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Safety of cephalosporins in penicillin class severe delayed hypersensitivity reactions

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To the Editor,

Currently, a significant driver of Ig-E-mediated cross-reactivity between penicillins/cephalosporins is thought to be the R1 side chain, with contemporary cephalosporin cross-reactivity with penicillin allergy occurring at a rate of < 2%¹. However, the extent to which there is cross-reactivity between drugs within the penicillin class in patients with severe delayed and presumed T-cell mediated reactions is unknown.

A prospective multicentre cohort study was performed at Austin Health and Peter MacCallum Cancer Centre in Melbourne Victoria, between 1st April 2015 and 24th February 2019. Study participants included patients referred for testing with a history of a severe T-cell mediated hypersensitivity associated with a penicillin. A penicillin was defined as any drug within the penicillin class and in our cohort included: penicillin VK, penicillin G, flucloxacillin, dicloxacillin, amoxicillin, amoxicillin-clavulanate, ampicillin or piperacillin-tazobactam.

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A severe T-cell mediated hypersensitivity syndrome was defined as drug reaction with eosinophilia and systemic symptoms (DRESS), acute generalized exanthematous pustulosis (AGEP) or severe maculopapular exanthem (MPE). Patients experiencing Stevens-Johnson syndrome and toxic epidermal necrolysis (SJS/TEN) associated with a penicillin were excluded. For phenotypes of DRESS and AGEP, a RegiSCAR score of ≥ 4 (probable or definite) and an AGEP score of ≥ 2 respectively were required^{2, 3}. Severe MPE was defined as an extensive cutaneous exanthem with more than 50% of body surface area and RegiSCAR score of 2-3 (possible)². All cases had at least one antibiotic that had been administered within 5 drug half-lives of onset of rash, a Naranjo score of ≥ 5 , phenotype confirmed by dermatologist or histopathology, and had at least three investigations to exclude common alternative causes such as infections or autoimmune diseases.

Both sites are tertiary referral testing centres with established drug and antibiotic allergy testing programs utilizing previously published *in vivo* (skin prick testing [SPT] and intradermal testing [IDT]) testing protocols including the highest non-irritating drug concentrations where possible^{4, 5}. As previously described, skin testing (SPT/IDT) and patch testing (PT) was performed no earlier than 6 weeks following the resolution of cutaneous manifestations utilizing the previously published method^{4, 5}. The routine IDT panel included: Normal Saline (0.9% solution), penicillin G (1000 IU/ml, 10,000IU/ml), DAP-major (benzylpenicilloyl poly-L-lysine; final concentration 1.07×10^{-2} mol/L), minor-determinate mixture (MDM; sodium benzylpenicillin, benzylpenicilloic acid, sodium benzylpenicilloate; 1.5 mg/mL; final concentration 1.5 mol/L), ampicillin (25mg/ml), flucloxacillin (2mg/ml), cefazolin (1mg/ml) and ceftriaxone (2.5mg/ml). Piperacillin-tazobactam (4.5mg/ml) IDT was performed in patients reporting a primary piperacillin-tazobactam allergy or were immunocompromised with a reported penicillin allergy. All test reagents (no excipients) were diluted in water or Normal Saline. Skin test positive was defined as per previous definitions, in brief a delayed IDT was considered positive when an infiltrated erythema with a diameter of greater than 5mm was present as per previous definition.⁵ Skin testing utilizing the aforementioned routine IDT panel was performed in 6 healthy controls and patients with a history of IgE mediated penicillin hypersensitivity (Supplementary Table E1).

Peripheral blood mononuclear cells (PBMCs) were isolated from whole heparinised blood of patients at time of IDT. PBMCs were stored at -80°C in 90% heat-inactivated fetal bovine serum and 10% DMSO until IFN- γ release Enzyme Linked ImmunoSpot (ELISpot) assay and DNA extraction was performed as per previously published methods.⁶ (Supplementary materials) Ethics approval was obtained from the Austin Health Research Ethics Committee (Approval 15/Austin/75).

During the study period 724 patients completed SPT/IDT or PT for a suspected antibiotic allergy. Among the 724 patients, 1163 antibiotic allergy labels were reported (905 [77.8%] beta-lactam; 680 [58.5%] penicillin). 602 patients (83%) reported penicillin-associated hypersensitivity, 216 with delayed hypersensitivity and 32 with a severe T-cell mediated hypersensitivity. Of these 32 patients with delayed and presumed T-cell mediated hypersensitivity, 14 (44%) were negative to all reagents, 6 (19%) positive to ≥ 2 tested

reagents (Supplementary Table E2) and 12 (38%) had a positive intradermal test documented to > 2 reagents from the routine IDT panel (Figure 1, Table 1).

The patient phenotypes and characteristics of the 12 patients positive to > 2 intradermal test reagents are demonstrated in Table 1. Briefly, the phenotypes were DRESS (3/12; 25%), AGEF (3/12; 25%) and severe MPE (6/12; 50%). The primary implicated penicillins were piperacillin-tazobactam (6, 50%); amoxicillin (4, 33%) and flucloxacillin (2, 17%). The median age was 52.5 years (IQR 36-48), 50% (6/12) female, and 5 (41.6%) immunocompromised (solid organ transplant recipient, cancer, autoimmune/connective tissue disorder requiring immunomodulating therapy). The median time between rash onset and intradermal testing and PBMC sampling was 395.5 days (IQR 195-1308). Positive reactions to IDT occurred as early as 6 hours post inoculation and all patients were positive by 24 hours with persistence of skin redness and induration for greater than 72 hours. No systemic adverse events to skin testing were reported. All patients were positive to tested IDT concentrations of ampicillin, penicillin G and flucloxacillin and negative to 0.9% *N. saline*, PPL (neat), MDM (neat), cefazolin and ceftriaxone (Figure 1). In patients with piperacillin-tazobactam as the primary implicated drug or immunocompromised, piperacillin-tazobactam IDT was performed and positive in all tested (8/8). A similar delayed pattern was not observed in healthy controls or in 255 patients with immediate IgE mediated hypersensitivity reactions to penicillins (Supplementary Table E1). Eleven of 12 (92%) patients tolerated an oral 1st or 2nd generation cephalosporin provocation after IDT (Table 1). One remaining patient has yet to undergo oral provocation. Five of 12 (41.6%) patients were positive to the primary implicated drug on IFN- γ ELISpot testing (Supplementary Figure E1).

We provide evidence for apparent cross-reactivity within penicillin class drugs by demonstrating penicillin IDT cross-reactivity in patients reporting a penicillin-associated severe T-cell mediated hypersensitivity. The vast majority of these patients subsequently tolerated oral cephalosporins. Prior studies have previously demonstrated cross-reactivity between R1-side chains of aminopenicillins (amoxicillin, ampicillin, amoxicillin-clavulanate) and aminocephalosporins (cefalexin, cefaclor, cefadroxil, cefprozil, cefatrizine, cefonicid, cefmandole) in delayed (T-cell-mediated) hypersensitivity, and the absence of cross-reactivity with non-cross reactive cephalosporins, carbapenems and monobactams.^{7, 8} Romano *et al.* in a cohort of 214 intradermal test positive patients reporting a penicillin T-cell-mediated hypersensitivity (8 with SCAR), 89 (42%) of patients were positive to benzylpenicillin, ampicillin and amoxicillin and only 8 (8.9%) also positive to the MDM or PPL, supporting the cross-reactivity pattern seen in our cohort of all severe T-cell mediated hypersensitivity⁷. Overall, cross-reactivity patterns in severe T-cell-mediated hypersensitivities have not been well-defined due to caution in performing IDT and patch testing in this population, the incomplete sensitivity and lack of widespread availability and validation of *ex vivo* and *in vitro* methods, and the inability to use oral ingestion challenge as the gold standard. Watts *et al.* previously described a single case of Penicillin G DRESS with IDT and PT positivity to both Penicillin G and amoxicillin,⁹ and similar to our cohort, the patient tolerated an oral cephalosporin. This finding may be under reported in the literature due to the prior general avoidance of performing IDT in patients reporting a severe T-cell mediated hypersensitivity. The pattern of cross-reactivity demonstrated in our study

and tolerance of similar R1 side-chain containing oral cephalosporins points towards the “penicillin ring” (thiazolidine) being an important component in generation of the primary antigen. Further, it highlights the importance of testing alternative penicillins in addition to PPL and MDM which were not useful reagents in documenting cross-reactivity between penicillins in patients with DRESS, AGEP and severe-MPE. We believe these are important lessons for skin testing in severe delayed and presumed T-cell mediated hypersensitivity and predicting beta-lactam tolerability in these patients.

Although a limitation to our findings includes a lack of confirmation of identified penicillin cross-reactivity with ingestion challenge, this would not be considered an ethical approach given the severity of the reported reactions and the presence of alternative therapeutic agents in these patients. Although, false positive reactions are possible, the absence of similar results in controls (Supplementary materials), reproducibility of the positive phenotypes on IDT and confirmatory patch testing with varied drug formulation (patient 1), are all supportive of true T-cell mediated responses. The dose-dependency of responses in the skin is in keeping with T-cell mediated responses where non-covalent binding of the drug or drug-altered peptide occurs in a concentration dependent fashion with an immune receptor. This could explain the positive responses we have seen with benzyl penicillin used at 1000 IU/mL and 10,000 IU/mL whereas MDM with a 0.5 mg/ml benzyl penicillin was consistently negative. The apparent lack of sensitivity of IFN- γ release ELISpot positivity to all penicillins is also noted, however this may reflect the variable sensitivity of the assay or that IFN- γ is not the relevant cytokine output for the phenotypes of delayed reactions tested. Further, the absence of IFN- γ positivity in all skin test positive patients may reflect the known absence of circulating drug-reactive effector memory T-cells despite durable long-lasting tissue-resident memory T cells responses being evident *in vivo*.¹⁰ This work confirms a previously infrequently described pattern of cross-reactivity between penicillins in severe T-cell mediated penicillin hypersensitivity and provides support for cephalosporin tolerability in these populations. Future work needs to be directed at understanding the antigenic structures and genomic predictors in these severe penicillin-associated T-cell mediated hypersensitivities.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Clinical implications

This study demonstrates the utility of intradermal testing to elucidate penicillin class cross-reactivity in patients with a history of severe presumed T-cell mediated hypersensitivity. The study suggests patients should avoid not just the inciting penicillin, yet demonstrates tolerance of oral cephalosporins and utilization of alternative narrow spectrum beta-lactams

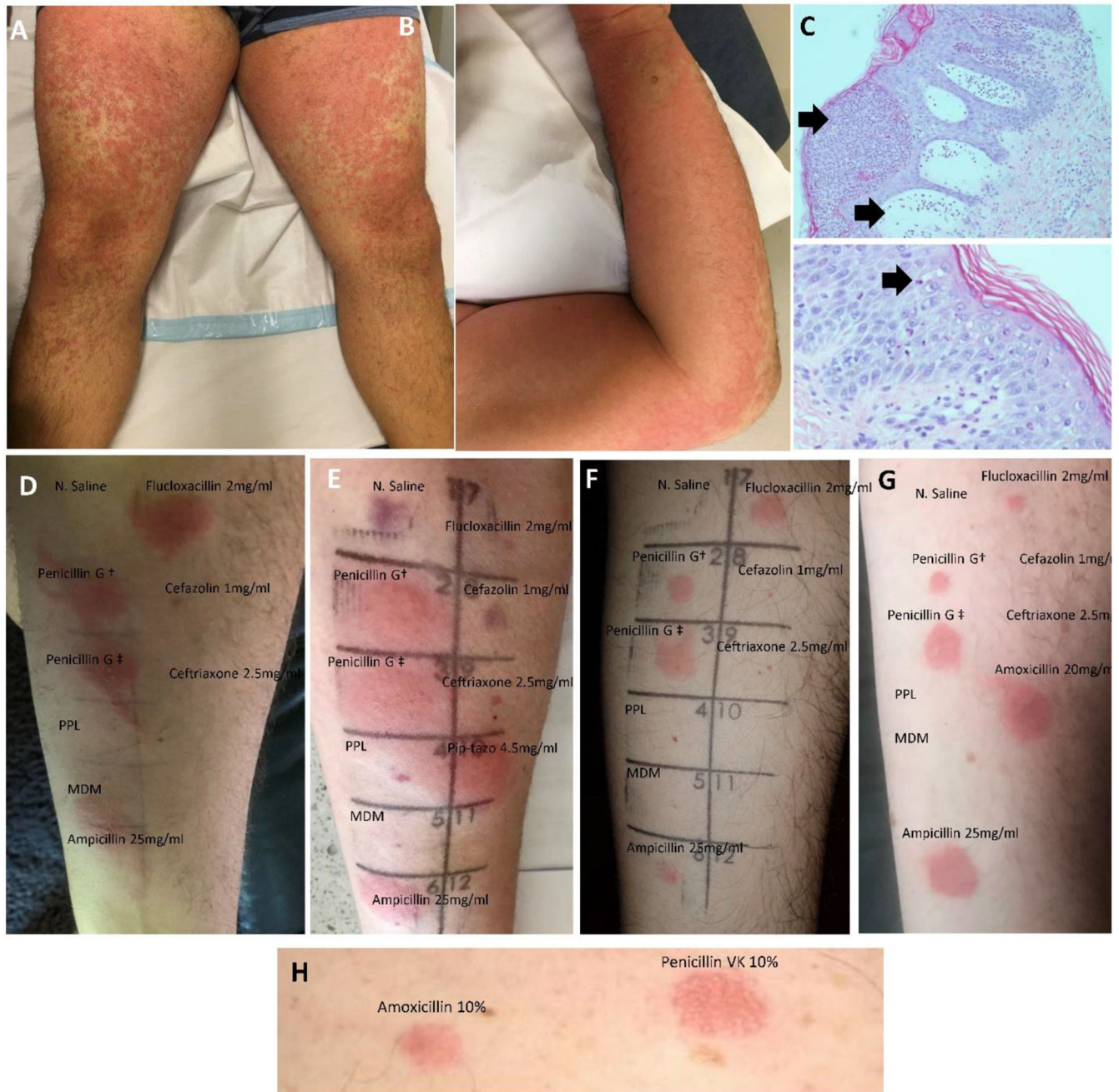


Figure 1. Pictorial representation of patients reporting penicillin-associated severe T-cell mediated hypersensitivity from tested cohort. Pustular exanthem of a patient with flucloxacillin-associated AGEP [Patient 11] (A-B) with corresponding histopathology demonstrating pustule formation (upper arrow) and upper dermal edema (lower arrow) (low power x10 magnification; upper image) and spongiosis and neutrophil migrating through epidermis [arrow] (high power x100 magnification; lower image) (C). Intradermal testing 24 hours post inoculation showing widespread penicillin cross-reactivity (pustule formation noted on penicillin 1000IU/ml IDT) (D). Further patients with identical pattern of intradermal test

cross-reactivity demonstrated in (**E** [Patient 4], **F** [Patient 9], **G** [Patient 1]). Please note that a bruise is noted in Patient **E** where the Normal Saline was inoculated. **Panel H** illustrates a Grade 3 positive patch test result from same patient with IDT demonstrated in **Panel G** (PT performed 6 months following positive IDT) [Patient 1]. Patient 1 IDT was performed with amoxicillin in addition to the standard panel (**Panel G**) to correlated with patch testing performed to amoxicillin (**Panel H**).

Abbreviations; PPL, DAP-major (benzylpenicilloyl poly-L-lysine; final concentration 1.07×10^{-2} mol/L), **MDM**, minor-determinate (sodium benzylpenicillin, benzylpenicilloic acid, sodium benzylpenicilloate), **Penicillin G[†]**, Penicillin G 1000 IU/mL; **PenicillinG[‡]**, Penicillin G 10000 IU/mL.

Table 1: Patients with penicillin-associated severe T-cell-mediated hypersensitivity with positive intradermal testing

No.	Sex/Age	Pre-existing skin disease or medical comorbidities	Phenotype	RegiSCAR score ¹	Biopsy Compatible*	Primary implicated drug	Time from reaction to testing (days) [†]	Positive IDT**	Time to positivity	Positive IFN- γ ELISpot	Oral provocation
1	43M	Nil	MPE	2	Not performed	Amoxicillin	6145	AMP, FLU, PEN, AMX ^{‡,^}	24 hours	No	Cephalexin
2	51F	Nil	DRESS	4	Yes	Piperacillin-tazobactam	473	AMP, FLU, PEN, PIP-TAZ	24 hours	No	Cefuroxime
3	38F	Diabetes mellitus	DRESS	7	Yes	Piperacillin-tazobactam	312	AMP, FLU, PEN, PIP-TAZ	24 hours	PIP-TAZ	Cefuroxime
4	42F	Hairy cell leukemia	MPE	2	No	Piperacillin-tazobactam	269	AMP, FLU, PEN, PIP-TAZ	24 hours	No [#]	Cephalexin
5	45F	Chronic myelocytic leukemia	MPE	2	No	Piperacillin-tazobactam	318	AMP, FLU, PEN, PIP-TAZ	24 hours	No [#]	Cefuroxime
6	64M	Liver transplant recipient for alcoholic liver disease	MPE	2	Not performed	Amoxicillin	1470	AMP, FLU, PEN, PIP-TAZ	24 hours	AMP	Cephalexin
7	38F	Metastatic melanoma ^{^,^}	MPE	3	Yes	Amoxicillin	121	AMP, FLU, PEN, PIP-TAZ	24 hours	AMP	Cefuroxime
8	34M	Hairy cell leukemia	MPE	2	Not performed	Piperacillin-tazobactam	1146	AMP, FLU, PEN, PIP-TAZ	24 hours	No	Cefuroxime
9	25M	Nil	AGEP	-	Yes	Flucloxacillin	101	AMP, FLU, PEN [^]	24 hours	No	Cephalexin
10	62M	Follicular lymphoma	DRESS	4	Yes	Piperacillin-tazobactam	1078	AMP, FLU, PEN, PIP-TAZ	24 hours	PIP-TAZ	Cephalexin
11	45M	Nil	AGEP	-	Yes	Flucloxacillin	90	AMP, FLU, PEN [^]	24 hours	No	Cephalexin
12	31F	Nil	AGEP	-	Yes	Amoxicillin	3097	AMP, FLU, PEN [^]	24 hours	AMP	Not yet performed

Abbreviations: F; female; M; male; DRESS, drug reaction with eosinophilia and systemic symptoms; AGEP, acute generalized exanthematous pustulosis; MPE, severe maculopapular exanthem; PT, patch testing; IDT, intradermal testing; AMX, amoxicillin; AMP, ampicillin; PEN, Penicillin G, FLU, flucloxacillin; PIP-TAZ, piperacillin-tazobactam; OP, oral provocation; pip-tazo, piperacillin-tazobactam.

¹ RegiSCAR score as per published definitions (<2, no DRESS; 2-3, possible DRESS; 4-5, probable DRESS; 6 definite DRESS)²

* Haemotology and Eosin (H & E) performed as per routine clinical practice

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** Tested IDT concentrations: Ampicillin 25mg/ml, benzylpenicillin 1000IU/ml, benzylpenicillin 10000IU/ml, flucloxacillin 2mg/ml, piperacillin-tazobactam 4.5mg/ml (amoxicillin 20mg/ml utilized only for Patient 1)

[‡] Latency period – time (days) from rash onset to intradermal testing. If year only known, date default 1st of January of implicated year.

[‡] Reproduced with patch testing to ampicillin 10%, benzylpenicillin 10%. Performed 6386 days post index reaction – 6 months following positive IDT test.

[^] Piperacillin-tazobactam not tested

[^] melanoma patient on check-point inhibitor

[#] Poor CD3 response reflecting recent cladribine (660) and cytarabine (859) chemotherapy