

# *Clostridium (Clostridioides) difficile* in animals

Journal of Veterinary Diagnostic Investigation  
2020, Vol. 32(2) 213–221  
© 2020 The Author(s)  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/1040638719899081  
jvdi.sagepub.com

J. Scott Weese<sup>1</sup> 

**Abstract.** *Clostridium (Clostridioides) difficile* is a gram-positive, spore-forming bacterium that is an important cause of disease in people, a variably important cause of disease in some animal species, and an apparently harmless commensal in others. Regardless of whether it is a known pathogen in a particular species, it can also be found in healthy individuals, sometimes at high prevalences and typically with higher rates of carriage in young individuals. As it is investigated in more animal species, it is apparent that this bacterium is widely disseminated in a diverse range of domestic and wild animal species. Although it can be found in most species in which investigations have been performed, there are pronounced intra- and inter-species differences in prevalence and clinical relevance. A wide range of strains can be identified, some that appear to be animal associated and others that are found in humans and animals. A large percentage of strains that cause disease in people can at least sporadically be found in animals. It is a potentially important zoonotic pathogen, but there is limited direct evidence of animal–human transmission. Although *C. difficile* has been studied extensively over the past few decades, it remains an enigmatic organism in many ways.

**Key words:** *Clostridioides*; *Clostridium*; diarrhea; epidemiology; gastrointestinal microbiome; microbiology; zoonoses.

## Introduction

*Clostridium (Clostridioides)*<sup>80</sup> *difficile* is a gram-positive, anaerobic, spore-forming bacterium that can be found in humans, a wide range of animal species, and the environment. It is the cause of *C. difficile* infection (CDI), a critically important disease in humans. Its role as an animal pathogen ranges from important to unclear to apparently irrelevant, depending on the animal species.

As a spore-former, the bacterium can be found in 2 states, the fastidious vegetative form that reproduces in the gastrointestinal tract, and as a hardy spore that is passed in feces and can survive for prolonged periods outside the host. The spore is the infectious form, whereas vegetative cells are the metabolically active form that can produce toxins, cause disease, and produce more spores. Virulence is classically attributed to 2 main toxins, toxin A (TcdA) and toxin B (TcdB), with some strains producing a binary toxin (CDT). Most strains produce TcdA and TcdB, or TcdA, TcdB, and CDT, but toxin-variant strains that produce some but not all of these toxins (e.g., TcdA–/TcdB+) can cause disease. Strains that lack genes encoding TcdA, TcdB, and CDT are considered nontoxicogenic and clinically irrelevant.

*C. difficile* is a genetically diverse bacterial species that is estimated to have diverged 1.1–85 million years ago.<sup>53</sup> Most isolates are able to be assigned to 1 of 6 main phylogenetic clades (Table 1), 5 of which (clades 1–5) contain toxigenic strains,<sup>127</sup> although classification has been evolving. Nontoxicogenic strains are intermixed with toxigenic strains in 3 clades (clades 1, 4, and 5). *PCR ribotyping*, a typing method that

involves PCR amplification of the spacer region between the 16S and 23S rRNA genes, and subsequent analysis of gel banding pattern or capillary PCR product,<sup>26,61</sup> is the most widely used typing method. Although > 300 ribotypes are formally classified and many others have been identified, a relatively small number of strains are of greatest concern. Clade 1 contains the most common human pathogens in most regions (ribotypes 001, 012, and 014). Clade 2 contains ribotype 027 (NAP1), a hypervirulent strain associated with widespread disease in humans internationally. Clade 4 contains the toxin variant (TcdA–, TcdB+) ribotype 017 and related strains, which are important causes of disease in humans in some regions. Most of these ribotypes can also be found in animals; however, the main animal concerns are livestock-associated strains in clade 5, particularly ribotype 078. This clade is highly divergent from other clades, and ribotype 078 has emerged as an important cause of community-associated CDI in humans, raising concerns about zoonotic or food origins.<sup>63,73,113</sup> *C. difficile* is an important cause of disease in some domestic animal species, a potential but unclear cause of disease, or apparently clinically relevant in others.

---

Department of Pathobiology and Centre for Public Health and Zoonoses, Ontario Veterinary College, University of Guelph, Guelph, Ontario, Canada (Weese).

<sup>1</sup>Corresponding author: J. Scott Weese, Department of Pathobiology and Centre for Public Health and Zoonoses, Ontario Veterinary College, University of Guelph, Guelph, ON N1G 2W1, Canada.  
jsweese@uoguelph.ca

**Table 1.** *Clostridium (Clostridioides) difficile* clades.

Clade	Common pathogenic PCR ribotypes	Comment
1	001, 012, 014	Most diverse clade; toxigenic and nontoxigenic strains.
2	027, 016, 036, 176	Contains the hypervirulent ribotype 027/NAP1 strain.
3	023	Uncommon clade dominated by one ribotype (023).
4	017	Toxigenic and nontoxigenic strains, includes toxin A-, B+ strains such as 017.
5	078, 126, 033	Toxigenic and nontoxigenic strains; evolutionarily highly divergent from other clades; contains the livestock-associated ribotype 078 (sequence type 11).
6		Nontoxigenic strains; highly diverged; consideration has been given to whether this should represent a separate species.

## Horses

Some of the earliest evidence of clinical relevance of *C. difficile* in domestic animals involved horses, and CDI is an important cause of disease in both adult horses and foals.<sup>25,138</sup>

Initial study of *C. difficile* in horses involved foals, in which the presence of the bacterium was associated with diarrhea.<sup>64</sup>

Subsequent studies demonstrated associations between the presence of TcdA and/or TcdB and diarrhea in adult horses and foals.<sup>42,138</sup> Enterocolitis was later reproduced experimentally through administration of *C. difficile* spores or vegetative cells to foals.<sup>12</sup> CDI is strongly associated with antimicrobial exposure in humans<sup>91</sup> and, although such an association has been suggested in horses,<sup>25</sup> antimicrobial exposure is an inconsistent risk factor and has not always been identified as a risk factor for CDI in horses.<sup>140</sup> Of particular interest have been older reports of severe CDI in mares whose foals were treated with erythromycin and rifampin,<sup>23,48</sup> with subsequent reproduction of CDI in mares given low oral doses of those drugs meant to mimic the concentrations they might be exposed to when their foals were treated.<sup>48</sup> Interestingly, this phenomenon has not been reported elsewhere, suggesting regional differences in susceptibility, perhaps because of differences in the gut microbiota. In contrast to human medicine, in which CDI is an important hospital-associated infection, hospital-associated CDI appears to be uncommon in horses, with only one report implicating it in a cluster of hospital-associated infections.<sup>87</sup> Anecdotally, outbreaks are not uncommon in foals on large breeding farms; however, objective data are lacking.

The clinical spectrum of CDI has not been well described in horses, but it can range from mild, self-limiting diarrhea to rapidly fatal, peracute hemorrhagic enterocolitis. It is most often manifest as diarrhea with various degrees of dehydration, toxemia, and abdominal pain.

The bacterium has also been implicated as a cause of duodenitis/proximal jejunitis.<sup>14</sup> Clinical and histologic changes consistent with that disease were reproduced experimentally by administration of a *C. difficile* toxin suspension.<sup>13</sup>

Although *C. difficile* has been clearly established as an equine pathogen, toxigenic strains can also be found commonly in healthy foals and adult horses. Shedding rates have

been reported of 0–29% in healthy foals,<sup>24,118,138</sup> 44% of antibiotic-treated foals,<sup>24</sup> 4.9–8.4% in hospitalized horses,<sup>65,93</sup> and 0–7.6% of healthy adult horses.<sup>24,93,132,138</sup> Risk factors for *C. difficile* shedding have been explored in adult horses and have focused on young age and antibiotic exposure.<sup>24,47</sup> Reasons why CDI develops in some individuals but not others are unknown, but the intestinal microbiota probably plays an important role.

Various strains have been found in horses. In some studies, the livestock-associated ribotype 078 has been present or predominant.<sup>93,96,117,119</sup> Other clade 5 strains found predominantly in livestock such as TcdA-, TcdB-, CDT+ ribotype 033<sup>65</sup> have also been found as common strains in some populations. More common human strains such as ribotypes 001, 012, 014, 017, and 027 have been identified in some regions.<sup>93,119,132</sup> Overall, most strains found in horses are also found in humans; however, the zoonotic disease risk is unclear.

## Pigs

As in horses, *C. difficile* is clearly a pathogen in pigs. However, in contrast, disease seems to predominantly, if not exclusively, affect piglets 10 d of age or younger.<sup>15,33,134,146,147</sup> Clinical manifestations are variable. Sudden death can occur, with or without evident diarrhea. In less severely affected animals, diarrhea, abdominal distension, decreased appetite, and poor growth may be evident. Pathologically, it is manifest predominantly as typhlocolitis and mesocolonic edema.<sup>147</sup>

Similar to clinical observations, disease has been reproduced experimentally in young piglets but not in older piglets.<sup>15</sup> Antimicrobial exposure is not required for development of CDI.

Although disease in piglets is the animal health concern, most studies of *C. difficile* in pigs has involved surveillance aimed at estimating prevalence and characteristics of *C. difficile* from pigs of various ages, with a focus on potential zoonotic risks. Shedding of *C. difficile* is common in many pig populations, with a clear impact of age. Shedding rates are high in young pigs and typically decline to lower levels by the time of slaughter. Rates in 1- to 14-d-old piglets of 27–100% have been reported,<sup>4,6,17,19,46,52,57,71,75,78,97,98,115,126,128,137</sup> with rates declining rapidly to 0–4% in finisher pigs.<sup>4,16,34,52,55,69,98,108</sup>

However, isolation rates from sows of 4–50% have been reported,<sup>46,57,69,78,97,98,128</sup> perhaps related to the stress of pregnancy and parturition. Reasons for the marked effect of age have not been investigated, but this pattern is consistent with various other species and is likely related to the protective gut microbiota, which becomes established in the first weeks to months of life.<sup>125</sup> Beyond age, no factors have been clearly associated with *C. difficile* shedding, including conventional versus organic production.<sup>67</sup>

A variety of strains can be found; however, a few livestock-associated strains predominate. At the forefront is ribotype 078, a toxinotype V, clade 5 strain that is strongly associated with pigs, and to a lesser degree the closely related ribotypes 126 and 066. These 3 ribotypes have been the sole or predominant strains identified in studies of livestock in North America, Europe, and Asia.<sup>4,6,9,17,56,66,67,69,78,98,115,128,137,141,148</sup> Yet, although ribotype 078 predominates, a wide range of strains can be identified, including other strains found in humans with CDI.<sup>6,8,97</sup> As in some other animal species, the ribotype distribution is different in Australia, where ribotype 078 is rare to nonexistent and various strains can be found, including some unusual strains such as ribotype 237.<sup>95,126</sup>

Further characterization of ribotype 078 isolates by multi-locus variable number tandem repeat analysis (MLVA) or whole genome sequencing has shown that human and porcine isolates are indistinguishable,<sup>40,68,70</sup> furthering concerns about zoonotic and foodborne exposure.

## Cattle

Increasing attention has been paid to *C. difficile* in cattle, but mainly because of zoonotic disease concerns rather than animal health concerns. The role of *C. difficile* in cattle health is unclear and probably minimal. In calves, an association between the presence of *C. difficile* or its toxins in feces and diarrhea has been reported.<sup>49,88,106</sup> However, disease could not be reproduced experimentally in neonatal calves.<sup>107</sup> There is no evidence of a role in disease in adult cattle.

As with other species, *C. difficile* can be found in a variable percentage of healthy individuals. There is a strong influence of age, with young calves having the highest shedding rates.<sup>20,21,37,58,72,88,109</sup> Shedding rates for adult dairy cattle (10%),<sup>20</sup> adult beef cattle (3.3–18%),<sup>38,50,109,110</sup> adult cattle of unknown type (1.5%),<sup>111</sup> and calves (2–60%)<sup>20,37,43,72,74,88,101</sup> have been reported. Farm management and health status factors, such as diet change, mastitis, number of calves, and antimicrobial exposure, have been associated with increased risk of *C. difficile* shedding.<sup>20,21,88</sup>

Strain data have been variable, and there may be pronounced regional differences in ribotype distribution. Some authors, mainly from North America and Europe, have reported a predominance or exclusive isolation of livestock-associated ribotype 078.<sup>37,38,49,88,110,114</sup> The closely related ribotype 126 has also been found in Australian cattle.<sup>72,74</sup> Other studies have reported more diversity, including various

common human strains such as ribotypes 033, 001, 014/020, and 027 as well as strains rarely or never recognized in humans.<sup>20,21,50,72,74,88</sup>

## Small ruminants

There has been very little study of sheep and goats, although *C. difficile* has been identified in a small (2–9.5%) percentage of healthy individuals.<sup>18,44,71,111</sup> As with other species, the prevalence may be higher in young individuals.<sup>71</sup> Strain data are very limited, but a variety of strains have been reported, including strains that have been found in humans and other animal species (e.g., ribotypes 014, 010, 045).<sup>18,71,111</sup> There is no evidence that CDI is a relevant disease in these species.

## Dogs

The role of *C. difficile* in enteric disease in dogs remains enigmatic. An association between the presence of *C. difficile* toxins in feces and diarrhea has been reported in some studies,<sup>139</sup> but not others.<sup>32,35</sup> Additionally, disease could not be reproduced experimentally.<sup>36</sup> Accurate diagnosis of CDI is a challenge in dogs because of limited information about optimal methods, interpretation of results, and the potentially high background colonization rate in some dog populations. Whether *C. difficile* is a common cause of disease, a rare cause of disease, a relevant coinfection, or a harmless commensal remains to be determined.

Regardless of its role (or lack thereof) in disease in dogs, *C. difficile* can be found in a small percentage of healthy individuals. Prevalences of 0–6% tend to be reported in healthy adult dogs,<sup>82,83,102,116,130,139,144</sup> but higher rates have also been identified (e.g., 17%).<sup>129</sup> Specific groups from which higher rates of shedding have been identified include hospitalized dogs (19–22%),<sup>35,130</sup> veterinary hospital outpatients (14–33%),<sup>60,130,133</sup> dogs that visit human healthcare facilities (41–58%),<sup>83,84</sup> and racing sled dogs (58%).<sup>92</sup> Colonization rates can be high in young puppies, as highlighted by a 62% prevalence in neonates<sup>30</sup> and a small longitudinal study that reported a cumulative prevalence of 100% in 2 litters.<sup>3</sup> Shedding rates of 6.7–12% have been reported in dogs with gastrointestinal disease,<sup>10,41,100,139</sup> similar to the range seen in healthy dog populations. Reported risk factors for *C. difficile* shedding include contact with human healthcare facilities,<sup>83</sup> exposure to groups of children,<sup>83</sup> antimicrobial treatment,<sup>35,60,82,83</sup> and administration of immunosuppressive drugs during hospitalization.<sup>35,60,82,83</sup> There may be a major human influence on *C. difficile* shedding in dogs, given that living with an owner that has been treated with antibiotics<sup>83</sup> and living with an immunocompromised owner<sup>136</sup> have been associated with increased risk of *C. difficile* shedding. These presumably are related to an increased risk of the owner shedding *C. difficile*, with subsequent direct or indirect transmission to their dogs. A study that combined dog and cat data identified proton pump inhibitor (PPI)

administration as a risk factor,<sup>102</sup> an interesting finding given increasing recognition of the association of PPI administration with *C. difficile* shedding in humans.<sup>11,89</sup> That study also reported antimicrobial exposure, and contact with a person with diarrhea as risk factors.

Strains identified in dogs tend to mimic those found in people, including some of the most common strains found in human CDI, such as ribotypes 001, 014, and 106.<sup>5,10,100,102,116,136</sup> The livestock-associated ribotype 078 has been reported less commonly,<sup>100,102</sup> and, although rare, the hypervirulent human pathogen ribotype 027 has also been found.<sup>81,102</sup> Whether colonization with the same strains represents human–dog transmission or common source exposure is unclear. Likely intra-house transmission of *C. difficile* from infected owners to their dogs was identified in 2 of 5 of dogs in a small study of households of people with CDI.<sup>85</sup> In that study, dogs were negative on baseline testing then acquired *C. difficile* that had an indistinguishable pulsed-field gel electrophoresis (PFGE) type compared to the infected person. However, in a study of households with healthy people and pets, the same strain of *C. difficile* was not identified in people and pets in any of 415 households.<sup>102</sup>

### Cats

*C. difficile* has been isolated from healthy and diarrheic cats.<sup>2,29,35,86,90,104,116,143</sup> Prevalence data are limited and sometimes hard to discern because some studies do not differentiate diarrheic versus healthy cats. Overall rates of 2.5–38.1% have been reported,<sup>35,86,102,116,135</sup> with the low end of that range (i.e., < 5%) likely representing the shedding rate for the general, healthy, homed cat population. Typing data are limited, but as for dogs, common human strains are often found, such as ribotypes 010, 106, and 014/020.<sup>102,116,122</sup> In a small study of people with CDI, likely human–animal transmission (based on the cat being negative on an initial sample then later harboring *C. difficile* with an indistinguishable PFGE type compared to the person) was identified in 1 of 8 cats.<sup>85</sup>

The clinical relevance of *C. difficile* in cats is unclear. Diagnosis of CDI has been made in diarrheic cats,<sup>143</sup> but the true role in disease is unclear because of the presence of the bacterium in healthy animals. Evidence of an association between *C. difficile* and diarrhea in cats is currently lacking.

### Poultry

There is no evidence that *C. difficile* is a relevant pathogen in poultry. Surveillance studies have investigated shedding of the bacterium in healthy birds, where, as in other species, it can be found in an age-dependent manner<sup>59,124</sup> and perhaps of greatest relevance from a potential zoonotic disease standpoint. Prevalences of 0–62% have been reported in healthy chickens.<sup>1,51,59,76,103,105,149</sup> High strain diversity has been identified, with perhaps a higher proportion of nontoxigenic strains compared to other species. Strains that have been

identified include common human strains (e.g., ribotypes 001, 014, 039), livestock-associated ribotype 078 or related strains, and various nontoxigenic strains.<sup>1,51,59,76,105</sup> As with some other livestock species, there appear to be regional differences in strain distribution.

### Wildlife

Given the estimate that *C. difficile* evolved well before humans (and perhaps before mammals),<sup>53</sup> it should be unsurprising that the bacterium can be found in wildlife. It has been isolated from an impressive array of wildlife species, including wild carnivores, omnivores, herbivores, and insectivores, including skunks, opossums, raccoons, coatis, bats, bears, rats, and other rodents, feral swine, various bird species, and wild canids.<sup>22,27,45,54,62,77,94,112,121,123,131,142</sup> It is likely that it has been isolated from most species for which a concerted effort has been made to identify it. Whether *C. difficile* is a pathogen in wildlife species is difficult to discern. Most of the available information is from surveillance of healthy animals or animals with unknown health status, something that precludes assessment of its role as a potential opportunistic (as opposed to obligate) pathogen. Sporadic case reports have implicated *C. difficile* in disease of individual animals (e.g., ocelot, captive Kodiak bear, harbor seal)<sup>7,99,120</sup>; however, it can be difficult to put results of detection of the organism or even its toxins into context without understanding the prevalence of the bacterium in healthy cohorts.

Strains found in wildlife often reflect their degree of exposure to human habitats. Three different types of wildlife can be considered, with potential differences in the epidemiology and ecology of the bacterium. These consist of urban wildlife, wildlife associated with farms, and wildlife more removed from humans and domestic animals.

Urban wildlife can have close contact with humans, human environments, human waste, and discarded food products, creating the potential for exposure to a variety of *C. difficile* strains. Most study of urban wildlife has involved rodents. Over 13% of rats from a low socioeconomic status area of Vancouver, Canada, were shedding *C. difficile* in one study, with a combination of known human strains (including ribotypes 001 and 014), livestock- (and food)-associated ribotype 078, and previously unknown strains,<sup>54</sup> suggesting exposure through contact with human environments or feces and food waste, as well as endemic rat- or wildlife-associated strains. A lower prevalence (1%), with no isolate characterization, was identified in rats from New York City.<sup>45</sup> However, a study of house mice in New York and mice caught in the Dutch city of Utrecht yielded prevalences of 4% and 35%, respectively, consisting predominantly of ribotypes that have been found in people.<sup>31,145</sup>

Animals living on or near farms can be exposed to pathogens from livestock, direct contact with farming areas, or indirectly through manure spread on fields or via runoff. Unsurprisingly, *C. difficile* can be isolated from species such

as raccoons and rats that live on or near farms, sometimes at high prevalences (up to 32%).<sup>9,39,62,77</sup> The livestock-associated ribotype 078 predominates in wildlife from farms; however, isolates more often associated with humans can also be found.<sup>9,28,39,62,77</sup> Birds also have contact with farm environments, as potentially highlighted by detection of *C. difficile*, including ribotype 078, in 4% of migrating European barn swallows.<sup>22</sup>

Although human or domestic animal contact is common for many wildlife populations, *C. difficile* has been identified in animals well removed from human influences. For example, it was isolated from 15% of polar bears in the Canadian Arctic, with isolates being strains that had not been previously identified in humans.<sup>142</sup> More study of remote wildlife is needed to help understand the natural (non-human-associated) ecology of this organism.

The role of wildlife in the epidemiology of human or domestic animal CDI is unclear. However, there is some potential for wildlife to be a reservoir of novel strains, to bridge agricultural and urban habitats, and to transmit *C. difficile* over long distances.

## Conclusion

*Clostridium difficile* is a genetically, evolutionarily, and ecologically diverse organism that can be found in a multitude of animal hosts. Why it can cause devastating disease in some species and be an innocuous commensal in others remains unclear and could offer insight into the pathophysiology of CDI.

Although there are marked differences in reported prevalences and strain distributions between studies, species, and regions, a few common themes emerge. The bacterium can be found in a variable and sometimes high prevalence, even in species in which it is a clear pathogen. Shedding is much more common in young animals, particularly neonates, likely as a result of a less well-developed protective gut microbiota, with rapid decreases in shedding with increasing age.

Strain distributions are variable between and within species. Of significant concern is the predominance of the livestock-associated ribotype 078 and related clade 5 strains because of increasing recognition of community-associated disease in people caused by this lineage.<sup>68,76,79</sup> However, a diverse range of other types can be found, including some of the most common human strains. The natural ecology of *C. difficile* and its movement between different species remains unclear. Similarity in strains found in some species (including humans and animals) supports some degree of interspecies movement or common source of exposure; however, the dynamics of interspecies (including zoonotic) transmission of this important and enigmatic bacterium remain to be clarified.


## Declaration of conflicting interests

The author declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The author received no financial support for the research, authorship, and/or publication of this article.

## ORCID iD

J. Scott Weese  <https://orcid.org/0000-0003-1896-1937>

## References

1. Abdel-Glil MY, et al. Presence of *Clostridium difficile* in poultry and poultry meat in Egypt. *Anaerobe* 2018;51: 21–25.
2. Álvarez-Pérez S, et al. Data from a survey of *Clostridium perfringens* and *Clostridium difficile* shedding by dogs and cats in the Madrid region (Spain), including phenotypic and genetic characteristics of recovered isolates. *Data Brief* 2017;14:88–100.
3. Álvarez-Pérez S, et al. Faecal shedding of antimicrobial-resistant *Clostridium difficile* strains by dogs. *J Small Anim Pract* 2015;56:190–195.
4. Alvarez-Perez S, et al. Prevalence of *Clostridium difficile* in diarrhoeic and non-diarrhoeic piglets. *Vet Microbiol* 2009;137:302–305.
5. Alvarez-Perez S, et al. Prevalence and characteristics of *Clostridium perfringens* and *Clostridium difficile* in dogs and cats attended in diverse veterinary clinics from the Madrid region. *Anaerobe* 2017;48:47–55.
6. Alvarez-Perez S, et al. High prevalence of the epidemic *Clostridium difficile* PCR ribotype 078 in Iberian free-range pigs. *Res Vet Sci* 2013;95:358–361.
7. Anderson CE, et al. Clinical and epidemiologic considerations of *Clostridium difficile* in harbor seals (*Phoca vitulina*) at a marine mammal rehabilitation center. *J Zoo Wildl Med* 2015;46:191–197.
8. Andino-Molina M, et al. Multidrug-resistant *Clostridium difficile* ribotypes 078 and 014/5-FLI01 in piglets from Costa Rica. *Anaerobe* 2019;55:78–82.
9. Andrés-Lasheras S, et al. Presence of *Clostridium difficile* in pig faecal samples and wild animal species associated with pig farms. *J Appl Microbiol* 2017;122:462–472.
10. Andrés-Lasheras S, et al. Preliminary studies on isolates of *Clostridium difficile* from dogs and exotic pets. *BMC Vet Res* 2018;14:77.
11. Anjewierden S, et al. Risk factors for *Clostridium difficile* infection in pediatric inpatients: a meta-analysis and systematic review. *Infect Control Hosp Epidemiol* 2019;40:420–426.
12. Arroyo LG, et al. Experimental *Clostridium difficile* enterocolitis in foals. *J Vet Intern Med* 2004;18:734–738.
13. Arroyo LG, et al. Duodenitis-proximal jejunitis in horses after experimental administration of *Clostridium difficile* toxins. *J Vet Intern Med* 2017;31:158–163.
14. Arroyo LG, et al. Potential role of *Clostridium difficile* as a cause of duodenitis-proximal jejunitis in horses. *J Med Microbiol* 2006;55:605–608.
15. Arruda PHE, et al. Effect of age, dose and antibiotic therapy on the development of *Clostridium difficile* infection in neonatal piglets. *Anaerobe* 2013;22:104–110.
16. Asai T, et al. *Clostridium difficile* isolated from the fecal contents of swine in Japan. *J Vet Med Sci* 2013;75:539–541.

17. Avbersek J, et al. Diversity of *Clostridium difficile* in pigs and other animals in Slovenia. *Anaerobe* 2009;15:252–255.
18. Avbersek J, et al. *Clostridium difficile* in goats and sheep in Slovenia: characterisation of strains and evidence of age-related shedding. *Anaerobe* 2014;28:163–167.
19. Baker AA, et al. Prevalence and diversity of toxigenic *Clostridium perfringens* and *Clostridium difficile* among swine herds in the midwest. *Appl Environ Microbiol* 2010;76:2961–2967.
20. Bandelj P, et al. Identification of risk factors influencing *Clostridium difficile* prevalence in middle-size dairy farms. *Vet Res* 2016;47:41.
21. Bandelj P, et al. Quantification of *Clostridioides (Clostridium) difficile* in feces of calves of different age and determination of predominant *Clostridioides difficile* ribotype 033 relatedness and transmission between family dairy farms using multilocus variable-number tandem-repeat analysis. *BMC Vet Res* 2018;14:298.
22. Bandelj P, et al. Prevalence and molecular characterization of *Clostridium difficile* isolated from European barn swallows (*Hirundo rustica*) during migration. *BMC Vet Res* 2014;10:40.
23. Båverud V, et al. *Clostridium difficile* associated with acute colitis in mares when their foals are treated with erythromycin and rifampicin for *Rhodococcus equi* pneumonia. *Equine Vet J* 1998;30:482–488.
24. Båverud V, et al. *Clostridium difficile*: prevalence in horses and environment, and antimicrobial susceptibility. *Equine Vet J* 2003;35:465–471.
25. Båverud V, et al. *Clostridium difficile* associated with acute colitis in mature horses treated with antibiotics. *Equine Vet J* 1997;29:279–284.
26. Bidet P, et al. Development of a new PCR-ribotyping method for *Clostridium difficile* based on ribosomal RNA gene sequencing. *FEMS Microbiol Lett* 1999;175:261–266.
27. Bondo KJ, et al. *Salmonella*, *Campylobacter*, *Clostridium difficile*, and anti-microbial resistant *Escherichia coli* in the faeces of sympatric meso-mammals in southern Ontario, Canada. *Zoonoses Public Health* 2019;66:406–416.
28. Bondo KJ, et al. Longitudinal study of *Clostridium difficile* shedding in raccoons on swine farms and conservation areas in Ontario, Canada. *BMC Vet Res* 2015;11:254.
29. Borriello SP, et al. Household pets as a potential reservoir for *Clostridium difficile* infection. *J Clin Pathol* 1983;36:84–87.
30. Buogo C, et al. Présence de *Campylobacter* spp., *Clostridium difficile*, *C. perfringens* et salmonelles dans des nichées de chiots et chez des chiens adultes d'un refuge [Presence of *Campylobacter* spp., *Clostridium difficile*, *C. perfringens* and salmonellae in litters of puppies and in adult dogs in a shelter]. *Schweiz Arch Tierheilkd* 1995;137:165–171. French.
31. Burt SA, et al. Wild mice in and around the city of Utrecht, the Netherlands, are carriers of *Clostridium difficile* but not ESBL-producing Enterobacteriaceae, *Salmonella* spp. or MRSA. *Lett Appl Microbiol* 2018;67:513–519.
32. Busch K, et al. *Clostridium perfringens* enterotoxin and *Clostridium difficile* toxin A/B do not play a role in acute haemorrhagic diarrhoea syndrome in dogs. *Vet Rec* 2015;176:253.
33. Chan G, et al. A retrospective study on the etiological diagnoses of diarrhea in neonatal piglets in Ontario, Canada, between 2001 and 2010. *Can J Vet Res* 2013;77:254–260.
34. Cho A, et al. Low prevalence of *Clostridium difficile* in slaughter pigs in Korea. *J Food Prot* 2015;78:1034–1036.
35. Clooten J, et al. Prevalence and risk factors for *Clostridium difficile* colonization in dogs and cats hospitalized in an intensive care unit. *Vet Microbiol* 2008;129:209–214.
36. Clooten JK, et al. Genotypic and phenotypic characterization of *Clostridium perfringens* and *Clostridium difficile* in diarrheic and healthy dogs. *J Vet Intern Med* 2003;17:123; author reply 123.
37. Costa MC, et al. Epidemiology of *Clostridium difficile* on a veal farm: prevalence, molecular characterization and tetracycline resistance. *Vet Microbiol* 2011;152:379–384.
38. Costa MC, et al. Prevalence and molecular characterization of *Clostridium difficile* isolated from feedlot beef cattle upon arrival and mid-feeding period. *BMC Vet Res* 2012;8:38.
39. de Oliveira CA, et al. Rodents are carriers of *Clostridioides difficile* strains similar to those isolated from piglets. *Anaerobe* 2018;51:61–63.
40. Debast SB, et al. *Clostridium difficile* PCR ribotype 078 toxinotype V found in diarrhoeal pigs identical to isolates from affected humans. *Environ Microbiol* 2009;11:505–511.
41. Diniz AN, et al. The incidence of *Clostridioides difficile* and *Clostridium perfringens* *netF*-positive strains in diarrheic dogs. *Anaerobe* 2018;49:58–62.
42. Donaldson MT, Palmer JE. Prevalence of *Clostridium perfringens* enterotoxin and *Clostridium difficile* toxin A in feces of horses with diarrhea and colic. *J Am Vet Med Assoc* 1999;215:358–361.
43. Doosti A, Mokhtari-Farsani A. Study of the frequency of *Clostridium difficile* *tcdA*, *tcdB*, *cdtA* and *cdtB* genes in feces of calves in south west of Iran. *Ann Clin Microbiol Antimicrob* 2014;13:21.
44. Esfandiari Z, et al. Isolation and characterization of *Clostridium difficile* in farm animals from slaughterhouse to retail stage in Isfahan, Iran. *Foodborne Pathog Dis* 2015;12:864–866.
45. Firth C, et al. Detection of zoonotic pathogens and characterization of novel viruses carried by commensal *Rattus norvegicus* in New York City. *MBio* 2014;5:e01933–14.
46. Fry PR, et al. Antimicrobial resistance, toxinotype, and genotypic profiling of *Clostridium difficile* isolates of swine origin. *J Clin Microbiol* 2012;50:2366–2372.
47. Gustafsson A, et al. Study of faecal shedding of *Clostridium difficile* in horses treated with penicillin. *Equine Vet J* 2004;36:180–182.
48. Gustafsson A, et al. The association of erythromycin ethylsuccinate with acute colitis in horses in Sweden. *Equine Vet J* 1997;29:314–318.
49. Hammitt MC, et al. A possible role for *Clostridium difficile* in the etiology of calf enteritis. *Vet Microbiol* 2008;127:343–352.
50. Hampikyan H, et al. The prevalence of *Clostridium difficile* in cattle and sheep carcasses and the antibiotic susceptibility of isolates. *Meat Sci* 2018;139:120–124.
51. Harvey RB, et al. *Clostridium difficile* in poultry and poultry meat. *Foodborne Pathog Dis* 2011;8:1321–1323.
52. Hawken P, et al. Longitudinal study of *Clostridium difficile* and methicillin-resistant *Staphylococcus aureus* associated with pigs from weaning through to the end of processing. *J Food Prot* 2013;76:624–630.

53. He M, et al. Evolutionary dynamics of *Clostridium difficile* over short and long time scales. *Proc Natl Acad Sci U S A* 2010;107:7527–7532.
54. Himsworth CG, et al. Carriage of *Clostridium difficile* by wild urban Norway rats (*Rattus norvegicus*) and black rats (*Rattus rattus*). *Appl Environ Microbiol* 2014;80:1299–1305.
55. Hoffer E, et al. Low occurrence of *Clostridium difficile* in fecal samples of healthy calves and pigs at slaughter and in minced meat in Switzerland. *J Food Prot* 2010;73:973–975.
56. Hopman NEM, et al. High occurrence of various *Clostridium difficile* PCR ribotypes in pigs arriving at the slaughterhouse. *Vet Q* 2011;31:179–181.
57. Hopman NEM, et al. Acquisition of *Clostridium difficile* by piglets. *Vet Microbiol* 2011;149:186–192.
58. Houser BA, et al. Prevalence of *Clostridium difficile* toxin genes in the feces of veal calves and incidence of ground veal contamination. *Foodborne Pathog Dis* 2012;9:32–36.
59. Hussain I, et al. Molecular characteristics of *Clostridium difficile* isolates from human and animals in the north eastern region of India. *Mol Cell Probes* 2016;30:306–311.
60. Hussain I, et al. Isolation and characterization of *Clostridium difficile* from pet dogs in Assam, India. *Anaerobe* 2015;36:9–13.
61. Indra A, et al. Characterization of *Clostridium difficile* isolates using capillary gel electrophoresis-based PCR ribotyping. *J Med Microbiol* 2008;57:1377–1382.
62. Jardine CM, et al. Detection of *Clostridium difficile* in small and medium-sized wild mammals in Southern Ontario, Canada. *J Wildl Dis* 2013;49:418–421.
63. Jhung MA, et al. Toxinotype V *Clostridium difficile* in humans and food animals. *Emerg Infect Dis* 2008;14:1039–1045.
64. Jones RL, et al. Isolation of *Clostridium difficile* and detection of cytotoxin in the feces of diarrheic foals in the absence of antimicrobial treatment. *J Clin Microbiol* 1987;25:1225–1227.
65. Kecerova Z, et al. *Clostridium difficile* isolates derived from Czech horses are resistant to enrofloxacin; cluster to clades 1 and 5 and ribotype 033 predominates. *Anaerobe* 2019;56:17–21.
66. Keel K, et al. Prevalence of PCR ribotypes among *Clostridium difficile* isolates from pigs, calves, and other species. *J Clin Microbiol* 2007;45:1963–1964.
67. Keessen EC, et al. The relation between farm specific factors and prevalence of *Clostridium difficile* in slaughter pigs. *Vet Microbiol* 2011;154:130–134.
68. Keessen EC, et al. *Clostridium difficile* infection associated with pig farms. *Emerg Infect Dis* 2013;19:1032–1034.
69. Kim HY, et al. High prevalence of *Clostridium difficile* PCR ribotype 078 in pigs in Korea. *Anaerobe* 2018;51:42–46.
70. Knetsch CW, et al. Whole genome sequencing reveals potential spread of *Clostridium difficile* between humans and farm animals in the Netherlands, 2002 to 2011. *Euro Surveill* 2014;19:20954.
71. Knight DR, Riley TV. Prevalence of gastrointestinal *Clostridium difficile* carriage in Australian sheep and lambs. *Appl Environ Microbiol* 2013;79:5689–5692.
72. Knight DR, et al. *Clostridium difficile* carriage in Australian cattle: a cross-sectional study reveals high prevalence of non-PCR ribotype 078 strains in veal calves at slaughter. *Appl Environ Microbiol* 2013;79:2630–2635.
73. Knight DR, et al. Evolutionary and genomic insights into *Clostridioides difficile* sequence type 11: a diverse zoonotic and antimicrobial-resistant lineage of global one health importance. *MBio* 2019;10:e00446–19.
74. Knight DR, et al. Contamination of Australian newborn calf carcasses at slaughter with *Clostridium difficile*. *Clin Microbiol Infect* 2016;22:266.e1–e7.
75. Knight DR, et al. Nationwide surveillance study of *Clostridium difficile* in Australian neonatal pigs shows high prevalence and heterogeneity of PCR ribotypes. *Appl Environ Microbiol* 2015;81:119–123.
76. Koene MGJ, et al. *Clostridium difficile* in Dutch animals: their presence, characteristics and similarities with human isolates. *Clin Microbiol Infect* 2012;18:778–784.
77. Krijger IM, et al. *Clostridium difficile* in wild rodents and insectivores in The Netherlands. *Lett Appl Microbiol* 2019;69:35–40.
78. Krutova M, et al. The emergence of *Clostridium difficile* PCR ribotype 078 in piglets in the Czech Republic clusters with *Clostridium difficile* PCR ribotype 078 isolates from Germany, Japan and Taiwan. *Int J Med Microbiol* 2018;308:770–775.
79. Kuijper EJ, van Dissel JT. Spectrum of *Clostridium difficile* infections outside health care facilities. *CMAJ* 2008;179:747–748.
80. Lawson PA, et al. Reclassification of *Clostridium difficile* as *Clostridioides difficile* (Hall and O’Toole 1935) Prévot 1938. *Anaerobe* 2016;40:95–99.
81. Lefebvre SL, et al. Epidemic *Clostridium difficile* strain in hospital visitation dog. *Emerging Infect Dis* 2006;12:1036–1037.
82. Lefebvre SL, et al. Evaluation of the risks of shedding *Salmonellae* and other potential pathogens by therapy dogs fed raw diets in Ontario and Alberta. *Zoonoses Public Health* 2008;55:470–480.
83. Lefebvre SL, et al. Incidence of acquisition of methicillin-resistant *Staphylococcus aureus*, *Clostridium difficile*, and other health-care-associated pathogens by dogs that participate in animal-assisted interventions. *J Am Vet Med Assoc* 2009;234:1404–1417.
84. Lefebvre SL, et al. Prevalence of zoonotic agents in dogs visiting hospitalized people in Ontario: implications for infection control. *J Hosp Infect* 2006;62:458–466.
85. Loo VG, et al. Household transmission of *Clostridium difficile* to family members and domestic pets. *Infect Control Hosp Epidemiol* 2016;37:1342–1348.
86. Madewell BR, et al. *Clostridium difficile*: a survey of fecal carriage in cats in a veterinary medical teaching hospital. *J Vet Diagn Invest* 1999;11:50–54.
87. Madewell BR, et al. Apparent outbreaks of *Clostridium difficile*-associated diarrhea in horses in a veterinary medical teaching hospital. *J Vet Diagn Invest* 1995;7:343–346.
88. Magistrali CF, et al. Prevalence and risk factors associated with *Clostridium difficile* shedding in veal calves in Italy. *Anaerobe* 2015;33:42–47.
89. Mallia G, et al. Examining the epidemiology and microbiology of *Clostridium difficile* carriage in elderly patients and residents of a healthcare facility in southern Ontario, Canada. *J Hosp Infect* 2018;99:461–468.

90. Marks SL, et al. Genotypic and phenotypic characterization of *Clostridium perfringens* and *Clostridium difficile* in diarrheic and healthy dogs. *J Vet Intern Med* 2002;16:533–540.
91. McDonald LC, et al. Recommendations for surveillance of *Clostridium difficile*-associated disease. *Infect Control Hosp Epidemiol* 2007;28:140–145.
92. McKenzie E, et al. Prevalence of diarrhea and enteropathogens in racing sled dogs. *J Vet Intern Med* 2010;24:97–103.
93. Medina-Torres CE, et al. Prevalence of *Clostridium difficile* in horses. *Vet Microbiol* 2011;152:212–215.
94. Miller MA, et al. Enteric bacterial pathogen detection in southern sea otters (*Enhydra lutris nereis*) is associated with coastal urbanization and freshwater runoff. *Vet Res* 2010;41:1.
95. Moono P, et al. Persistence of *Clostridium difficile* RT 237 infection in a Western Australian piggery. *Anaerobe* 2016;37:62–66.
96. Niwa H, et al. Postoperative *Clostridium difficile* infection with PCR ribotype 078 strain identified at necropsy in five Thoroughbred racehorses. *Vet Rec* 2013;173:607.
97. Norén T, et al. *Clostridium difficile* PCR ribotype 046 is common among neonatal pigs and humans in Sweden. *Clin Microbiol Infect* 2014;20:O2–O6.
98. Norman K, et al. Varied prevalence of *Clostridium difficile* in an integrated swine operation. *Anaerobe* 2009;15:256–260.
99. Orchard JL, et al. Antibiotic-associated colitis due to *Clostridium difficile* in a Kodiak bear. *Am J Vet Res* 1983;44:1547–1548.
100. Orden C, et al. Isolation of *Clostridium difficile* from dogs with digestive disorders, including stable metronidazole-resistant strains. *Anaerobe* 2017;43:78–81.
101. Pirs T, et al. Isolation of *Clostridium difficile* from food animals in Slovenia. *J Med Microbiol* 2008;57:790–792.
102. Rabold D, et al. The zoonotic potential of *Clostridium difficile* from small companion animals and their owners. *PLoS One* 2018;13:e0193411.
103. Razmyr J, et al. Toxigenic *Clostridium difficile* in retail packed chicken meat and broiler flocks in northeastern Iran. *Iran J Vet Res* 2017;18:271–274.
104. Riley TV, et al. Gastrointestinal carriage of *Clostridium difficile* in cats and dogs attending veterinary clinics. *Epidemiol Infect* 1991;107:659–665.
105. Rodriguez-Palacios A, et al. Three-week summer period prevalence of *Clostridium difficile* in farm animals in a temperate region of the United States (Ohio). *Can Vet J* 2014;55:786–789.
106. Rodriguez-Palacios A, et al. *Clostridium difficile* PCR ribotypes in calves, Canada. *Emerging Infect Dis* 2006;12:1730–1736.
107. Rodriguez-Palacios A, et al. Natural and experimental infection of neonatal calves with *Clostridium difficile*. *Vet Microbiol* 2007;124:166–172.
108. Rodriguez C, et al. Presence of *Clostridium difficile* in pigs and cattle intestinal contents and carcass contamination at the slaughterhouse in Belgium. *Int J Food Microbiol* 2013;166:256–262.
109. Rodriguez C, et al. *Clostridium difficile* in beef cattle farms, farmers and their environment: assessing the spread of the bacterium. *Vet Microbiol* 2017;210:183–187.
110. Rodriguez C, et al. *Clostridium difficile* in young farm animals and slaughter animals in Belgium. *Anaerobe* 2012;18:621–625.
111. Romano V, et al. Prevalence and genotypic characterization of *Clostridium difficile* from ruminants in Switzerland. *Zoonoses Public Health* 2012;59:545–548.
112. Rothenburger JL, et al. Livestock-associated methicillin-resistant *Staphylococcus aureus* and *Clostridium difficile* in wild Norway rats (*Rattus norvegicus*) from Ontario swine farms. *Can J Vet Res* 2018;82:66–69.
113. Rupnik M. Is *Clostridium difficile*-associated infection a potentially zoonotic and foodborne disease? *Clin Microbiol Infect* 2007;13:457–459.
114. Schneeberg A, et al. Presence of *Clostridium difficile* PCR ribotype clusters related to 033, 078 and 045 in diarrhoeic calves in Germany. *J Med Microbiol* 2013;62:1190–1198.
115. Schneeberg A, et al. *Clostridium difficile* genotypes in piglet populations in Germany. *J Clin Microbiol* 2013;51:3796–3803.
116. Schneeberg A, et al. Prevalence and distribution of *Clostridium difficile* PCR ribotypes in cats and dogs from animal shelters in Thuringia, Germany. *Anaerobe* 2012;18:484–488.
117. Schoster A, et al. Longitudinal study of *Clostridium difficile* and antimicrobial susceptibility of *Escherichia coli* in healthy horses in a community setting. *Vet Microbiol* 2012;159:364–370.
118. Schoster A, et al. Effect of a probiotic on prevention of diarrhea and *Clostridium difficile* and *Clostridium perfringens* shedding in foals. *J Vet Intern Med* 2015;29:925–931.
119. Schoster A, et al. Presence and molecular characterization of *Clostridium difficile* and *Clostridium perfringens* in intestinal compartments of healthy horses. *BMC Vet Res* 2012;8:94.
120. Silva RO, et al. *Clostridium difficile*-associated diarrhea in an ocelot (*Leopardus pardalis*). *Anaerobe* 2013;20:82–84.
121. Silva RO, et al. Carriage of *Clostridium difficile* in free-living South American coati (*Nasua nasua*) in Brazil. *Anaerobe* 2014;30:99–101.
122. Silva RO, et al. *Clostridium difficile* ribotypes in humans and animals in Brazil. *Mem Inst Oswaldo Cruz* 2015;110:1062–1065.
123. Silva ROS, et al. *Clostridium difficile* and *Clostridium perfringens* from wild carnivore species in Brazil. *Anaerobe* 2014;28:207–211.
124. Skraban J, et al. Changes of poultry faecal microbiota associated with *Clostridium difficile* colonisation. *Vet Microbiol* 2013;165:416–424.
125. Slifierz MJ, et al. Longitudinal study of the early-life fecal and nasal microbiotas of the domestic pig. *BMC Microbiol* 2015;15:184.
126. Squire MM, et al. Novel molecular type of *Clostridium difficile* in neonatal pigs, Western Australia. *Emerg Infect Dis* 2013;19:790–792.
127. Stabler RA, et al. Comparative phylogenomics of *Clostridium difficile* reveals clade specificity and microevolution of hypervirulent strains. *J Bacteriol* 2006;188:7297–7305.
128. Stein K, et al. PCR-ribotype distribution of *Clostridium difficile* in Irish pigs. *Anaerobe* 2017;48:237–241.
129. Stone NE, et al. More than 50% of *Clostridium difficile* isolates from pet dogs in Flagstaff, USA, carry toxigenic genotypes. *PLoS One* 2016;11:e0164504.



130. Struble AL, et al. Fecal shedding of *Clostridium difficile* in dogs: a period prevalence survey in a veterinary medical teaching hospital. *J Vet Diagn Invest* 1994;6:342–347.
131. Thakur S, et al. Detection of *Clostridium difficile* and *Salmonella* in feral swine population in North Carolina. *J Wildl Dis* 2011;47:774–776.
132. Thean S, et al. *Clostridium difficile* in horses in Australia—a preliminary study. *J Med Microbiol* 2011;60:1188–1192.
133. Usui M, et al. Distribution and characterization of *Clostridium difficile* isolated from dogs in Japan. *Anaerobe* 2016;37:58–61.
134. Waters EH, et al. Typhlocolitis caused by *Clostridium difficile* in suckling piglets. *J Vet Diagn Invest* 1998;10:104–108.
135. Weber A, et al. Untersuchungen zum Vorkommen von *Clostridium difficile* in Kotproben von Hunden und Katzen [The occurrence of *Clostridium difficile* in fecal samples of dogs and cats]. *Zentralbl Veterinarmed B* 1989;36:568–576. German.
136. Weese JS, et al. Evaluation of *Clostridium difficile* in dogs and the household environment. *Epidemiol Infect* 2010;138:1100–1104.
137. Weese JS, et al. *Clostridium difficile* and methicillin-resistant *Staphylococcus aureus* shedding by slaughter-age pigs. *BMC Vet Res* 2011;7:41.
138. Weese JS, et al. A prospective study of the roles of *Clostridium difficile* and enterotoxigenic *Clostridium perfringens* in equine diarrhoea. *Equine Vet J* 2001;33:403–409.
139. Weese JS, et al. The roles of *Clostridium difficile* and enterotoxigenic *Clostridium perfringens* in diarrhea in dogs. *J Vet Intern Med* 2001;15:374–378.
140. Weese JS, et al. *Clostridium difficile* associated diarrhoea in horses within the community: predictors, clinical presentation and outcome. *Equine Vet J* 2006;38:185–188.
141. Weese JS, et al. Longitudinal investigation of *Clostridium difficile* shedding in piglets. *Anaerobe* 2010;16:501–504.
142. Weese JS, et al. *Clostridium (Clostridioides) difficile* shedding by polar bears (*Ursus maritimus*) in the Canadian Arctic. *Anaerobe* 2019;57:35–38.
143. Weese JS, et al. Suspected *Clostridium difficile*-associated diarrhea in two cats. *J Am Vet Med Assoc* 2001;218:1436–1439.
144. Wei Y, et al. Prevalence, genotype and antimicrobial resistance of *Clostridium difficile* isolates from healthy pets in Eastern China. *BMC Infect Dis* 2019;19:46.
145. Williams SH, et al. New York City house mice (*Mus musculus*) as potential reservoirs for pathogenic bacteria and antimicrobial resistance determinants. *MBio* 2018;9:e00624–18.
146. Yaeger M, et al. A survey of agents associated with neonatal diarrhea in Iowa swine including *Clostridium difficile* and porcine reproductive and respiratory syndrome virus. *J Vet Diagn Invest* 2002;14:281–287.
147. Yaeger MJ, et al. A prospective, case control study evaluating the association between *Clostridium difficile* toxins in the colon of neonatal swine and gross and microscopic lesions. *J Vet Diagn Invest* 2007;19:52–59.
148. Zhang LJ, et al. The first isolation of *Clostridium difficile* RT078/ST11 from pigs in China. *PLoS One* 2019;14:e0212965.
149. Zidaric V, et al. High diversity of *Clostridium difficile* genotypes isolated from a single poultry farm producing replacement laying hens. *Anaerobe* 2008;14:325–327.