MOLECULAR BIOLOGY

Polycomb regulation is coupled to cell cycle transition in pluripotent stem cells

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When self-renewing pluripotent cells receive a differentiation signal, ongoing cell duplication needs to be coordinated with entry into a differentiation program. Accordingly, transcriptional activation of lineage specifier genes and cell differentiation is confined to the G_1 phase of the cell cycle by unknown mechanisms. We found that Polycomb repressive complex 2 (PRC2) subunits are differentially recruited to lineage specifier gene promoters across cell cycle in mouse embryonic stem cells (mESCs). Jarid2 and the catalytic subunit Ezh2 are markedly accumulated at target promoters during S and G_2 phases, while the transcriptionally activating subunits EPOP and EloB are enriched during G_1 phase. Fluctuations in the recruitment of PRC2 subunits promote changes in RNA synthesis and RNA polymerase II binding that are compromised in Jarid2 −/− mESCs. Overall, we show that differential recruitment of PRC2 subunits across cell cycle enables the establishment of a chromatin state that facilitates the induction of cell differentiation in G_1 phase.

INTRODUCTION

Deciphering the molecular mechanisms regulating pluripotent stem cell differentiation is of fundamental importance to understand mammalian development and for safe application of pluripotent stem cell–based therapies (1). Self-renewing mouse embryonic stem cells (mESCs) can be derived from the developing mouse blastocyst and provide a well-established system to study the molecular basis of pluripotency and early development. mESCs can differentiate into all cell types of the adult organism. However, the ability of individual mESCs within the population to respond to differentiation stimuli can be markedly different (2–4). This reveals a key aspect of the regulation of pluripotent cell differentiation and poses an important handicap for the application of stem cell–based therapies to humans. The features determining the responsiveness of individual cells to differentiation cues are mostly unknown and are currently a subject of intense debate. Accumulated evidence indicates that transcriptional activation of lineage specifier genes and cell differentiation in pluripotent cells is confined to the G_1 phase of the cell cycle (5–8). This observation partly explains the observed functional heterogeneity of pluripotent cell populations and highlights a very important regulatory feature of stem cell differentiation. Despite its obvious relevance, very little is known about the molecular mechanisms underlying this type of regulation (9).

Polycomb group (PcG) proteins are a hallmark of epigenetic control in eukaryotes and key regulators of mammalian development, cancer progression, and stem cell differentiation (10, 11). In mESCs, PcG proteins associate to form Polycomb repressive complexes 1 and 2 (PRC1 and PRC2) that catalyze H2AK119 monoubiquitination and H3K27 methylation, respectively (11). PRCs bind and repress hundreds of developmental regulator genes that will be activated later during cell differentiation (12, 13). In mESCs, PRC target repressed genes display nucleosomes modified with functionally op-posing histone modifications including H3K4me3 and H3K27me3 (hence, the designation of bivalent chromatin), leaky production of transcripts, and binding of poised RNAPII (RNA polymerase II) phosphorylated on Ser^5 (Ser^5-RNAPII) (14–17). PRC2 is typically composed of core subunits Eed, Suz12, Rbbp4/7, and Ezh1/2 of which the last one harbors the histone methyltransferase catalytic function (11). PRC2 function is regulated by nonstoichiometric accessory subunits that are differentially expressed during development (11). mESCs express high levels of Jarid2 that recruits and enhances PRC2 activity (11, 18–22). In addition, mESCs express the PRC2-interacting protein EPOP, which mediates recruitment of EloB and promote low-level transcription of bivalent genes (23). It has been proposed that Jarid2 and EPOP form mutually exclusive complexes (23). In agreement, studies of PcG proteins interactome have shown the existence of at least two types of PRC2 subcomplexes, one containing EPOP and PCL1-3 (Polycomb-like proteins 1 to 3) and another one containing Jarid2 and Aebp2 (11). They have been proposed to be termed PRC2.1 and PRC2.2, respectively (24). The functional relevance of PRC2 subcomplexes specialization remains unknown.

In this study, we asked whether chromatin regulation by PRC2 was linked to the preference of pluripotent cells to enter differentiation in G_1 phase. We found that recruitment of PRC2.1 complex to target promoters is favored in G_1 phase, while binding of PRC2.2 complex increases during S and G_2-M phases, leading to gradual accumulation of the catalytic subunit Ezh2 at bivalent promoters. This is accompanied by enhanced gene repression and accumulation of paused Ser^5-RNAPII at bivalent promoters during S and G_2-M phases. This cell cycle–dependent regulation is particularly evident at pioneering lineage specifiers, whose tight regulation is hindered in Jarid2 −/− mESCs. Together, our results strongly suggest that differential recruitment of PRC2 complexes across cell cycle is key to establish a chromatin state that facilitates the induction of cell differentiation in G_1 phase.

RESULTS

Recruitment of Ezh2 to bivalent promoters increases during S and G_2-M phases

We established wild-type mESCs that stably express the fluorescent ubiquitination-based cell cycle indicator (FUCCI) reporter system

(FUCCI-mESCs) (fig. S1A) (*25*)and used flow cytometry to isolate highly enriched populations of cells in G_1 (83 ± 2%), S (55 ± 1%), and G_2-M phases (81 ± 1%) (see Methods and table S1) (Fig. 1A). Genome-wide analysis of Ezh2 binding by chromatin immuno-

precipitation followed by sequencing (ChIP-seq) readily revealed the prevalence of recruitment of Ezh2 to chromatin in S and G_2-M phases compared to G_1 phase (Ezh2 binding peaks: 6316 in G_1; 9630 in S; and 15,967 in G_2-M) (table S2). Using published data, we established
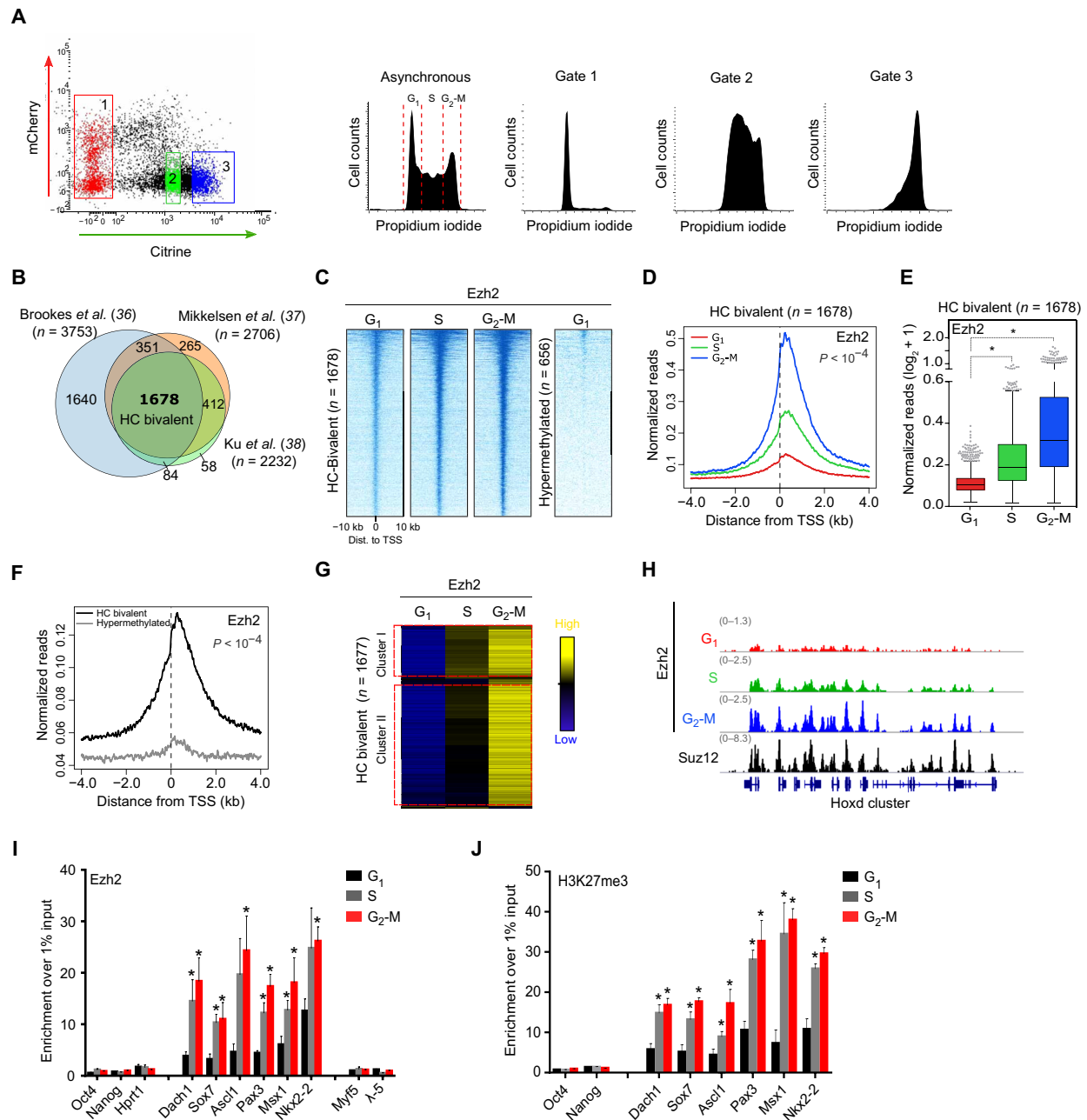

Fig. 1. Recruitment of the PRC2 core subunit Ezh2 to bivalent genes increases during S and G_2-M phases of the cell cycle. (**A**) Flow cytometry dot plot analysis of FUCCI-mESCs indicating sorting gates used to obtain cell populations enriched in G_1 (Gate 1), S (Gate 2), and G_2-M (Gate 3) cell cycle phases (left). Sorted cells were stained with propidium iodide and analyzed by flow cytometry (right). (**B**) Venn diagrams of bivalent genes previously published in (*36–38*). (**C**) Heatmaps of normalized Ezh2 ChIP-seq reads around the TSS of HC bivalent promoters at different cell cycle phases. Heatmap of hypermethylated promoters is shown as a negative control. (**D**) Average binding profile of Ezh2 around the TSS of HC bivalent promoters in G_1 (red), S (green), and G_2-M (blue). (**E**) Quantification of Ezh2-binding signal at the promoter regions (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes in indicated cell cycle phases. (**F**) Average binding profile of Ezh2 around the TSS of HC bivalent (black) and hypermethylated (gray) promoters in G_1 phase. (**G**) Hierarchical clustering analysis of binding of Ezh2 to the promoter region (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes at indicated phases of the cell cycle. Binding relative to the average is presented. (**H**) Genome browser view of Ezh2 ChIP-seq data across cell cycle at the *Hoxd* gene cluster. Suz12 binding was analyzed using published data (*19*). (**I** and **J**) Histogram showing enrichment of Ezh2 (I) or H3K27me3 (J) at PRC2 target promoter regions (*Dach1*, *Sox7*, *Ascl1*, *Pax3*, *Msx1*, and *Nkx2-2*) in G_1 (black), S (gray), and G_2-M (red) assayed by ChIP-qPCR. Active (*Oct4*, *Nanog*, and *Hprt1*) and hypermethylated (*Myf5* and *λ-5*) gene promoters were used as negative controls. Means ± SEM of three (I) or four (J) experiments is shown. (E, I, and J) Asterisk (*) marks statistically significant differences.

a list of high confidence Polycomb target bivalent genes [High confidence (HC) bivalent, $n = 1678$] (Fig. 1B). As controls, we used transcriptionally active ($n = 1557$) and hypermethylated ($n = 656$) genes that are not targeted by PRC2 (see Methods and table S3). Heatmap analysis of Ezh2 binding to HC bivalent genes showed that recruitment of Ezh2 was increased as cells exit G_1 and transit into S and G_2-M phases (Fig. 1C). Comparison of Ezh2 binding at HC bivalent gene promoters showed that, although Ezh2 accumulates around the transcription start site (TSS) of bivalent genes at all cell cycle phases, the amount of Ezh2 bound gradually increases as cells exit G_1 phase and transit through the cell cycle (Fig. 1, D and E, and fig. S1, B and C). Recruitment of Ezh2 in G_1 phase appeared weak compared to G_2-M (Fig. 1, D and E), but it was evident when compared to hypermethylated promoters known to be devoid of PRC2 (Fig. 1F and fig. S1D). Analysis of Ezh2 binding at individual promoters revealed a very consistent and gradual accumulation of Ezh2 during S and G_2-M phases in most (1576 of 1677; 93.9%) HC bivalent gene promoters (see clusters I and II in Fig. 1G and fig. S1E) including the archetypical *Hoxd* gene cluster (Fig. 1H). These observations were confirmed by ChIP–quantitative polymerase chain reaction (qPCR) for Ezh2 and analysis of a subset of well-characterized (*18*) PRC2 target promoters (Fig. 1I). Increased binding of Ezh2 during S and G_2-M phases was particularly obvious at bivalent promoters, but it could also be detected at bivalent gene bodies and other Ezh2-bound genomic regions (fig. S1F). Enhanced recruitment of Ezh2 upon G_1 phase exit resulted in concomitant accumulation of H3K27me3 at bivalent promoters during S and G_2-M phases (Fig. 1J). Thus, we concluded

that binding of Ezh2 to target promoters is markedly enhanced upon G_1 phase exit in mESCs.

Inverse binding patterns of Jarid2 and EPOP across cell cycle

We next asked whether accumulation of Ezh2 at bivalent promoters during S and G_2-M phases reflected increased binding of PRC2.1, PRC2.2, or both. ChIP-seq analysis of Jarid2 binding in cell cycle–sorted FUCCI-mESCs demonstrated that recruitment of Jarid2 to the promoter region of HC bivalent genes was augmented in S and G_2-M phases compared to G_1 phase (Fig. 2, A and B, and fig. S2, A and B). Increased recruitment of Jarid2 to target genes in S and G_2-M phases was evident but quantitatively less accused than changes found in binding of Ezh2 across cell cycle (Fig. 1D). Notwithstanding, clustering analysis showed a very consistent tendency to accumulate Jarid2 in G_2-M at individual HC bivalent genes (1404 of 1677; 83.7%) (see clusters I and II in Fig. 2C). Jarid2 was bound to most of the Ezh2-bound HC promoters in G_2-M (1188 of 1262; 94%) (fig. S2C), and binding of Jarid2 and Ezh2 around the TSS of HC bivalent gene promoters in G_2-M displayed concordant distributions (Fig. 2D). Correlation analysis demonstrated that accumulation of Jarid2 at target genes in S and G_2-M phases correlates with increased recruitment of Ezh2 to the same promoter regions (Fig. 2E). Coordinated recruitment of Jarid2 and Ezh2 to target genes upon G_1 phase exit was also observed at individual candidate regions (i.e., gene *Adra2c*) (Fig. 2F). While increased binding of Jarid2 to HC bivalent promoters was clear in G_2-M compared to G_1 phase (Fig. 2C), changes in recruitment of Jarid2 in S phase were less homogeneous. Most of
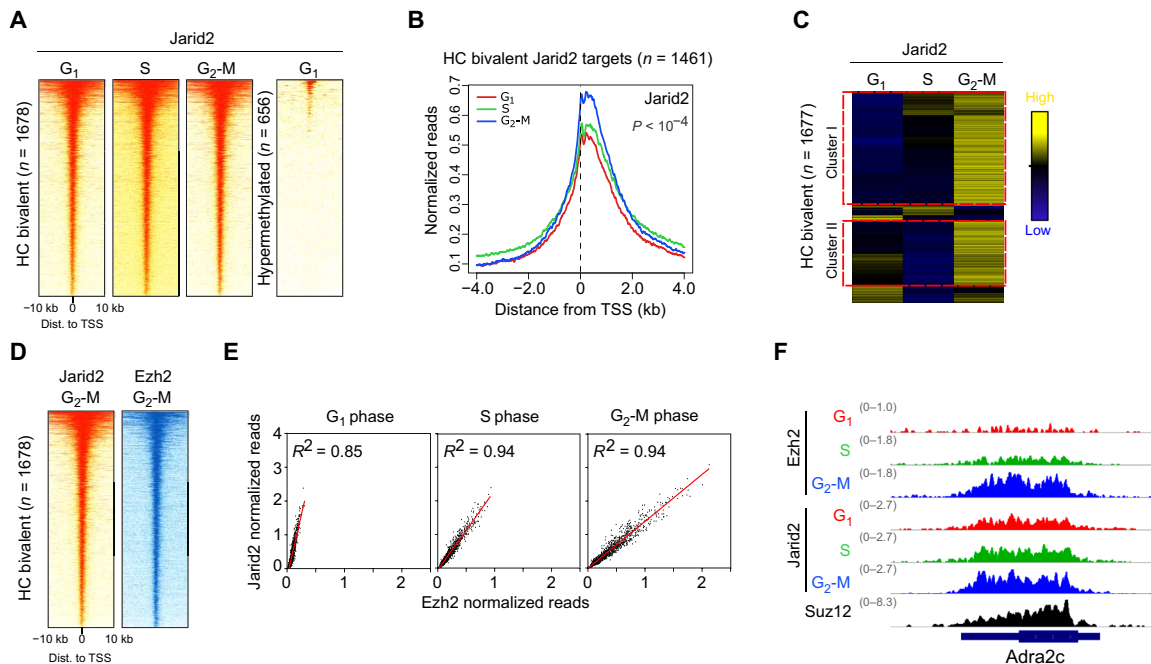
Fig. 2. Binding of Jarid2 to target promoters increases during S and G_2-M phases of the cell cycle. (**A**) Heatmaps of normalized ChIP-seq reads showing the binding of Jarid2 around the TSS of HC bivalent promoters at different cell cycle phases. Heatmap of hypermethylated promoters is shown as a negative control. (**B**) Average binding profile of Jarid2 around the TSS of HC bivalent promoters in G_1 (red), S (green), and G_2-M (blue). (**C**) Hierarchical clustering analysis of binding of Jarid2 to the promoter region (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes at indicated phases of the cell cycle. Binding relative to the average is presented. (**D**) Heatmaps comparing the binding of Jarid2 and Ezh2 around the TSS of HC bivalent promoters in G_2-M. (**E**) Linear regression analysis showing the correlation between the binding signals of Jarid2 and Ezh2 at HC bivalent promoters (−0.5 to +1.5 kb relative to TSS) at indicated cell cycle phases. (**F**) Genome browser view of Ezh2 and Jarid2 binding across cell cycle at the *Adra2c* bivalent gene. Suz12 binding was analyzed using published data (*19*).

the HC bivalent genes (cluster I, 899 genes; 64%) showed increased binding of Jarid2 in S phase compared to G_1 phase; however, a smaller proportion of genes (cluster II, 505 genes; 36%) showed reduced binding of Jarid2 in this cell cycle phase (Fig. 2C). This pattern is in contrast with the homogeneous increase of Ezh2 binding at HC bivalent promoters in S phase (Fig. 1G) and suggests that recruitment of Ezh2 to target genes in S phase might be Jarid2 independent for this subset of bivalent genes (cluster II).

We next analyzed binding of the PRC2.1 subunit EPOP. In stark contrast to the binding pattern observed for Ezh2 and Jarid2, binding of EPOP around the TSS of HC bivalent genes was reduced in cells in G_2-M (Fig. 3, A to C, and fig. S3, A and B). Clustering analysis showed that increased binding of EPOP in G_1 phase compared to G_2-M can be observed in most (1426 of 1677; 85%) bivalent gene promoters (see cluster I in fig. S3C). Increased recruitment of EPOP to target promoters in G_1 phase was also observed by ChIP-qPCR for a representative set of PRC2 target promoters (Fig. 3D). We next addressed whether increased binding of EPOP in G_1 phase led to augmented recruitment of EloB in this cell cycle phase. As expected, ChIP-seq analysis showed that EloB is recruited to HC bivalent genes more profoundly in G_1 phase compared to S and G_2-M phases in average (Fig. 3, E to G, and fig. S3, D and E) and individual gene (fig. S3F) analyses. Global levels of Ezh2, Jarid2, and EPOP across cell cycle were unchanged, indicating that differential recruitment to target genes is regulated by changes in protein interactions rather than changes in protein abundance (Fig. 3H and fig. S3, G and H). Together, these results

demonstrate that Jarid2 and Ezh2 recruitment is increased during S and G_2-M phases, while EPOP and EloB are preferentially bound to chromatin in G_1 phase. This indicates that PRC2.1 complex is preferentially recruited to target promoters during G_1 phase while PRC2.2 complex is favored during S and G_2-M phases.
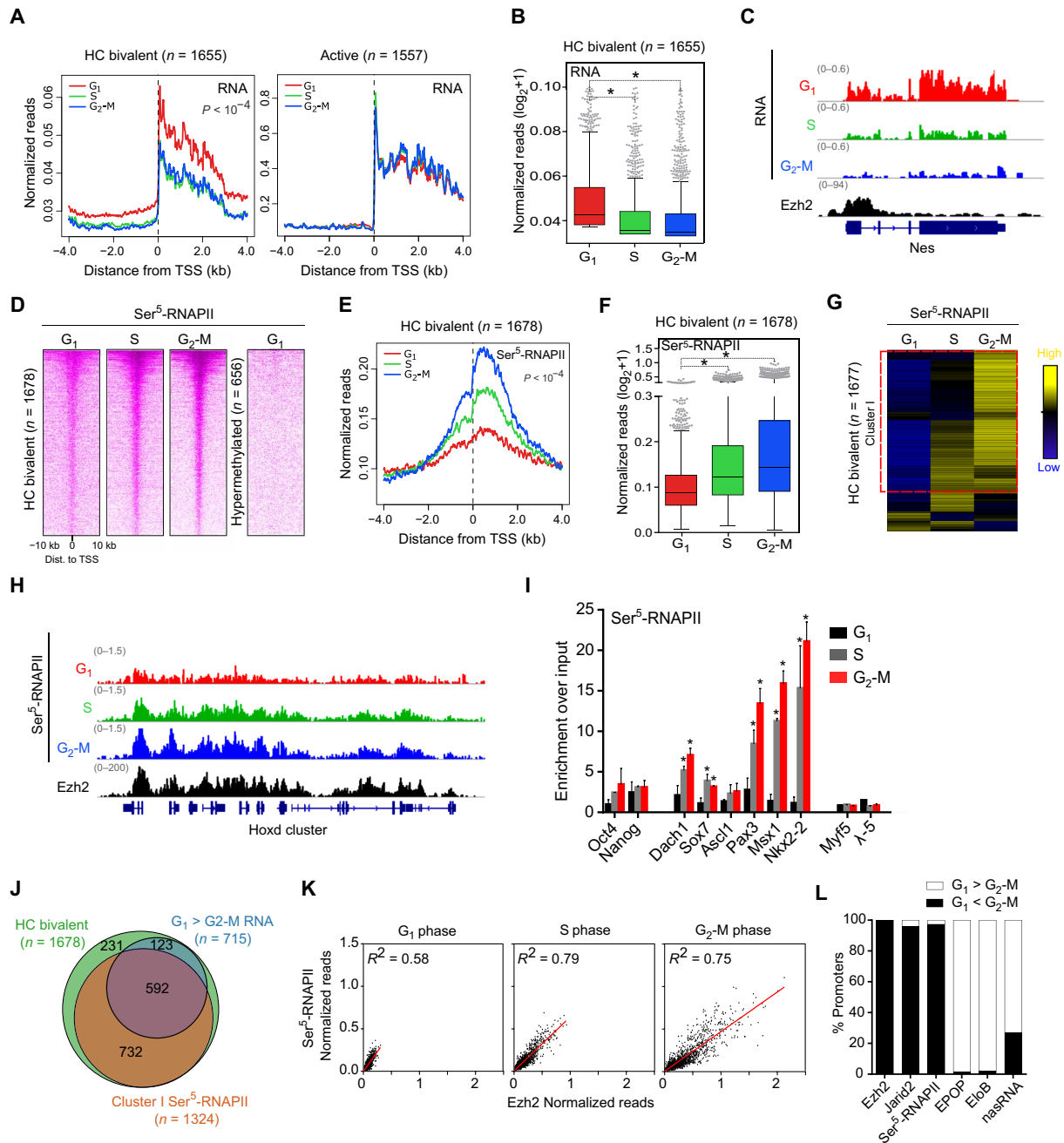
RNA synthesis is reduced and Ser5-RNAPII is accumulated at PRC2 target genes during S and G_2-M phase

We next questioned whether changes in recruitment of PRC2 subunits across cell cycle had functional consequences in the transcriptional regulation of target genes. To analyze low-level RNA transcription typically found at bivalent promoters, we used 4-thiouridine (4sU) tagging followed by high-throughput sequencing (4sU-seq). 4sU-seq permits the analysis of newly transcribed RNA (fig. S4A) (26, 27) and thus allowed us to minimize contamination of RNA molecules produced in preceding phases of the cell cycle. We found increased production of RNA in G_1 phase compared to S-G_2-M at HC bivalent genes but not at promoters of active genes (Fig. 4, A and B, and fig. S4, B and C). Of 1655 HC bivalent genes, 974 genes showed differences in RNA production between G_1 and G_2-M. Among these, most of them (715; 73.4%) were down-regulated in G_2-M compared to G_1 phase (fig. S4D), revealing a consistent tendency of bivalent genes to be more strictly repressed during S and G_2-M phases than in G_1 phase. In agreement, analysis of individual candidate genes confirmed that bivalent promoters (i.e., *Nes*) are overtly repressed in S and G_2-M phases
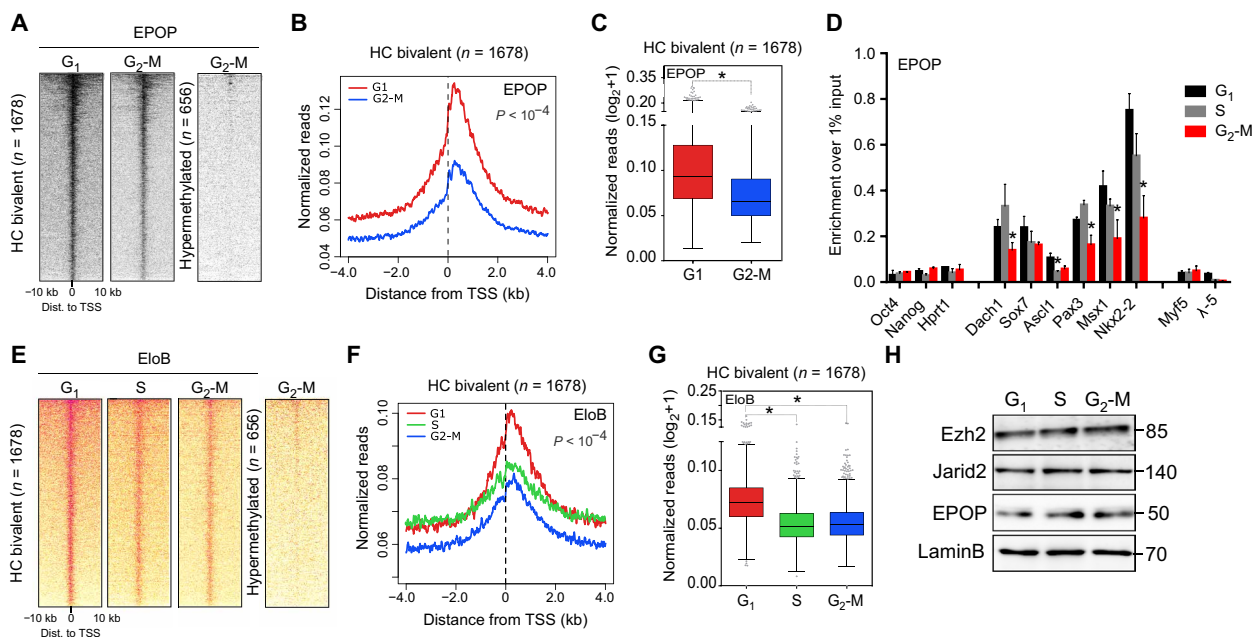
Fig. 3. Binding of EPOP and EloB to bivalent promoters is enhanced in G_1 phase. (**A**) Heatmaps showing the binding of EPOP around the TSS of HC bivalent promoters in G_1 and G_2-M. Heatmap of hypermethylated promoters is shown as a negative control. (**B**) Average binding profile of EPOP around the TSS of HC bivalent promoters in G_1 (red) and G_2-M (blue). (**C**) Boxplot of EPOP-binding signal at the promoter regions (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes in indicated cell cycle phases. (**D**) Histogram showing enrichment of EPOP to PRC2 target promoter regions (*Dach1*, *Sox7*, *Ascl1*, *Pax3*, *Msx1*, and *Nkx2-2*) in G_1 (black), S (gray), and G_2-M (red) assayed by ChIP-qPCR. Active (*Oct4*, *Nanog*, and *Hprt1*) and hypermethylated (*Myf5* and λ-5) gene promoters were used as negative controls. Means ± SEM of three experiments is shown. (**E**) Heatmaps showing the binding of EloB around the TSS of HC bivalent promoters in G_1, S, and G_2-M. Heatmap of hypermethylated promoters is shown as a negative control. (**F**) Average binding profile of EloB around the TSS of HC bivalent promoters in G_1 (red), S (green), and G_2-M (blue). (**G**) Quantification of EloB-binding signal at the promoter regions (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes in indicated cell cycle phases. (**H**) Whole-cell lysate Western blots comparing Ezh2, Jarid2 and EPOP protein levels in G_1, S, and G_2-M. Lamin B was used as a loading control. (C, D, and G) Asterisk (*) marks statistically significant differences.

Fig. 4. RNA synthesis is down-regulated and Ser5-RNAPII is accumulated at PRC2 target promoters during S and G2-M phase. (**A**) Average RNA production from HC bivalent (left) and active (right) promoters in G_1 (red), S (green), and G_2-M (blue). (**B**) Boxplot comparing 4sU-seq reads mapped to the proximal promoter region (TSS to +3Kb) of HC bivalent genes in indicated cell cycle phases. (**C**) Genome browser view of RNA synthesis at indicated cell cycle phases at the bivalent gene *Nes*. Ezh2 binding was analyzed using published data (*46*). (**D**) Heatmaps showing the binding of Ser5-RNAPII around the TSS of HC bivalent promoters in G_1, S, and G_2-M. Heatmap of hypermethylated promoters is shown as a negative control. (**E**) Average binding profile of Ser5-RNAPII around the TSS of HC bivalent gene promoters in G_1 (red), S (green), and G_2-M (blue). (**F**) Quantification of Ser5-RNAPII–binding signal at the promoter regions (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes in indicated cell cycle phases. (**G**) Hierarchical clustering analysis of binding of Ser5-RNAPII to the promoter region (−0.5 to +1.5 kb relative to TSS) of HC bivalent genes at indicated phases of the cell cycle. Binding relative to the average is presented. (**H**) Genome browser view of the binding of Ser5-RNAPII across cell cycle at the *Hoxd* gene cluster. Ezh2 binding was analyzed using published data (*46*). (**I**) Analysis by ChIP-qPCR of Ser5-RNAPII binding at PRC2 target promoter regions (*Dach1*, *Sox7*, *Ascl1*, *Pax3*, *Msx1*, and *Nkx2-2*) in G_1 (black), S (gray), and G_2-M (red) assayed by ChIP-qPCR. Active (*Oct4* and *Nanog*) and hypermethylated (*Myf5* and *λ-5*) gene promoters were used as controls. Means ± SEM of three experiments is shown. (**J**) Venn diagram showing the overlap between HC bivalent genes overtly repressed in G_2-M (FC > 1.5) and genes displaying accumulation of Ser5-RNAPII at their promoter region in G_2-M (cluster I in Fig. 3G). (**K**) Linear regression analysis of the binding signals of Ezh2 and Ser5-RNAPII at HC bivalent promoters (−0.5 to +1.5 kb relative to TSS) at indicated cell cycle phases. (**L**) Histogram displaying the percentage of genes showing changes ($G_1 > G_2$-M or $G_1 < G_2$-M) of binding for Ezh2 (FC > 2), Jarid2 (FC > 1.3), Ser5-RNAPII (FC > 2), EPOP (FC > 1.5), EloB (FC > 1.5), and RNA production (FC > 1.5) during cell cycle transition. Genes that showed no difference between analyzed phases were excluded to calculate the percentage. (B, F, and I) Asterisk (*) marks statistically significant differences. FC, fold change.

(Fig. 4C). The level of RNA synthesized from bivalent promoters in G_1 phase was still very low compared to the level of RNA produced at active promoters (compare scale of y axis in HC bivalent and Active plots; Fig. 4A), indicating that transient alleviation of PRC2 repression in G_1 results in increased leaky transcription rather than full activation of bivalent genes.

To address whether recruitment of Jarid2-Ezh2 and firmer repression of bivalent genes during S-G_2-M were associated with changes in the activity of RNAPII, we analyzed binding of Ser[5]-RNAPII, typically associated with transcriptionally pausing and gene poising of bivalent genes in mESCs (28). We found augmented accumulation of Ser[5]-RNAPII around the TSS of HC bivalent genes in S and G_2-M phases as compared to G_1 phase (Fig. 4, D to F, and fig. S4, E and F). Increased binding of Ser[5]-RNAPII was evident for most bivalent promoters (1324 of 1677; 78.9%) genes (see cluster I in Fig. 4G) and at individual candidate bivalent genomic domains (i.e., *Hoxd* gene cluster) (Fig. 4H). ChIP-qPCR analysis of a subset of PRC2 target genes further confirmed gradual accumulation of Ser[5]-RNAPII during S and G_2-M phases compared to G_1 phase (Fig. 4I). Most of HC bivalent genes (592 of 715; 82.7%) that showed reduced RNA synthesis in G_2-M displayed accumulation of Ser[5]-RNAPII at their promoter region (Fig. 4J), suggesting that reduced production of RNA is coupled to RNAPII pausing at the promoters of bivalent genes during S and G_2-M phases. Increased recruitment of Ezh2 (or Jarid2) during S and G_2-M phases correlated with accumulation of Ser[5]-RNAPII at target promoters at all phases of the cell cycle, and it became more evident during S and G_2-M phases (Fig. 4K and fig. S4G). Together (Fig. 4L and fig. S4H), these observations indicate that increased recruitment of Jarid2-Ezh2 to target genes is associated with reduced binding of EPOP and EloB, pausing of transcription by RNAPII, and reduced production of leaky RNA during S and G_2-M phases.

Cell cycle–dependent regulation of PRC2 is more evident at the promoter of developmental transcription factors

We reasoned that, because cell cycle–dependent regulation of bivalent genes involved changes in recruitment of Jarid2, Ezh2, EPOP, EloB, and Ser[5]-RNAPII, bivalent genes that are common targets of these proteins in asynchronous mESCs might display more accused regulation across cell cycle. Notably, we found that a set of 390 bivalent genes cobound by these factors (common target genes) (Fig. 5A) is very significantly enriched for transcription factors and DNA binding proteins (162 of 390 genes; $P = 7.6 \times 10^{-94}$), in contrast to the remaining 991 genes that are enriched for protein binding and transmembrane transporters (Fig. 5B). Common target genes included key pioneering factors involved in mesoderm, ectoderm, and endoderm differentiation (fig. S5A), suggesting that cell cycle–dependent regulation of Polycomb recruitment modulates differentiation to the three germ layers. In fitting, bivalent genes that showed more accused cell cycle–dependent changes in PRC2 binding were also enriched in a similar subset of transcription factors (fig. S5, B to E). Comparative analysis of PRC2 binding revealed that common targets are more profoundly bound by PRC2 subunits (Ezh2, Jarid2, and EPOP) and Ser[5]-RNAPII than remaining genes and that their differential recruitment of PRC2 subunits, Ser[5]-RNAPII, and RNA production across cell cycle are more accused (Fig. 5C and fig. S6, A and B). In agreement, analysis using published data of PRC2 binding in asynchronous populations of mESCs showed that recruitment of Ezh2, Jarid2, EPOP, and EloB, as well as

Eed, Suz12, Mtf2, and H3K27me3 was higher at the promoters of common targets than at the promoters of remaining genes (fig. S6C). In contrast, common targets were not enriched for trithorax protein MLL2 and H3K4me3 (fig. S6C). Together, these results indicate that the promoter regions of transcription factors that regulate cell differentiation recruit higher levels of PRC2 subunits that are prominently regulated across cell cycle.

Cell cycle–dependent regulation of PRC2 is hindered in Jarid2 −/− mESCs

We next tested how the lack of the PRC2-recruiter Jarid2 affected the cell cycle–dependent regulation of PRC2. We derived FUCCI-*Jarid2* −/− by introducing the FUCCI reporter system into previously derived *Jarid2* −/− mESCs (fig. S7A) (20). Jarid2-depleted mESCs have been reported to display reduced binding of core PRC2 subunits and Ser[5]-RNAPII (18, 20) to bivalent promoters, but they show unchanged levels of EPOP binding at target genes (23). In fitting, depletion of Jarid2 hinders accumulation of H3K27me3 at target promoters during S and G_2-M phases (Fig. 6A and fig. S7, B and C) where binding of Ezh2 and H3K27me3 is more accused in wild-type cells (Fig. 1). Analysis of RNA expression and binding of Ser[5]-RNAPII revealed that depletion of Jarid2 and concomitant decrease in H3K27me3 at bivalent promoters results in derepression of RNA synthesis and loss of Ser[5]-RNAPII during S and G_2-M phases (Fig. 6, B and C, and fig. S7, D and E). Analysis by ChIP-qPCR further confirmed that accumulation of paused Ser[5]-RNAPII at bivalent genes in S and G_2-M phases is visibly reduced in Jarid2 −/− compared to parental mESCs (Fig. 6D and fig. S7, F and G). Depletion of Jarid2 had no detectable effect on the level of H3K27me3, Ser[5]-RNAPII binding, or RNA synthesis in cells in G_1 phase (Fig. 6B and fig. S7B), suggesting that the function of Jarid2 and PRC2.2. might be specific of S and G_2-M phases and dispensable in G_1 phase. Overall, these results demonstrate that increased recruitment of Jarid2 and Ezh2 to bivalent genes during S and G_2-M phases results in enhanced H3K27me3 deposition, pausing of RNAPII, and reduced production of RNA transcripts.

DISCUSSION
Our results show that recruitment of PRC2 complexes to target promoters is regulated across cell cycle and indicate that PRC2.1 complexes are preferentially recruited in G_1 phase, while binding of PRC2.2 is favored in S and G_2-M phases (Fig. 6E). This is supported by our ChIP-seq analysis revealing augmented binding of EPOP and EloB (part of PRC2.1) to target promoters in G_1 phase as opposed to increased binding of Jarid2 (part of PRC2.2) during S and G_2-M phases. We found that binding of the catalytic subunit Ezh2 is markedly increased in S and G_2-M phases compared to G_1 phase, indicating that regulation across cell cycle is not exclusive of accessory subunits but is also happening for the core catalytic subunit of the complex. This is in fitting with previous reports, suggesting a cell cycle–dependent regulation of Ezh2 binding by CDK1 and CDK2 phosphorylation (29). Jarid2 activity is known to favor high levels of PRC2 binding (18–22), while EPOP activity promotes the opposite (23). Thus, reduced recruitment of Ezh2 in G_1 phase is probably a consequence of the accumulation of EPOP-PRC2 (PRC2.1) at the expense of Jarid2-PRC2 (PRC2.2) complexes in this cell cycle phase. We showed that nascent RNA produced in G_1 phase is still in the range of leaky transcription rather than full activation of target genes, suggesting that
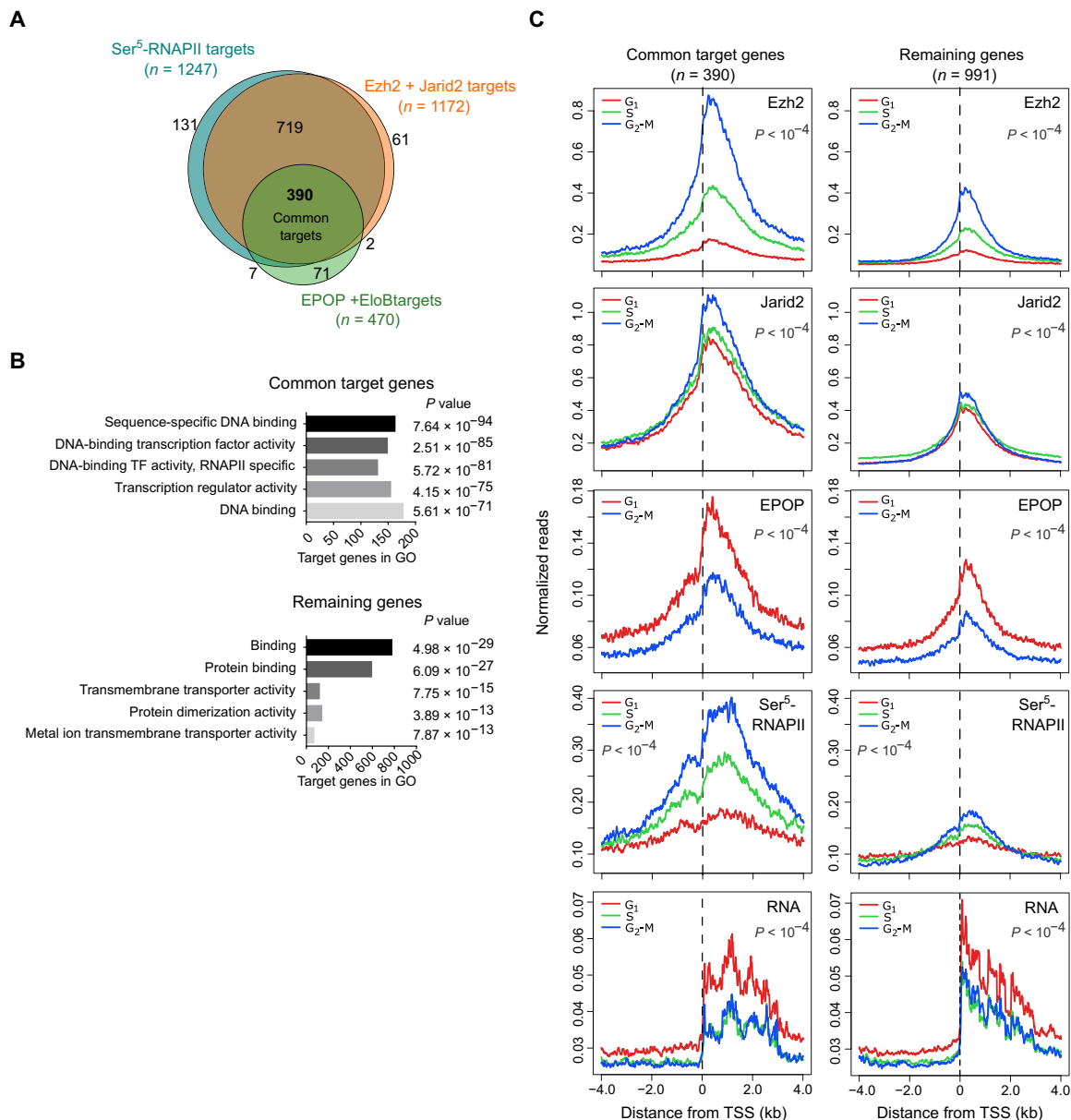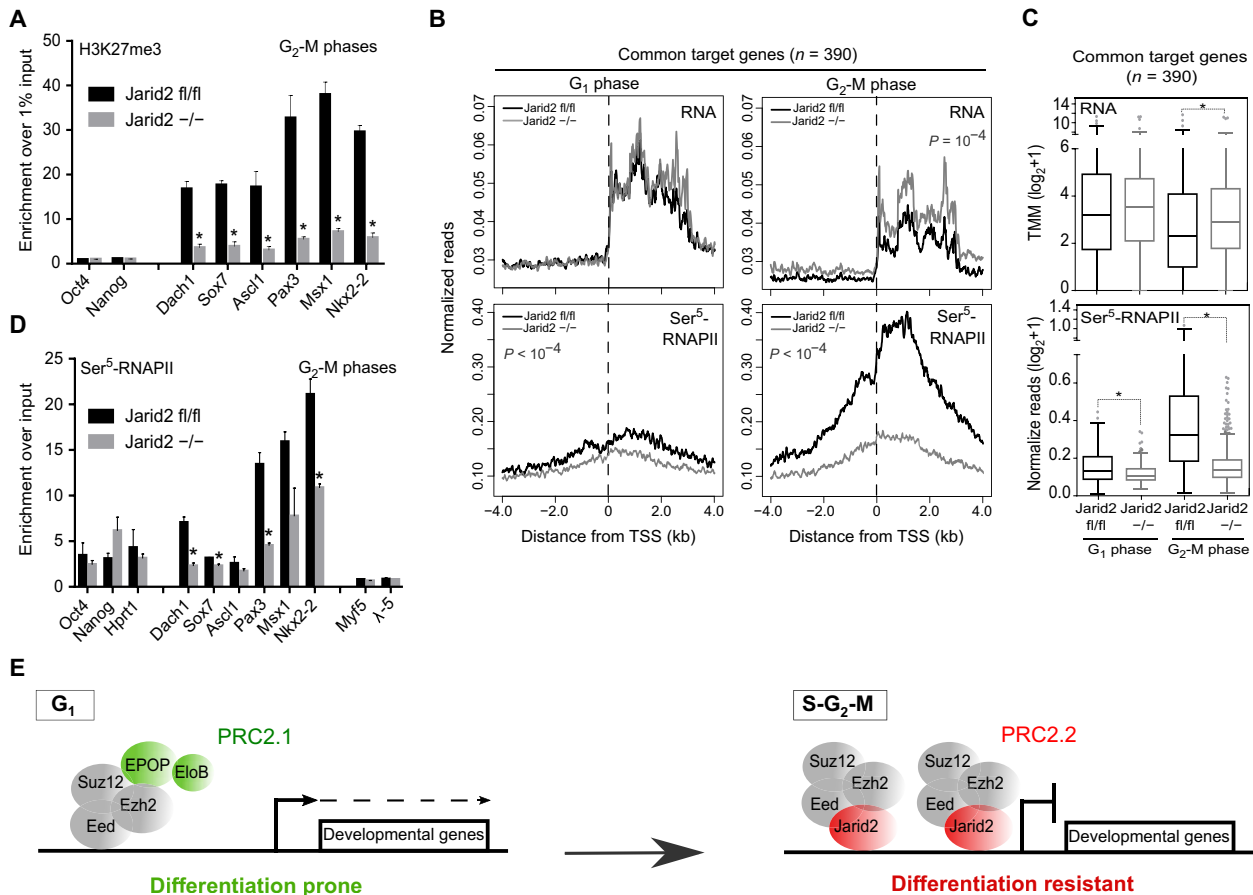
Fig. 5. Cell cycle–dependent regulation of PRC2 is more accused at the promoter of developmental transcription factors. (**A**) Venn diagram showing overlap between HC bivalent genes that are targets of Ser[5]-RNAPII, Ezh2, Jarid2, EPOP, and EloB in asynchronous populations using published (*23*, *36*) and our (Jarid2) datasets. (**B**) Gene Ontology (GO) analysis of common target and remaining genes. Bars represent the number of genes that fall into indicated GO categories. *P* value is shown next to each category. (**C**) Average binding profile of Ezh2, Jarid2, EPOP, Ser[5]-RNAPII, and RNA synthesis around the TSS of HC bivalent genes in G_1 (red), S (green), and G_2-M (blue) comparing common target and remaining HC bivalent genes (as defined in Fig. 5A).

although the amount of Ezh2 bound to promoters of bivalent genes in G_1 phase is lower than in S and G_2-M phases, it is probably enough to maintain substantial gene repression of target genes. Notwithstanding, repression of Ezh2 target genes in G_1 phase might also be dependent on Ezh2-independent mechanisms (*11*), including the activity of Ezh1 or PRC1 complexes.

We found that recruitment of Jarid2-PRC2 (PRC2.2) during S and G_2-M phases leads to more robust gene repression of target bivalent genes; reduced production of RNA coupled with increased binding of paused Ser[5]-RNAPII in S and G_2-M phases that is lost upon Jarid2 depletion. These observations, together with the known

role of EPOP in recruiting EloB and promoting leaky transcription at bivalent genes (*23*, *30*), support that recruitment of EPOP-PRC2 (PRC2.1) to bivalent promoters in G_1 phase favors a transcriptional permissive chromatin setup. Accumulation of Ser[5]-RNAPII is temporally coincident with recruitment of PRC2.2 during S and G_2-M phases. Because simultaneous binding of PRCs and Ser[5]-RNAPII at bivalent gene promoters is a key defining feature of transcriptional priming, our results highlight a previously unanticipated regulation of gene priming within the pluripotent cell cycle. In particular, our results demonstrate that gene priming is evident during S and G_2-M phases (high levels of Ezh2 and Ser[5]-RNAPII), but it is dismantled during

Fig. 6. *Jarid2* −/− mESCs display altered cell cycle–dependent regulation of bivalent genes. (**A**) Analysis by ChIP-qPCR comparing the enrichment of H3K27me3 at PRC2 target promoter regions (*Dach1*, *Sox7*, *Ascl1*, *Pax3*, *Msx1*, and *Nkx2-2*) in *Jarid2* fl/fl (black bars) and *Jarid2* −/− (gray bars) in G_2-M phase. Active (*Oct4* and *Nanog*) gene promoters were used as negative controls. Means ± SEM of four experiments is shown. (**B**) Average RNA synthesis (top) and binding of Ser[5]-RNAPII (bottom) around the TSS of common target bivalent genes in G_1 (left) and G_2-M (right) in *Jarid2* fl/fl (black lines) and *Jarid2* −/− (gray lines) mESCs. (**C**) Quantification of RNA synthesis (top) and Ser[5]-RNAPII binding (bottom) in *Jarid2* fl/fl and *Jarid2* −/− cells in G_1 and G_2-M at common target genes. (**D**) Analysis by ChIP-qPCR comparing the binding of Ser[5]-RNAPII to PRC2 target promoter regions (*Dach1*, *Sox7*, *Ascl1*, *Pax3*, *Msx1*, and *Nkx2-2*) in *Jarid2* fl/fl (black bars) and *Jarid2* −/− (gray bars) in G_2-M phase. Active (*Oct4*, *Nanog*, and *Hprt*) and hypermethylated (*Myf5* and *λ-5*) gene promoters were used as controls. Means ± SEM of three experiments are shown. (**E**) Schematic diagram of observations described here. (A, C, and D) Asterisk (*) marks statistically significant differences.

G_1 phase (low level of Ezh2 and Ser[5]-RNAPII), rather than being maintained until the exit of the pluripotent cell cycle.

Asynchronous populations of pluripotent cells display heterogenous expression of genes and cell differentiation ability (*2*–*4*). This is partly because G_1 phase cells display elevated expression of developmental regulators (*8*), and they are more prone to activate lineage-specific genes in response to differentiation cues (*5*, *6*). We found that mESCs in G_1 phase display a proactivation chromatin setup characterized by enhanced binding of the PRC2 activating subunits EPOP and EloB, together with reduced binding of Jarid2 and Ezh2. We showed that this leads to alleviation of transcriptional repression in G_1 phase. In fitting, it has been reported that human ESCs display increased H3K4me3 at the promoter of bivalent genes during G_1 phase as compared to S and G_2-M phases (*31*). Thus, given that derepression of PRC2 target genes is a critical early event during cell differentiation (*32*), transient alleviation of Polycomb repression in G_1 phase is a key observation to explain why pluripotent cells in G_1 phase are more sensitive to differentiation signals.

Overall, in this study, we showed that Polycomb complexes subunit configuration is controlled by cell cycle–dependent mechanisms in mESCs and support that this type of regulation will be functionally relevant in the context of pluripotent cell differentiation. Our observations are in fitting with the established role of Polycomb as a regulator of pluripotent cell differentiation; however, to fully establish the relevance of cell cycle–mediated regulation of Polycomb function, future studies will need to address how the lack of Polycomb proteins at particular cell cycle phases affect cell differentiation. Because the general principles underlying Polycomb function are widely conserved from flies to humans (*11*), our discovery will probably be relevant for other model systems in which the regulation of gene expression by Polycomb needs to be coordinated with DNA replication and cell division, including adult stem cells and tumor cells (*33*). In addition, PRCs can also regulate gene activity in somatic G_1 phase–arrested cells (*33*) and thus, in the future, it will be interesting to see to what extent differences in the regulation by Polycomb proteins in different model systems are a consequence of their different cell cycle configurations.

METHODS

ESCs growth, FUCCI, and flow cytometry
Stable FUCCI mES cell lines (background 129/Sv/C57BL/C6) were generated for parental and *Jarid2* knockout mESCs (20) by transfecting the ES-FUCCI plasmid (Addgene repository no. 62451) (34). mESCs expressing mCherry:hCdt and Citrine:Geminin were cultured in 5% CO_2 at 37°C on 0.1% gelatin–coated dishes in Dulbecco's modified Eagle's medium knockout (Gibco) media supplemented with 10% fetal calf serum (FCS), leukemia-inhibiting factor (LIF), penicillin/streptomycin (Gibco), L-glutamine (Gibco), 2-mercaptoethanol (Gibco), and hygromycin B (InvivoGen), as described previously (35). Upon trypsinization and resuspension in sorting buffer [phosphate-buffered saline (PBS), 2% FCS, 2 mM EDTA, and LIF] at 4°C, FUCCI-mESCs were cell sorted in an Aria Fusion flow cytometer equipped with 488- and 561-mm lasers to discriminate cells expressing citrine (516 nm/529 nm) and mCherry (587 nm/610 nm). Sorted cells were counted and 1 million cells were used typically for downstream genome-wide analysis. Purity and cell cycle profile of sorted cell populations were routinely checked by propidium iodide staining, followed by flow cytometry. While G_1 and G_2-M fractions were highly enriched for G_1 and G_2-M cells, respectively, S fraction was composed by less homogeneous cell populations; typically, 15% in G_1, 55% in S, and 30% in G_2-M (table S1).

Gene promoter classification
The list of bivalent genes (promoter positive for H3K4me3 and H3K27me3) (3753 genes) was described previously (36). HC bivalent gene list (1678 genes) was obtained by extracting bivalent genes consistently described in three different studies (36–38). Active genes (1557 genes) were determined using published data (36): genes positive for H3K4me3, bound by phosphorylated (Ser[5], Ser2, and Ser7), hypophosphorylated RNAPII (8WG16 antibody), with RNA expression higher than 20 Fragments Per Kilobase of transcript per Million mapped reads (FPKM) and negative for PRC2 binding (Ezh2, Suz12, and H3K27me3). Hypermethylated promoters in mESCs (more than 80% CpG methylation) were identified using published bisulfite sequencing data (39) and cross-analyzed with published data (36) to identify hypermethylated promoters transcriptionally silent (less than 1 FPKM) and not bound by PRC2 (Ezh2, Suz12, and H3K27me3) (656 genes). See table S3 for complete gene lists.

ChIP-qPCR and sequencing
ChIP assays for H3K27me3, Ser[5]-RNAPII, Ezh2, Jarid2, and EPOP (table S4) were performed as described previously (36) with minor modifications: Typically, 1 million cell cycle–sorted cells were resuspended in 37°C complete media (200 μl per million cells) and incubated in a rotating platform for 12 min with 1% formaldehyde at room temperature. To stop the reaction, glycine was added to a final concentration of 125 mM. Swelling and sonication buffers (protease and phosphatase inhibitors supplemented) were used at 4°C in a proportion of 0.5 ml per million cells. Ezh2, H3K27me3, Jarid2, and EPOP ChIPs were carried without using bridge antibody. Chromatin and antibody were incubated in a rotating wheel at 4°C overnight. Protein G magnetic beads (Dynabeads, Invitrogen) were then added and incubated for 5 hours. Washes were carried out for 5 min at 4°C with 1 ml of the following buffers: 1× sonication buffer, 1× wash buffer A [50 mM Hepes (pH 7.9), 500 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.1% Na-deoxycholate, and 0.1% SDS], 1× wash buffer B [20 mM tris (pH 8.0), 1 mM EDTA, 250 mM LiCl, 0.5% NP-40, and 0.5% Na-deoxycholate], and 2× TE buffer (pH 8). ChIP of EloB was performed as above but adding a chromatin double–cross-link step as described in (23) with minor modifications: Cells were resuspended in PBS at 4°C after flow cytometry sorting and incubated with ChIP Cross-link Gold (Diagenode no. C01019027, 0.8 μl in 200 μl of PBS per 1 million cells) in a rotating platform for 30 min at room temperature. After one wash, step chromatin was cross-linked with 1% formaldehyde for 10 min at room temperature. ChIP-qPCR of Ezh2, H3K27me3, EPOP, and Ser[5]-RNAPII and quality control of all immunoprecipitated DNA samples were tested by qPCR using GoTaq qPCR Master Mix (Promega) with a QuantStudio 6 Flex Real-time PCR System (Applied Biosystems). Enrichment was calculated relative to 1% input for all ChIPs except for Ser[5]-RNAPII that was normalized by loading the same amount of DNA as described previously (36). Details of antibodies and primers used are available in table S5.

Libraries of immunoprecipitated DNA in Ezh2, Jarid2, EPOP, and EloB ChIP-seqs were generated from 1 to 5 ng of starting DNA with the NEBNext Ultra DNA Library Prep kit for Illumina (no. 7370) according to the manufacturer's instructions at Centre for Genomic Regulation (CRG) Genomics Core Facility (Barcelona) and sequenced using HiSeq 2500 Illumina technology. Library of Ser[5]-RNAPII ChIP-seq was performed using the NextFlex ChIP-Seq kit (Bioo Scientific no. NOVA-5143-01), starting with 4 ng of immunoprecipitated DNA and sequenced at Centre for Genomics and Oncological Research (GENYO, Granada) using Illumina technology (NextSeq 500) according to the manufacturer's instructions. Twenty million to 30 million reads [50–base pair (bp) single reads] were obtained for each library.

Reads were aligned and quantified using STAR 2.5.2 (40) against GENCODE NCBI m37 (mm9) genome. SAMtools 1.3.1 (41) was used to discard alignments with a quality score <200 to remove multi-mapping reads. Last, we used BamCompare from deepTools suite (42) to create bigwig files with the signal normalized by reads per million (RPM) and against an input sample. Peak calling was performed with MACS2. Data mining of publicly available ChIP-seq datasets (Eed, Suz12, Ezh2, H3K27me3, Jarid2, EPOP, EloB, Mtf2, Mll2, and H3K4me3) were treated the same way. CoverageView (Coverage visualization package for R. R package version 1.20.0.) was used to calculate coverage around TSS.

Average normalized reads (RPM) for a genomic window of −0.5 to +1.5 kb relative to TSS for each analyzed promoter was calculated and represented as boxplots and were subjected to clustering using Cluster 3.0, followed by the Java TreeViewer software. Log_2 of binding values relative to the average were used. Average binding plots were generated by counting normalized reads every 10 bp. In heatmap analyses of reads density in ChIP-seq experiments, we log_2-transformed RPMs and trimmed these values between the minimum 5th percentile and the maximum 95th percentile. To compare different samples, genes were ranked according to G_2-M (Ezh2, Jarid2, and Ser[5]-RNAPII) or G_1 phase (EPOP and EloB). Gene Ontology analysis was performed using the Gene Ontology knowledge database (geneontology.org).

4sU-tagging sequencing
4sU-seq experiments were carried out as described in (27) with minor modifications: Cells were treated with 4sU (500 μM) during 1 hour 37°C before trypsinization and flow cytometry sorting. One million sorted cells were resuspended in 500 μl of TRIzol

(Invitrogen). Isolated RNA was bioanalyzed (Agilent) and was subjected to qPCR analysis as quality control. 4sU-RNA and total RNA samples were retrotranscribed using the RevertAid RT Reverse Transcription kit (Thermo Fisher Scientific no. K1691), treated with deoxyribonuclease I (Thermo Fisher Scientific 18068015) and were subjected to qPCR using primers designed to specifically amplify unspliced RNA (contiguous exon-intron sequences) and total RNA (exon sequences). Strand-specific 4sU-seq libraries were generated using 75 ng of 4sU-RNA and the NextFlex Rapid Directional RNA-seq kit (Bioo Scientific no. NOVA-5138-07) according to the manufacturer's instructions. Libraries were quantified by NanoDrop 2000 (Thermo Fisher Scientific) and bioanalyzed. Fifty million 75-bp paired-end reads per sample were sequenced using Illumina technology (NextSeq 500) at GENYO.

After quality control, we used SortMeRNA 2.1 software (43) to filter out ribosomal RNA reads. We aligned and quantified filtered reads with STAR 2.5.2 (40) using GENCODE NCBI m37 (mm9) as the reference genome. We used SAMtools 1.3.1 (41) to remove alignments with a quality score <200 to discard multimapping reads. Last, we used BamCompare from deepTools suite (42) to create bigwig files with normalized signal in RPM and with positive values for forward strand and negative values in reverse strand. To normalize gene expression values, we applied trimmed mean of M values (44) method with NOISeq package (45).

Western blot analysis and cell fractionation

Whole extracts were prepared for 1 million cells after flow cytometry sorting. Cells were pelleted and resuspended in 50 µl of PBS and 50 µl of 2× Laemmli buffer [0.1 M tris (pH 6.8), 2% SDS, and 5% glycerol] supplemented with protease and phosphatase inhibitors [1× EDTA-free inhibitor cocktail (Roche), 1 mM phenylmethylsulfonyl fluoride, 5 mM NaF, and 2 mM Na3VO4]. Alternatively, 1 million cell cycle–sorted FUCCI wild-type cells subjected to cell fractionation as previously described in (23). Equivalent amount of cells was loaded for Western blots. Western blotting was carried out using standard procedures. Quantification of band intensity and normalization with LaminB was carried out using ImageJ.

Statistical analysis

Statistical analyses were carried out using R 3.5.1. In boxplots, whiskers denote the interval within 1.5× the interquartile range, and P values were calculated using Mann-Whitney test (significant differences $P < 0.0001$). Average mapped reads around the TSS was carried out using analysis of variance (ANOVA), comparing all samples in a window of −0.5 to +1.5 kb from TSS (significant differences $P < 0.0001$). ChIP-qPCR statistical analysis was carried out for triplicates and using Student's t test (significant differences $P < 0.05$).

SUPPLEMENTARY MATERIALS

Supplementary material for this article is available at http://advances.sciencemag.org/cgi/content/full/6/10/eaay4768/DC1

Fig. S1. Differential recruitment of the PRC2 catalytic subunit Ezh2 across cell cycle.
Fig. S2. Cell cycle–dependent regulation of Jarid2.
Fig. S3. Binding of EPOP and EloB to bivalent promoters is regulated across cell cycle.
Fig. S4. Transcriptional activity at PRC2 target genes is attenuated during S and G2-M phases.
Fig. S5. Cell cycle–regulated PRC2 targets are enriched in developmental transcription factors.
Fig. S6. PRC2 target promoters that are regulated across cell cycle display higher levels of PRC2 binding.
Fig. S7. Jarid2 −/− mESCs display abnormal regulation of H3K27me3, Ser5-RNAPII, and RNA synthesis during S phase of the cell cycle.

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