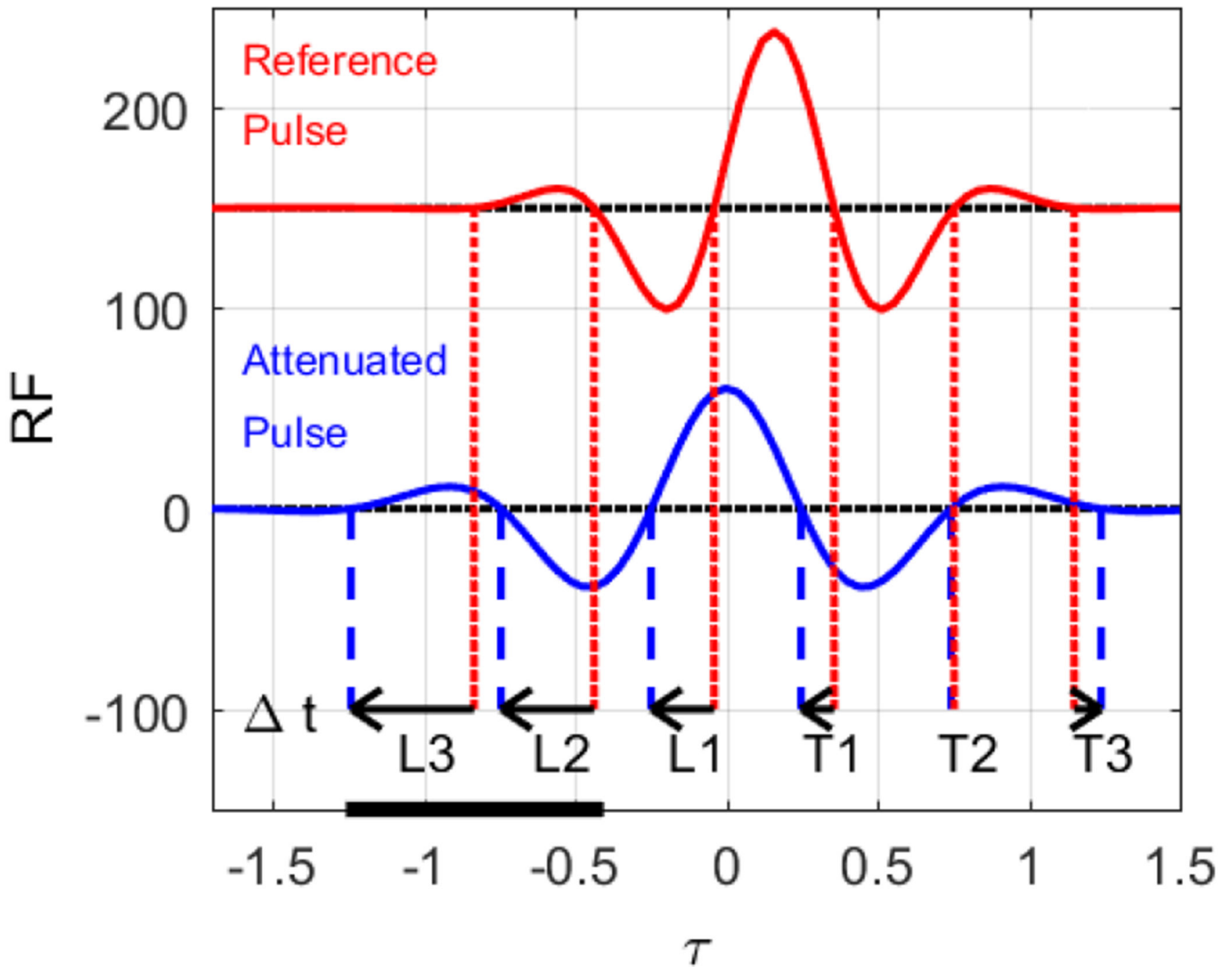


**Fig. 7.** Log strength vs log nBUA in human calcaneus *in vitro*. Reprinted with permission from *Bone*, 18, C. M. Langton *et al.*, Prediction of mechanical properties of the human calcaneus by broadband ultrasonic attenuation, 495-503, Copyright (1996), with permission from Elsevier.



**Fig. 8.**

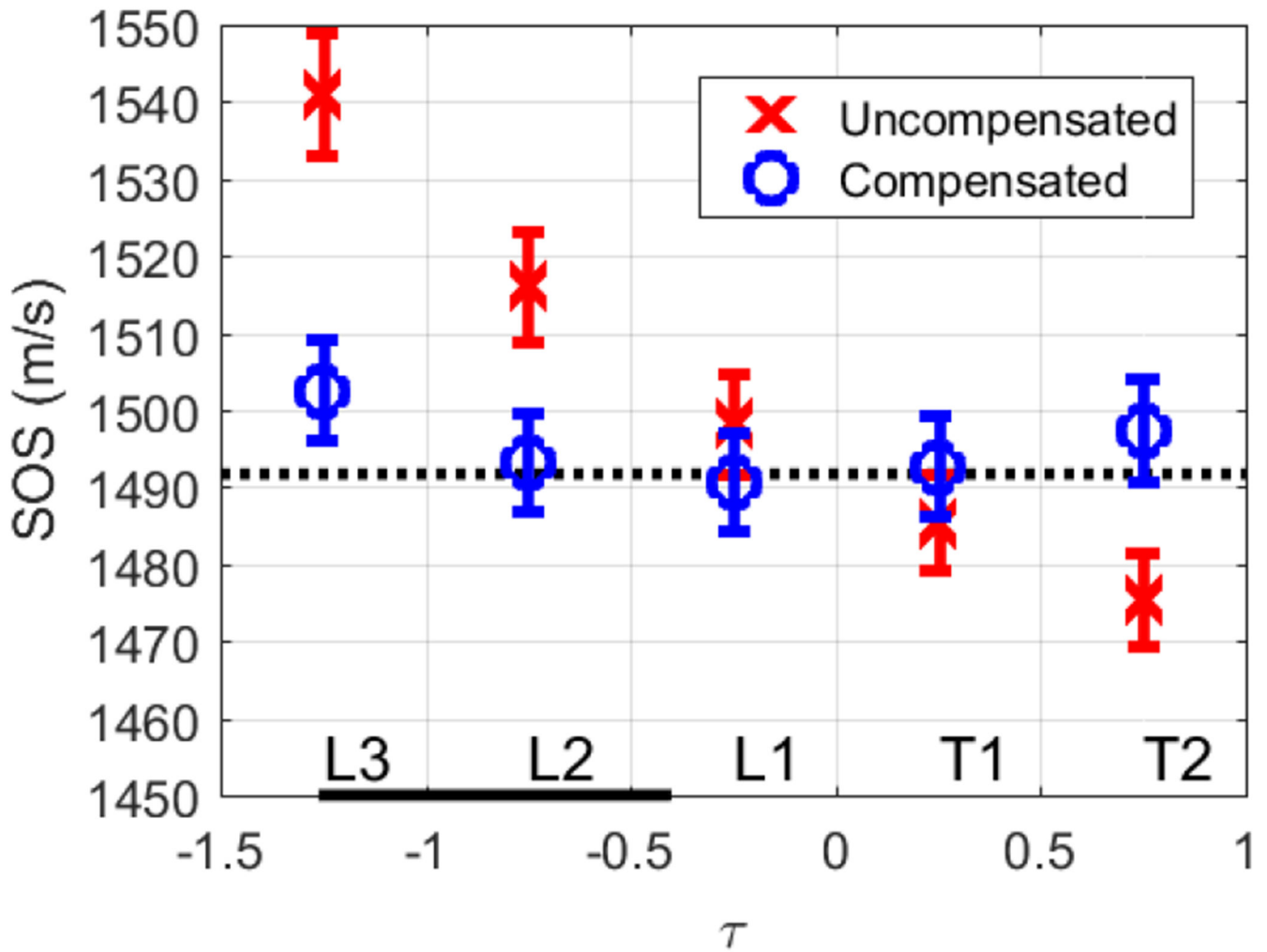
Phase insensitive (PI) vs. phase sensitive (PS) BUA in 73 women. © 2007 IEEE. Reprinted, with permission, from K. A. Wear, The effect of phase cancellation on estimates of calcaneal broadband ultrasound attenuation in vivo, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 54(7), 1353-1359, 2007.



**Fig. 9.**

The effect of frequency-dependent attenuation and transit-time marker location on transit-time differential,  $\Delta t$ . Zero-crossing markers are labeled with an L for leading half or a T for trailing half and are numbered outward from the pulse center. The values for  $\Delta t$  for each marker location are shown by the black arrows. The variation in  $\Delta t$  with marker location is due to the fact that the attenuated pulse is stretched in time as a consequence of the low-pass filtering effect of frequency-dependent attenuation. The black bar on the time axis represents the mean  $\pm$  one standard deviation of marker locations used in 43 papers. See Table I. © 2008 IEEE. Reprinted, with permission, from K. A. Wear, A method for improved standardization of in vivo calcaneal time-domain speed-of-sound measurements, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 55(7), 1473-1479, 2008.





**Fig. 10.**

Average values for SOS from 73 women computed from 5 different transit-time markers.

The x's were not compensated. The o's were compensated using (3). The black bar on the

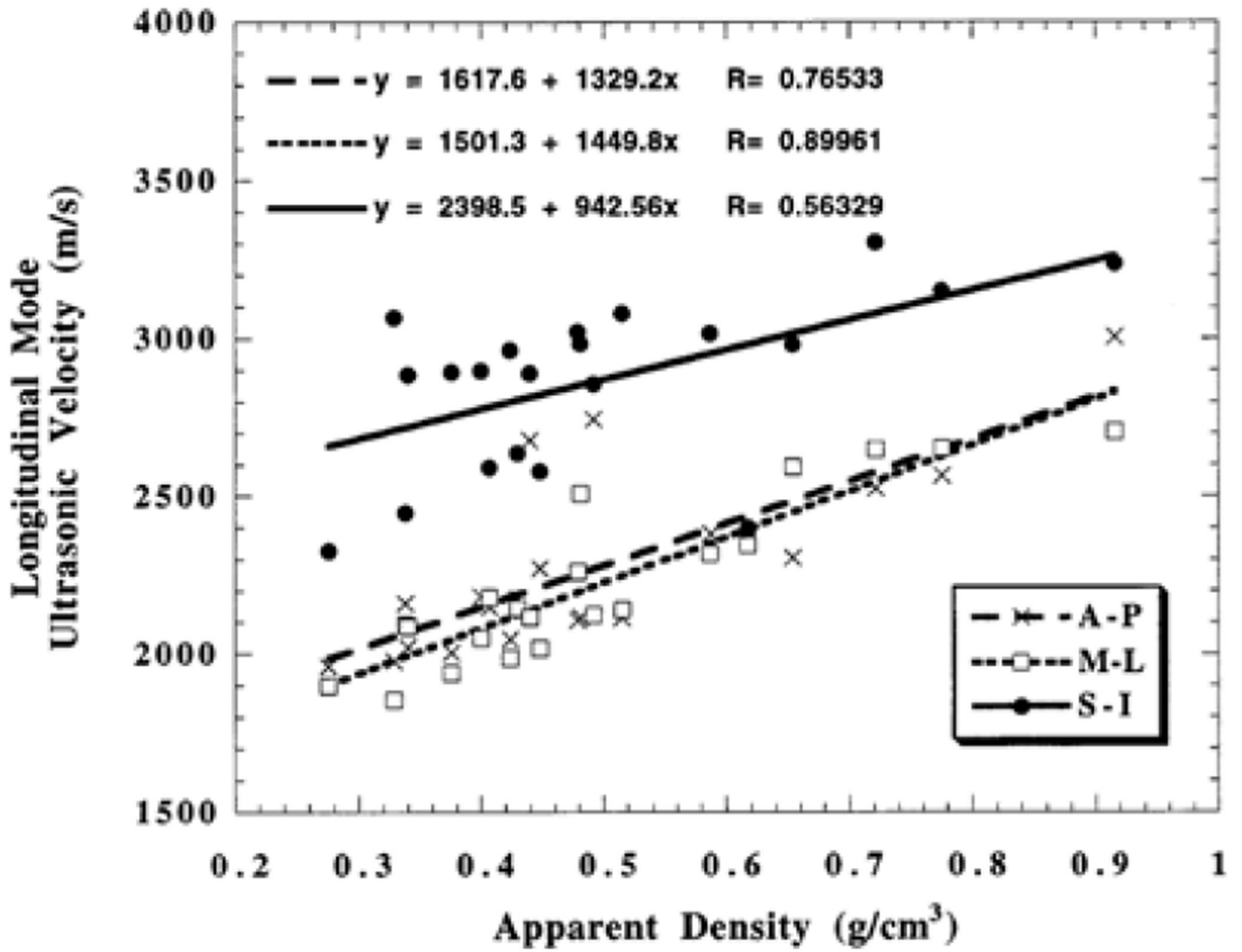
time axis represents the mean  $\pm$  one standard deviation of marker locations used in 43

papers. See Table I. © 2008 IEEE. Reprinted, with permission, from K. A. Wear, A method

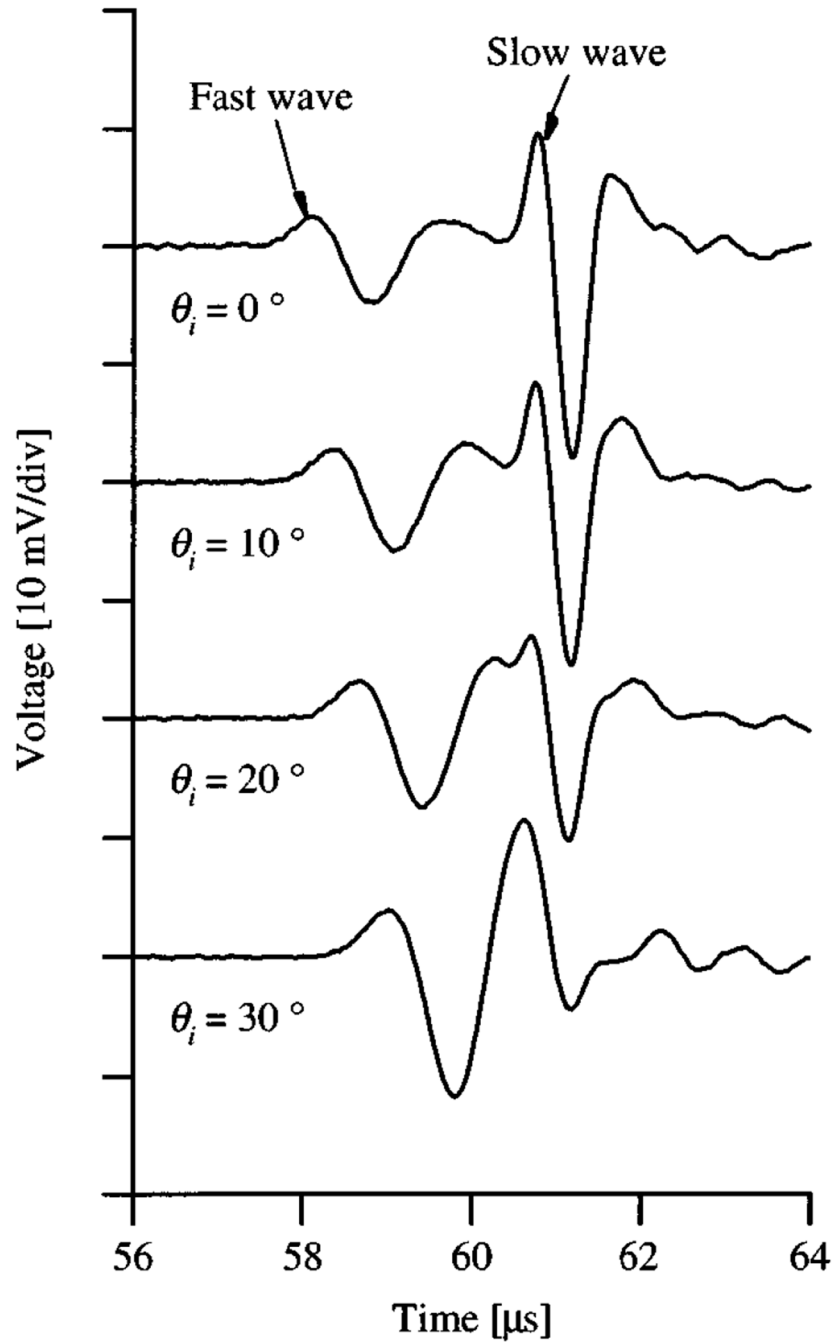
for improved standardization of in vivo calcaneal time-domain speed-of-sound

measurements, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 55(7), 1473-1479.

2008.

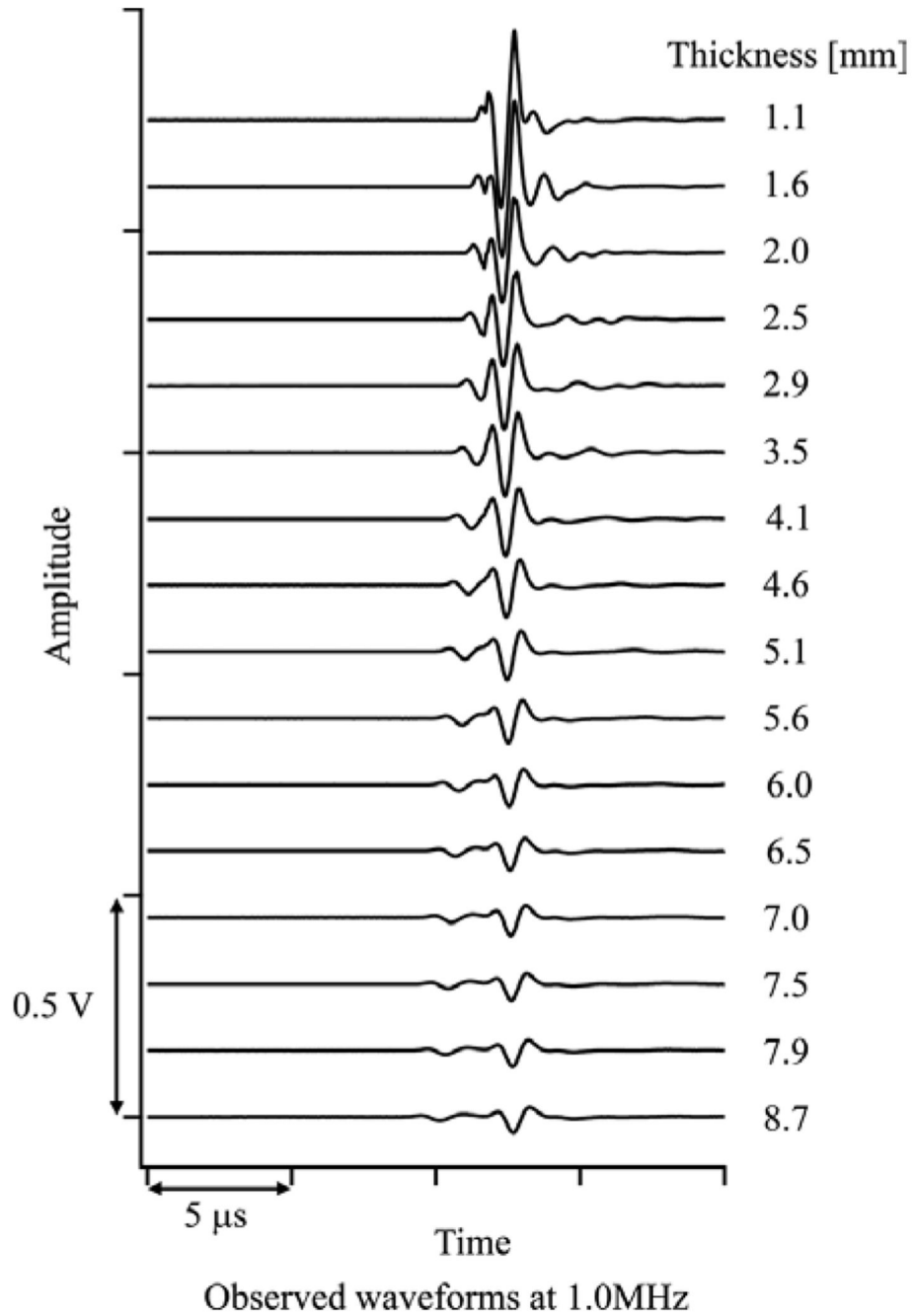


**Fig. 11.** Speed of sound vs. apparent density of bovine cancellous tibia *in vitro* in three orientations: anteroposterior (AP), mediolateral (ML), and superoinferior (SI). Reprinted from *Bone*, 26, B. K. Hoffmeister *et al.*, Low-megahertz ultrasonic properties of bovine cancellous bone, 635-642 Copyright (2000), with permission from Elsevier.



**Fig. 12.**

Fast and slow waves as a function of the angle between ultrasound propagation and main trabecular direction. Reprinted with permission from A. Hosokawa and T. Otani, *Acoustic anisotropy in bovine cancellous bone*, *J. Acoust. Soc. Am.*, 103(5), 2718-2722, 1998. Copyright 1998, Acoustical Society of America.

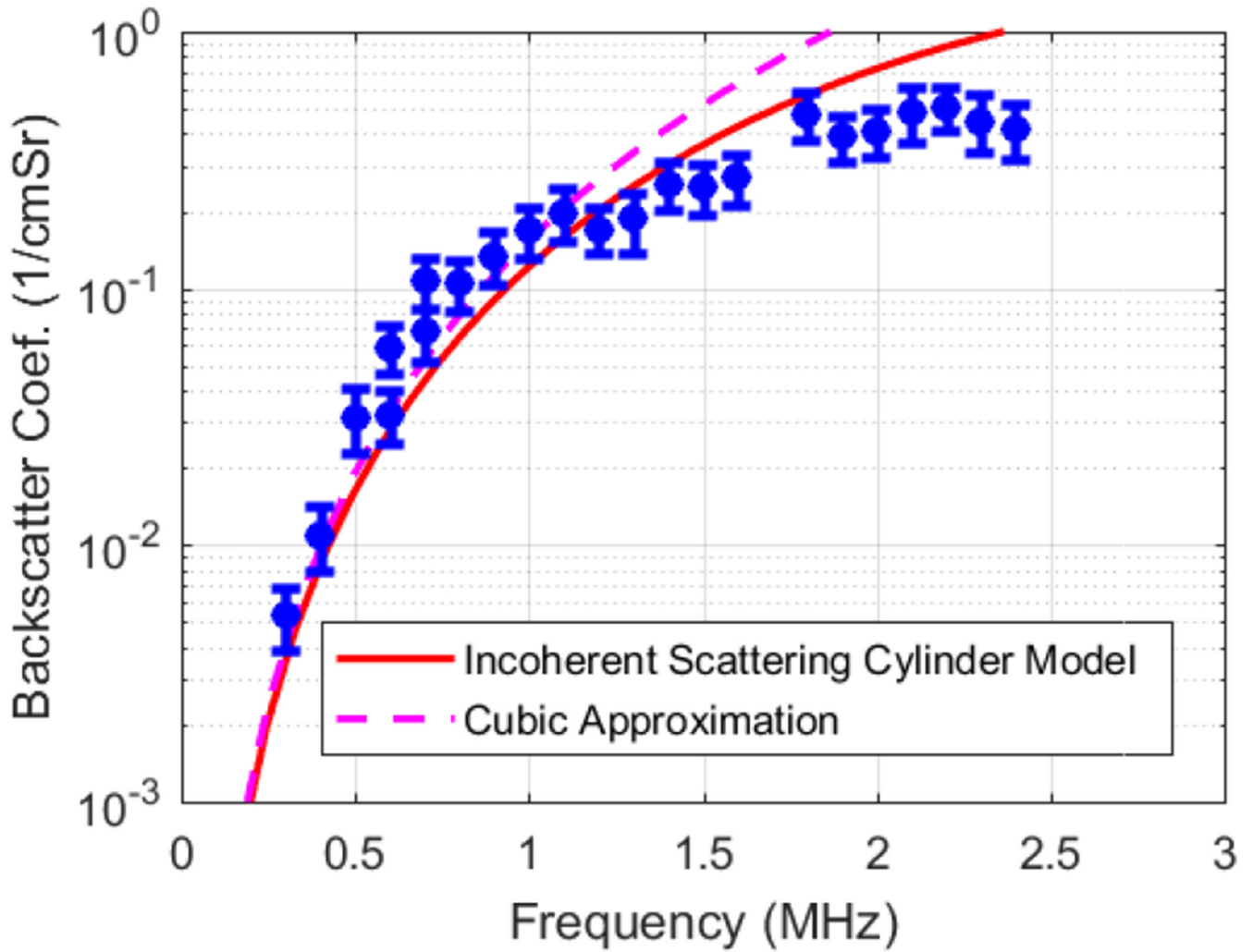


**Fig. 13.**

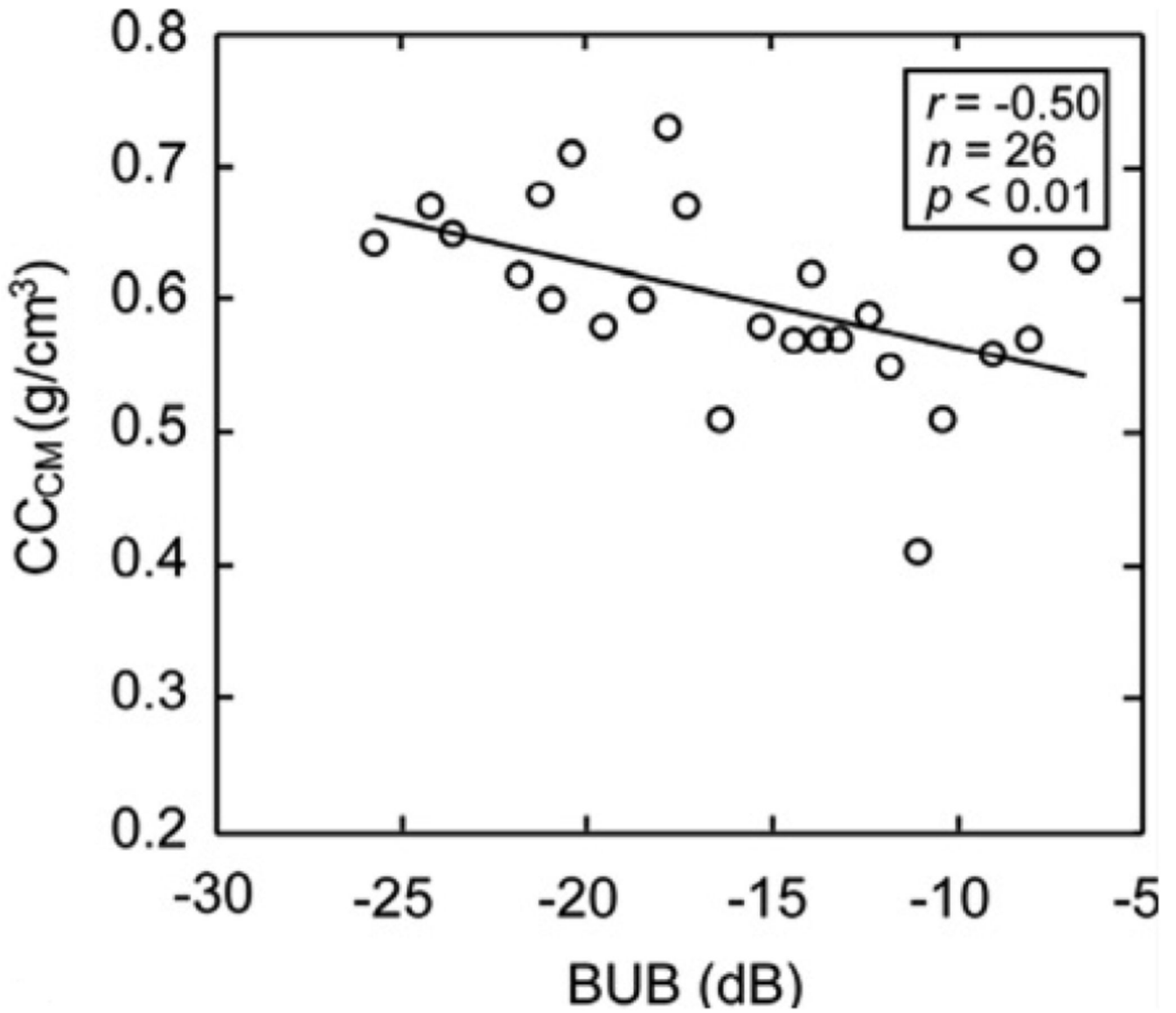
Through-transmitted signal from equine cancellous bone as a function of sample thickness from 1.1 to 8.7 mm. As the sample gets thicker, fast and slow wave magnitudes decrease due to attenuation and temporal separation between fast and slow waves increases due to longer propagation paths at different velocities. Reprinted with permission from F. Fujita *et al.*, An experimental study on the ultrasonic wave propagation in cancellous bone: Waveform changes during propagation, *J. Acoust. Soc. Am.*, 134(6), 4775-4781, 2013. Copyright 2013, Acoustical Society of America.



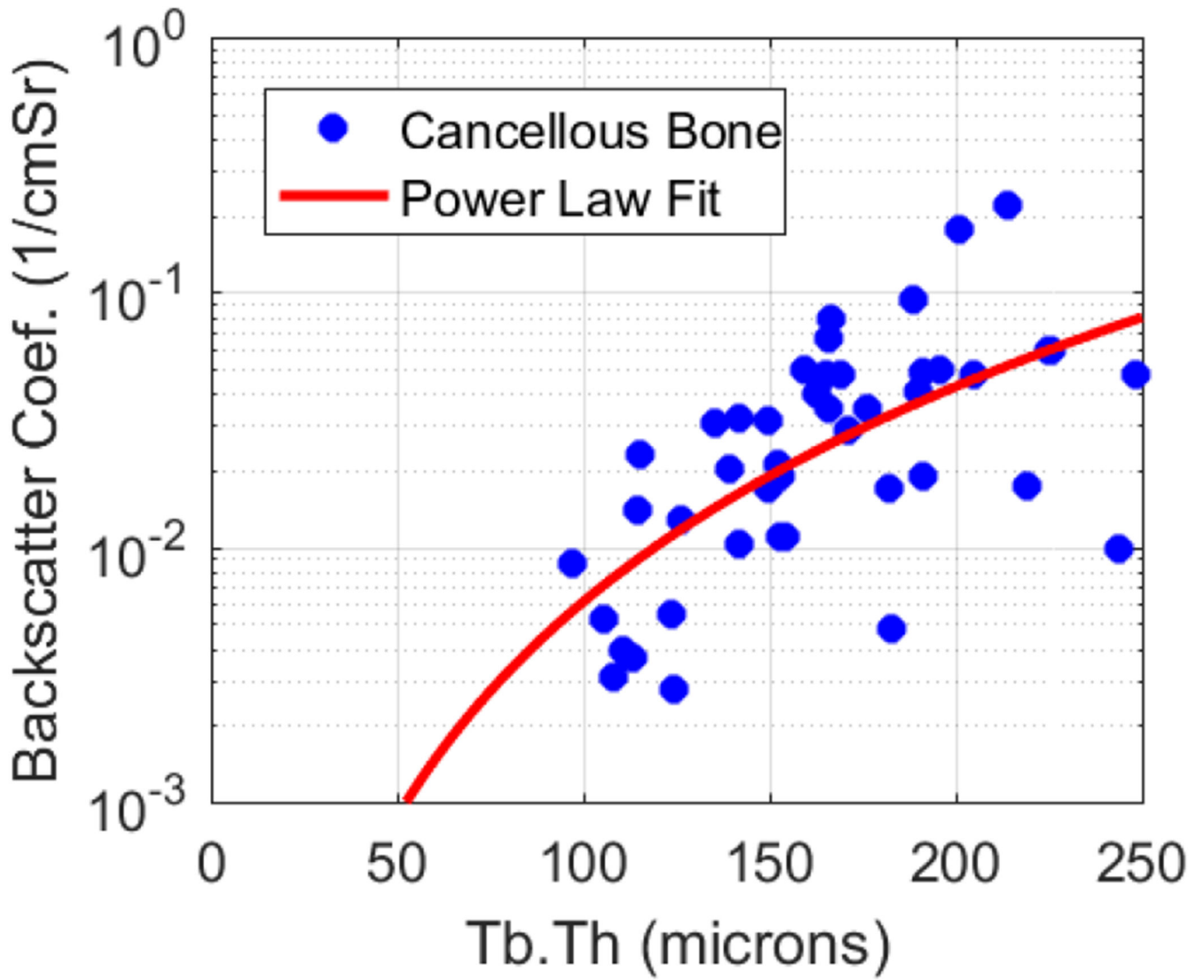
**Fig. 14.** Acquisition of backscatter data from human calcaneus *in vivo* using a standoff pad. Reprinted with permission from K. A. Wear and D. W. Armstrong III, Relationships among calcaneal backscatter, attenuation, sound speed, hip bone mineral density, and age in normal adult women, *J. Acoust. Soc. Am.*, 110(1), 573-578, 2001. Copyright 2001, Acoustical Society of America.



**Fig. 15.** Backscatter coefficient vs. frequency measured from 16 human calcaneus samples *in vitro*. Reprinted with permission from K. A. Wear, Frequency dependence of ultrasonic backscatter from human trabecular bone: theory and experiment, *J. Acoust. Soc. Am.*, 106(6), 3659-3664, 1999. Copyright 1999, Acoustical Society of America.

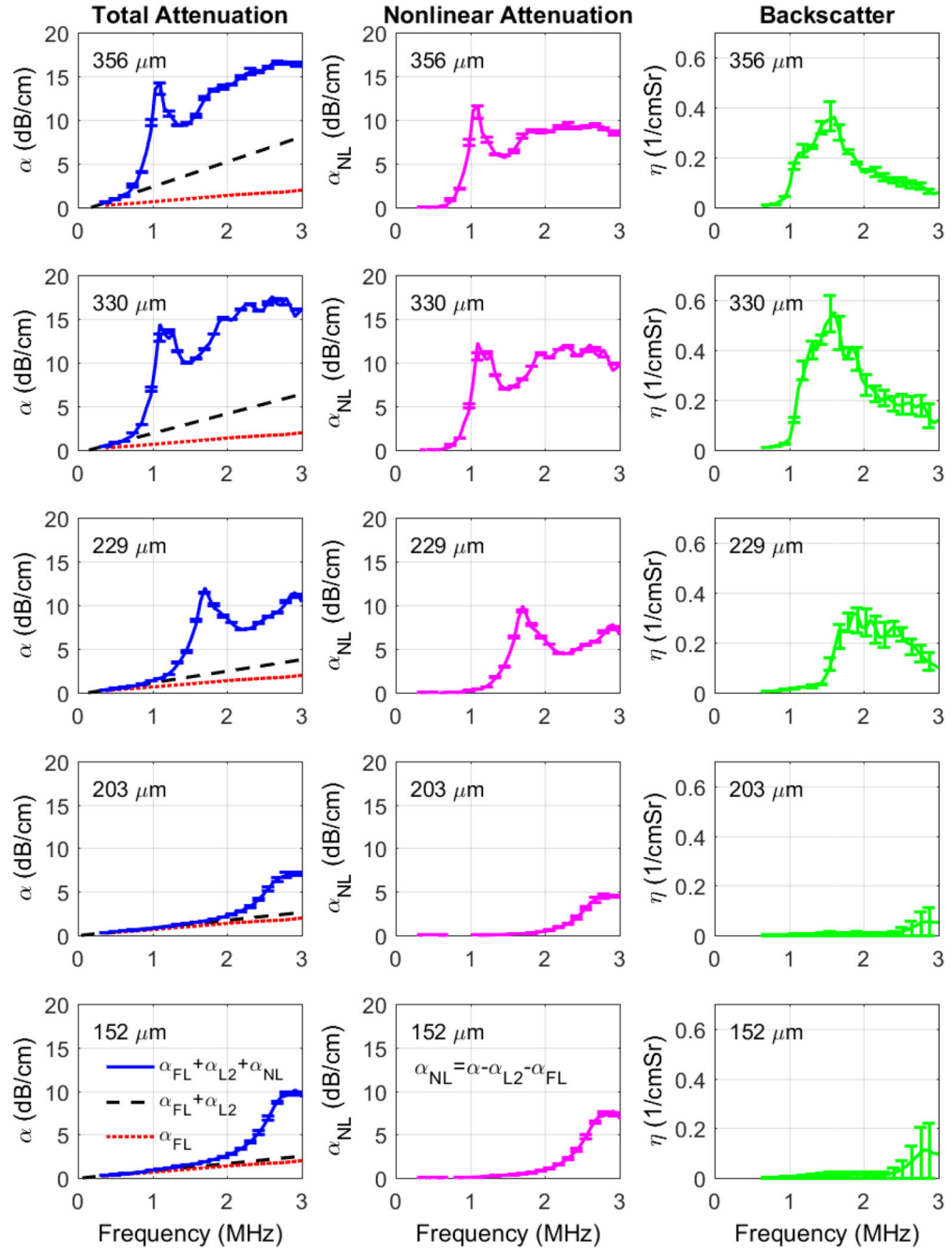


**Fig. 16.** Collagen content (CC) vs. broadband ultrasonic backscatter (BUB). Reprinted from *Ultrasound in Med. & Biol.*, 33, Riekkinen *et al.*, Acoustic properties of trabecular bone—relationships to tissue composition, 1438-1444, Copyright (2007), with permission from Elsevier.



**Fig. 17.** Backscatter coefficient vs. mean trabecular thickness Tb.Th (measured with  $\mu$ CT) measured from 43 human cancellous femur samples *in vitro*. © 2003 IEEE. Reprinted, with permission, from K. A. Wear and A. Laib, The dependence of ultrasonic backscatter on trabecular thickness in human calcaneus, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 50(8), 979-986. 2003.





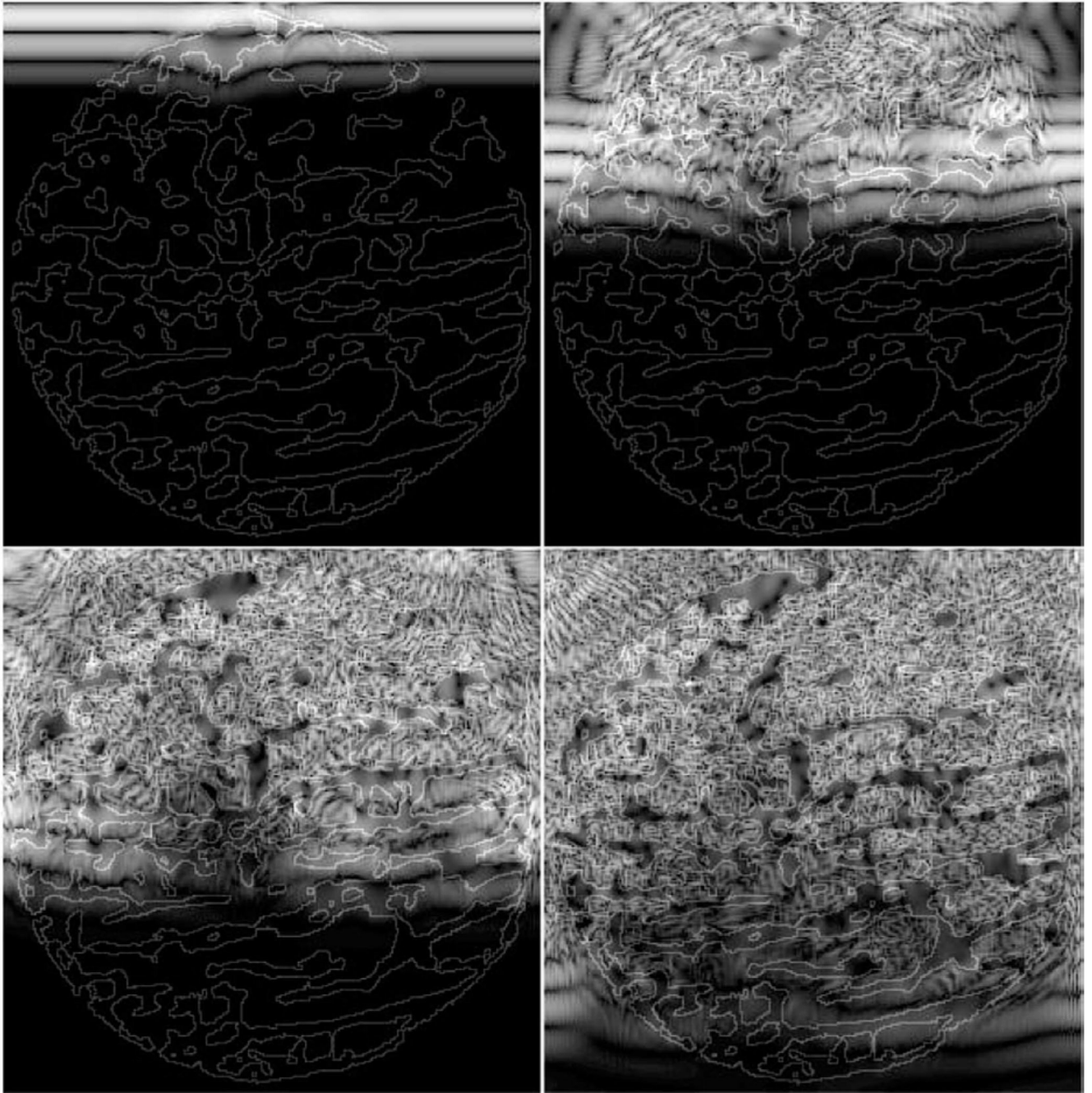
**Fig. 18.**

Total attenuation (left column), nonlinear component of attenuation (middle column) and backscatter coefficient (right column) for 5 phantoms containing nylon filaments (simulating trabeculae) suspended in a soft-tissue-mimicking fluid (simulating marrow). The left panel shows measurements of attenuation coefficient vs. frequency. The red dotted lines correspond to frequency-dependent attenuation coefficients measured from the reference phantom (i.e., phantom without nylon filaments). The black dashed lines correspond to linear fits of attenuation coefficient vs. frequency at low frequencies. The middle panel

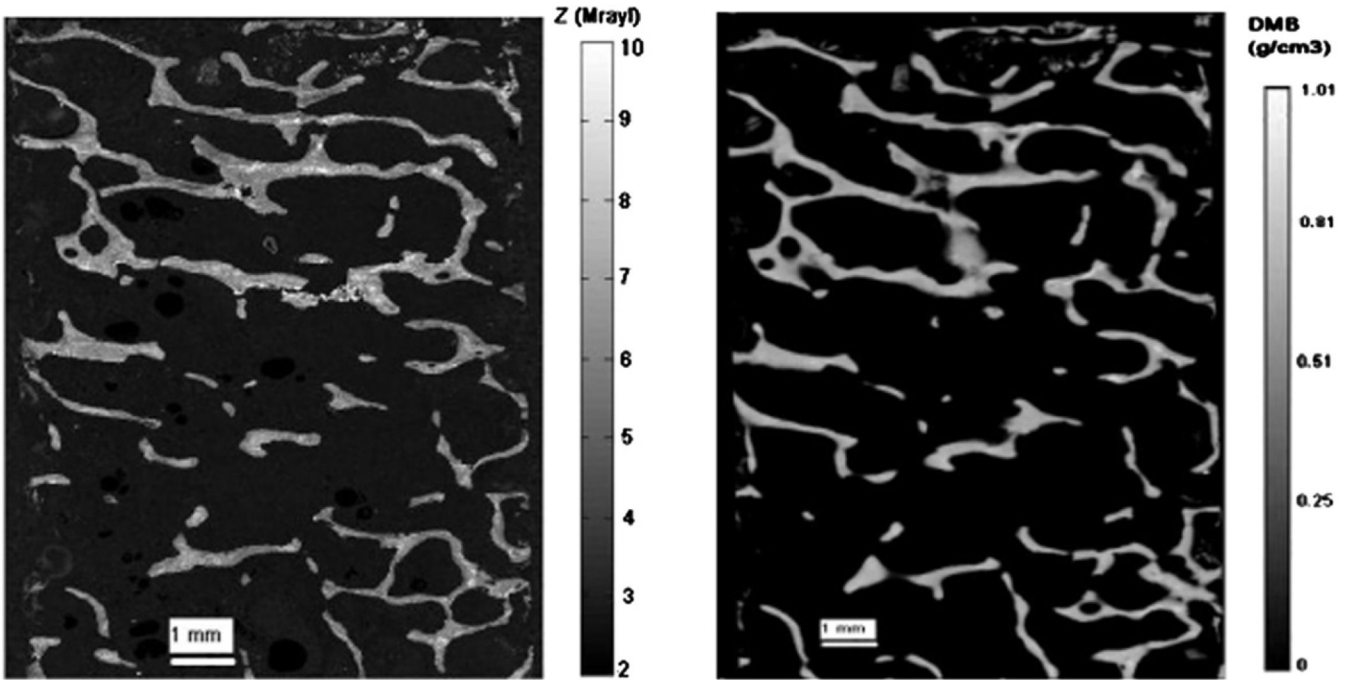
shows the nonlinear component of attenuation coefficient,  $\alpha_{NL}(f)$  which is the difference between  $\alpha(f)$  (left panel) and the low-frequency linear fit to  $\alpha(f)$  (left panel, black dashed line). The right panel shows measurements of backscatter coefficient,  $\eta(f)$ . © 2008 IEEE. Reprinted, with permission, from K. A. Wear, Mechanisms for attenuation in cancellous-bone-mimicking phantoms, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 55(11), 2418-2425. 2008.



**Fig. 19.** Aluminum foam sample to mimic cancellous bone. Reprinted with permission from C. Zhang *et al.*, Measurements of ultrasonic phase velocities and attenuation of slow waves in cellular aluminum foams as cancellous bone-mimicking phantoms, *J. Acoust. Soc. Am.*, 129(5), 3317-3326, 2011. Copyright 2011, Acoustical Society of America.



**Fig. 20.** Snapshots of waves propagating through cancellous bone, showing ballistic and scattered components. © 2008 IEEE. Reprinted, with permission, from J. J. Kaufman, Ultrasound Simulation in Bone, *IEEE Trans Ultrason., Ferroelectr., and Freq. Contr.*, 55(6), 1205-1218, 2008.



**Fig. 21.** Site-matched acoustic impedance measured with acoustic microscopy (left) and degree of mineralization of bone measured with synchrotron radiation  $\mu$ CT (right) from bovine cancellous sternum. Reprinted from *Ultrasound in Med. & Biol.*, 36, Rupin *et al.* Adaptive remodeling of trabecular bone core cultured in 3-D bioreactor providing cyclic loading: an acoustic microscopy study, 999-1007, Copyright (2010), with permission from Elsevier.

TABLE I.

## ACRONYMS

ABTF	Apparent Backscatter Transfer Function (dB)
AIB	Apparent Integrated Backscatter (dB)
AP	Anteroposterior
BMD	Bone Mineral Density ( $\text{g}/\text{cm}^2$ for areal or projection methods like DXA; $\text{g}/\text{cm}^3$ for volumetric methods like QCT)
BSCS	Backscattered Spectral Centroid Shift (MHz)
BV/TV	Bone Volume Fraction = Bone Volume / Total Volume
BUA	Broadband Ultrasound Attenuation (dB/MHz)
BUB	Broadband Ultrasound Backscatter ( $1/\text{cmSr}$ )
DXA	Dual-energy X-ray Absorptiometry
EDTA	Ethylenediaminetetraacetic acid
FMA	Frequency Modulated Attenuation (dB/cm)
FSAB	Frequency Slope of Apparent Backscatter (dB/MHz)
IRC	Integrated Reflection Coefficient
ML	Mediolateral
MLSP	Modified Least Squares Prony's (method)
MLSP+CF	MLSP plus Curve Fitting
NTD	Net time delay ( $\mu\text{s}$ )
$\mu\text{CT}$	Micro Computed Tomography
nBUA	Normalized BUA= $\text{BUA}/\text{sample thickness}$ (dB/cmMHz)
SOS	Speed of Sound (m/s)
QCT	Quantitative Computed Tomography
QUS	Quantitative Ultrasound
SAGE	Space Alternating Generalized Expectation maximization
SI	Superoinferior
Tb.Th	Mean trabecular thickness ( $\mu\text{m}$ )

**TABLE II.**

## WAVEFORM MARKERS FOR SOS MEASUREMENTS

Marker	$\tau_n$	# References
First detectable deviation from zero (L3)	-1.25	13
3 times noise standard deviation	$\sim -1.2$	1
10% of first rising half cycle	-1.19	4
15% of first rising half cycle	-1.18	1
20% of first rising half cycle	-1.17	3
10% of maximum amplitude	-1.08	1
First maximum	-1	3
"First" zero crossing (L2)	-0.75	17
"Second" zero crossing (L1)	-0.25	3
Envelope maximum	0	7
Mean $\pm$ standard deviation	$-0.83 \pm 0.42$	

$\tau_n$  = offset of waveform time marker from envelope maximum divided by the oscillation period. On the leading half of the pulse,  $\tau_n < 0$ . On the trailing half of the pulse,  $\tau_n > 0$ .