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TGF β and activin A in the tumor microenvironment in colorectal cancer

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Abstract

Although overall survival in colorectal cancer (CRC) is increasing steadily due to progress in screening, therapeutic options and precise diagnostic tools remain scarce. As the understanding of CRC as a complex and multifactorial condition moves forward, the tumor microenvironment has come into focus as a source of diagnostic markers and potential therapeutic targets. The role of TGF β in shifting the epithelial cancer compartment towards invasiveness and a pro-migratory phenotype via stromal signaling has been widely investigated. Accordingly, recent studies have proposed that CRC patients could be stratified into distinct subtypes and have identified one poor prognosis subset of CRC that is characterized by high stromal activity and elevated levels of TGF β . The TGF β superfamily member activin A is crucial for the pro-metastatic properties of the TGF β pathway, yet it has been under-researched in CRC carcinogenesis. In this review, we will elucidate the signaling network and interdependency of both ligands in the context of the tumor microenvironment in CRC.

Keywords

TGF β ; activin A; tumor microenvironment; colorectal cancer

Introduction

Colorectal cancer (CRC) is among the deadliest cancers worldwide, approximately 50,000 annual cancer-associated deaths in the United States alone are attributed to CRC (1). Mortality is mainly due to metastatic disease, with 5-year survival rates as low as 14% in patients diagnosed at advanced stages (2). This circumstance has driven researchers to search for predictive biomarkers that indicate a tumor's propensity to metastasize and ideally to develop therapeutic strategies that specifically combat metastasis-prone tumors. Recent advances have been made in the understanding of colorectal cancer in its entirety, where the

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paradigm of carcinogenesis being only attributed to faulty epithelial cells is being questioned. Investigation of the tumor microenvironment (TME) has given us important insights into the complex metastasis-driven forces that do not originate from but are potentiated by the epithelial axis.

The TME is seen as the non-malignant entity which is comprised of cellular components (fibroblasts, mesenchymal stem cells, osteoblasts, fat cells, blood and immune cells) as well as the extracellular matrix (ECM) scaffold surrounding the cancerous cells (3, 4). The TME has gained a lot of attention as a provider of growth factors, cytokines and other pro-metastatic and anti-apoptotic molecules that orchestrate the malignant cell's ability to migrate. A body of evidence has emerged indicating that the interplay between the tumor and surrounding stroma is pivotal for a cancer to spread to distant sites, as first described by Paget's 'seed and soil theory' (5). Rather than regarding the TME as a stationary compartment, it should be considered as equally prone to changes like its surrounding cancer cells, and there is likely a parallel co-evolution of cancer cells and the TME (6). To co-opt and promote the pro-tumorigenic effects of the TME is necessary for cancer cells to progress, and the more stroma-rich a tumor is, the more aggressive the tumor becomes. Therefore, gene signatures and overexpressed proteins in the cancer tissue that are associated with stromal activity provide potential biomarkers that are indicative of patient prognosis and disease progression (7–11).

Transforming Growth Factor β (TGF β) is major player in tumorigenic stroma-cancer interactions, and the tumor stroma is well known to be a rich source of TGF β (12, 13). Given the abundance of the molecule and its pro-metastatic effects, TGF β was intensively studied and targeted in many clinical trials, although with mixed results (14). The underlying problem of its underperformance as a drug target may be due to its highly context-dependent behavior as a tumor suppressor and promoter, and many aspects of its signaling network are still unclear. TGF β superfamily member activin A is involved in and necessary for some TGF β effects. As such, it has been shown that the invasive, pro-metastatic CRC phenotype induced by TGF β is activin A-dependent. Furthermore, the fact that TGF β and activin A expression correlates on the mRNA level in colorectal tumors and co-occurring mutations in their receptors are frequent, suggests a relationship between these signaling pathways (15). TGF β and activin A are indispensable players in CRC metastasis, and assessment of the two molecules could therefore yield important prognostic information. Since they are not only structurally related, but their signaling pathways are entwined, TGF β and activin A pathways should be viewed as a network, and the ligands should be assessed together (15, 16). Especially in settings of uncertainty of disease relapse (high-risk patients in stage II), TGF β and activin A could serve as biomarkers to stratify patients into tailored chemotherapy regimens, and given their pro-metastatic effects, TGF β and activin A could be exploited as dual drug targets in advanced CRC.

1. The bigger picture: The colorectal cancer microenvironment

The initiative to define a molecular signature for colorectal cancer was undertaken to provide prognostic and predictive information that goes beyond the classical TNM (primary tumor, lymph node, metastasis) staging. However, aside from *KRAS* testing in metastatic

CRC (mCRC) and assessment for microsatellite instability (MSI), a diagnostic tool that encompasses mutational characterization of the epithelium has not been implemented in the clinic. The TME has become a focus of interest in CRC, as it is becoming evident that much of the prognostic information in fact lies in the composition of the environmental factors supporting cancer cell growth and metastases. For example, a high stromal fraction in tumor tissues is associated with poor prognosis and higher tumor staging (10, 17), as evidenced by a correlation between tumor-infiltrating immune cells and patient outcome (18).

The tumor microenvironment initially acts in a tumor-suppressive manner by default; however, at some point in carcinogenesis the TME fails this task and starts promoting pro-tumorigenic pathways. Furthermore, metastatic spread would not be possible without a favorable TME. Thus, researchers in the field have endeavored to identify factors that prompt the TME to switch to a tumor-promoting milieu with pro-metastatic functions. Recent studies indicate that TGF β and activin A are intimately involved in this process (14, 19, 20).

2. TGF β

TGF β , a member of the TGF β superfamily, plays a role in a spectrum of physiologic processes including growth, differentiation, and migration, but is also associated with fibrosis, immune suppression, and carcinogenesis (21–24). Other members of the TGF β superfamily include activins, bone morphogenetic proteins (BMPs), nodals and growth and differentiation factors (GDFs) (25). In the healthy colon epithelium, TGF β is crucial for homeostasis, as increasing gradients from crypt to surface control enterocyte growth. Furthermore, it is a mediator of intestinal immunity (26). There are three isoforms of TGF β : TGF β 1, TGF β 2, TGF β 3, with TGF β 1 being the most prominent. TGF β signals through receptors that are serine-threonine kinases that comprise a heterotetramer of two type I and two type II receptor subunits (TGFBR1 and TGFBR2). A cellular response is elicited through binding of TGF β ligand that initiates the type II receptor phosphorylation of the type I receptor, which in turn allows association of the receptor SMADs (R-SMADs) SMAD2 and SMAD3 (27). After dimerization with SMAD4, the complex translocates to the nucleus to initiate a transcriptional response. Other stimulated pathways that are not SMAD-dependent are considered non-canonical. This heterogeneous group includes PI3K/Akt, MAPK/Erk, WNT/ β -catenin, Rho-like GTPases and JNK/p38 pathways (28, 29). TGF β signaling is highly context-dependent, where the cellular response may be influenced by tissue type, concentration of ligands and mutations in pathway components (30).

Germline mutations in components of TGF β superfamily signaling pathways have been linked to increased risk of developing CRC. For example, increased susceptibility to CRC is observed in individuals harboring germline mutations in the *BMPRI* and *SMAD4* genes, leading to a condition known as juvenile polyposis syndrome (JPS) (31). Patients with JPS develop juvenile hamartomatous lesions in the stomach, small intestine, and colon and have a 50% lifetime risk of developing GI cancer (32). Germline variations in the TGF β receptors are also associated with higher risk of developing CRC, although the extent of the effect is likely modest (32).

TGF β signaling has been described as tumor-suppressive in early stages of carcinogenesis, based on observations of their SMAD-dependent growth inhibition through p21 activation and induction of apoptosis (33–35). However, various reports of TGF β in metastatic CRC add to the complexity of its framework. Inactivating mutations in pathway components such as SMAD4 are seen in 30% of cancers and are typically considered a late stage event associated with metastatic CRC (36). Loss of SMAD4 seems to be an almost exclusive event in microsatellite stable (MSS) cancers. In a study of protein expression of SMAD4 in sporadic colorectal neoplasia, only 4% of MSI carcinomas showed depletion of SMAD4 expression (37). Loss of canonical SMAD expression may be the main factor to co-opt the TGF β signaling network and circumvent the tumor-suppressing effects, leaving only the non-canonical pro-tumorigenic and pro-metastatic functions, such as enhanced cell migration, cell growth and resistance to apoptosis (24, 34, 38, 39). Interestingly, in a study aiming at identifying high risk stage I and II patients, patients with high tumor stroma and loss of SMAD4 had the most unfavorable prognosis compared to stroma low and SMAD4 intact patients (40).

The non-canonical mitogenic MAPK/Erk and the survival-promoting PI3K/Akt pathway are two prominent targets of TGF β , and are known to drive cancer cell malignancy (28). One of the main factors by which TGF β facilitates metastatic spread is by the induction of EMT, a process by which cancer cells lose their epithelial polarity and tight junctions and express mesenchymal proteins such as vimentin. EMT is a hallmark of metastasis and is necessary for cancer cells to acquire mesenchymal characteristics that endow them with the ability to migrate and invade distant tissues (41). In a study of a 5-FU-resistant colorectal cancer cell line, drug resistance was associated with upregulation of EMT markers and a change in cellular morphology. As TGF β is a prominent inducer of EMT, this suggests another mechanism as to how TGF β can render CRC more aggressive (42).

2.1. TGF β in the TME as master regulator of CRC malignancy—TGF β has been extensively studied and is now recognized a main driver of metastasis in CRC. Although many of the mechanisms that alter TGF β 's behavior from tumor suppressor to promoter remain elusive, the field has reached consensus that TGF β is predominantly over-represented in a late stage setting, and in combination with the TME, promotes disease progression and increases the likelihood of metastatic spread (43). TGF β can be used as a prognostic biomarker in CRC, as it is indicative of survival and disease relapse (44). A recent meta-analysis concluded that CRC patients with either elevated TGF β serum levels or high TGF β protein or mRNA expression in the primary tumor have worse overall survival (with a hazard ratio (HR) of 1.68) compared to low TGF β -expressing patients (44). This analysis strongly suggests that TGF β is pivotal for cancer progression. However, mutations of TGF β pathway components are frequent in CRC, such as inactivating mutations in TGFBR2 (45). What may seem paradoxical at first can be explained by the notion that TGF β exerts its pro-metastatic functions through effects on the TME rather than the epithelium. Calon et al. showed that epithelial cancer cells are able to initiate metastasis through a stromal TGF β -guided response (12). According to their study, TGF β response signatures (TBRS) in TME cell types (T-cells, macrophages, and fibroblasts) were predictors of disease relapse in stage I-III patients. This group also developed an *in vivo* model to

investigate the metastatic action of TGF β in the stroma. TGF β signaling was inhibited in a CRC epithelial cell line by mutating TGF β receptor 2. TGF β ligand was then overexpressed in these cells and inoculated in the caecum of nude mice. Compared to mice with non-overexpressing tumors, mice with TGF β -overexpressing tumors developed significantly more metastases. Thus, this study points to the importance of TGF β signaling in the stroma specifically to promote metastasis and underscores the need for further study of the TME in CRC progression (12).

2.2. The immune landscape and its modulation by TGF β —Suppressing antitumor immunity is a hallmark for cancer progression and is controlled by TGF β on many levels. The cytokine represses cytolytic activity of CD8 $^+$ cytotoxic T-lymphocytes (CTL) by inhibiting granzyme B, perforin and FAS-L (46), inhibits T-cell proliferation, reduces antigen spreading (47), reduces T-cell activation, and mediates Treg induction and activity (48–50). The interplay between TGF β and the adaptive immunity in CRC has been demonstrated in a study of an *in vivo* model of the metastatic consensus molecular subtype 4 (CMS4) which is defined by a TGF β -rich stroma and first described by Guinney et al. (51). In this study, upon depletion of CD8 $^+$ CTL and CD4 $^+$ T-helper cells, metastatic tumors were no longer responsive to TGF β RI inhibitor Galunisertib, suggesting that the anti-tumor effect of TGF β pathway inhibition is dependent on a functional adaptive immune system. Furthermore, mice receiving Galunisertib in combination with anti-PD-L1 antibodies had more remissions and longer disease-free survival than mice treated with Galunisertib alone. This finding might be crucial for combatting metastasis, as dual targeting of TGF β and PD-1/PD-L1 might have synergistic effects (51, 52). The authors further demonstrated that in an MSS patient cohort, TGF β 1, 2 and 3 expression negatively correlated with the ratio of TH1/TH-naïve cells, implying that TGF β might suppress T-cell maturation (51, 53). Another report showed that dysregulation of the ECM through TGF β overexpression in ‘immunogenically hot’ tumors predicted failure of treatment with PD-1 inhibitors, again implying TGF β to be a pivotal modulator of the adaptive immunity, and substantiating the rationale of dual targeting of TGF β and PD-1/PD-L1 (54).

The effects by which TGF β regulates the innate immunity are just as meaningful to promote cancer progression (55). As such, TGF β acts as a chemoattractant for neutrophils and polarizes them to the pro-tumorigenic N2-phenotype (56). Subsequently, N2 TANs are able to undermine the antitumor immunity (57). Furthermore, cancer-associated fibroblast (CAFs), which are primarily induced by TGF β , have been shown to shift macrophage populations in CRC towards M2 (58). Taken together, a main mode of action for TGF β is fine-tuning the tumor-promoting effects of the immune landscape in colorectal cancer.

2.3. Desmoplastic tumor stroma and TGF β —In wound healing, epithelial cells are physiologically able to migrate through the tissue by undergoing EMT. That, along with activated contractile myofibroblasts, is necessary for wound closure (59). Desmoplasia, a phenomenon that creates a rigid microenvironment that ‘forces’ tumor cells to undergo EMT and metastasize, is sustained by the pro-fibrotic effects of TGF β (60). As such, TGF β activates EMT in tumor cells, induces α -SMA expression in fibroblasts to transform them to the CAF phenotype, and promotes deposition of ECM components, ultimately leading to

increased tissue rigidity through positive feedback loops (61, 62). As tumors progress, tissue density increases, and cancer cells are confronted with remarkable mechanical forces exerted by the contractile abilities of TGF β -activated CAFs. A stiffening microenvironment further promotes CAF differentiation (63, 64) and increases TGF β secretion (65), creating a vicious cycle and a self-sustained imbalance that ultimately increases the cancer cell's propensity to metastasize.

3. The role of activin A in the TGF β signaling network

Activin A, a member of the TGF β superfamily, was originally described as a multifunctional protein in embryonic development as well as gonadal and pituitary physiology (66). The cytokine has been studied in the context of esophageal (67), skin (68), ovarian (69), lung (70), breast (71, 72), pancreatic (73) and colorectal cancer (74, 75) and may be critical in cancer cachexia (76). Similar to the TGF β pathway, activin A binds to serine/threonine kinase receptors. Three type I receptors (ACVRIA, ACVRAIB, ACVRIC) and two type II receptors (ACVRIIA, ACVRIIB) exist (77). Dimerization of a type I and type II receptor after ligand binding allows phosphorylation of the type I receptor and leads to activation of the canonical SMAD2/3 cascade to elicit a transcriptional response (78).

In the context of CRC, inactivating mutations of ACVRIIA alongside TGFBR2 mutations are very common in patients with MSI CRC. Microsatellite instability causes frequent frameshift mutations within the polyadenine tracts of exon 10 in ACVRIIA and exon 3 in TGFBR2 (79, 80). It has been reported that stage III and high-risk stage II patients with MSI receiving adjuvant chemotherapy harboring defective TGF β receptors have a better 5-year disease-free survival compared to non-TGFBR2 mutated MSI patients (81). However, this survival advantage has not been shown for ACVR2 (82, 83). One study observed mutations in ACVR2 to be associated with metastasis and decreased survival (84). Conflicting with these reports, it has consequently been shown that overexpression of activin A in CRC tissues is associated with stage IV tumors and indicates lower overall survival (74, 85), and serum activin A levels positively correlate with disease stage (86). These discrepancies can be explained by the fact activin A signaling, similar to TGF β signaling, is context dependent. As with TGF β , activin A plays a much larger role in the TME than in the epithelial cells. Therefore, assessing the mutational status of activin pathway components in the epithelial compartment might not yield qualitative prognostic information.

The activation of canonical SMAD2/3 and subsequent dimerization with SMAD4 is the core similarity between activin A and TGF β signaling pathways (87). Despite distinct receptors, activin A and TGF β pathways in CRC are often seen as redundant due to shared canonical SMAD signaling. However, it has been reported that canonical as well as non-canonical signaling patterns are divergent. p21, a primary transcriptional target of activated SMADs, carries out different cellular effects depending on whether the activin A or TGF β signaling pathway is activated. For example, activin A-associated p21 signaling induces apoptosis, whereas TGF β promotes growth suppression (34).

Non-canonical pathways associated with activin A and TGF β are also distinct. It has been shown that activin A induces a downstream PI3K/Akt response, and TGF β engages the MAP/Erk pathway to enable EMT (88). Interestingly, activated Erk is able to phosphorylate

SMAD2 and 3 on alternative sites, which impedes canonical, tumor-suppressive transcriptional activity (89). As concerning non-canonical activin A signaling, the beforementioned loss of SMAD4 causes upregulation of Akt, and in the clinical context, patients with SMAD4 mutations are more likely to show increased protein expression of phosphorylated Akt, which in turn predicts poor prognosis (90).

Activin A is increasingly recognized as a player in the metastatic process in CRC, as it carries out many of the malignant effects of TGF β . As such, in a study of a CRC cell line, the pro-metastatic phenotype of TGF β -treated cells was found to be activin A-dependent (15). The authors of the same study also report that activin A and TGF β should be assessed together, as combined activin A and TGF β protein expression scores in stage II patients yield a better prognostic information than either ligand alone (15). Acknowledging the differences in activin A and TGF β while recognizing them as functionally intertwined is an important milestone and justifies continued investigation.

3.1. Activin A in the TME—Not only is activin A an important component in TGF β signaling, the cytokine itself has many effects on cells of the TME to promote metastasis. Activin A is one of the first responders in wound healing, elevated mRNA expression of *INHBA*, the gene encoding for the β A subunit of activin A, is observed within 24 hours of wound infliction (91). However, perpetual activin A activity in tumors might add to the desmoplastic process, as overexpression of activin A is associated with excessive scarring and fibrosis (92). Strengthening the concept of activin A as a mediator in desmoplasia, CAFs secrete activin A in contrast to non-activated fibroblasts (93, 94), and activin A release can be potentiated by increased tissue stiffness (95). Strikingly, fibroblasts are a richer source of activin A than epithelial cells, and treatment with TGF β leads to increased activin A release in epithelial and stromal cells, again underscoring a close reciprocal relationship of both ligands in the desmoplastic process (15).

The upregulation of MMPs by activin A provides another mechanistic insight into its function in facilitating metastasis. Induction of MMP-7 by activin A is necessary for dissolving the basement membrane and other components of the ECM to enable metastatic spread (96). Early increase of activin A in wounds and inflammatory processes establishes a logical link to its capacity to regulate inflammation and the innate immunity (97). This notion is corroborated by a study of LPS-stimulated mice, where the cytokines TNF- α and IL-1 β were decreased after treatment with activin A antagonist follistatin (98).

It is still under debate whether activin A is considered a pro- or anti-inflammatory cytokine. The answer, as in many cases, may lie in the conditions where the molecule is active. Activin A was shown to have suppressive effects on pro-inflammatory IL-6 in a study of rheumatoid arthritis (99). Interestingly IL-6 regulation in amnion cells seems to be activin A dose-dependent; low quantities of activin A lead to a decrease in IL-6, whereas high amounts lead to a IL-6 increase, highlighting its complex role in coordinating inflammatory processes (100). *INHBA* is overexpressed in patients with inflammatory bowel diseases (101). The role of activin A in CRC-associated inflammation remains elusive. It has been reported that activin A is able to induce both the inflammatory M1 and the pro-tumorigenic

M2 phenotype in macrophages (102, 103). Activin A modulates neutrophil function (104), but further studies are needed to investigate whether activin A influences their polarization.

Activin A has been demonstrated to influence the function of T-cells, as such it is able to induce Treg cells, which suppress antitumor immunity (105–107). Intriguingly, activin A is able to convert CD4+CD25- T-cells into iTreg that express FOXP3+ in a TGF β -dependent manner (108), again hinting at a close relationship between activin A and TGF β signaling. In line with these findings, a recent report showed that Treg induced by TGF β increases mRNA expression of activin-receptor 1 (ACVRI) and activin A ligand (50). This suggests that TGF β and activin A synergize to promote Treg activity and act in tandem to suppress anti-cancer immune responses.

3.2. TGF β and activin A as drug targets—The context-dependent behavior and the various physiologic functions exerted by activin A and TGF β yield challenges to targeting the pathways for cancer treatment. Nevertheless, the prominence of both molecules in advanced CRC makes them attractive targets in a late stage setting, or in situations with high likelihood of metastatic spread. However, patients need to be carefully selected for treatment to specifically combat pro-metastatic effects and ensure tumor-suppressive functions are not abolished. One approach could be to assess for activin A and TGF β in tumor samples with emphasis on the stromal levels of the ligands. Another tool could be to use established fibroblast and T-cell response signatures (F-TBRS, T-TBRS) of TGF β (12, 109) and activin A to help identify patients who would benefit the most from anti-activin A or anti-TGF β treatment. Since the TGF β and activin A signaling networks are intertwined and respective receptors are structurally related, small molecule inhibitors that target both activin A and TGF β receptors could be the most effective option (110). Thus far, targeting the TGF β pathway has been conducted by four different approaches: small molecule inhibitors, neutralizing antibodies, fusion proteins, and vaccines, whereas small molecule inhibitors and one ligand trap are the most promising tools for targeting activin A (Table 1). Current clinical trials are exploring the synergistic effects of drugs targeting the PD-L1 and TGF β pathway, as the combination has been shown to be efficacious in various preclinical modalities of advanced cancer (51, 111). Two ongoing clinical trials are targeting TGF β and PD-L1/PD-1 with separate drugs, and one utilizes a combined PD-L1/TGF β ligand trap (<https://clinicaltrials.gov> : NCT02734160; NCT02423343; NCT03620201). As activin A has a similar potency in suppressing antitumor immunity as TGF β , future studies will show whether combined inhibition of immune checkpoints and activin A can be effective in selected patient populations (68).

Conclusion

Since the discovery of the multistep adenoma-to-carcinoma sequence, it took many years to uncover that cancer cells cooperate with and are controlled by various cues of the TME. This paradigm shift helped us understand the metastasis-driving forces outside the epithelium and has led to the discovery of predictive and prognostic biomarkers, as well as potential therapeutic targets. For successfully combatting CRC, combination therapies that entail specific targets for cancer cells as well as the TME will be the most efficient solution. Investigating activin A and TGF β , key players in the TME, has provided insights into how

these molecules are able to orchestrate the pro-metastatic actions in the TME. Their tumor-promoting behavior in late stage cancers is an excellent example how progressing cancer cells co-opt a signaling network that is tumor-suppressive by nature. Activin A and TGF β could serve as biomarkers for risk stratification in early and advanced tumor stages (high - risk stage II and stage III) and may be promising drug targets in patients where cancers have already metastasized. Future research will elucidate the precise mechanisms by which activin A and TGF β influence functions of all cell types in the TME and will help us understand the crosstalk and synergism of both molecules in CRC. Furthermore, high throughput screening of colorectal tumors will aid to distinguish between a specific subtype of CRC with an activin A/TGF β enriched TME by identifying response signatures of both molecules. Taken together, activin A and TGF β are promising markers and targets in CRC, and their further investigation will help us develop necessary individual tailored therapies.

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Abbreviations

CAF	cancer-associated fibroblast
CMS	consensus molecular subtype
CRC	colorectal cancer
CTL	cytotoxic T-lymphocyte
ECM	extracellular matrix
EMT	epithelial to mesenchymal transition
IL	interleukin
mCRC	metastatic colorectal cancer
MMP	matrix metalloprotease
MSI	microsatellite instability
MSS	microsatellite stable
PD1	programmed cell death protein 1
PD-L1	programmed death-ligand 1
TAM	tumor-associated macrophage
TAN	tumor-associated neutrophil

TBR5	TGF β response signature
TME	tumor microenvironment
TNF-α	tumor necrosis factor α
VEGF	Treg, regulatory T cell, vascular endothelial growth factor
5-FU	fluorouracil

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. *CA: A Cancer Journal for Clinicians*. 2018;68(1):7–30.doi:10.3322/caac.21442 [PubMed: 29313949]
2. Siegel RL, Miller KD, Fedewa SA, Ahnen DJ, Meester RGS, Barzi A, et al. Colorectal cancer statistics, 2017. *CA: A Cancer Journal for Clinicians*. 2017;67(3):177–93.doi:10.3322/caac.21395 [PubMed: 28248415]
3. Peddareddigari VG, Wang D, Dubois RN. The tumor microenvironment in colorectal carcinogenesis. *Cancer Microenvironment*. 2010;3:149–66.doi:10.1007/s12307-010-0038-3 [PubMed: 21209781]
4. Valkenburg KC, De Groot AE, Pienta KJ. Targeting the tumour stroma to improve cancer therapy. *Nature Reviews Clinical Oncology*. 2018;15:366–81.doi:10.1038/s41571-018-0007-1
5. Stephen P The distribution of secondary growths in cancer of the breast. *The Lancet*. 1889;133:571–3.doi:10.1016/S0140-6736(00)49915-0
6. Sleeman JP, Christofori G, Fodde R, Collard JG, Berx G, Decraene C, et al. Seminars in Cancer Biology Concepts of metastasis in flux : The stromal progression model. *Seminars in Cancer Biology*. 2012;22:174–86.doi:10.1016/j.semcancer.2012.02.007 [PubMed: 22374376]
7. Sund M, Kalluri R. Tumor stroma derived biomarkers in cancer. *Cancer and Metastasis Reviews*. 2009;28(1):177–83.doi:10.1007/s10555-008-9175-2 [PubMed: 19259624]
8. Sipos F, Germann TM, Wichmann B, Galamb O, Spisák S, Krenács T, et al. MMP3 and CXCL1 are potent stromal protein markers of dysplasia-carcinoma transition in sporadic colorectal cancer. *European Journal of Cancer Prevention*. 2014;23:336–43.doi:10.1097/CEJ.000000000000058 [PubMed: 24999605]
9. Bhome R, Goh R, Pillar N, Shomron N, Sayan E, Mirnezami A. ExomiRs can distinguish tumor-associated from normal stroma: Potential biomarkers in colorectal cancer. *Cancer Research*. 2018;78:5397 LP - doi:10.1158/1538-7445.AM2018-5397
10. Danielsen HE, Hveem TS, Domingo E, Pradhan M, Kleppe A, Syvertsen RA, et al. Prognostic markers for colorectal cancer: estimating ploidy and stroma. *Annals of Oncology* 2018;29:616–23.doi:10.1093/annonc/mdx794 [PubMed: 29293881]
11. Cammarota R, Bertolini V, Pennesi G, Bucci EO, Gottardi O, Garlanda C, et al. The tumor microenvironment of colorectal cancer: stromal TLR-4 expression as a potential prognostic marker. *Journal of Translational Medicine*. 2010;8(1):112.doi:10.1186/1479-5876-8-112 [PubMed: 21059221]
12. Calon A, Espinet E, Palomo-Ponce S, Tauriello DVF, Iglesias M, Céspedes MV, et al. Dependency of Colorectal Cancer on a TGF- β -Driven Program in Stromal Cells for Metastasis Initiation. *Cancer Cell*. 2012.doi:10.1016/j.ccr.2012.08.013
13. Achyut BR, Yang L. Transforming Growth Factor- β in the Gastrointestinal and Hepatic Tumor Microenvironment. *Gastroenterology*. 2011;141(4):1167–78.doi:10.1053/j.gastro.2011.07.048 [PubMed: 21839702]
14. Neuzillet C, Tijeras-Raballand A, Cohen R, Cros J, Faivre S, Raymond E, et al. Targeting the TGF β pathway for cancer therapy. *Pharmacology and Therapeutics*. 2015;147:22–31.doi:10.1016/j.pharmthera.2014.11.001 [PubMed: 25444759]

15. Staudacher JJ, Bauer J, Jana A, Tian J, Carroll T, Mancinelli G, et al. Activin signaling is an essential component of the TGF- β induced pro-metastatic phenotype in colorectal cancer. *Scientific Reports*. 2017;7:1–9.doi:10.1038/s41598-017-05907-8 [PubMed: 28127051]
16. Loomans HA, Andl CD, Andl CD, Andl CD, Andl CD, Andl CD. Intertwining of activin a and TGF β signaling: Dual roles in cancer progression and cancer cell invasion. *Cancers*. 2014;7:70–91.doi:10.3390/cancers7010070 [PubMed: 25560921]
17. van Pelt GW, Sandberg TP, Morreau H, Gelderblom H, van Krieken JHJM, Tollenaar RAEM, et al. The tumour–stroma ratio in colon cancer: the biological role and its prognostic impact. *Histopathology*. 2018;73(2):197–206.doi:10.1111/his.13489 [PubMed: 29457843]
18. Type Galon J., Density, and Location of Immune Cells Within Human Colorectal Tumors Predict Clinical Outcome. *Science*. 2006;313(5795):1960–4.doi:10.1126/science.1129139 [PubMed: 17008531]
19. Desmouliere A Transforming growth factor-beta 1 induces alpha-smooth muscle actin expression in granulation tissue myofibroblasts and in quiescent and growing cultured fibroblasts. *1993;122(1):103–11.doi:10.1083/jcb.122.1.103*
20. Calon A, Tauriello DVF, Batlle E. TGF-beta in CAF-mediated tumor growth and metastasis. *Seminars in Cancer Biology*. 2014;25:15–22.doi:10.1016/j.semcancer.2013.12.008 [PubMed: 24412104]
21. Weiss A, Attisano L. The TGFbeta superfamily signaling pathway. *Wiley Interdisciplinary Reviews: Developmental Biology*. 2013;2:47–63.doi:10.1002/wdev.86 [PubMed: 23799630]
22. Meng X-M, Nikolic-Paterson DJ, Lan HY. TGF- β : the master regulator of fibrosis. *Nature Reviews Nephrology*. 2016;12(6):325–38.doi:10.1038/nrneph.2016.48 [PubMed: 27108839]
23. Wrzesinski SH, Wan YY, Flavell RA. Transforming Growth Factor- and the Immune Response: Implications for Anticancer Therapy. 2007;13(18):5262–70.doi:10.1158/1078-0432.ccr-07-1157
24. Morikawa M, Derynck R, Miyazono K. TGF- β and the TGF- β Family: Context-Dependent Roles in Cell and Tissue Physiology. *Cold Spring Harbor perspectives in biology*;8(5):a021873.doi:10.1101/cshperspect.a021873
25. Wrana JL. Signaling by the TGF Superfamily. 2013;5(10):a011197–a.doi:10.1101/cshperspect.a011197
26. John A.Barnard, Ginger J.Warwick, Gold I, L. Localization of Transforming Growth Factor β Isoforms in the Normal Murine Small Intestine and Colon *Gastroenterology*. 1993;105(Issue 1):67–73 [PubMed: 8514063]
27. Massague J TGF-beta signal transduction. *Annu Rev Biochem*. 1998;67:753–91.doi:10.1146/annurev.biochem.67.1.753 [PubMed: 9759503]
28. Zhang YE. Non-Smad pathways in TGF- β signaling. *Cell Research*. 2009;19:128–39.doi:10.1038/cr.2008.328 [PubMed: 19114990]
29. Zhang L, Zhou F, Ten Dijke P. Signaling interplay between transforming growth factor- β receptor and PI3K/AKT pathways in cancer. *Trends in Biochemical Sciences*. 2013;38(12):612–20.doi:10.1016/j.tibs.2013.10.001 [PubMed: 24239264]
30. Massagué J TGF β signalling in context. *Nature Reviews Molecular Cell Biology*. 2012;13:616.doi:10.1038/nrm3434 [PubMed: 22992590]
31. Elena M, Stoffel M and Kastanos Fay. Familial CRC—Beyond the Lynch Syndrome. *J Am Coll Surg*. 2011;212:1049–60.doi:10.1016/j.jamcollsurg.2011.02.017.Cost-Effective [PubMed: 21444220]
32. Jung B, Staudacher JJ, Beauchamp D. Transforming Growth Factor β Superfamily Signaling in Development of Colorectal Cancer. *Gastroenterology*. 2017;152:36–52.doi:10.1053/j.gastro.2016.10.015 [PubMed: 27773809]
33. Schuster N, Krieglstein K. Mechanisms of TGF- β -mediated apoptosis. *Cell and Tissue Research*. 2002;307:1–14.doi:10.1007/s00441-001-0479-6 [PubMed: 11810309]
34. Bauer J, Sporn JC, Cabral J, Gomez J, Jung B. Effects of Activin and TGF β on p21 in Colon cancer. *PLoS ONE*. 2012;7:1–11.doi:10.1371/journal.pone.0039381
35. Jakowlew SB. Transforming growth factor- β in cancer and metastasis. *Cancer and Metastasis Reviews*. 2006;25:435–57.doi:10.1007/s10555-006-9006-2 [PubMed: 16951986]

36. Woodford-Richens KL, Rowan AJ, Gorman P, Halford S, Bicknell DC, Wasan HS, et al. SMAD4 mutations in colorectal cancer probably occur before chromosomal instability, but after divergence of the microsatellite instability pathway. 2001;98(17):9719–23.doi:10.1073/pnas.171321498
37. Salovaara R, Roth S, Loukola A, Launonen V, Sistonen P, Avizienyte E, et al. Frequent loss of SMAD4/DPC4 protein in colorectal cancers. 2002;51(1):56–9.doi:10.1136/gut.51.1.56
38. Zhao M, Mishra L, Deng C-X. The role of TGF- β /SMAD4 signaling in cancer. International journal of biological sciences. 2018;14(2):111–23.doi:10.7150/ijbs.23230 [PubMed: 29483830]
39. Muñoz NM, Baek JY, Grady WM. TGF- β has paradoxical and context dependent effects on proliferation and anoikis in human colorectal cancer cell lines. Growth Factors. 2008;26:254–62.doi:10.1080/08977190802291667 [PubMed: 18651288]
40. Mesker WE, Liefers G-J, Junggeburst JMC, van Pelt GW, Alberici P, Kuppen PJK, et al. Presence of a high amount of stroma and downregulation of SMAD4 predict for worse survival for stage I-II colon cancer patients. Cellular oncology : the official journal of the International Society for Cellular Oncology. 2009;31(3):169–78.doi:10.3233/CLO-2009-0478 [PubMed: 19478385]
41. Ye X, Weinberg RA. Epithelial – Mesenchymal Plasticity : A Central Regulator of Cancer Progression. Trends in Cell Biology. 2015;25:675–86.doi:10.1016/j.tcb.2015.07.012 [PubMed: 26437589]
42. Kim AY, Kwak JH, Je NK, Lee Yh, Jung YS. Epithelial-mesenchymal transition is associated with acquired resistance to 5-fluorouracil in HT-29 colon cancer cells. Toxicological Research. 2015;31:151–6.doi:10.5487/TR.2015.31.2.151 [PubMed: 26191381]
43. Calon A, Lonardo E, Berenguer-Llergo A, Espinet E, Hernando-Momblona X, Iglesias M, et al. Stromal gene expression defines poor-prognosis subtypes in colorectal cancer. Nature genetics. 2015;47:320–9.doi:10.1038/ng.3225 [PubMed: 25706628]
44. Chen X-L, Chen Z-Q, Zhu S-L, Liu T-W, Wen Y, Su Y-S, et al. Prognostic value of transforming growth factor-beta in patients with colorectal cancer who undergo surgery: a meta-analysis. BMC Cancer. 2017;17(1).doi:10.1186/s12885-017-3215-7
45. Markowitz SD, Bertagnolli MM. Molecular Basis of Colorectal Cancer. New England Journal of Medicine. 2009;361(25):2449–60.doi:10.1056/nejmra0804588 [PubMed: 20018966]
46. Thomas DA, Massagué J. TGF- β directly targets cytotoxic T cell functions during tumor evasion of immune surveillance. Cancer Cell. 2005;8:369–80.doi:10.1016/j.ccr.2005.10.012 [PubMed: 16286245]
47. Holmgaard RB, Schaer DA, Li Y, Castaneda SP, Murphy MY, Xu X, et al. Targeting the TGF β pathway with galunisertib, a TGF β RI small molecule inhibitor, promotes anti-tumor immunity leading to durable, complete responses, as monotherapy and in combination with checkpoint blockade. Journal for ImmunoTherapy of Cancer. 2018;6(1).doi:10.1186/s40425-018-0356-4
48. Kim R, Emi M, Tanabe K. Cancer immunoediting from immune surveillance to immune escape. Immunology. 2007;121(1):1–14.doi:10.1111/j.1365-2567.2007.02587.x [PubMed: 17386080]
49. Mougiakakos D Regulatory T Cells in Colorectal Cancer: From Biology to Prognostic Relevance. 2011;3(4):1708–31.doi:10.3390/cancers3021708
50. Ni X, Tao J, Barbi J, Chen Q, Park BV, Li Z, et al. YAP Is Essential for Treg-Mediated Suppression of Antitumor Immunity. Cancer Discovery. 2018;8(8):1026–43.doi:10.1158/2159-8290.cd-17-1124 [PubMed: 29907586]
51. Tauriello DVF, Palomo-Ponce S, Stork D, Berenguer-Llergo A, Badia-Ramentol J, Iglesias M, et al. TGF β drives immune evasion in genetically reconstituted colon cancer metastasis. Nature. 2018;554:538–43.doi:10.1038/nature25492 [PubMed: 29443964]
52. Knudson KM, Hicks KC, Luo X, Chen J-Q, Schlom J, Gameiro SR. M7824, a novel bifunctional anti-PD-L1/TGF β Trap fusion protein, promotes anti-tumor efficacy as monotherapy and in combination with vaccine. OncoImmunology. 2018;7(5):e1426519.doi:10.1080/2162402x.2018.1426519 [PubMed: 29721396]
53. Tauriello DVF, Battle E. Targeting the Microenvironment in Advanced Colorectal Cancer. Trends in Cancer. 2016;2:495–504.doi:10.1016/J.TRECAN.2016.08.001 [PubMed: 28741478]
54. Chakravarthy A, Khan L, Bensler NP, Bose P, De Carvalho DD. TGF- β -associated extracellular matrix genes link cancer-associated fibroblasts to immune evasion and immunotherapy failure. Nature Communications. 2018;9(1).doi:10.1038/s41467-018-06654-8

55. Monteleone G A Failure of Transforming Growth Factor- 1 Negative Regulation Maintains Sustained NF- B Activation in Gut Inflammation. 2003;279(6):3925–32.doi:10.1074/jbc.m303654200
56. Mizuno R, Kawada K, Itatani Y, Ogawa R, Kiyasu Y, Sakai Y. The Role of Tumor-Associated Neutrophils in Colorectal Cancer. *International Journal of Molecular Sciences*. 2019;20(3):529.doi:10.3390/ijms20030529
57. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of Tumor-Associated Neutrophil Phenotype by TGF-OI: “N1” versus “N2” TAN. *Cancer Cell*. 2009;16(3):183–94.doi:10.1016/j.ccr.2009.06.017
58. Zhang R, Qi F, Zhao F, Li G, Shao S, Zhang X, et al. Cancer-associated fibroblasts enhance tumor-associated macrophages enrichment and suppress NK cells function in colorectal cancer. *Cell Death & Disease*. 2019;10(4).doi:10.1038/s41419-019-1435-2
59. Sorg H, Tilkorn DJ, Hager S, Hauser J, Mirastschijski U. Skin Wound Healing: An Update on the Current Knowledge and Concepts. *European Surgical Research*. 2017;58(1–2):81–94.doi:10.1159/000454919 [PubMed: 27974711]
60. Wei SC, Yang J. Forcing through Tumor Metastasis : The Interplay between Tissue Rigidity and Epithelial – Mesenchymal Transition. 2016;26:111–20
61. Leask A Targeting the TGFβ, endothelin-1 and CCN2 axis to combat fibrosis in scleroderma. 2008;20(8):1409–14.doi:10.1016/j.cellsig.2008.01.006
62. Ling E, Robinson DS. Transforming growth factor-bbeta1: its anti-inflammatory and pro-fibrotic effects. *Clinical Experimental Allergy*. 2002;32(2):175–8.doi:10.1046/j.1365-2222.2002.01287.x [PubMed: 11929477]
63. Hinz B Tissue stiffness, latent TGF-β1 Activation, and mechanical signal transduction: Implications for the pathogenesis and treatment of fibrosis. 2009;11(2):120–6.doi:10.1007/s11926-009-0017-1
64. Johnson LA, Rodansky ES, Haak AJ, Larsen SD, Neubig RR, Higgins PDR. Novel Rho/MRTF/SRF Inhibitors Block Matrix-stiffness and TGF-β-Induced Fibrogenesis in Human Colonic Myofibroblasts. 2014;20(1):154–65.doi:10.1097/01.mib.0000437615.98881.31
65. Pang M, Teng Y, Huang J, Yuan Y, Lin F, Xiong C. Substrate stiffness promotes latent TGF-β1 activation in hepatocellular carcinoma. 2016.doi:10.1016/j.bbrc.2016.12.107
66. Xia Y, Schneyer AL. The biology of activin: Recent advances in structure, regulation and function. *Journal of Endocrinology*. 2009;202:1–12.doi:10.1677/JOE-08-0549 [PubMed: 19273500]
67. Taylor C, Loomans HA, Bras GFL, Koumangoye RB, Romero-morales AI, Quast LL, et al. Activin a signaling regulates cell invasion and proliferation in esophageal adenocarcinoma. *Oncotarget*. 2015;6.doi:10.18632/oncotarget.5349
68. Donovan P, Dubey OA, Kallioinen S, Rogers KW, Muehlethaler K, Müller P, et al. Paracrine Activin-A Signaling Promotes Melanoma Growth and Metastasis through Immune Evasion. *Journal of Investigative Dermatology*. 2017;137(12):2578–87.doi:10.1016/j.jid.2017.07.845 [PubMed: 28844941]
69. Dean M, Davis DA, Burdette JE. Activin A stimulates migration of the fallopian tube epithelium, an origin of high-grade serous ovarian cancer, through non-canonical signaling. *Cancer letters*. 2017;391:114–24.doi:10.1016/j.canlet.2017.01.011 [PubMed: 28115208]
70. Hoda MA, Rozsas A, Lang E, Klikovits T, Lohinai Z, Torok S, et al. High circulating activin A level is associated with tumor progression and predicts poor prognosis in lung adenocarcinoma. *Oncotarget*. 2016;7.doi:10.18632/oncotarget.7796
71. Bashir M, Damineni S, Mukherjee G, Kondaiah P. Activin-A signaling promotes epithelial-mesenchymal transition, invasion, and metastatic growth of breast cancer. *NPJ breast cancer*. 2015;1:15007-.doi:10.1038/npjbcancer.2015.7 [PubMed: 28721365]
72. Xie D, Liu Z, Wu J, Feng W, Yang K, Deng J, et al. The effects of activin A on the migration of human breast cancer cells and neutrophils and their migratory interaction. *Experimental Cell Research*. 2017;357:107–15.doi:10.1016/J.YEXCR.2017.05.003 [PubMed: 28479070]
73. Togashi Y, Kogita A, Sakamoto H, Hayashi H, Terashima M, de Velasco MA, et al. Activin signal promotes cancer progression and is involved in cachexia in a subset of pancreatic cancer. *Cancer Letters*. 2015;356:819–27.doi:10.1016/J.CANLET.2014.10.037 [PubMed: 25449777]

74. Wildi S, Kleeff J, Maruyama H, Maurer CA, Büchler MW, Korc M. Overexpression of activin A in stage IV colorectal cancer. *Gut*. 2001;49:409–17.doi:10.1136/gut.49.3.409 [PubMed: 11511564]
75. Refaat B, El-Shemi AG, Mohamed AM, Kensara OA, Ahmad J, Idris S. Activins and their related proteins in colon carcinogenesis: Insights from early and advanced azoxymethane rat models of colon cancer. *BMC Cancer*. 2016;16:1–16.doi:10.1186/s12885-016-2914-9
76. Loumaye A, de Barsy M, Nachit M, Lause P, van Maanen A, Trefois P, et al. Circulating Activin A predicts survival in cancer patients. *Journal of Cachexia, Sarcopenia and Muscle*. 2017;8:768–77.doi:10.1002/jcsm.12209
77. Tsuchida K, Nakatani M, Uezumi A, Murakami T, Cui X. Signal Transduction Pathway through Activin Receptors as a Therapeutic Target of Musculoskeletal Diseases and Cancer. 2008;55(1):11–21.doi:10.1507/endocrj.kr-110
78. Attisano L, Wrana JL, Montalvo E, Massagué J. Activation of signalling by the activin receptor complex. *Molecular and cellular biology*. 1996;16:1066–73 [PubMed: 8622651]
79. Jung B, Doctolero RT, Tajima A, Nguyen AK, Keku T, Sandler RS, et al. Loss of Activin Receptor Type 2 Protein Expression in Microsatellite Unstable Colon Cancers. *Gastroenterology*. 2004;126:654–9.doi:10.1053/j.gastro.2004.01.008 [PubMed: 14988818]
80. Biswas S, Trobridge P, Romero-Gallo J, Billheimer D, Myeroff LL, Willson JKV, et al. Mutational inactivation of TGFBR2 in microsatellite unstable colon cancer arises from the cooperation of genomic instability and the clonal outgrowth of transforming growth factor β resistant cells. 2008;47(2):95–106.doi:10.1002/gcc.20511
81. Watanabe T, Wu T-T, Catalano PJ, Ueki T, Satriano R, Haller DG, et al. Molecular Predictors of Survival after Adjuvant Chemotherapy for Colon Cancer. *New England Journal of Medicine*. 2001;344(16):1196–206.doi:10.1056/nejm200104193441603 [PubMed: 11309634]
82. Yuza K, Nagahashi M, Ichikawa H, Nakajima M, Hanyu T, Ishikawa T, et al. Association of activin type II receptor mutation with microsatellite instability in gastric cancer. *Journal of Clinical Oncology*. 2017;35(15_suppl):e23191–e.doi:10.1200/JCO.2017.35.15_suppl.e23191
83. Wodzi ski D, Wosiak A, Pietrzak J, wiechowski R, Jele A, Balcerzak E. Does the expression of the ACVR2A gene affect the development of colorectal cancer? *Genetics and molecular biology*. 2019;42(1):32–9.doi:10.1590/1678-4685-GMB-2017-0332 [PubMed: 30856244]
84. Zhuo C, Hu D, Li J, Yu H, Lin X, Chen Y, et al. Downregulation of Activin A Receptor Type 2A Is Associated with Metastatic Potential and Poor Prognosis of Colon Cancer. *Journal of Cancer*. 2018;9(19):3626–33.doi:10.7150/jca.26790 [PubMed: 30310521]
85. Okano M, Yamamoto H, Ohkuma H, Kano Y, Kim H, Nishikawa S, et al. Significance of INHBA expression in human colorectal cancer. *Oncology Reports*. 2013;30:2903–8.doi:10.3892/or.2013.2761 [PubMed: 24085226]
86. Wu S, Qi Y, Niu LM, Xie DX, Cui XL, Liu ZH. Activin A as a novel biomarker for colorectal adenocarcinoma in humans. *European Review for Medical and Pharmacological Sciences*. 2015;19:4371–8 [PubMed: 26636525]
87. Zhou S, Buckhaults P, Zawel L, Bunz F, Riggins G, Le Dai J, et al. Targeted deletion of Smad4 shows it is required for transforming growth factor and activin signaling in colorectal cancer cells. *Proceedings of the National Academy of Sciences*. 1998;95:2412–6.doi:10.1073/pnas.95.5.2412
88. Bauer J, Ozden O, Akagi N, Carroll T, Principe DR, Staudacher JJ, et al. Activin and TGF β use diverging mitogenic signaling in advanced colon cancer. *Molecular Cancer*. 2015;14:1–14.doi:10.1186/s12943-015-0456-4 [PubMed: 25560632]
89. Kretschmar M, Doody J, Timokhina I, Massagué J. A mechanism of repression of TGFbeta/Smad signaling by oncogenic Ras. *Genes & development*. 1999;13(7):804–16.doi:10.1101/gad.13.7.804 [PubMed: 10197981]
90. Zhang B, Zhang B, Chen X, Bae S, Singh K, Washington MK, et al. Loss of Smad4 in colorectal cancer induces resistance to 5-fluorouracil through activating Akt pathway. 2014;110(4):946–57.doi:10.1038/bjc.2013.789
91. Hübner G, Hu Q, Smola H, Werner S. Strong Induction of Activin Expression after Injury Suggests an Important Role of Activin in Wound Repair. 1996;173(2):490–8.doi:10.1006/dbio.1996.0042
92. Werner S, Alzheimer C. Roles of activin in tissue repair, fibrosis, and inflammatory disease. 2006;17(3):157–71.doi:10.1016/j.cytogfr.2006.01.001

93. Gascard P, Tlsty TD. Carcinoma-associated fibroblasts: orchestrating the composition of malignancy. *Genes & Development*. 2016;30(9):1002–19.doi:10.1101/gad.279737.116 [PubMed: 27151975]
94. Sobral LM, Bufalino A, Lopes MA, Graner E, Salo T, Coletta RD. Myofibroblasts in the stroma of oral cancer promote tumorigenesis via secretion of activin A. 2011;47(9):840–6.doi:10.1016/j.oraloncology.2011.06.011
95. Emon B, Bauer J, Jain Y, Jung B, Saif T. Biophysics of Tumor Microenvironment and Cancer Metastasis - A Mini Review. *Computational and Structural Biotechnology Journal*. 2018;16:279–87.doi:10.1016/j.csbj.2018.07.003 [PubMed: 30128085]
96. Yoshinaga K, Mimori K, Inoue H, Kamohara Y, Yamashita K, Tanaka F, et al. Activin A enhances MMP-7 activity via the transcription factor AP-1 in an esophageal squamous cell carcinoma cell line. *International Journal of Oncology*. 1992;33:453–9.doi:10.3892/ijo_00000027
97. Sideras P, Apostolou E, Stavropoulos A, Sountoulidis A, Gavriil A, Apostolidou A, et al. Activin, neutrophils, and inflammation: just coincidence? 2013;35(4):481–99.doi:10.1007/s00281-013-0365-9
98. Jones KL, Mansell A, Patella S, Scott BJ, Hedger MP, De Kretser DM, et al. Activin A is a critical component of the inflammatory response, and its binding protein, follistatin, reduces mortality in endotoxemia. 2007;104(41):16239–44.doi:10.1073/pnas.0705971104
99. Yu Dolter, Shao Yu. Suppression of IL-6 biological activities by activin A and implications for inflammatory arthropathies. 1998;112(1):126–32.doi:10.1046/j.1365-2249.1998.00522.x
100. Keelan JA, Zhou RL, Mitchell MD. Activin A Exerts both Pro- and Anti-inflammatory Effects on Human Term Gestational Tissues. 2000;21(1):38–43.doi:10.1053/plac.1999.0451
101. Hübner G, Brauchle M, Gregor M, Werner S. Activin A: a novel player and inflammatory marker in inflammatory bowel disease? *Laboratory investigation; a journal of technical methods and pathology*. 1997;77(4):311–8 [PubMed: 9354766]
102. Ogawa K, Funaba M, Chen Y, Tsujimoto M. Activin A Functions as a Th2 Cytokine in the Promotion of the Alternative Activation of Macrophages. 2006;177(10):6787–94.doi:10.4049/jimmunol.177.10.6787
103. Sierra-Filardi E, Puig-Kroger A, Blanco FJ, Nieto C, Bragado R, Palomero MI, et al. Activin A skews macrophage polarization by promoting a proinflammatory phenotype and inhibiting the acquisition of anti-inflammatory macrophage markers. *Blood*. 2011;117(19):5092–101.doi:10.1182/blood-2010-09-306993 [PubMed: 21389328]
104. Qi Y, Ge J, Ma C, Wu N, Cui X, Liu Z. Activin A regulates activation of mouse neutrophils by Smad3 signalling. *Open Biology*;7(5):160342.doi:10.1098/rsob.160342
105. Antsiferova M, Werner S. The bright and the dark sides of activin in wound healing and cancer. *J Cell Sci*. 2012;125(Pt 17):3929–37.doi:10.1242/jcs.094789 [PubMed: 22991378]
106. Semitekolou M, Alissafi T, Aggelakopoulou M, Kourepini E, Kariyawasam HH, Kay AB, et al. Activin-A induces regulatory T cells that suppress T helper cell immune responses and protect from allergic airway disease. *The Journal of experimental medicine*. 2009;206(8):1769–85.doi:10.1084/jem.20082603 [PubMed: 19620629]
107. Tousa S, Semitekolou M, Morianos I, Banos A, Trochoutsou AI, Brodie TM, et al. Activin-A co-opts IRF4 and AhR signaling to induce human regulatory T cells that restrain asthmatic responses. *Proceedings of the National Academy of Sciences*. 2017;114(14):E2891–E900.doi:10.1073/pnas.1616942114
108. Huber S, Stahl FR, Schrader J, Lüth S, Presser K, Carambia A, et al. Activin A Promotes the TGF- β -Induced Conversion of CD4⁺CD25⁻ T Cells into Foxp3⁺ Induced Regulatory T Cells. *The Journal of Immunology*. 2009;182(8):4633.doi:10.4049/jimmunol.0803143 [PubMed: 19342638]
109. Mariathasan S, Turley SJ, Nickles D, Castiglioni A, Yuen K, Wang Y, et al. TGF β attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. *Nature*. 2018;554(7693):544–8.doi:10.1038/nature25501 [PubMed: 29443960]
110. Fu K, Corbley MJ, Sun L, Friedman JE, Shan F, Papadatos JL, et al. SM16, an Orally Active TGF- β Type I Receptor Inhibitor Prevents Myofibroblast Induction and Vascular Fibrosis in the

- Rat Carotid Injury Model. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 2008;28(4):665–71. doi:doi:10.1161/ATVBAHA.107.158030
111. Lan Y, Zhang D, Xu C, Hance KW, Marelli B, Qi J, et al. Enhanced preclinical antitumor activity of M7824, a bifunctional fusion protein simultaneously targeting PD-L1 and TGF- β .
112. Simonelli M, Zucali P, Santoro A, Thomas MB, De Braud FG, Borghaei H, et al. Phase I study of PF-03446962, a fully human monoclonal antibody against activin receptor-like kinase-1, in patients with hepatocellular carcinoma. 2016;27(9):1782–7. doi:10.1093/annonc/mdw240
113. Keedy VL, Bauer TM, Clarke JM, Hurwitz H, Baek I, Ha I, et al. Association of TGF- β responsive signature with anti-tumor effect of vactosertib, a potent, oral TGF- β receptor type I (TGFBR1) inhibitor in patients with advanced solid tumors. *Journal of Clinical Oncology*. 2018;36(15_suppl):3031-. doi:10.1200/JCO.2018.36.15_suppl.3031 [PubMed: 30199311]
114. Formenti SC, Lee P, Adams S, Goldberg JD, Li X, Xie MW, et al. Focal irradiation and systemic TGF β blockade in metastatic breast cancer. *Clinical Cancer Research*. 2018;24:2493–504. doi:10.1158/1078-0432.CCR-17-3322 [PubMed: 29476019]
115. Raftopoulos H, Laadem A, Puccio M, Knight RD. A phase II/III study of sotatercept (ACE-011), an activin antagonist, for chemotherapy-induced anemia in patients with metastatic non-small cell lung cancer treated with first-line platinum-based chemotherapy. *Journal of Clinical Oncology*. 2011;29(15_suppl):TPS235–TPS. doi:10.1200/jco.2011.29.15_suppl.tps235
116. Tao JJ, Cangemi NA, Makker V, Cadoo KA, Liu JF, Rasco DW, et al. First-in-Human Phase I Study of the Activin A Inhibitor, STM 434, in Patients with Granulosa Cell Ovarian Cancer and Other Advanced Solid Tumors. *Clinical Cancer Research*. 2019. doi:10.1158/1078-0432.CCR-19-1065

Highlights

- The tumor microenvironment is a pivotal driver of colorectal cancer metastasis
- TGF β and activin A are the main determinants of a stroma-rich colorectal cancer subtype with poor prognosis
- Assessing activin A and TGF β in tumors can identify a metastasis-prone subset of colorectal cancer and guide individual-tailored therapies

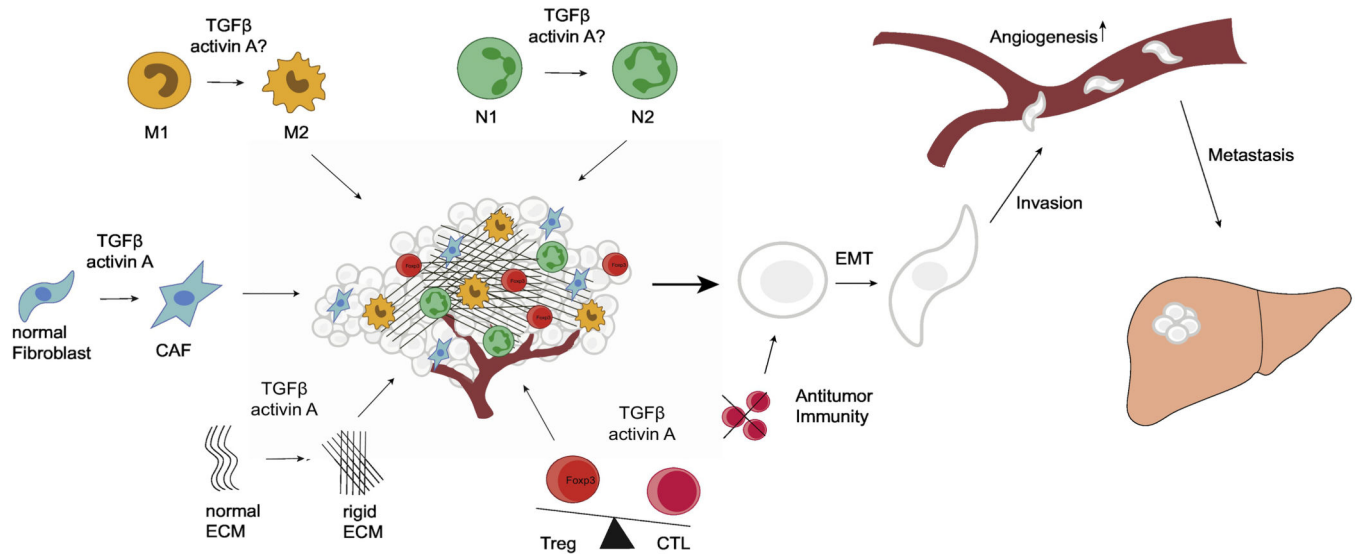


Figure 1:

The metastasis-promoting effects on the tumor microenvironment by activin A and TGFβ. Whereas TGFβ has been shown to shift neutrophil and macrophage populations towards N2/M2, the effects of activin A on those leukocytes are not as clear. Both molecules can induce the CAF-phenotype in fibroblasts, and reciprocally participate in the desmoplastic process in tumors. Desmoplasia is potentiated by a rigid ECM and the abundance of CAFs. Activin A and TGFβ are efficient suppressors of antitumor immunity and are able to induce immunosuppressive regulatory T-cells. The effects of activin A and TGFβ on the tumor microenvironment lead to increased cancer cells migration through induction of EMT, as well as angiogenesis and cancer cell invasiveness to ultimately accelerate the metastatic process.

(N1 classically activated neutrophils, N2 pro-tumor neutrophils, M1 classically activated macrophages, M2 pro-tumor macrophages, CAF cancer-associated fibroblasts CTL cytotoxic T-cells, Treg regulatory T-cells, EMT epithelial-to-mesenchymal transition)

Table 1:

Therapeutic approaches to inhibit components of the activin A and TGF β pathways in solid tumors. (ALK-1 activin receptor-like kinase, TGFBR1 TGF β receptor 1, ACVR1 activin receptor 1)

Name of drug	Target	Clinical phase	Type of cancer	Reference	Trial registration number	Status/Main outcome
TGFβ targeting agents						
<u>Small molecule inhibitors</u>						
PF-03446962	ALK-1	Phase I	Hepatocellular carcinoma (HCC)	(112)	NCT00557856	Completed/No complete or partial responses. 50% had stable disease
TEW-7197 (Vactosertib)	TGFBR1	Phase I	Advanced stage solid tumors	(113)	NCT02160106	Completed/Well tolerated, Vactosertib showed increased efficacy in patients with high fibroblast TGF β response signature (F-TBR5)
LY3200882	TGFBR1	Phase I	Solid tumors		NCT02937272	recruiting
LY2157299 (Galunisertib)	TGFBR1	Phase I/II	Metastatic colorectal cancer (mCRC)		NCT03470350	recruiting
		Phase II	Metastatic Prostate cancer		NCT02452008	recruiting
<u>Ligand traps</u>						
AVID200	TGF β 1, TGF β 3	Phase I	Metastatic solid tumors		NCT03834662	recruiting
<u>Vaccines</u>						
bi-shRNA ^{furin} /GMCSF DNA/ Autologous Tumor Cell Vaccine (FANG)	short hairpin RNAi (bi-shRNAi) targeting furin convertase to block TGF β 1 and TGF β 2 activation	Phase II	Metastatic colorectal cancer (mCRC)		NCT01505166	terminated
<u>Antibodies</u>						
GC1008 (Fresolimumab)	TGFBR1	Phase II	Metastatic breast cancer	(114)	NCT01401062	Completed/ Fresolimumab + radiation therapy, patients receiving 10mg/kg had significantly higher median overall survival than patients with 1mg/kg
Activin A targeting Agents						
<u>Small molecule inhibitors</u>						
ACE-011 (Sotatercept)	ACVR1I	Phase II	Non-small cell lung cancer (NSCLC)	(115)	NCT01284348	terminated
		Phase II	Metastatic breast cancer		NCT00931606	terminated
<u>Ligand traps</u>						
STM 434	receptor-Fc fusion protein	Phase I	Advanced solid tumors	(116)	NCT02262455	Completed/53.5% had stable disease

Name of drug	Target	Clinical phase	Type of cancer	Reference	Trial registration number	Status/Main outcome
	against activin A					

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