

A Lysosome-Targetable Fluorescence Probe Based on L-Cysteine-Polyamine-Morpholine-Modified Quantum Dots for Imaging in Living Cells

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Purpose: Quantum dots (QDs) are used as fluorescent probes due to their high fluorescence intensity, longevity of fluorescence, strong light-resistant bleaching ability and high light stability. Therefore, we explore a more precise probe that can target an organelle.

Methods: In the current study, a new class of fluorescence probes were developed using QDs capped with 4 different L-cysteine-polyamine-morpholine linked by mercapto groups. Ligands were characterised by Electrospray ionization mass spectrometry (ESI-MS), H-Nuclear Magnetic Resonance (^1H NMR) spectroscopy, and ^{13}C NMR spectroscopy. Modified QDs were characterized by Transmission Electron Microscope (TEM), Ultraviolet and visible spectrophotometry (UV-Vis), and fluorescence microscopy. And the biological activity of modified QDs was explored by using MTT assay with HeLa, SMMC-7721 and HepG2 cells. The fluorescence imaging of modified QDs was obtained by confocal laser scanning fluorescence microscopy (CLSM).

Results: Synthesized QDs ranged between 4 to 5 nm and had strong optical emission properties. UV-Vis and fluorescence spectra demonstrated that the cysteine-polyamine-morpholine were successfully incorporated into QD nanoparticles. The MTT results demonstrated that modified QDs had lesser cytotoxicity when compared to unmodified QDs. In addition, modified QDs had strong fluorescence intensity in HeLa cells and targeted lysosomes of HeLa cells.

Conclusion: This study demonstrates the modified QDs efficiently entered cells and could be used as a potential lysosome-targeting fluorescent probe.

Keywords: quantum dots, fluorescence probe, lysosome-targetable, cells imaging

Introduction

Quantum dots (QDs) have range of applications as nano-fluorescent probes and antitumor drug carriers, due to their unique optical properties and electronic structure.¹⁻⁶ Although QDs have strong fluorescence intensity, long fluorescence lifetime and high light stability, QDs generally have poor water solubility, high toxicity and poor cell targeting ability.⁷⁻¹³ It is well known that hydrophobic QDs can be engineered to have great water solubility through surface-exchange of hydrophobic surfactant molecules for hydrophilic ligands.¹⁴⁻¹⁸ In addition, the cytotoxicity of QDs can be reduced and targeting of tumor cells can be enhanced through modification of QD surfaces using biomolecules.¹⁹⁻²³ Therefore, when interfaced with biological molecules including proteins, peptides, carbohydrates or DNA, the resulting QD-biocomposites have widespread applicability in areas ranging from

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cellular fluorescence imaging to in vivo fluorescence imaging.^{24–30} However, following surface modification of QDs, it is important to retain its fundamental properties, such as uniform dispersion and to ensure that functional groups remain available for biological applications.^{31–35} As a result, development of targetable fluorescent probe QDs with fundamental properties of bio-molecules.

Polyamines are widely distributed in the body, and are associated with cell proliferation and differentiation process.^{36–40} In recent years, polyamine derivatives have attracted increasing attention due to their ability to enhance the selectivity and bioactivity of compounds.^{41–46} Protein primary structure consists of amino acids linked by peptide bonds. Because amino acids are generally water-soluble and inexpensive, they have been used as capping agents for QDs.^{47–51} Amino acid contain an amino and carboxyl group, therefore they can be directly linked to the surface of QDs and other drugs with the antitumor activity or targetable groups.^{52–57}

In this paper, we synthesized a new class of modified QDs with small size, good water solubility, disperse uniformly and strong optical emission properties. In such models, four types of amino acid-polyamine-morpholine derivatives were used and the bio-activities of modified QDs were greatly improved. Since the polyamine or its derivatives are able to mimic the environment of the second coordination sphere in the active site of the corresponding enzyme. The modified QDs were characterized by transmission electron micrographs (TEM), UV-Visible Spectrophotometer (UV-Vis), and fluorescence spectrophotometer. Biological activity of modified QDs was explored using the MTT assay with HeLa, SMMC-7721 and HepG2 cells. Results demonstrated

that modified QDs were less toxic to these cells. Fluorescence imaging of modified QDs with confocal laser scanning fluorescence microscopy (CLSM) demonstrated strong fluorescence intensity in targeted lysosomes of HeLa cells.

Materials and Methods

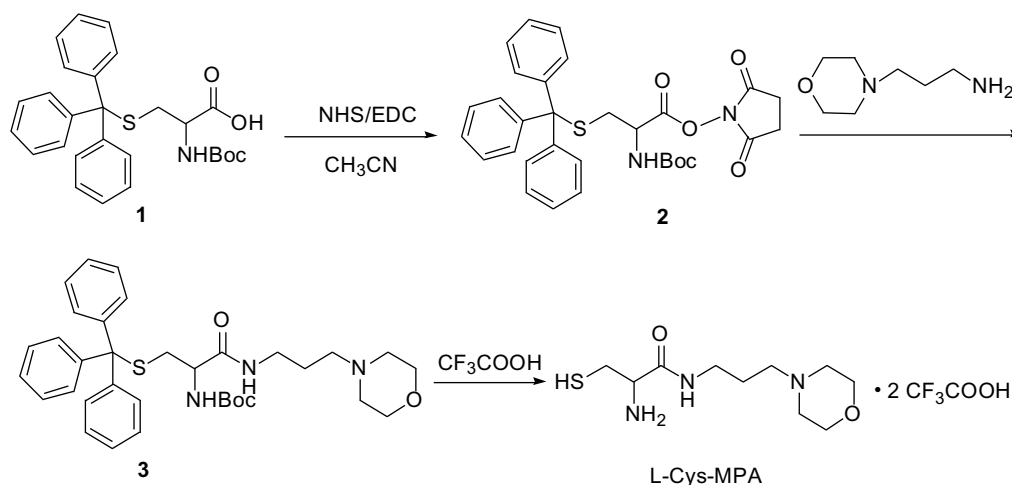
Materials

All reagents were purchased from commercial suppliers and used without further purification. Solvents were purified by standard methods prior to use. Cervical cancer cells (HeLa), human hepatoma cells (SMMC-7721) and human hepatoma cells (HepG2) were obtained from the Shanghai Cell Bank of the Chinese Academy of Sciences.

¹H NMR spectra were recorded on a Bruker AV-400 spectrometers. ESI-MS spectra were performed on a Thermo LCQ-DECA-XP spectrometer. UV-Vis spectra were monitored using a Varian Cary 300 UV-Vis spectrophotometer. Fluorescence spectra were measured using a Varian Cary Eclipse fluorescence spectrophotometer (American, Agilent, Co.). Confocal microscopy images were obtained using a confocal laser scanning fluorescence microscope (Leica SP8, Germany). The cell fluorescence intensity was measured using a FACSCalibur flow cytometer (Becton Dickinson & Co., Franklin Lakes, NJ).

Synthesis of L-Cysteine-2-Amino-3-Mercapto-N-(3-Morpholinopropyl)propanamide (L-Cys-MPA) (Scheme 1)

A 250 mL round bottomed flask equipped with a magnetic stirring bar was charged with Boc-Cys(trt)-OH (2.0 g, 3.5 mmol) (**1**) dissolved in 100 mL acetonitrile and 0.59 g (3.5



Scheme 1 Schematic illustration of the formation of the L-Cys-MPA.

mmol) NHS. Next, EDC (0.89 g, 3.5 mmol) was added dropwise and stirred at 0°C for 2 hrs. Then the reaction mixture was stirred at room temperature overnight. After completion, the precipitate was filtered, extracted with chloroform and vacuum dried to produce a slightly yellow solid.

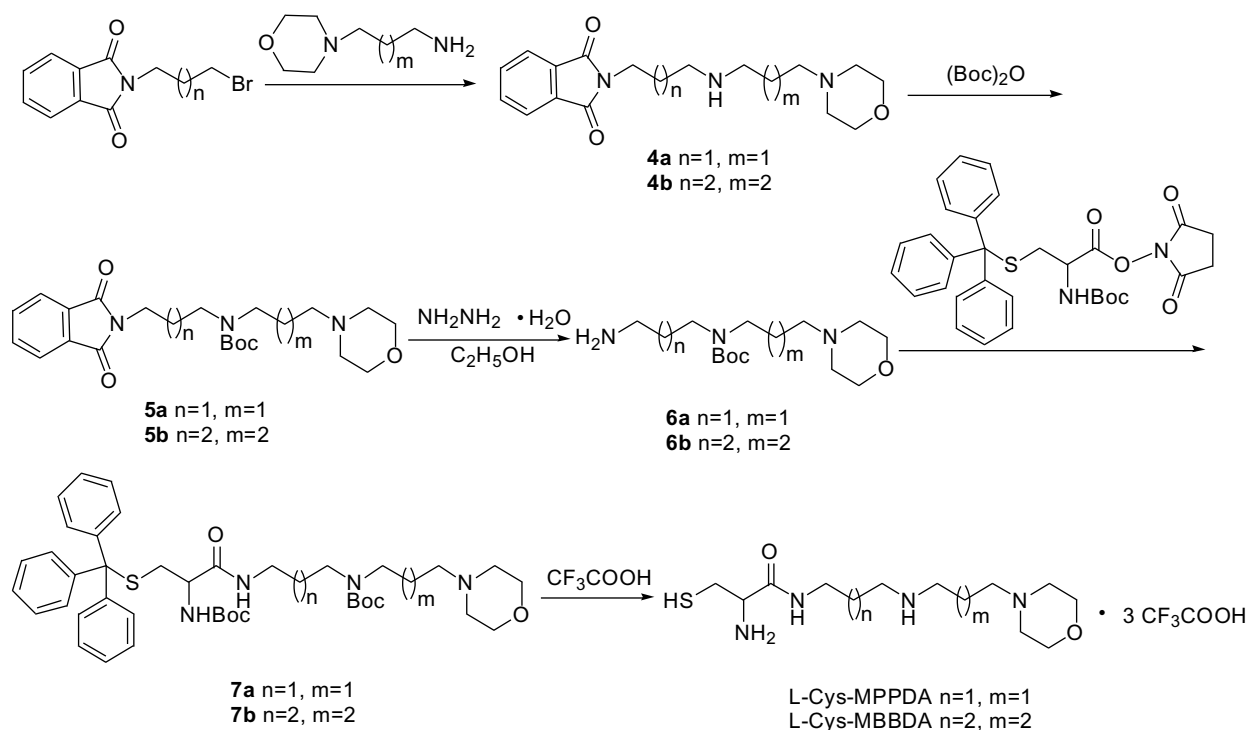
N-aminopropylmorpholine (0.5 g, 3.5 mmol), compound **2**, and NaHCO₃ (0.35 g, 4.2 mmol) were placed in a 100 mL round-bottomed flask with a magnetic stirrer, and stirred at 60°C for 12 hrs. Next, the reaction mixture was evaporated to dryness under reduced pressure and extracted with chloroform. After vacuum drying, the solid was purified by silica gel column chromatography using a CH₂Cl₂-CH₃OH (v:v = 20:1) mixture as the eluent to obtain the yellow grease compound **3**.

Compound **3** was dissolved in 10 mL CH₂Cl₂ in a 50 mL three-necked flask, and then trifluoroacetic acid (TFA) (9.5 mL) and triethyl silane 0.5 mL were injected sequentially under nitrogen protection. The resulting solution was stirred at 25°C for 6 hrs. After completion, the reaction mixture was evaporated to dryness under reduced pressure, and the residues were extracted with dichloromethane and distilled water. Removal of the solvent from the aqueous phase resulted in a solid compound 2-amino-3-mercapto-N-(3-morpholinopropyl) propanamide (L-Cys-MPA).

L-Cys-MPA, yield: 75%; ¹H NMR (300 MHz, D₂O) δ 3.98 (dd, J=7.4, 4.5 Hz, 2H), 3.92 (d, J=1.5 Hz, 1H), 3.71–3.55 (m, 2H), 3.41–3.30 (m, 2H), 3.20 (q, J=7.2 Hz, 2H), 3.04 (dt, J=12.3, 4.5 Hz, 4H), 2.88 (dd, J=5.9, 2.1 Hz, 2H), 1.90–1.76 (m, 2H); ¹³C NMR (75 MHz, D₂O) δ 168.03, 163.35, 162.87, 162.40, 161.93, 121.97, 118.11, 114.25, 110.39, 63.66, 54.67, 54.42, 51.67, 51.65, 36.38, 24.62, 22.98; ESI-MS, m/z: 248.09 [M+H]⁺.

Synthesis of L-Cysteine-N¹-(3-Morpholinopropyl) Propane-1,3-Diamine (L-Cys-MPPDA), L-Cysteine-N¹-(4-Morpholinobutyl) Butane-1,4-Diamine (L-Cys-MBBDA) (Scheme 2)

A 100 mL round-bottom flask equipped with a magnetic stirring bar was charged with N-aminopropylmorpholine (1.4 g, 10 mmol) dissolved in 50 mL acetonitrile and K₂CO₃ (2.1 g, 15 mmol). Then N-(3-Bromopropyl) phthalimide (2.7 g, 10 mmol) or N-(4-Bromobutyl) phthalimide (2.8 g, 10 mmol) was added and stirred at 25°C for 12 hrs. After completion, the reaction mixture was evaporated to dryness under reduced pressure, and the residues were extracted with dichloromethane and 10% Na₂CO₃. Removal of the solvent in the organic phase resulted in



Scheme 2 Schematic illustration of the formation of the L-Cys-MPPDA and L-Cys-MBBDA.

a yellow compound **4a** or **4b**. Then compound **4a** or **4b** was dissolved in 50 mL methyl alcohol and (Boc)₂O (3.3 g, 15 mmol) was added and the solution was stirred at 25°C for 12 hrs. After completion, the reaction mixture was evaporated to dryness under reduced pressure. After vacuum drying, the resulting solid was purified on a silica gel column using a CH₂Cl₂-CH₃OH (v:v = 20:1) mixture as the eluent to obtain the grease compound **5a** or **5b**. Compound **5a** or **5b** was then dissolved in 50 mL absolute ethyl alcohol, and hydrazine hydrate (NH₂NH₂) (5 mL) was added and stirred for at 25°C for 12 hrs. The resulting mixture was evaporated to dryness under reduced pressure, and the residues extracted with CH₂Cl₂ and 10% Na₂CO₃. Following removal of the solvent, the solid compound **6a** or **6b** was produced. Synthesis of **7a** or **7b**, L-Cys-MPPDA and L-Cys-MBBDA was completed using a similar method but with compound **3** and L-Cys-MPA.

L-Cys-MPPDA, yield: 76%; ¹H NMR (300 MHz, D₂O) δ 4.10 (d, J=3.0 Hz, 2H), 4.05 (d, J=3.0 Hz, 1H), 3.78 (t, J=13.5Hz, 2H), 3.50 (d, J=12.0 Hz, 2H), 3.32 (dd, J=12.1, 5.2 Hz, 1H), 3.27–3.18 (m, 3H), 3.18–3.00 (m, 6H), 2.20–2.05 (m, 2H), 1.89 (p, J=7.0 Hz, 2H). ¹³C NMR (75 MHz, D₂O) δ 168.12, 63.64, 54.41, 53.60, 51.73, 45.31, 44.26, 36.41, 25.33, 24.64, 20.32. ESI-MS, m/z: 305.20 [M+H]⁺.

L-Cys-MBBDA, yield: 79%; ¹H NMR (300 MHz, D₂O) δ 4.08 (d, J=6.0 Hz, 2H), 4.04 (t, J=4.5 Hz, 1H), 3.74 (t, J=12 Hz, 2H), 3.45 (d, J=6.0Hz, 2H), 3.33–3.09 (m, 4H), 3.09–2.94 (m, 6H), 1.83–1.47 (m, 8H). ¹³C NMR (75 MHz, D₂O) δ 167.73, 163.49, 163.02, 162.55, 162.08, 122.07, 118.21, 114.35, 110.49, 63.70, 56.22, 54.48, 51.61, 47.09, 46.61, 38.84, 25.37, 24.76, 22.97, 22.59, 20.32. ESI-MS, m/z: 333.29 [M+H]⁺.

Synthesis of L-Cysteine-N¹-(3-Aminopropyl)-N³-(3-Morpholinopropyl)propane-1,3-Diamine (L-Cys-AMPDA) (Scheme 3)

A 100 mL round-bottomed flask equipped with a magnetic stirring bar was charged with **6a** (3.0 g, 10 mmol) dissolved in 50 mL acetonitrile solution and K₂CO₃ (2.1 g, 15 mmol). Then, N-(3-Bromopropyl) phthalimide (2.7 g, 10 mmol) was added and stirred at 25°C for 12 hrs. After completion, the reaction mixture was evaporated to dryness under reduced pressure, and the residues extracted with CH₂Cl₂ and 10% Na₂CO₃. Removal of the solvent in the organic phase resulted in a yellow greasy compound, termed compound

8. Then the compound **8** was dissolved in 50 mL methyl alcohol solution. (Boc)₂O (3.3 g, 15 mmol) was added and the mixture was stirred at 25°C for 12 hrs. The resulting reaction mixture was concentrated under vacuum and further purified by silica gel column chromatography to obtain a greasy compound, termed compound **9**. Next, the compound **9** was dissolved in 50 mL ethyl alcohol and the hydrazine hydrate (NH₂NH₂) (5 mL) was added. The solution stirred at 25°C for 12 hrs. After completion, the reaction mixture was evaporated to dryness under reduced pressure, and the residues were extracted with CH₂Cl₂ and 10% Na₂CO₃. The organic phases were then dried with Na₂SO₄ under vacuum to yield compound **10**. Synthesis of compound **11** and L-Cys-AMPDA can refer to the synthetic method of compound **3** and L-Cys-MPA.

L-Cys-AMPDA, yield: 70%; ¹H NMR (300 MHz, Deuterium Oxide) δ 4.18 (d, J=6.0 Hz, 2H), 4.12 (s, 1H), 3.83 (t, J=12.0 Hz, 2H), 3.56 (d, J=12.0 Hz, 2H), 3.45–3.22 (m, 6H), 3.22–3.04 (m, 10H), 2.25–2.04 (m, 4H), 1.96 (q, J=7.3 Hz, 2H). ¹³C NMR (75 MHz, D₂O) δ 163.10, 162.63, 118.18, 114.32, 63.68, 54.45, 53.60, 51.77, 45.32, 44.57, 44.39, 36.46, 29.74, 25.36, 24.67, 22.60, 20.32. ESI-MS, m/z: 364.32 [M+H]⁺.

Synthesis of Water-Soluble Amino Acid-Polyamine-Morpholine-Modified QDs

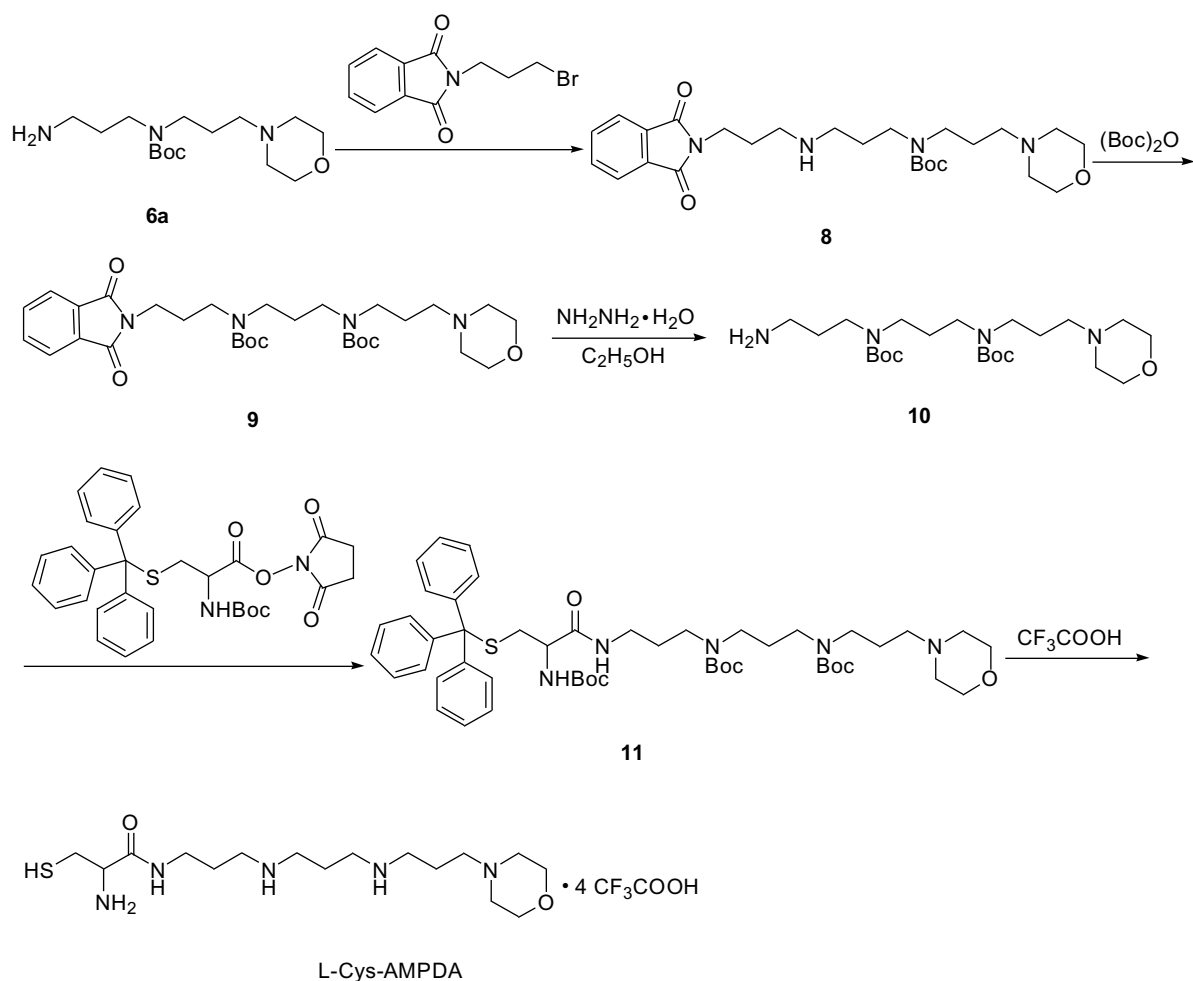
QDs (1 mL) (CdSe or CdSe/ZnS or CdSe/CdS) in 1 mL of chloroform, amino acid-polyamine-morpholine (300 mg) in 3 mL triple distilled water and 10 drops triethanolamine were placed in a bottle, and sonicated for 4 hrs. After completion, ethyl acetate was added to precipitate nanocrystal complexes to purify nanocrystals. The solid was filtered and dried under vacuum to obtain the water-soluble amino acid-polyamine-morpholine derivatives modified QDs.

Cell Culture

Three types of cells (HeLa, SMMC-7721, HepG2) were cultured in RPMI-1640 medium at 37°C under 5% CO₂. These cells were allowed to grow in a monolayer in a tissue culture flask.

Cell Cytotoxicity Test

Antitumor activity of modified QDs was monitored using a modified MTT method. Briefly, cells were seeded into 96-well plates at 5×10⁴ cells per well. After 12 hrs, the modified QDs (50, 100, 200, 500 and 1000 μg/mL) were



Scheme 3 Schematic illustration of the formation of the L-Cys-AMPDA.

subsequently added and incubated for 48 hrs. Next, 20 μL MTT (2.5 mg/mL) was added to each well and after 4 hrs, supernatant was removed and 100 μL DMSO was added to each well for 10–15 mins. The absorbance was measured at a wavelength of 570 nm and the inhibition of cell growth was calculated for each concentration of the samples.

Cellular Uptake of Modified QDs

HeLa cells were seeded in a 35 mm diameter utensil containing 15 mm diameter glass cover-slips, in culture medium containing 10% FBS incubated for 24 hrs at 37°C and 5% CO_2 . The modified QDs were then added at the indicated time points (1, 2, 6, 12, and 24 hrs). Then, cells were washed twice with PBS buffer, and incubated with Hoechst 33342 (10 $\mu\text{g}/\text{mL}$) for 30 mins at 37°C. The cells were washed with PBS, and viewed using a confocal microscope (Leica-SP8) at an excitation wavelength of

488 nm and emission wavelength between 580 and 650 nm. Cell nuclei stained with Hoechst 33342 were observed at a wavelength of 405 nm, and emission wavelength between 430 and 480 nm.

Targeted Localization of Modified QDs in Cells

HeLa cells were seeded in a 35 mm diameter utensil containing 15 mm diameter glass cover-slips, in culture medium containing 10% FBS and incubated for 24 hrs at 37°C and 5% CO_2 . Then the modified QDs (200 $\mu\text{g}/\text{mL}$) were added to the utensil and incubated for 6 hrs. The cells were thoroughly washed with PBS buffer to remove QDs which were not absorbed. Then the cells were incubated for 30 mins at 37°C with the green fluorescent probe LysoTracker Green DND-26 (1 μM), and then rinsed twice with PBS. The images of the cells were obtained using a fluorescence microscope. The excitation

wavelength of the QDs was 488 nm, and the emission wavelength was 580–650 nm. The excitation wavelength of LysoTracker green DND-26 was 488 nm. The band path for the marker imaging was between 500 and 560 nm.

Results and Discussion

Synthesis and Characterization

In this work, three kinds of QDs (CdSe, CdSe/ZnS, and CdSe/CdS) were prepared using an organic phase synthesis method. And four kinds of amino acid-polyamine-morpholine derivatives, L-Cysteine-2-amino-3-mercapto-N-(3-morpholinopropyl) propanamide (L-Cys-MPA), L-Cysteine-N¹-(3-morpholinopropyl) propane-1,3-diamine (L-Cys-MPPDA), L-Cysteine-N¹-(4-morpholinobutyl)

butane-1,4-diamine (L-Cys-MBBDA), and L-Cysteine-N¹-(3-aminopropyl)-N³-(3-morpholinopropyl) propane-1,3-diamine (L-Cys-AMPDA), were connected to the surface of QDs. Synthesized ligands were characterised by ESI-MS, ¹H NMR spectroscopy, and ¹³C NMR spectroscopy. In addition, we characterized the size and dispersion of original QDs and cysteine-polyamine-morpholine-modified QDs using TEM. As shown in Figure 1, the cysteine-polyamine-morpholine-modified QDs maintained uniform dispersion and a spherical shape when suspended in PBS. Namely, the modified QDs had good water solubility and dispersed evenly.

QDs had improved better optical properties when measured using UV/Vis spectra and fluorescence spectroscopy. Figure 2 presents absorption spectra of original QDs and the cysteine-polyamine-morpholine modified QDs, where

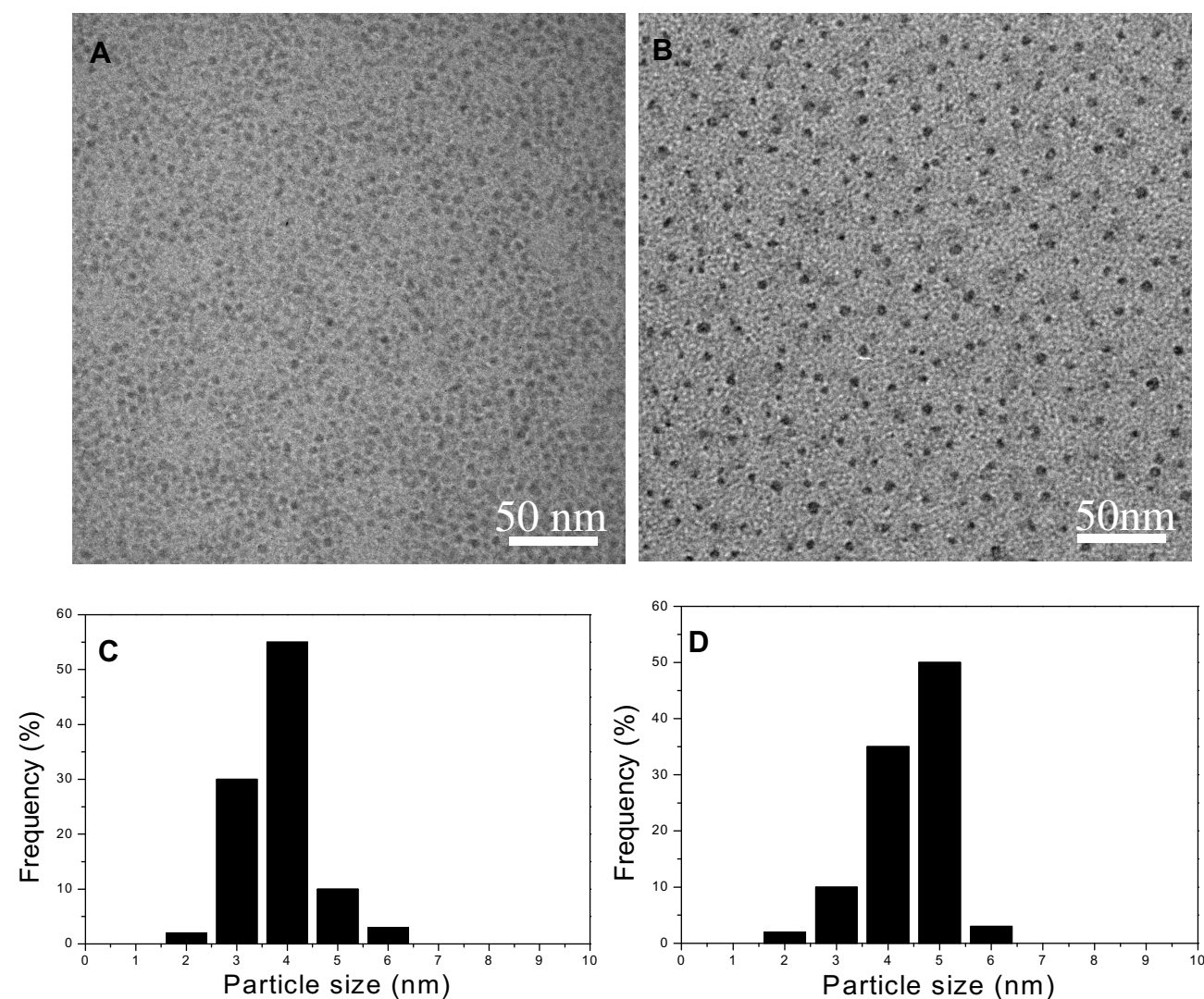


Figure 1 TEM images and size distribution of CdSe/ZnS QDs (**A, C**) and CdSe/ZnS@L-Cys-MPPDA QDs (**B, D**) (scale bars = 50 nm).

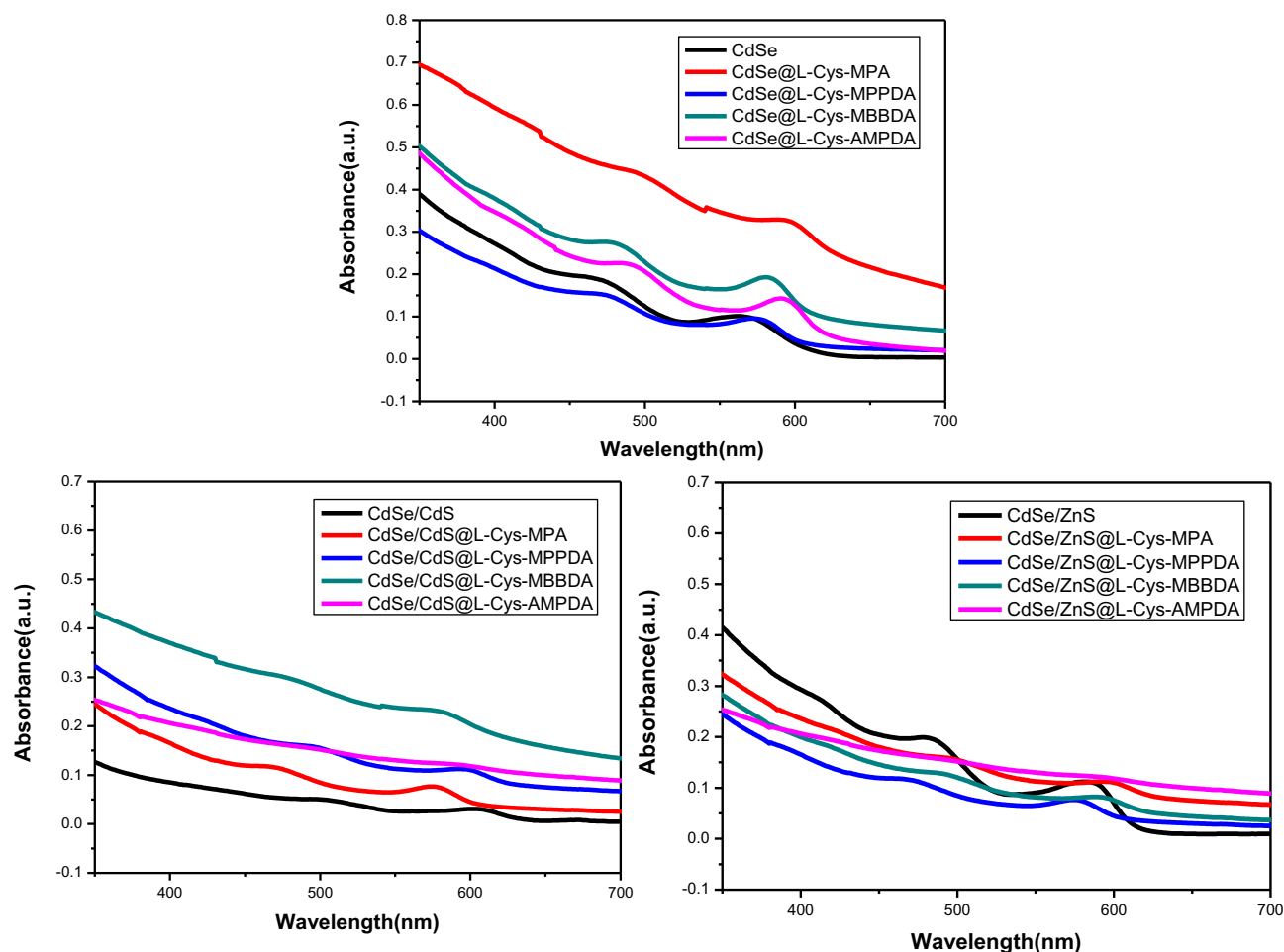


Figure 2 UV-Vis spectra of original QDs and cysteine-polyamine-morpholine-modified QDs.

it is clear the UV-Vis absorption spectra of QDs had absorption properties typical of CdSe, CdSe/CdS, and CdSe/ZnS QDs. The shell would not affect peak position of the absorption spectra of QDs due to the high energy band gap of ZnS in comparison to that of either CdS or CdSe.⁵⁸ Absorbance peak of modified QDs has a slight red-shift relative to that of the original QD nanoparticles. Therefore, it is likely that the cysteine-polyamine-morpholine has been successfully incorporated into QD nanoparticles.

The original QDs and cysteine-polyamine-morpholine-modified QDs were further characterized by fluorescence spectroscopy. As shown in Figure 3, the fluorescence peak of cysteine-polyamine-morpholine modified QDs exhibited a slight red-shifted emission spectra when compared with that of corresponding QDs, suggesting that the elicited wavelengths arose due to recombination of cysteine-polyamine-morpholine derivatives on the

surface of QDs. Bands of fluorescence of modified QDs were broad and significantly more intense when compared to that of original QDs. Differences in bands of fluorescence among QDs were likely due to recombination of cysteine-polyamine-morpholine derivatives on their surface.

Cell Cytotoxicity Assay

In order to explore the cell toxicity of original QDs, amino acid-polyamine-morpholine-modified QDs, HeLa, SMMC-7721 and HepG2 cells were treated with different concentrations of samples for 48 hrs. Cytotoxicity was measured using the MTT assay. The IC_{50} values of original QDs and cysteine-polyamine-morpholine-modified QDs in HeLa, SMMC-7721 and HepG2 cells are presented in Table 1. Calculated IC_{50} values demonstrate that original QDs have greater cytotoxicity when compared with cysteine-polyamine-morpholine-modified

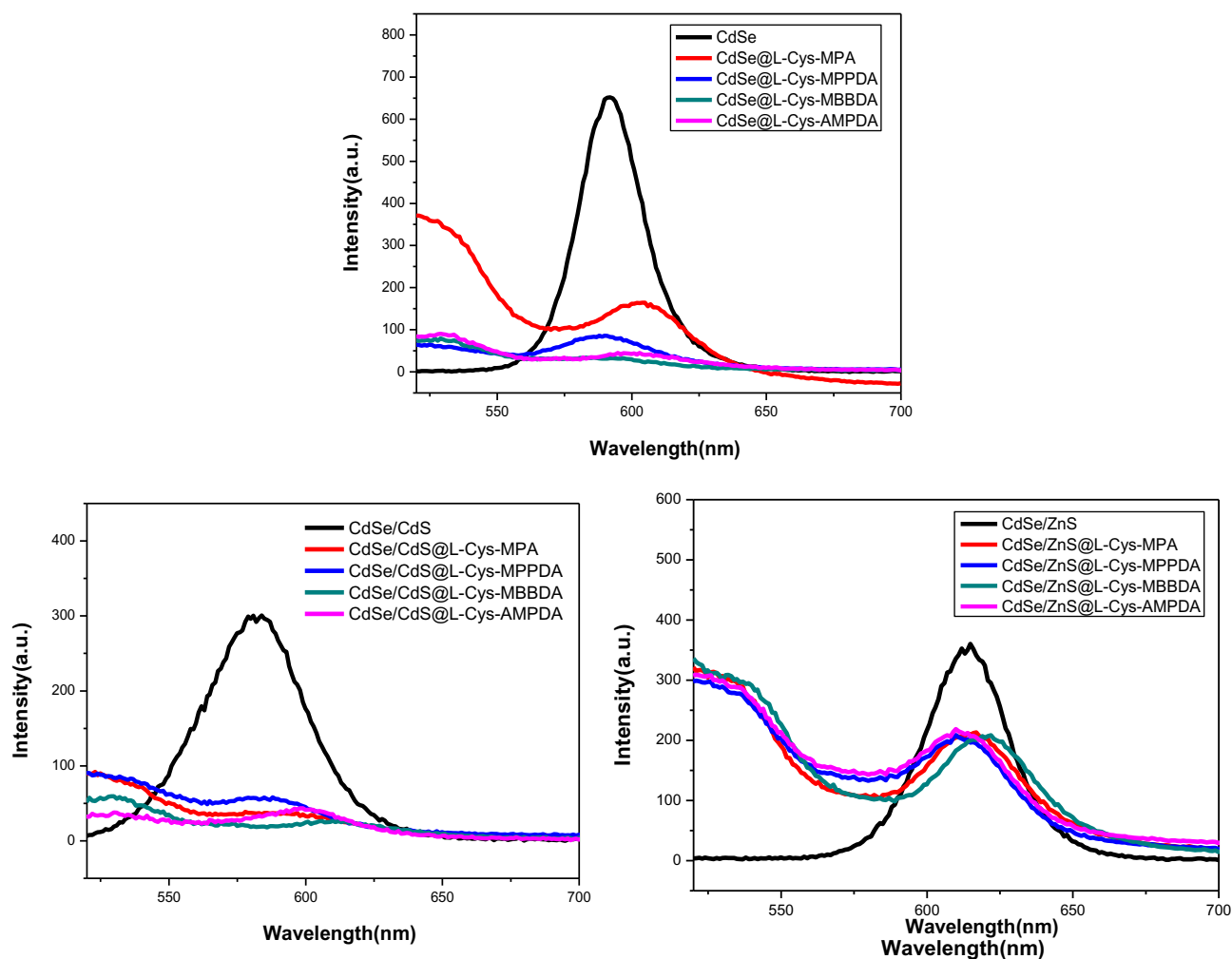


Figure 3 Fluorescence spectra of original QDs and cysteine-polyamine-morpholine-modified QDs.

QDs. Because CdSe QDs may slowly release the toxic Cd^{II} ions into solution, it is expected that particles would be inert for a short period of time during in vitro application. The release of Cd^{II} from the particle surface can be reduced using core/shell particles or coating of the particles with biomolecules or a polymer. Results of this study indicate that the developed cysteine-polyamine-morpholine-modified QDs reduce release of toxic Cd^{II} ions from QDs.

Fluorescent Labeling of Cells

The unique optical properties of QDs make them useful as biological fluorescent probes.^{59,60} Laser scanning confocal microscopy was used to study fluorescence intensity of cysteine-polyamine-morpholine-modified QDs at different concentrations across time. Since CdSe/ZnS@L-Cys-MPPDA QDs showed good optical properties and minimal

toxicity in this work, they were used as a representative QDs in subsequent experiments. Figure 4 presents changes in fluorescence intensity of CdSe/ZnS@L-Cys-MPPDA QDs at different concentrations in cells. As concentration increased, more CdSe/ZnS@L-Cys-MPPDA QDs entered the cell and a concomitant increase in fluorescence intensity was observed. Furthermore, it was observed that QDs selectively accumulated in the cytoplasm of cells. Accumulation of CdSe/ZnS@L-Cys-MPPDA QDs (200 µg/mL) in HeLa cells over differing incubation periods was monitored (Figure 5). As presented in Figure 5, fluorescence intensity of QDs in cells gradually increased with incubation time and reached a plateau in the cytoplasm after 6 hrs. Remarkably, the fluorescence intensity of QDs in cells did not decrease following 24 hrs incubation. Thus, the fluorescence intensity of CdSe/ZnS@L-Cys-MPPDA QDs treated cells increased with increasing

Table 1 IC₅₀ Values of Modified and Unmodified QDs to HeLa, SMMC-7721 and HepG2 Cells

Compounds	IC ₅₀ (µg/mL)		
	HeLa	SMMC-7721	HepG2
CdSe@L-Cys-MPA	798.2 ± 1.9	767.9 ± 2.7	703.1 ± 3.2
CdSe/ZnS@L-Cys-MPA	816.5 ± 2.5	775.6 ± 2.9	823.8 ± 2.0
CdSe/CdS@L-Cys-MPA	820.3 ± 4.3	817.0 ± 3.5	757.4 ± 2.9
CdSe@L-Cys-MPPDA	779.0 ± 1.8	796.1 ± 2.4	775.0 ± 2.3
CdSe/ZnS@L-Cys-MPPDA	830.7 ± 3.2	858.2 ± 2.8	869.1 ± 1.8
CdSe/CdS@L-Cys-MPPDA	798.4 ± 3.7	803.6 ± 1.2	827.8 ± 2.1
CdSe@L-Cys-MBBDA	512.9 ± 2.5	686.9 ± 0.9	700.6 ± 1.4
CdSe/ZnS@L-Cys-MBBDA	771.7 ± 4.0	798.0 ± 3.1	815.9 ± 3.3
CdSe/CdS@L-Cys-MBBDA	657.3 ± 1.6	754.6 ± 2.6	768.0 ± 2.0
CdSe@L-Cys-AMPDA	767.5 ± 3.0	779.0 ± 2.3	796.2 ± 2.6
CdSe/ZnS@L-Cys-AMPDA	823.4 ± 1.1	812.6 ± 0.8	826.9 ± 2.4
CdSe/CdS@L-Cys-AMPDA	779.8 ± 2.7	795.2 ± 1.8	814.0 ± 1.3
CdSe	15.5 ± 2.5	13.9 ± 3.1	14.5 ± 3.4
CdSe/ZnS	18.4 ± 2.2	18.2 ± 1.9	18.7 ± 2.9
CdSe/CdS	16.2 ± 3.1	15.4 ± 2.7	16.8 ± 2.8

concentrations and incubation times. So the cysteine-polyamine-morpholine-modified QDs can act as potential fluorescent probes for cells.

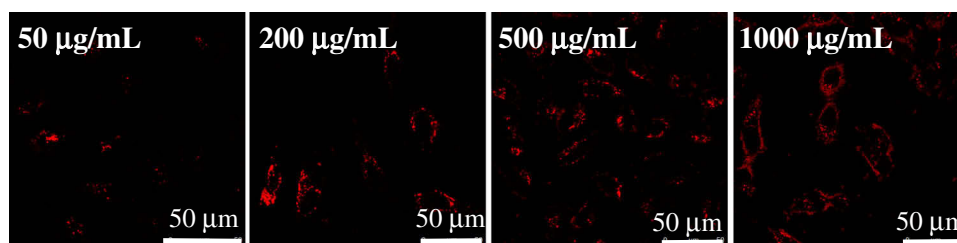
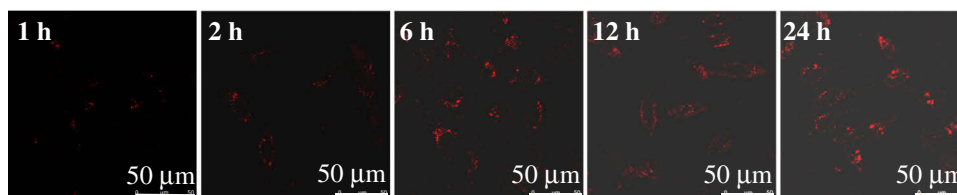
To confirm the cell selectivity of the QDs in vitro, HT-29, HepG2, SMMC-7721 and HeLa cells were incubated with CdSe/ZnS@L-Cys-MPPDA QDs. After incubation for 6 hrs with QDs, the cells were washed. And the images of cells were collected under CLSM. As shown in Figure 6, the fluorescence intensity of CdSe/ZnS@L-Cys-MPPDA QDs in SMMC-7, HepG2, and HT-29 cells was weak and little to no fluorescence was observed. However, the

fluorescence intensity of CdSe/ZnS@L-Cys-MPPDA QDs in HeLa cells was strong. These results provide strong evidence to suggest CdSe/ZnS@L-Cys-MPPDA QDs have strong HeLa cells targeting ability.

In addition, labeling of subcellular compartments by QDs was of particular importance. To further study targeting and localization of modified QDs in cells, we stained HeLa cells with the lysosome green fluorescent probe LysoTracker Green DND-26, and observed distribution of QDs in HeLa cells using laser confocal microscopy. As can be seen from Figure 7, the fluorescence of CdSe/ZnS@L-Cys-MPPDA QDs demonstrated complete co-localization with lysosomes in HeLa cells. Evaluation of the staining patterns and co-localization demonstrated that CdSe/ZnS@L-Cys-MPPDA QDs remained intact following uptake and had a perinuclear distribution; therefore, these results suggest that QDs are predominantly confined within lysosomes. This evidence indicates that nearly all cysteine-polyamine-morpholine-modified QDs were transported inside lysosomes without release to the cytoplasm or nucleus. Lysosome targeting of modified QDs may be attributed to the morpholine group of ligand cysteine-polyamine-morpholine.

Conclusion

In conclusion, a new class of fluorescence probes, L-cysteine-polyamine-morpholine-modified QDs was synthesized in this work. These modified QDs had good water solubility and dispersed evenly. And the modified QDs had lower cytotoxicity to cells in vitro when

**Figure 4** Confocal images of HeLa cells treated with CdSe/ZnS@L-Cys-MPPDA for 6 hrs at 37°C with different concentrations.**Figure 5** Confocal images of HeLa cells treated with CdSe/ZnS@L-Cys-MPPDA (200 µg/mL) at 37°C in the medium at different time.

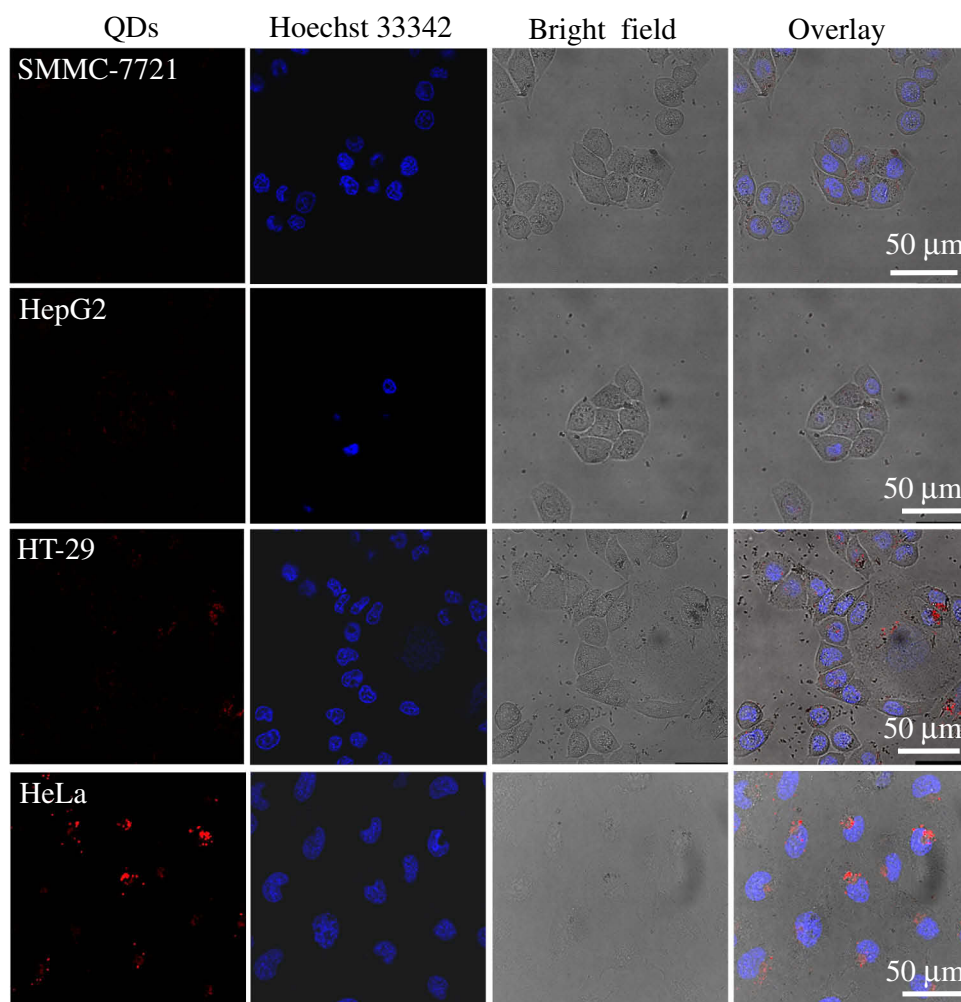


Figure 6 Confocal images of different cells treated with CdSe/ZnS@L-Cys-MPPDA (200 µg/mL) for 6 hrs at 37°C.

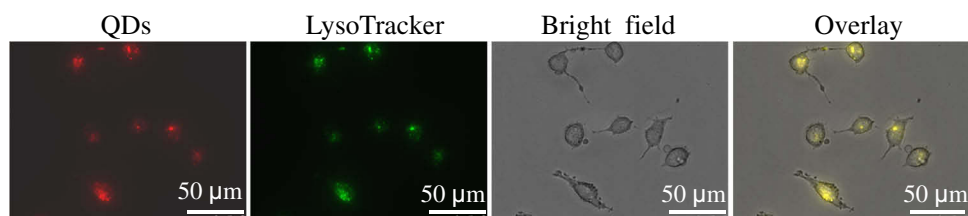


Figure 7 The CdSe/ZnS@L-Cys-MPPDA QDs display co-localization with lysosomes in HeLa cells.

compared to unmodified QDs. In addition, modified QDs had strong fluorescence intensity in HeLa cells and effectively targeted lysosome of HeLa cells. Therefore, modified QDs can efficiently enter cells and could be used as lysosome-targetable fluorescent probes.

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Author Contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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