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Review

Infectome: A platform to trace infectious triggers of autoimmunity



Dimitrios P. Bogdanos^{a,b,*}, Daniel S. Smyk^a, Pietro Invernizzi^c, Eirini I. Rigopoulou^b, Miri Blank^d, Shideh Pouria^e, Yehuda Shoenfeld^d

^a Institute of Liver Studies, King's College London School of Medicine at King's College Hospital, Denmark Hill Campus, London, UK

^b Department of Medicine, University of Thessaly Medical School, Thessaly, Mezourlo, Larissa, Greece

^c Center for Autoimmune Liver Diseases, Humanitas Clinical and Research Center, Rozzano (MI), Italy

^d The Zabłudowicz Center for Autoimmune Diseases, Sheba Medical Center, Israel

^e Nutritional Sciences Division, King's College School of Medicine, Franklin Wilkins Building, London, UK

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ABSTRACT

The “exposome” is a term recently used to describe all environmental factors, both exogenous and endogenous, which we are exposed to in a lifetime. It represents an important tool in the study of autoimmunity, complementing classical immunological research tools and cutting-edge genome wide association studies (GWAS). Recently, environmental wide association studies (EWAS) investigated the effect of environment in the development of diseases. Environmental triggers are largely subdivided into infectious and non-infectious agents. In this review, we introduce the concept of the “infectome”, which is the part of the exposome referring to the collection of an individual's exposures to infectious agents. The infectome directly relates to geoepidemiological, serological and molecular evidence of the co-occurrence of several infectious agents associated with autoimmune diseases that may provide hints for the triggering factors responsible for the pathogenesis of autoimmunity. We discuss the implications that the investigation of the infectome may have for the understanding of microbial/host interactions in autoimmune diseases with long, pre-clinical phases. It may also contribute to the concept of the human body as a superorganism where the microbiome is part of the whole organism, as can be seen with mitochondria which existed as microbes prior to becoming organelles in eukaryotic cells of multicellular organisms over time. A similar argument can now be made in regard to normal intestinal flora, living in symbiosis within the host. We also provide practical examples as to how we can characterise and measure the totality of a disease-specific infectome, based on the experimental approaches employed from the “immunome” and “microbiome” projects.

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Contents

1. Introduction	727
2. The exposome: what is it, and how do we measure it?	727
3. Exposome, infectome and autoimmunity	728
4. SLE as an infectome model: known infections, stronger links	729
5. Multiple sclerosis as an infectome model for relapse remittance clinical states	730
6. Primary biliary cirrhosis as an infectome model disease	731
7. How to study the infectome	732
8. Lessons that can be learned from the microbiome and immunome projects	732
9. Who to screen?	732
10. How to screen?	733
11. Where to screen?	733

Abbreviations: AMA, anti-mitochondrial antibody; ANA, anti-nuclear antibody; CMV, cytomegalovirus; CSF, cerebrospinal fluid; EBV, Epstein–Barr virus; EWAS, environmental-wide association study; FDR, first degree relatives; GWAS, genome wide association study; HHV6, human herpes virus 6; LC–MS/MS, liquid chromatography–tandem mass spectrometry (LC–MS/MS); MS, multiple sclerosis; IBS, irritable bowel syndrome; PBC, primary biliary cirrhosis; PCR, polymerase chain reaction; PDC, pyruvate dehydrogenase complex; SLE, systemic lupus erythematosus.

* Corresponding author at: Institute of Liver Studies, Division of Transplantation Immunology and Mucosal Biology, King's College London Medical School at King's College London Hospital, Denmark Hill Campus, London SE5 9RS, UK. Tel./fax: +44 2032993397, +44 302410555138.

E-mail addresses: dimitrios.bogdanos@kcl.ac.uk, bogdanos@med.uth.gr (D.P. Bogdanos). URL: <http://www.bogdanoslab.com> (D.P. Bogdanos).

12. Conclusion	733
Take-home messages	734
Acknowledgements	734
References	734

1. Introduction

It is widely accepted that the vast majority of diseases develop from the interaction between genes and the environment [1–3]. This concept has formed the basis for studying the pathogenesis of many diseases including autoimmune diseases [1,2,4]. There are now almost 100 categories of autoimmune diseases, both organ specific or systemic in nature. Although individual autoimmune diseases are relatively rare in any population that has been investigated so far, projected data estimate that approximately 5–20% of North Americans are affected by at least one autoimmune disease [5]. Some of the best known autoimmune diseases are type 1 diabetes, rheumatoid arthritis, multiple sclerosis, Grave's disease, Hashimoto's thyroiditis, myasthenia gravis, systemic lupus erythematosus, Sjögren's syndrome, scleroderma and autoimmune liver diseases such as autoimmune hepatitis, primary sclerosing cholangitis, and primary biliary cirrhosis (PBC) [5]. The observation that many autoimmune diseases may affect one individual, has led to the concept of the Mosaic of Autoimmunity [6–13].

The study of genetic and epigenetic factors linking to autoimmunity is the focus of ongoing research [14]. Also, the exact signalling cascades that govern the perpetuation of inflammatory processes responsible for tissue destruction are poorly understood [15]. In recent years, genome wide association studies (GWAS) have identified numerous gene–disease associations, many of which include autoimmune diseases [2]. Although these associations are connected to genetic susceptibility, the genetic 'dosage' or number of associated genes required for disease development is not well defined [16]. Although GWAS have been instrumental in unlocking a pathogenetic starting point, exposure to environmental factors is also likely to contribute to the actual development of most diseases, and work with genetics in the induction of autoimmune disease [1,2,17]. For example, smoking has been indicated to increase the risk of developing MS in individuals with HLA-DRB1*1501 [18]. Epidemiological studies using toxicological, microbiological, biochemical, and immunological testing are important in order to identify these environmental agents, which include infectious organisms, xenobiotics, chemical compounds, heavy metals from prostheses and dental materials, radiation, vaccines, and foods to name but a few [1,19–22]. Heavy metals and vaccinations have also been implicated in the pathogenesis of autoimmune (auto-inflammatory) syndrome induced by adjuvants (ASIA) [23–32]. In fact, silicosis, Gulf war syndrome (GWS), macrophagic myofasciitis syndrome (MMF) and post-vaccination phenomena were linked with past exposure to adjuvants.

Since GWAS underlined the view that multiple genes are needed to induce autoimmunity [2], it is also likely that several environmental triggers either complement or substitute each other to provoke immune mediated processes which then lead to autoimmunity. The additive effects of these triggers, and their interaction with susceptible genes, remain poorly defined.

In recent years, the concept of an “exposome” has been introduced, as a means of collating, and possibly measuring the effects of environmental factors. The exposome takes into account all internal and external stimuli associated with a disease, and provides a potentially quantifiable way for the evaluation of environmental factors (Fig. 1). This review will examine the role of the exposome in relation to major exemplary autoimmune disease where genetics and environment most likely play key roles in pathogenesis. The role of the “infectome” as the infectious component of the microbiome/

exposome that takes part (directly or indirectly) in the development of autoimmunity will also be introduced (Fig. 2).

2. The exposome: what is it, and how do we measure it?

The exposome represents the totality of all environmental exposures, both exogenous and endogenous, which begins from conception, and extends throughout our lives [33–37]. This differs from previous epidemiological studies, which have largely concentrated on the external environment, and specifically to air, water, soil and food [35]. In contrast to approaches limited to external triggers, the study of the exposome needs to address endogenous factors directly or indirectly related to the environment [36,37]. Endogenous sources include by-products of inflammation, lipid peroxidation, as well as oxidative stress [35]. Some of these components may act as nucleophiles or electrophiles, and as such, would be capable of DNA and protein modification [38]. Indeed, bacterial alterations in glycosylation patterns of immunoglobulins have been noted in several studies [39–41]. The collation of all these exposures may provide a fingerprint of a particular disease, which would likely complement data from GWAS [38].

The predominant problems in examining environmental exposures are the difficulties of quantification and normalisation as well as the determination of exposure sequences. This information is required, as alterations induced by one environmental agent may be necessary for a subsequent exposure to have its effect [2]. Moreover, environmental exposures within a population, as well as within an individual, are greatly varied. Given the myriad of causative possibilities in one individual, it may be useful clinically to look at causes within an individual in terms of exposure, sequence of exposure, hierarchy of exposure events, genetic and other risk factors that initiate or provoke exposure to various stimuli. Examined individually, these pieces may comprise the jigsaw puzzle that is loss of immunological tolerance, development of autoimmune disease and progression of

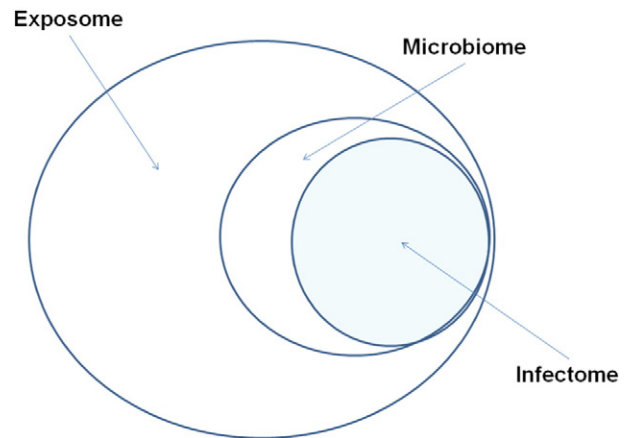


Fig. 1. From exposome to infectome via microbiome. “Exposome” describes all environmental factors which we are exposed to in a lifetime, both exogenous and endogenous, infectious and non-infectious. Environmental exposures are basically subdivided into infectious and non-infectious agents. The concept of “infectome” that we introduce, describes the part of the exposome which refers to the collection of an individual's exposures to infectious agents participating in the pathogenesis of autoimmune disease. The infectome can be considered a part of “microbiome”, the collection of the microbial products which the human body is exposed to at a given time.

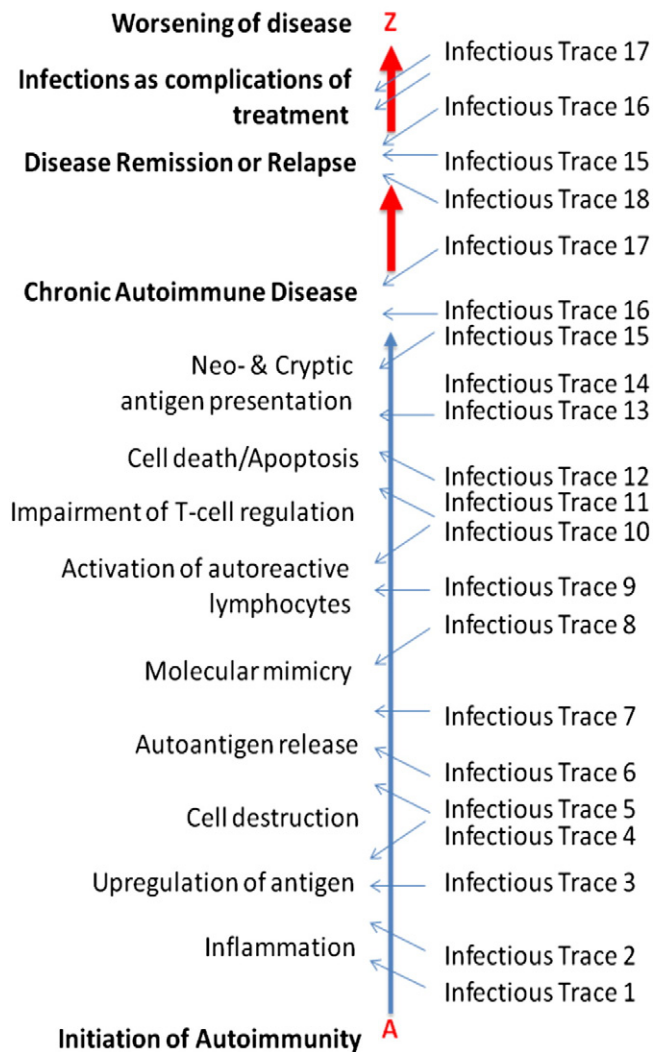
Infectome: From A to Z

Fig. 2. Tracing infectious triggers of autoimmunity: the infectome from A to Z. Infectious agents participating in a series of events critical for the initiation of autoimmunity and the development of autoimmune disease can be traced at various time points. The traces of these infectious agents may help us to understand the extent of their involvement in the loss of immunological breakdown and/or the maintenance of autoreactive immune responses leading to self destruction. Infections, at times different of those responsible for the chain reaction of events that led to autoimmune disease, may participate in the remission/relapse clinical patterns seen after the onset of the disease.

clinically over disease and its complications. Although great variability may be seen from patient to patient, the overall collation of well-defined disease-related exposures individually may provide a picture of disease development in that patient. The exposome provides a potential platform by which these exposures may be measured. It has been suggested that their by-products in an individual, such as DNA and protein modification, can serve as biomarkers that are potentially measurable [38] in the blood or body fluids of patients [3,36–38] via technologies such as liquid chromatography–tandem mass spectrometry (LC–MS/MS) [42], DNA adducts [43], functional measurements of antioxidant capacity, and breath analysis [34].

Rappaport suggests two methods in which the exposome may be measured: the 'bottom-up' method measures chemicals in air, water and the external environment, but is limited in that it does not take into account the actual uptake of those substances nor the internal environment of a subject [35]. As an alternative, the 'top-down' method measures biomarkers in the patient's blood, however, this method

does not indicate a potential source of the toxicant. It is likely that both complementary methods, in conjunction with refined questionnaires, will define the exposome in individuals, and larger populations [35–37]. Frequent sampling could demonstrate changes of these markers over time, especially during critical phases in the development of a disease [35]. A study by Pleil and colleagues [34] indicates that breath analysis for particular biomarkers is capable of identifying whether one has been exposed, what the dosage was, how rapidly the body is eliminating the toxicant, as well as possibly identifying the short and long term effects [34]. Finally, a recent study by Patel and colleagues [44] has provided evidence that the exposome can indeed be measured and characterised. That study conducted an environmental-wide association study (EWAS) on type 2 diabetes mellitus, where epidemiological data was comprehensively interpreted in a manner similar to GWAS [44]. Associations were found with 37 environmental factors, including organochlorine, pesticides, nutrients/vitamins, polychlorinated biphenyls, and dioxins [44]. Other studies have shown a potential crossreactivity between antigens within the pancreatic islet cells and cow's milk casein in siblings of type 1 diabetics [45], and multiple sclerosis's myelin oligodendrocyte glycoprotein with milk butyrophilin [46], highlighting the potential role for food antigens as triggers of autoimmune disease. Similar approaches can be used in future studies on the role of the exposome in the development of autoimmune diseases, especially those which develop in genetically susceptible individuals several years or decades following the initial insult.

One major obstacle to the complete analysis of all these triggers is their heterogeneity. It is unlikely to cover all such components by homogeneous technology platforms, as all human biomarker measurements are subject to inter- and intra-subject variance. This includes the composition of the received data into one model, which appears to be a Sisyphean task even in the age of ultra-fast computing. Therefore, the break-down of this multiversity of components into easier accessible, homogeneous realms of markers seems to be a reasonable next step. Techniques that can be utilised to detect infectious agents like multi-parametric immunoassays for antibodies of various isotypes specific to bacterial or viral antigens, urine and stool cultures and polymerase chain reaction (PCR) can be considered to be robust and integrateable [36]. Routine screening to detect the presence of an infectious causative agent is used clinically in a small scale and for individual microbial agents on an everyday basis [36].

3. Exposome, infectome and autoimmunity

Infectious and non-infectious environmental agents have long been considered important for the development of autoimmunity [4,47–51]. The list of non-infectious environmental factors which can trigger autoimmunity is vast, and includes tobacco smoke, pharmaceuticals, oestrogens, ultraviolet radiation, silica solvents, dietary components, heavy metals, dental materials, vaccines, and collagen/silicone implants (Table 1) [47,52–80]. Infectious triggers implicated in autoimmunity are also numerous and include bacteria, viruses, parasites and fungi. An infection burden corresponding to geo-epidemiological and serological evidence of the co-occurrence of anti-infectious agents has been observed and it is of interest that such an infection burden may differ from one autoimmune disease to another [4,81,82]. An analogous, autoantibody burden in non-autoimmune individuals during infection with various agents, further points towards the close relation of exposure to several infectious agents and the development of autoantibody reactivities [83]. From our point of view, it is helpful to define this subgroup of triggers as the "infectome" which is the part of the exposome referring to the collection of an individual's infectious exposures which are associated with disease, and in our case specific autoimmune diseases. It may also demonstrate the alteration in disease states, which are affected by alterations of the flora within the microbiome. Examples include

Table 1

Non-infectious environmental agents associated with the development of autoimmunity. This table provides examples of several non-infectious agents from a variety of sources, which have been associated with the development of autoimmune disease. AIH, autoimmune hepatitis; AiLD, autoimmune liver disease; COPD, chronic obstructive pulmonary disease; DM, dermatomyositis; MG, myasthenia gravis; rheumatoid arthritis; SLE, systemic lupus erythematosus; SSc, systemic sclerosis.

Non-infectious environmental triggers	Disease	Reference
<i>Occupational exposures</i>		
Silica	RA, SLE, SSc, glomerulonephritis, small vessel vasculitis.	[52–56]
Solvents	SLE, AIH	[54,55,57,58]
Pesticides	Autoimmune thyroid, RA, SLE, SSc	[59–65]
Ultraviolet radiation	SLE, RA, DM, PM, MS, type 1 diabetes mellitus	[54,66–69]
<i>Drugs</i>		
Allopurinol	Immune haemolytic anaemia	[70]
Captopril	Autoimmune thrombocytopenia	[71]
Chlorpromazine	Anti-phospholipid syndrome, haemolytic anaemia, SLE, AiLD	[72–78]
Estrogens	PBC, SLE, RA	[155–158,316–320]
Halothane	AIH	[77,321,322]
Iodine	Autoimmune thyroid	[62]
Penicillins	AiLD, immune haemolytic anaemia	[77,323]
Rifampicin	AIH, autoimmune thyroid, immune haemolytic anaemia	[324–326]
Tetracyclines	AIH, DM, SLE	[79,327–336]
<i>Miscellaneous</i>		
Vaccines	PBC, AIH, SLE, RA, MS, MG, DM, polyarteritis nodosa,	[279,337–349]
Cigarette smoke	PBC, COPD, RA, autoimmune thyroid	[155–158,350–355]
Collagen/silicone implants	SLE, Sjögren's, SSc	[353,356–362]

treatment with antibiotics [84,85], or exposure to toxic metals, silicone or other xenobiotics [30,86,87]. These exposures may very well alter the disease course or progression, by altering the flora within the microbiome, or by increasing the likelihood of infection with a disease causing infection. They may also increase the burden of oxidative stress. While the hygiene hypothesis underlines the protective role played by infections [88–92], clinical and experimental data in animal models implicate infections in the development of autoimmunity and autoimmune disease [4]. One of the best-studied examples of infection-induced autoimmunity is that of acute rheumatic fever presenting several weeks after infection with *Streptococcus pyogenes*. It is now well established that molecular mimicry between the bacterial M-protein and human lysoganglioside is responsible for the loss of immunological tolerance, and the development of autoimmunity in genetically susceptible individuals [93]. Other examples include the associations between *Helicobacter pylori* and autoimmune gastritis [94], as well as between *Trypanosoma cruzi* and Chagas' cardiomyopathy [95], and *Mycoplasma* with rheumatoid arthritis [96].

Although experimental models of autoimmune diseases and studies like those mentioned above provide data to support the role played by a single infectious trigger, the prevailing idea is that a multitude of infections from birth, in our term the infectome, contributes to the induction of autoimmunity [4]. In fact, Rolf Zinkernagel's hypothesis is that the increasing predisposition to autoimmune disease in the developing world may be due to the overall host–infection balance, beginning as early as the transfer of maternal antibodies via the placenta or via breast milk in the gastrointestinal tract. The question which then arises is how one can identify those specific infectious agents responsible for the initiation of an autoimmune disease. Studies in animals have provided some clues, but their resemblance to the human setting is poor in most cases.

Ideally, investigations have to be carried out in affected individuals. However, when work is performed on biological material obtained from patients with a given autoimmune disease, it is difficult to single out disease-triggering microbes. Undoubtedly, information from the collection of the microbial genes in a particular region (such as the gut or the oral cavity), also known as “microbiome”, can provide important hints for those potentially involved in the development of or protection against autoimmunity. However, this by itself cannot differentiate those with a pathogenic potential from the non-harmful ones, the latter representing the great majority. In fact, most of the isolated microbes have little to do with the initiation of immune-mediated inflammatory processes leading to a given disease. Moreover, induction of autoimmunity by viruses or bacteria is probably done by a ‘hit-and-run’ mechanism when the causative agent has been cleared from circulation by the time of diagnosis. Tracking down each individual's exposure to infectious agents as well as anti-microbial immune responses may be important for the establishment of a causative link between infection and autoimmunity.

Studies attempting to investigate the role of the “infectome” in the induction or remission/relapse state of autoimmune disease may need to focus on autoimmune diseases such as multiple sclerosis or rheumatoid arthritis, as patients with these diseases experience frequent remissions and relapses as well as fluctuations in disease activities that are poorly understood. Systemic lupus erythematosus, multiple sclerosis and rheumatoid arthritis are three of the best studied diseases so far for which specific viral agents have been considered to be important for the development of the disease and the breakdown of immunological tolerance [4]. As well, exposure to metals and various other chemical components have also been linked with these two conditions, and it would be interesting to see how these exposures alter the presence of infectious agents. It may be the case that exposure to certain xenobiotics increases susceptibility to disease causing microorganisms, or possibly eliminates organisms that are determined to be protective.

Arguably, the ideal clinical setting for the study of the role of the “infectome” in autoimmunity would involve typical autoimmune diseases with long prodromal stages [97]. Serial testing of biological material from individuals who progress from an asymptomatic sub-clinical phase to clinical disease would be optimal for the study of the “infectome”. Examples of this include systemic lupus erythematosus (SLE), multiple sclerosis (MS) and primary biliary cirrhosis (PBC) to name a few. All of those are characterised by highly specific antibodies that may appear years or even decades before the onset of symptoms [98–100]. These diseases also have long prodromal stages and their course highly varies among individuals. Relapse and remission states can be seen, but the reasons for this remain unknown. The characteristic features of these diseases and their reported pathogenic relationship with infectious agents make them ideal candidates for the study of the infectome.

4. SLE as an infectome model: known infections, stronger links

Systemic lupus erythematosus (SLE) is often characterised by a prodromal stage of antibody positivity (predominantly anti-Sm and anti-Ro) with no symptomatology [101,102]. A recent study has also found that a significant proportion of first degree relatives of SLE patients are positive for several autoantibodies, namely ANA (mostly anti-dsDNA), anti-Ro/SSA, and many other specificities [103–105]. In fact, ANA titre $\geq 1:160$, anti-dsDNA, anti-Ro/SSA and anti-chromatin may have a high predictive value for SLE diagnosis [103]. The reasons underlying the development of and/or the progression to clinical disease, as well as the mechanisms underlying disease flares, remain elusive [19]. However, several infectious agents have been implicated including EBV, cytomegalovirus (CMV) and parvovirus B19 [48,101, 102,104–110]. These features make SLE an ideal model for the

infectome, given the strong evidence for an infectious trigger, in addition to its unpredictable clinical course.

EBV appears to have an association with SLE, with variations and susceptibility based on demographical and genetic characteristics [111]. Harley and colleagues [101] propose a progression of EBV infection followed by Epstein–Barr virus nuclear antigen 1 (EBNA-1) antibody production, which predisposes to the development of cross-reactive autoantibodies, with progression to clinical SLE. Harley and James [102] indicated that the first lupus specific antibodies are directed against EBNA-1, which interestingly also binds lupus specific antigens such as Sm and Ro. As with many implicated viruses, the exact mechanisms in which they induce autoimmunity are not yet clear. However, one group which has indicated molecular mimicry as a mechanism with regard to EBV and SLE, notes similarities between the EBV peptide PPPGRRP and the PPPGMRPP peptide of Sm [109]. That same group tested 196 ANA positive adult SLE patients, and two age, race, and sex matched controls per patient, for evidence of previous infection with EBV, CMV, herpes simplex virus-1 (HSV-1), HSV-2, and varicella zoster virus (VZV) by ELISA [109]. Among the SLE patients, 195 out of 196 had previous EBV infection compared to 370 controls [109]. No differences were found in regard to the rate of infection with other viruses [109]. A study by Wang et al. [112] found that the EBV-encoded latent membrane protein 2A induced a heightened sensitivity to toll-like receptor (TLR) ligand stimulation, which increased proliferation of anti-Sm B-cells, and/or increased antibody secreting cell differentiation.

Parvovirus B19 has also been found to be associated with SLE, although the evidence is not as clear cut as that of EBV. The group of Bengtsson [106] tested 99 SLE patients and 99 age and sex matched controls for parvovirus IgG antibodies. No evidence of parvovirus B19 was found in SLE compared to controls, and in one analysis, the controls had a higher positivity than the SLE group [106]. As the symptoms of SLE and parvovirus B19 infection may be similar, a prospective study involving 42 patients with acute parvovirus B19 infection attempted to determine whether the symptoms persisted, and whether infection contributed to the development of SLE [110]. Clinical and laboratory investigations were performed at 1, 2, 6, 12, and 24 months from initial infection. Arthralgias persisted for 2–6 months in three female patients, for greater than two years in one female, but resolved in the remaining cases within 2 weeks [110]. One female with persistent arthralgia over two years was ANA positive and had hypercomplementaemia, but did not fulfil the diagnostic criteria for SLE or RA [110]. Despite the lack of evidence linking parvovirus B19 to the induction of SLE, one case report of a 26 year old female with SLE, who had a disease flare-up following parvovirus B19 infection, suggests that parvovirus B19 causes disease flares in SLE patients [107].

As with parvovirus B19, the evidence linking CMV to SLE is scarce. Hrycek and colleagues found that 100% of female SLE patients have evidence of CMV infection, compared to 75% of controls [108]. As well, another study had found that the CMV pp65 antigen triggers humoral immunity in SLE patients and autoimmune prone mice [113]. The lack of evidence linking particular viruses to SLE does not infer non-involvement, but reflects the difficulty of detecting infectious organisms, which may be transient, in these patients. It is likely that diagnosis of SLE and/or SLE flares occurs after an infection. The infectome serves as a model in such cases. At-risk patients, such as first degree relatives of SLE patients, who are found to be positive for autoantibodies, may be screened at regular intervals. Likewise, SLE patients may be screened, and changes in clinical course, such as flares, may be correlated with infection. Several other diseases with a well defined connection such as that of *H. pylori*-induced idiopathic thrombocytopenic purpura could be explored. These methods are useful in that they not only indicate the who and when, but also provide a more narrowed-down list of organisms to evaluate in regard to the mechanism in which they induce autoimmunity.

5. Multiple sclerosis as an infectome model for relapse remittance clinical states

Multiple sclerosis (MS) serves as another example in which the infectome model may be applied. MS is a chronic autoimmune neurological syndrome characterised by chronic inflammation, demyelination and gliosis in the central nervous system (CNS) [114,115]. It is characterised by periods of relapse and remission [114,115], the reasons for which are not understood. Current evidence suggests that the risk for acquiring MS is spread over a long period of time, and is not limited to childhood or early adult life [116–118].

The most prevalent form of the syndrome is the so called relapsing–remitting MS, characterised by flares whereby pre-existing symptoms become more severe, or new symptoms develop [119]. These flares are followed by phases of complete or partial recovery and their duration is highly variably among affected individuals. A considerable proportion of these patients acquire progressive disability with or without disease relapses (secondary progressive MS). Another form of MS is characterised by a stable progression of the disease with worsening of the symptomatology over the course of the disease. This form is also known as primary progressive MS (PPMS). Two further forms of the disease are also found with totally contradictory outcomes. The benign form of MS usually presents with minimal or mild progression of disability, and is clinically characterised by full recovery of sporadic sensory episodes [119]. On the other hand, the Marburg variant of MS is characterised by rapidly progressive disease which leads to accumulating disability and eventually to death. The mechanisms responsible for the development of clinically distinct disease phenotypes are poorly understood [119].

The pathological hallmark of MS is inflammatory lesions (areas of demyelination) in the CNS composed of mononuclear cell infiltrates in the perivascular spaces that develop into plaques [114,120]. These mononuclear cells are largely composed of T and B lymphocytes, plasma cells, macrophages and microglia [114,120]. Positive staining for IgG is found in the peripheral regions of the plaques [114,120]. Additionally, approximately 90% of MS patients show intrathecal IgG synthesis in the cerebrospinal fluid (CSF) [114].

Environmental and genetic components are clearly involved in the aetiology of MS, with geographical and twin data indicating a greater environmental component [114,115,121,122]. However, identical twins are 100 times more likely to develop MS if their co-twin is affected, whereas non-twin siblings are 20 times more likely [123,124]. Genetic and GWAS have implicated several alleles, with the strongest association being with HLA-DRB1*1501 [125]. Seven GWAS studies have been conducted, which included nearly 10,000 MS patients and 15,000 controls [115]. Implicated genes include IL7R, IL12RA, CLEC16, and CD226 [126,127]. Nonetheless, implicated genes have only demonstrated an odds ratio of less than 1.3 [115], and MS twins only demonstrate a 30% concordance [114], indicating more of an environmental influence in the disease pathogenesis.

Non-infectious and infectious components have been implicated to be involved in the development of MS. Within the group of non-infectious agents, vitamin D is of particular interest, not only in MS but also in several other autoimmune diseases [115,128–140]. Vitamin D appears to play a role in the modulation of pro-inflammatory pathways and T cell regulation [115]. Increased distance from the equator has been correlated with low vitamin D, and interestingly, MS rates increase as distance from the equator increases [114]. As well, populations with increased dietary vitamin D intake have lower rates of MS [115]. An Australian study found a decreased risk of a first demyelinating event in those with increased sun exposure, who also had increased levels of serum vitamin D [141]. This has also been found in other studies [142]. As well, the effect of month of birth on MS development was more apparent in familial MS groupings, suggesting an influence on prenatal vitamin D levels, as well as an interaction between genes and environment [115]. Like many

conditions, smoking has also been associated with MS development, and a recent meta-analysis overwhelmingly associated smoking with MS [143]. Gene–environment interactions have been suggested in regard to smoking and the presence of HLA-DRB1*15 with the absence of HLA-A*02 [144]. Smokers with this genetic combination appeared to have an increased risk of developing MS [144]. Whether this is due to the effect of the nitrosamines or the heavy metals in cigarette smoke is not clear. There is limited evidence for the role of heavy metals in some patients with MS [145]. Infectious agents investigated in regard to MS have included bacteria and viruses (Table 3) [114,115,121,122,146,147], and implicated viruses include EBV [114,115,121,122] and human herpes virus 6 (HHV6). The relapse–remittance pattern of the human herpes virus 6 (HHV6) is similar to the clinical pattern of MS [115] and HHV6 reservoirs have been found in CNS tissue [148,149], in addition to the CSF and serum of MS patients [150,151]. Molecular mimicry has been indicated, as sequence homology has been found between myelin basic protein and HHV6 encoded U24, and cross-reactive T cells responding to both protein types are found to be elevated in MS patients [152]. Varicella zoster virus, torque teno virus, retroviruses, coronaviruses and JC virus have also been implicated, but with limited evidence [114,115, 121,122]. Reactivity to several viral peptides was found [153], which has led to the speculation that continual exposure to a variety of viruses can lead to T cell expansion reactive against highly conserved proteins, including self-peptides.

An infectome for the above conditions (and others) may clarify questions regarding aetiology, but may also identify the cause of certain disease characteristics, such as variable presentation and progression among PBC patients, disease flares in patients with SLE, or relapse–remittance among MS patients. As such, these conditions serve as models for an infectomal analysis.

6. Primary biliary cirrhosis as an infectome model disease

PBC can be used as a model disease to investigate the role of the exposome, and indeed of the infectome [154], as A) it is an autoimmune disease and it is not as rare as many other such diseases, B) it has a long silent phase in which well-established biomarkers can be determined in high-risk populations, C) it presents with subgroups of patients who have differing disease progressions and/or concomitant other autoimmune diseases, D) it is not treated with immunosuppressive drugs that might deteriorate the immunological assessment of the infectome, and E) there is growing evidence in support of genetic, environmental, and infectious factors involved in the pathogenesis of the disease such as recurrent urinary tract infection (UTI), cigarette smoke, and oestrogen deficiency [155–163]. A plethora of experimental and clinical data clearly indicate that the disease

Table 2

Examples of infectious agents implicated in multiple sclerosis. Several organisms have been implicated in the pathogenesis and clinical course of multiple sclerosis (MS), the vast majority of which are viruses.

Viruses		Bacteria	
Epstein Barr virus	[363–378]	<i>Chlamydia pneumoniae</i>	[379]
Human herpes virus 6	[148–152,380,381]	<i>Borrelia burgdorferi</i>	[382]
Varicella zoster virus	[383–385]	<i>Mycobacterium tuberculosis</i>	[386]
Human cytomegalovirus	[387]		
Retroviruses	[388,389]		
Coronavirus	[390,391]		
Torque teno virus	[153]		
JC virus	[392,393]		
Rubella virus	[394]		
Parainfluenza virus I	[395]		
Measles virus	[396,397]		
Mumps virus	[398]		

Table 3

Examples of infectious agents implicated in primary biliary cirrhosis. This table provides several examples of infectious organisms which have been implicated in the pathogenesis of primary biliary cirrhosis (PBC). Strong evidence exists for some organisms such as *Escherichia coli*, while weaker evidence exists for others. This may be due to the lack of investigation into the prevalence of some organisms in PBC.

Bacteria	<i>Escherichia coli</i>	[176,178,265,267,399,400]
	<i>Chlamydia pneumoniae</i>	[263,401]
	<i>Mycobacterium gordonae</i>	[402–404]
	<i>Novosphingobium aromaticivorans</i>	[178,399]
	<i>Pseudomonas aeruginosa</i>	[257,405]
	<i>Lactobacillus delbrueckii</i> subsp <i>bulgaricus</i>	[259,279]
	<i>Yersinia enterocolitica</i>	[260,406]
	<i>Salmonella typhimurium</i>	[407]
	<i>Salmonella minnesota</i>	[408]
	<i>Haemophilus influenzae</i>	[257,260]
	<i>Streptococcus intermedius</i>	[409]
	<i>Paracoccus dentrificans</i>	[410]
	<i>Borrelia burgdorferi</i>	[411]
	<i>Propionibacterium acnes</i>	[412]
<i>Mycoplasma pneumoniae</i>	[413]	
<i>Mycoplasma gallisepticum</i>	[414]	
Viruses	Betaretrovirus	[272,276,415,416]
	Cytomegalovirus	[257]
	Epstein Barr virus	[417]
Parasites	Trypanosomes	[418,419]
	<i>Ascaridia galli</i>	[419]
Other	<i>Saccharomyces cerevisiae</i>	[420]

is indeed autoimmune in nature [98,159,164–184]. Infectious agents and xenobiotics mimicking or modifying the core epitopic region of PDC-E2, the major autoantigen in PBC, appear to induce immune-mediated destruction of the bile ducts. As PBC affects the liver, access to the affected organ for research purposes is possible at the time of diagnosis via liver biopsy, and over the course of the disease from early to advanced stages. This is not possible in diseases such as diabetes mellitus type I.

In line with the already discussed ideal situation of a long prodromal period, PBC has a long pre-clinical phase, characterised by an asymptomatic period with evidence of autoimmune markers in the form of autoantibodies, followed by biochemical evidence of liver damage and finally clinically-evident disease. The progression from asymptomatic to symptomatic PBC may take years or decades. Virtually, all patients with PBC have high-titre anti-mitochondrial antibodies (AMA) at the time of diagnosis [99,175,184–210]. Also, the presence of AMA is predictive of eventual disease development [211]. PBC patients with disease-specific anti-nuclear autoantibodies (ANA) appear to have more advanced disease and poorer prognosis [212–227]. Individuals at the highest risk of developing PBC are family members of PBC patients. [228–234]. The vast majority of patients with PBC experience concomitant autoimmune diseases such as *Sicca* syndrome, autoimmune thyroiditis, rheumatoid arthritis, autoimmune hepatitis and systemic sclerosis. The reasons behind the fast pace of progression seen in some patients with PBC are unknown, and attempts to predefine those individuals which will progress faster than others have been largely unsuccessful.

PBC does not appear to respond to immunosuppressive treatment, making the disease perfect to study the involvement of infectious agents over the course of the disease without the need to consider the effects of immunosuppressive therapy on relapse/remission states. PBC patients are instead treated with ursodeoxycholic acid (UDCA) at adequate doses of 13–15 mg/kg/day [235,236]. It is not clear as to whether UDCA has immunomodulatory properties or not.

A number of environmental factors have been implicated in PBC by 'bottom-up' approaches in various epidemiological studies examining associated risk factors [155–158] including numerous xenobiotics and infectious agents [159–162,237]. Animal models have further supported this notion [159,238–242]. The genetics of PBC appear to include HLA [243,244], non-HLA [245–251] and sex-linked

genes [252–254]. Particular allelic associations may be indirectly relevant to the pathogenesis of the disease, as they may influence the penetrance of infectious agents. For example HLA-DRB1*11 and HLA-DRB1*13 have a negative association with PBC, and are protective for several viruses that affect the liver, such as hepatitis B and C. The lack of HLA alleles protective for PBC, which are associated with resistance to specific pathogens, may lead to susceptibility to infection responsible for the induction of the disease [244,255]. These findings underline the urgent need to investigate the infectome in parallel with GWAS. An infection burden involving EBV, CMV and *Toxoplasma* has been recently reported in patients with PBC.

However, the list of pathogens involved in PBC is vast, with some such as *Escherichia coli*, having a relatively strong evidence base (Table 2) [164,182,252,256–279]. Molecular mimicry has been considered a likely mechanism that could account for the initiation of liver autoimmune diseases, including PBC [164,182,256–260,264,274,275,279–285].

Much as the exposome reflects the collation of all exposures, an infectome may reflect all those bacterial, viral, or parasitic exposures which may contribute to the development of an autoimmune disease.

7. How to study the infectome

The study of the infectome needs to be customised taking into account the disease to be investigated. The type of the sample to be collected largely depends on the disease under investigation.

A generic, step-wise approach of infectome analysis at presentation would be the following:

- 1) Determination of HLA class I and II is performed in all individuals under investigation. Ideally, this could be performed at birth, or at the baseline when the individual/patient presents for the first time in the clinic. This information is important in order to sub-group the individual into high or low risk.
- 2) Collect urine, oro-nasal swabs, saliva, faecal material, and blood (for isolation of plasma, serum, and peripheral blood mononuclear cells – PBMCs).
- 3) Regular follow-up (once yearly) with collection of samples;
- 4) Meticulous recording of clinical data and collection of samples are needed when anti-infective treatment is applied for incidental/casual infections at the pre-clinical stages of the disease, as this may affect the final outcome of the underlying processes.
- 5) Store samples until patient has laboratory and/or clinical features related to the disease;
- 6) Analysis of collected samples for infectious agents with a known association with the disease, and “out of the box” analysis using multiplex technology for other infectious agents (see section “How to screen?” below). The analysis will provide information regarding the infection burdens at various-time points;
- 7) Associations are analysed, providing evidence for known/unknown infectious agents in the development of symptomatic disease.
- 8) Continuous analysis over the course of the disease, as it may reveal a close link between a specific agent and the clinical phenotypes of the disease.

The study of the role of infectious agents in the development of an autoimmune disease, whether RA, SLE, MS or PBC, may reveal which agents are involved in the development of the disease as well as other concomitant autoimmune diseases. It is likely that the combination of particular genes and particular infectious exposures is responsible for the development of an autoimmune disease, with or without a particular concomitant autoimmune disease. In other words, there may be several infectomes for SLE, some of which define SLE with a particular concomitant autoimmune disease. This of course applies to other autoimmune diseases.

8. Lessons that can be learned from the microbiome and immunome projects

The recently described microbiome concept can be separated from that of the infectome. The microbiome defines the collection of microbial genes in a particular region, such as the gut, inguinal crease, oral cavity, or virtually any body site [286–295]. The microbiome may be reflective of the normal or pathological profile of organisms. For example, the microbiome of the gut has been analysed in healthy individuals [288–291,294], which identified three distinct profiles or clusters, known as enterotypes [287,293]. Similar studies have been performed for the oral cavity as well [286,295]. The microbiome may also be applied to a disease-affected body site, such as the gastrointestinal tract in children with irritable bowel syndrome (IBS) [292]. A study by Saulnier and colleagues [292] obtained 71 faecal samples from 22 children with IBS and 22 healthy children, all aged 7–12 years. These samples were analysed by 16S rRNA gene sequencing, which showed an elevated percentage of γ -proteobacteria in the gut flora of children with IBS, with *Haemophilus parainfluenza* being a prominent component [292].

So how does the infectome differ from this? First, the infectome relates to those infectious organisms which are associated with the disease in question, as opposed to a totality of all organisms, both pathological and non-pathological within the human microbiome. Second, the infectome reflects all sites of infection, as opposed to one body site usually represented by the microbiome. As well, microbiome studies to date have largely concentrated on bacterial species, whereas the infectome takes into account all pathological bacteria, viruses, parasites, and fungi. The infectome is not limited to the affected site or organ but includes biological fluids as well as sampling of various body-sites including the oral cavity. Some argue that the organisms of the human microbiome do not usually induce antigen specific systemic humoral and cellular immune responses provoking local or systemic inflammatory response, while others believe that these organisms may not be directly pathogenic, but create a dysbiosis of the gut flora and participate in the induction of autoimmunity. This is a key difference between the microbiome and the infectome as the former appreciates the direct or indirect effect of immune responses against infectious agents as pivotal for the initiation of autoaggression and immune-mediated, self-targeting pathology. For the infectome, monitoring of the microbial/host immunity is as important as the isolation of potentially harmful bacterial products, since the host/microbe immunological interaction is the likely cause of the self-destruction in the case of non-cytopathic viruses and microbes.

Although the infectome and microbiome are distinct entities, they may be used symbiotically to provide a micropathological profile (or profiles) of a particular disease. As well, the microbiome is essential to define what “normal” actually is. Recent microbiome studies have been able to provide an idea of what “normal” may be in the gut by performing metagenomic screens of bacterial populations in these regions [288–291,294]. For example, in the case of PBC it may be pertinent to understand the normal microbiome of the urinary bladder and vagina, as infections in these sites have been associated with PBC [155–158,296,297]. Other body sites may also need investigation, as inhalation or consumption of potentially pathogenic organisms has also been implicated [275,298]. All associated organisms located in all potential body-sites, would comprise the infectome. The establishment of what is normal as well as infectious may contribute to the understanding of the network of events or exposures which lead to disease development. It is possible that a series of exposures leads to increased susceptibility to further exposures, which could be defined by the infectomal model.

9. Who to screen?

Unlike GWAS and the microbiome, it is unlikely that population screening of infectomal components could be performed due to the

lack of integrated analysis platforms. It may be more reasonable to screen particular groups of individuals who are at risk of developing autoimmune disease, like family members or individuals with an HLA profile conferring risk for a given disease. Further monitoring may reveal whether additional infections play a role in the progression of the disease to a symptomatic stage. Additionally, particular exposome/infectome profiles may be found among patients with rapidly progressive diseases (as in a subset of PBC patients), or in relapse remittance states in MS, and flares in SLE. This approach may delineate which infectious agents are responsible for disease development and/or progression, as well as identifying those that are associated with rapid versus slow progression, remittance, and flares.

10. How to screen?

The establishment of the infectome would have to rely on the detection of microbiological materials in patients, favourably in ease-to-sample material like blood, urine or saliva. There are several methods by which this may be done, and some of those have already been used for research purposes. Serological detection of IgM, IgA and IgG antibody responses to microbes, viruses, and fungi is likely to be the most common, fastest, and most cost effective method and independent of the actual presence of a respective infectious agent. Monitoring for seroconversion of antibody responses and isotype class switching from IgM to IgG is imperative for the identification of newly acquired infectious triggers over time. To this end, the part of the immunome which relates to infections is an integral part of the infectome and its primary goal to identify microbial triggers of autoimmunity. Immunoassays for the determination of antibodies against most of them, e.g. ELISA, are broadly available, have a high degree of robustness and standardisation and are comparably cheap. Furthermore in some diagnostic scenarios, multi-parametric immunoassays, e.g. lineblots for the detection of antibodies against tick-borne diseases exist that facilitate the determination of antibodies against several or even multiple entities at the same time with a limited amount of sample. The highest complexity up-to-date can be reached by using protein microarrays that allow for the screening of several thousand antibody entities at the same time [299,300]. However, such platforms have so far only been used to screen for immune profiles of individuals, and small numbers of similar infectious agents and commercial solutions with a similar degree of standardisation as those containing human proteins are not available. A major breakthrough might result from the development of high-density peptidome libraries of relevant microbes and viruses similar to the concepts of peptidome libraries covering human proteins, e.g. T7-Pep library for the human peptidome [301].

A large German study used a PCR approach to detect a variety of organisms in archived liver tissues of PBC patients [302]. This approach has also been adopted in several other studies examining the role of mycobacteria in PBC [303–305], as well as in another group which used this method to detect beta-retroviral material in the liver and lymph node tissue from PBC patients [276,277]. Similar studies have been conducted in other autoimmune diseases. Multi-parametric detection of viral and bacterial genetic material in tissues may be another promising approach. In recent years, DNA sequencing technology has undergone an outstanding upgrade, and the so-called ‘high-throughput DNA sequencers’ can determine hundreds of megabases of DNA sequences per run, enabling the analysis of a broad range of infectious agents [306–309]. Massive, parallel sequencing might be the next step of this approach for that it is the most sensitive procedure available and allows for the detection of a multitude of infectious agents at the same time [310]. However, the considerable costs of this testing limit its wide use. Immunohistochemistry detecting several microbial agents in tissue samples can be applied [311], though it is likely that tissue based methods are not ideal in the establishment of the infectome, as they are time

consuming, and tissue of the affected organ may not be readily available from all patients. The analysis of blood, faecal material, urine, or saliva is more plausible, and can be used for screening, reflecting the ‘top-down’ approach suggested by other researchers [35]. As mentioned, multiplex PCR is a useful tool for evaluating the presence of microorganisms from a variety of sample types. Microbiome studies have also highlighted the use of 16S/18S rRNA gene sequencing, which allows for the mass-analysis of samples [290,292]. However, one drawback of this approach in comparison to immune profiles is the risk of missing the right time or site of sampling.

11. Where to screen?

The question also remains as to which body sites should be sampled. In addition to the affected organ, general or systemic infections should be an indication for sampling. This may include urine and stool samples in urinary and gastrointestinal tract infections respectively, or blood cultures in febrile patients. In addition to clinically overt infections, many patients with autoimmunity may have latent infections such as Lyme disease or *Mycoplasma*. Examples of this include hidden chronic low grade infection associated with dental treatments. In fact, the oral cavity is not commonly examined for infection in patients with autoimmune disease, and it would most likely be useful to examine and sample the oral cavity for such overt infections [312–314]. Accumulating evidence indicates that oral hygiene practices may induce alterations in the flora of the oral mucosa, which leads to a dysbiosis in the gut microbiome, and thereby contributes to the pathogenesis of IBD [312]. On the other hand, the increased frequency of dental problems in IBD patients may be due to alterations in oral flora. Complementing these findings, a study by the group of Helenius [314] indicates that patients with rheumatic conditions had various alterations in salivary flow and composition, and oral health. It remains to be seen whether the increased incidence of bacterial infections is common among all autoimmune diseases, and if so, which oral microorganisms are common among those conditions.

The proposed model of screening is not only useful in characterising the components which lead to autoimmune disease development/progression, but may also provide further evidence as to preventative measures. For example, if it is demonstrated that progression of an autoimmune disease is dependent upon infectious triggers, the proper use of antibiotic or even antiviral therapy may be initiated early on in the management of these patients. Antibiotic treatment for infectious agents associated with autoimmune disease may slow the progression of the disease in some individuals, and possibly prevent the development of the disease in individuals found to be at risk. If exposure to a particular microbe is proven to be critical in a particular autoimmune disease development, measures may be taken to reduce the exposure to this microbe, regardless of whether it is exogenous or endogenous. Such treatments can only be incorporated in the routine management of these diseases through consensus statements and authoritative position papers/guidelines; otherwise are potentially dangerous for the wellbeing of the patient [272,276,277,315].

12. Conclusion

Genetic and environmental components are involved in the pathogenesis of most diseases. The GWAS have contributed greatly to the understanding of the genetic basis of disease. The theory of the exposome complementing GWAS is now evolving. The complexity of the exposome may require that it be broken down to several sub-components such as the infectome, which reflects the characterisation and measurement of all infectious organisms that we are exposed to. The relationship of the exposome/infectome to the development of a disease, in individuals with particular genetic characteristics, may also help us characterise disease initiation and progression.

Given the growing knowledge of the genetic basis of disease, it may one day be possible to screen newborns for HLA and genetic characteristics which infer susceptibility to autoimmune disease, which may be done as routinely as the current newborn screening. Infants with a particular susceptibility profile can then be monitored closely for exposure to environmental triggers that may contribute to the development of a particular autoimmune disease. It is therefore essential that we establish not only the genetic basis of the disease (via GWAS), but also the environmental component of the disease, which must be established through an exposomal model. The covalent binding of xenobiotics to genomic DNA has been extensively explored and should be re-visited in this context in the light of more recent findings. As there is a clear delineation between organic and inorganic materials, the establishment of the infectome will represent the infectious component, comprised of all potential infections that initiate the disease, or alter the disease course. It is important that the subgroups of the exposome are considered together alongside the detailed investigation of each component part. It is likely that the methods used to establish and analyse the infectome will be simple modifications of technology already in use. It remains to be seen how this approach may be applied to other potential subgroups of the exposome, such as xenobiotics.

Take-home messages

- The infectious/host immunological interactions in autoimmunity are poorly understood.
- A direct link between infection and autoimmunity is difficult to obtain.
- Infectome refers to the collection of an individual's exposures to infectious agents.
- Infectome can be characterised with approaches employed for the “immunome” and “microbiome” projects.
- The study of the infectome can help us to delineate the triggers of autoimmunity.

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References

- [1] Gourley M, Miller FW. Mechanisms of disease: environmental factors in the pathogenesis of rheumatic disease. *Nat Clin Pract Rheumatol* 2007;3:172–80.
- [2] Lettre G, Rioux JD. Autoimmune diseases: insights from genome-wide association studies. *Hum Mol Genet* 2008;17:R116–21.
- [3] Rappaport SM, Smith MT. Epidemiology. Environment and disease risks. *Science* 2010;330:460–1.
- [4] Kivity S, Agmon-Levin N, Blank M, Shoenfeld Y. Infections and autoimmunity—friends or foes? *Trends Immunol* 2009;30:409–14.
- [5] Jacobson DL, Gange SJ, Rose NR, Graham NM. Epidemiology and estimated population burden of selected autoimmune diseases in the United States. *Clin Immunol Immunopathol* 1997;84:223–43.
- [6] Blank M, Gershwin ME. Autoimmunity: from the mosaic to the kaleidoscope. *J Autoimmun* 2008;30:1–4.
- [7] Brickman CM, Shoenfeld Y. The mosaic of autoimmunity. *Scand J Clin Lab Invest Suppl* 2001;235:3–15.
- [8] Rahamim-Cohen D, Shoenfeld Y. The mosaic of autoimmunity. A classical case of inhalation of a polyclonal activating factor in a genetically and hormonally susceptible patient leading to multiple autoimmune diseases. *Isr Med Assoc J* 2001;3:381–2.
- [9] Shepshelovich D, Shoenfeld Y. Prediction and prevention of autoimmune diseases: additional aspects of the mosaic of autoimmunity. *Lupus* 2006;15:183–90.
- [10] Shoenfeld Y, Blank M, Abu-Shakra M, Amital H, Barzilai O, Berkun Y, et al. The mosaic of autoimmunity: prediction, autoantibodies, and therapy in autoimmune diseases—2008. *Isr Med Assoc J* 2008;10:13–9.

- [11] Shoenfeld Y, Gilburd B, Abu-Shakra M, Amital H, Barzilai O, Berkun Y, et al. The mosaic of autoimmunity: genetic factors involved in autoimmune diseases—2008. *Isr Med Assoc J* 2008;10:3–7.
- [12] Asherson RA, Gunter K, Daya D, Shoenfeld Y. Multiple autoimmune diseases in a young woman: tuberculosis and splenectomy as possible triggering factors? Another example of the “mosaic” of autoimmunity. *J Rheumatol* 2008;35:1224–6.
- [13] de Carvalho JF, Pereira RM, Shoenfeld Y. The mosaic of autoimmunity: the role of environmental factors. *Front Biosci (Elite Ed)* 2009;1:501–9.
- [14] Costenbader KH, Gay S, Alarcon-Riquelme ME, Iaccarino L, Doria A. Genes, epigenetic regulation and environmental factors: which is the most relevant in developing autoimmune diseases? *Autoimmun Rev* 2012;11:604–9.
- [15] Mavropoulos A, Orfanidou T, Liaskos C, Smyk DS, Billinis C, Blank M, et al. p38 mitogen-activated protein kinase (p38 MAPK)-mediated autoimmunity: lessons to learn from ANCA vasculitis and pemphigus vulgaris. *Autoimmun Rev* Nov 30 2012, <http://dx.doi.org/10.1016/j.autrev.2012.10.019>. [pii: S1568-9972(12)00277-7. Electronic publication ahead of print PMID:23207287].
- [16] Karlson EW, Deane K. Environmental and gene–environment interactions and risk of rheumatoid arthritis. *Rheum Dis Clin North Am* 2012;38:405–26.
- [17] Villeda SA, Wyss-Coray T. The circulatory systemic environment as a modulator of neurogenesis and brain aging. *Autoimmun Rev* Nov 29 2012, <http://dx.doi.org/10.1016/j.autrev.2012.10.014> [pii: S1568-9972(12)00272-8. Electronic publication ahead of print PMID: 23201925].
- [18] Simon KC, van der Mei IA, Munger KL, Ponsonby A, Dickinson J, Dwyer T, et al. Combined effects of smoking, anti-EBNA antibodies, and HLA-DRB1*1501 on multiple sclerosis risk. *Neurology* 2010;74:1365–71.
- [19] Gatto M, Zen M, Ghirardello A, Bettio S, Bassi N, Iaccarino L, et al. Emerging and critical issues in the pathogenesis of lupus. *Autoimmun Rev* Sep 18 2012, <http://dx.doi.org/10.1016/j.autrev.2012.09.003> [pii: S1568-9972(12)00213-3. Electronic publication ahead of print PMID: 23000207].
- [20] Selmi C, Leung PS, Sherr DH, Diaz M, Nyland JF, Monestier M, et al. Mechanisms of environmental influence on human autoimmunity: a national institute of environmental health sciences expert panel workshop. *J Autoimmun* 2012;39:272–84.
- [21] Bogdanos DP, Smith H, Ma Y, Baum H, Mieli-Vergani G, Vergani D. A study of molecular mimicry and immunological cross-reactivity between hepatitis B surface antigen and myelin mimics. *Clin Dev Immunol* 2005;12:217–24.
- [22] Smyk DS, Rigopoulou EI, Muratori L, Burroughs AK, Bogdanos DP. Smoking as a risk factor for autoimmune liver disease: what we can learn from primary biliary cirrhosis. *Ann Hepatol* 2012;11:7–14.
- [23] Israeli E, Agmon-Levin N, Blank M, Shoenfeld Y. Adjuvants and autoimmunity. *Lupus* 2009;18:1217–25.
- [24] Nancy AL, Shoenfeld Y. Chronic fatigue syndrome with autoantibodies—the result of an augmented adjuvant effect of hepatitis-B vaccine and silicone implant. *Autoimmun Rev* 2008;8:52–5.
- [25] Perricone C, Agmon-Levin N, Valesini G, Shoenfeld Y. Vaccination in patients with chronic or autoimmune rheumatic diseases: the ego, the id and the super-ego. *Joint Bone Spine* 2012;79:1–3.
- [26] Rosenblum H, Shoenfeld Y, Amital H. The common immunogenic etiology of chronic fatigue syndrome: from infections to vaccines via adjuvants to the ASIA syndrome. *Infect Dis Clin North Am* 2011;25:851–63.
- [27] Shoenfeld Y, Agmon-Levin N. ‘ASIA’ — autoimmune/inflammatory syndrome induced by adjuvants. *J Autoimmun* 2011;36:4–8.
- [28] Agmon-Levin N, Hughes GR, Shoenfeld Y. The spectrum of ASIA: ‘Autoimmune (Auto-inflammatory) Syndrome induced by Adjuvants’. *Lupus* 2012;21:118–20.
- [29] Katzav A, Kivity S, Blank M, Shoenfeld Y, Chapman J. Adjuvant immunization induces high levels of pathogenic antiphospholipid antibodies in genetically prone mice: another facet of the ASIA syndrome. *Lupus* 2012;21:210–6.
- [30] Kivity S, Katz M, Langevitz P, Eshed I, Olchovski D, Barzilai A. Autoimmune syndrome induced by adjuvants (ASIA) in the Middle East: morphea following silicone implantation. *Lupus* 2012;21:136–9.
- [31] Zafir Y, Agmon-Levin N, Paz Z, Shilton T, Shoenfeld Y. Autoimmunity following hepatitis B vaccine as part of the spectrum of ‘Autoimmune (Auto-inflammatory) Syndrome induced by Adjuvants’ (ASIA): analysis of 93 cases. *Lupus* 2012;21:146–52.
- [32] Blank M, Israeli E, Shoenfeld Y. When APS (Hughes syndrome) met the autoimmune/inflammatory syndrome induced by adjuvants (ASIA). *Lupus* 2012;21:711–4.
- [33] Arlt VM, Schwerdtle T. UKEMS/Dutch EMS-sponsored workshop on biomarkers of exposure and oxidative DNA damage & 7th GUM-32P-postlabelling workshop, University of Munster, Munster, Germany, 28–29 March 2011. *Mutagenesis* Sep 2011;26(5): 679–85, <http://dx.doi.org/10.1093/mutage/ger036> [Electronic publication ahead of print 2011 Jun 21].
- [34] Pleil JD, Stiegel MA, Sobus JR, Liu Q, Madden MC. Observing the human exposome as reflected in breath biomarkers: heat map data interpretation for environmental and intelligence research. *J Breath Res* 2011;5:037104.
- [35] Rappaport SM. Implications of the exposome for exposure science. *J Expo Sci Environ Epidemiol* 2011;21:5–9.
- [36] Wild CP. Complementing the genome with an “exposome”: the outstanding challenge of environmental exposure measurement in molecular epidemiology. *Cancer Epidemiol Biomarkers Prev* 2005;14:1847–50.
- [37] Wild CP. Environmental exposure measurement in cancer epidemiology. *Mutagenesis* 2009;24:117–25.
- [38] Smith MT, Zhang L, McHale CM, Skibola CF, Rappaport SM. Benzene, the exposome and future investigations of leukemia etiology. *Chem Biol Interact* 2011;192:155–9.
- [39] Collin M, Shannon O, Björck L. IgG glycan hydrolysis by a bacterial enzyme as a therapy against autoimmune conditions. *Proc Natl Acad Sci U S A* 2008;105: 4265–70.

- [40] Allhorn M, Olin AI, Nimmerjahn F, Collin M. Human IgG/Fc gamma R interactions are modulated by streptococcal IgG glycan hydrolysis. *PLoS One* 2008;3:e1413.
- [41] McCulloch J, Zhang YW, Dawson M, Harkiss GD, Peterhans E, Vogt HR, et al. Glycosylation of IgG during potentially arthritogenic lentiviral infections. *Rheumatol Int* 1995;14:243–8.
- [42] Polacco BJ, Purvine SO, Zink EM, Lavoie SP, Lipton MS, Summers AO, et al. Discovering mercury protein modifications in whole proteomes using natural isotope distributions observed in liquid chromatography–tandem mass spectrometry. *Molecular & Cellular Proteomics* 2011 Aug;10(8) [M110.004853].
- [43] McLaren Howard J. The detection of DNA adducts (risk factors for DNA damage). A method for genomic DNA, the results and some effects of nutritional intervention. *J Nutr Environ Med* 2002;12:19–31 [ΔEN TO BPHKA ΣTO PUBMED].
- [44] Patel CJ, Bhattacharya J, Butte AJ. An Environment-Wide Association Study (EWAS) on type 2 diabetes mellitus. *PLoS One* 2010;5:e10746.
- [45] Karges W, Hammond-McKibben D, Cheung RK, Visconti M, Shibuya N, Kemp D, et al. Immunological aspects of nutritional diabetes prevention in NOD mice: a pilot study for the cow's milk-based IDDM prevention trial. *Diabetes* 1997;46:557–64.
- [46] Guggenmos J, Schubart AS, Ogg S, Andersson M, Olsson T, Mather IH, et al. Antibody cross-reactivity between myelin oligodendrocyte glycoprotein and the milk protein butyrophilin in multiple sclerosis. *J Immunol* 2004;172:661–8.
- [47] Molina V, Shoenfeld Y. Infection, vaccines and other environmental triggers of autoimmunity. *Autoimmunity* 2005;38:235–45.
- [48] Pordeus V, Szyper-Kravitz M, Levy RA, Vaz NM, Shoenfeld Y. Infections and autoimmunity: a panorama. *Clin Rev Allergy Immunol* 2008;34:283–99.
- [49] Doria A, Sarzi-Puttini P, Shoenfeld Y. Infections, rheumatism and autoimmunity: the conflicting relationship between humans and their environment. *Autoimmun Rev* 2008;8:1–4.
- [50] Tozzoli R, Barzilai O, Ram M, Villalta D, Bizzaro N, Sherer Y, et al. Infections and autoimmune thyroid diseases: parallel detection of antibodies against pathogens with proteomic technology. *Autoimmun Rev* 2008;8:112–5.
- [51] Corthesy B. Role of secretory IgA in infection and maintenance of homeostasis. *Autoimmun Rev* Nov 29 2012, <http://dx.doi.org/10.1016/j.autrev.2012.10.012> [pii: S1568-9972(12)00270-4. Electronic publication ahead of print PMID: 23201924].
- [52] Brown JM, Pfau JC, Pershouse MA, Holian A. Silica, apoptosis, and autoimmunity. *J Immunotoxicol* 2005;1:177–87.
- [53] Otsuki T, Hayashi H, Nishimura Y, Hyodo F, Maeda M, Kumagai N, et al. Dysregulation of autoimmunity caused by silica exposure and alteration of Fas-mediated apoptosis in T lymphocytes derived from silicosis patients. *Int J Immunopathol Pharmacol* 2011;24:115–65.
- [54] Cooper GS, Wither J, Bernatsky S, Claudio JO, Clarke A, Rioux JD, et al. Occupational and environmental exposures and risk of systemic lupus erythematosus: silica, sunlight, solvents. *Rheumatology* 2010;49:2172–80.
- [55] Finckh A, Cooper GS, Chibnik LB, Costenbader KH, Watts J, Pankey H, et al. Occupational silica and solvent exposures and risk of systemic lupus erythematosus in urban women. *Arthritis Rheum* 2006;54:3648–54.
- [56] Parks CG, Cooper GS. Occupational exposures and risk of systemic lupus erythematosus: a review of the evidence and exposure assessment methods in population- and clinic-based studies. *Lupus* 2006;15:728–36.
- [57] Griffin JM, Gilbert KM, Lamps LW, Pumford NR. CD4(+) T-cell activation and induction of autoimmune hepatitis following trichloroethylene treatment in MRL +/- mice. *Toxicol Sci* 2000;57:345–52.
- [58] Khan MF, Kaphalia BS, Prabhakar BS, Kanz MF, Ansari GA. Trichloroethene-induced autoimmune response in female MRL +/- mice. *Toxicol Appl Pharmacol* 1995;134:155–60.
- [59] Duntas LH. Environmental factors and thyroid autoimmunity. *Ann Endocrinol (Paris)* 2011;72:108–13.
- [60] Parks CG, Walitt BT, Pettinger M, Chen JC, de Roos AJ, Hunt J, et al. Insecticide use and risk of rheumatoid arthritis and systemic lupus erythematosus in the Women's Health Initiative Observational Study. *Arthritis Care Res (Hoboken)* 2011;63:184–94.
- [61] Fortes C. Lupus erythematosus. Are residential insecticides exposure the missing link? *Med Hypotheses* 2010;75:590–3.
- [62] Burek CL, Talor MV. Environmental triggers of autoimmune thyroiditis. *J Autoimmun* 2009;33:183–9.
- [63] Gold LS, Ward MH, Dosemeci M, De Roos AJ. Systemic autoimmune disease mortality and occupational exposures. *Arthritis Rheum* 2007;56:3189–201.
- [64] Sobel ES, Gianini J, Butfiloski EJ, Croker BP, Schiffenbauer J, Roberts SM. Acceleration of autoimmunity by organochlorine pesticides in (NZB×NZW)F1 mice. *Environ Health Perspect* 2005;113:323–8.
- [65] Mayes MD. Epidemiologic studies of environmental agents and systemic autoimmune diseases. *Environ Health Perspect* 1999;107(Suppl. 5):743–8.
- [66] Artukovic M, Ilic M, Kustelega J, Artukovic IN, Kaliterna DM. Influence of UV radiation on immunological system and occurrence of autoimmune diseases. *Coll Antropol* 2010;34(Suppl. 2):175–8.
- [67] Handunnetthi L, Ramagopalan SV. UV radiation, vitamin D, and multiple sclerosis. *Proc Natl Acad Sci U S A* 2010;107:E130 [author reply E1].
- [68] Prieto S, Grau JM. The geoepidemiology of autoimmune muscle disease. *Autoimmun Rev* 2010;9:A330–4.
- [69] Kuhn A, Beissert S. Photosensitivity in lupus erythematosus. *Autoimmunity* 2005;38:519–29.
- [70] Tungor G, Balkan C, Arikian K, Kavakli K, Aydogdu S. Immune haemolytic anaemia induced by allopurinol after liver transplantation. *Acta Paediatr* 2006;95:762–3.
- [71] Pujol M, Duran-Suarez JR, Martin Vega C, Sanchez C, Tovar JL, Valles M. Autoimmune thrombocytopenia in three patients treated with captopril. *Vox Sang* 1989;57:218.
- [72] Gharavi AE, Sammaritano LR, Wen J, Miyawaki N, Morse JH, Zarrabi MH, et al. Characteristics of human immunodeficiency virus and chlorpromazine induced antiphospholipid antibodies: effect of beta 2 glycoprotein I on binding to phospholipid. *J Rheumatol* 1994;21:94–9.
- [73] Canoso RT, de Oliveira RM. Chlorpromazine-induced anticardiolipin antibodies and lupus anticoagulant: absence of thrombosis. *Am J Hematol* 1988;27:272–5.
- [74] Stein PB, Inwood MJ. Hemolytic anemia associated with chlorpromazine therapy. *Can J Psychiatry* 1980;25:659–61.
- [75] Hadnagy C. Letter: Coombs-positive haemolytic anaemia provoked by chlorpromazine. *Lancet* 1976;1:423.
- [76] Berglund S, Gottfries CG, Gottfries I, Stormby K. Chlorpromazine-induced antinuclear factors. *Acta Med Scand* 1970;187:67–74.
- [77] Liu ZX, Kaplowitz N. Immune-mediated drug-induced liver disease. *Clin Liver Dis* 2002;6:755–74.
- [78] Yung RL, Richardson BC. Drug-induced lupus. *Rheum Dis Clin North Am* 1994;20:61–86.
- [79] Czaja AJ. Drug-induced autoimmune-like hepatitis. *Dig Dis Sci* 2011;56:958–76.
- [80] Chighizola C, Meroni PL. The role of environmental estrogens and autoimmunity. *Autoimmun Rev* 2012;11:A493–501.
- [81] Shapira Y, Agmon-Levin N, Renaudineau Y, Porat-Katz BS, Barzilai O, Ram M, et al. Serum markers of infections in patients with primary biliary cirrhosis: evidence of infection burden. *Exp Mol Pathol* 2012;93:386–90.
- [82] Shapira Y, Agmon-Levin N, Shoenfeld Y. Defining and analyzing geo-epidemiology and human autoimmunity. *J Autoimmun* 2010;34:J168–77.
- [83] Berlin T, Zandman-Goddard G, Blank M, Matthias T, Pfeiffer S, Weis I, et al. Autoantibodies in nonautoimmune individuals during infections. *Ann N Y Acad Sci* 2007;1108:584–93.
- [84] Saad R, Rizkallah MR, Aziz RK. Gut pharmacomicrobiomics: the tip of an iceberg of complex interactions between drugs and gut-associated microbes. *Gut Pathog* 2012;4:16.
- [85] Buccigrossi V, Nicastro E, Guarino A. Functions of intestinal microflora in children. *Curr Opin Gastroenterol* 2013;29:31–8.
- [86] Gielda LM, DiRita VJ. Zinc competition among the intestinal microbiota. *MBio* 2012;3:e00171–12.
- [87] Lidar M, Agmon-Levin N, Langevitz P, Shoenfeld Y. Silicone and scleroderma revisited. *Lupus* 2012;21:121–7.
- [88] Shoenfeld Y, Toubi E. Protective autoantibodies: role in homeostasis, clinical importance, and therapeutic potential. *Arthritis Rheum* 2005;52:2599–606.
- [89] Toubi E, Shoenfeld Y. Predictive and protective autoimmunity in cardiovascular diseases: is vaccination therapy a reality? *Lupus* 2005;14:665–9.
- [90] Ram M, Anaya JM, Barzilai O, Izhaky D, Porat Katz BS, Blank M, et al. The putative protective role of hepatitis B virus (HBV) infection from autoimmune disorders. *Autoimmun Rev* 2008;7:621–5.
- [91] Meroni PL, Shoenfeld Y. Predictive, protective, orphan autoantibodies: the example of anti-phospholipid antibodies. *Autoimmun Rev* 2008;7:585–7.
- [92] Plot L, Amital H, Barzilai O, Ram M, Nicola B, Shoenfeld Y. Infections may have a protective role in the etiopathogenesis of celiac disease. *Ann N Y Acad Sci* 2009;1173:670–4.
- [93] Guilherme L, Oshiro SE, Fae KC, Cunha-Neto E, Renesto G, Goldberg AC, et al. T-cell reactivity against streptococcal antigens in the periphery mirrors reactivity of heart-infiltrating T lymphocytes in rheumatic heart disease patients. *Infect Immun* 2001;69:5345–51.
- [94] Amedei A, Bergman MP, Appelmelk BJ, Azzurri A, Benagiano M, Tamburini C, et al. Molecular mimicry between *Helicobacter pylori* antigens and H+, K+-adenosine triphosphatase in human gastric autoimmunity. *J Exp Med* 2003;198:1147–56.
- [95] Cunha-Neto E, Coelho V, Guilherme L, Fiorelli A, Stolf N, Kalil J. Autoimmunity in Chagas' disease. Identification of cardiac myosin-B13 *Trypanosoma cruzi* protein crossreactive T cell clones in heart lesions of a chronic Chagas' cardiomyopathy patient. *J Clin Invest* 1996;98:1709–12.
- [96] da Rocha Sobrinho HM, Jarach R, da Silva NA, Shio MT, Jancsar S, Timenetsky J, et al. Mycoplasma lipid-associated membrane proteins and *Mycoplasma arthritidis* mitogen recognition by serum antibodies from patients with rheumatoid arthritis. *Rheumatol Int* 2011;31:951–7.
- [97] Tobon JC, Pers JO, Canas CA, Rojas-Villarraga A, Youinou P, Anaya JM. Are autoimmune diseases predictable? *Autoimmun Rev* 2012;11:259–66.
- [98] Jones DE. Pathogenesis of primary biliary cirrhosis. *Gut* 2007;56:1615–24.
- [99] Kaplan MM, Gershwin ME. Primary biliary cirrhosis. *N Engl J Med* 2005;353:1261–73.
- [100] Arbuckle MR, McClain MT, Rubertone MV, Scofield RH, Dennis GJ, James JA, et al. Development of autoantibodies before the clinical onset of systemic lupus erythematosus. *N Engl J Med* 2003;349:1526–33.
- [101] Harley JB, Harley IT, Guthridge JM, James JA. The curiously suspicious: a role for Epstein–Barr virus in lupus. *Lupus* 2006;15:768–77.
- [102] Harley JB, James JA. Epstein–Barr virus infection induces lupus autoimmunity. *Bull NYU Hosp Jt Dis* 2006;64:45–50.
- [103] Navarra SV, Ishimori MI, Uy EA, Hamijoyo L, Sama J, James JA, et al. Studies of Filipino patients with systemic lupus erythematosus: autoantibody profile of first-degree relatives. *Lupus* 2011;20:537–43.
- [104] Zandman-Goddard G, Berkun Y, Barzilai O, Boaz M, Blank M, Ram M, et al. Exposure to Epstein–Barr virus infection is associated with mild systemic lupus erythematosus disease. *Ann N Y Acad Sci* 2009;1173:658–63.
- [105] Barzilai O, Sherer Y, Ram M, Izhaky D, Anaya JM, Shoenfeld Y. Epstein–Barr virus and cytomegalovirus in autoimmune diseases: are they truly notorious? A preliminary report. *Ann N Y Acad Sci* 2007;1108:567–77.
- [106] Bengtsson A, Widell A, Elmstahl S, Sturfelt G. No serological indications that systemic lupus erythematosus is linked with exposure to human parvovirus B19. *Ann Rheum Dis* 2000;59:64–6.

- [107] Hemauer A, Beckenlehner K, Wolf H, Lang B, Modrow S. Acute parvovirus B19 infection in connection with a flare of systemic lupus erythematosus in a female patient. *J Clin Virol* 1999;14:73–7.
- [108] Hrycek A, Kusmierz D, Mazurek U, Wilczok T. Human cytomegalovirus in patients with systemic lupus erythematosus. *Autoimmunity* 2005;38:487–91.
- [109] James JA, Neas BR, Moser KL, Hall T, Bruner GR, Sestak AL, et al. Systemic lupus erythematosus in adults is associated with previous Epstein–Barr virus exposure. *Arthritis Rheum* 2001;44:1122–6.
- [110] Seishima M, Oyama Z, Yamamura M. Two-year follow-up study after human parvovirus B19 infection. *Dermatology* 2003;206:192–6.
- [111] Parks CG, Cooper GS, Hudson LL, Dooley MA, Treadwell EL, St Clair EW, et al. Association of Epstein–Barr virus with systemic lupus erythematosus: effect modification by race, age, and cytotoxic T lymphocyte-associated antigen 4 genotype. *Arthritis Rheum* 2005;52:1148–59.
- [112] Wang H, Nicholas MW, Conway KL, Sen P, Diz R, Tisch RM, et al. EBV latent membrane protein 2A induces autoreactive B cell activation and TLR hypersensitivity. *J Immunol* 2006;177:2793–802.
- [113] Chang M, Pan MR, Chen DY, Lan JL. Human cytomegalovirus pp 65 lower matrix protein: a humoral immunogen for systemic lupus erythematosus patients and autoantibody accelerator for NZB/W F1 mice. *Clin Exp Immunol* 2006;143:167–79.
- [114] Gildden DH. Infectious causes of multiple sclerosis. *Lancet Neurol* 2005;4:195–202.
- [115] Kakalacheva K, Lunemann JD. Environmental triggers of multiple sclerosis. *FEBS Lett* 2011;23:3724–9 [PMID: 21486562].
- [116] Hammond SR, English DR, McLeod JG. The age-range of risk of developing multiple sclerosis: evidence from a migrant population in Australia. *Brain* 2000;123(Pt 5):968–74.
- [117] Kurtzke JF, Delasnerie-Laupretre N, Wallin MT. Multiple sclerosis in North African migrants to France. *Acta Neurol Scand* 1998;98:302–9.
- [118] Kurtzke JF, Hyllested K. Multiple sclerosis in the Faroe Islands: I. Clinical and epidemiological features. *Ann Neurol* 1979;5:6–21.
- [119] Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. *N Engl J Med* 2000;343:938–52.
- [120] Frohman EM, Racke MK, Raine CS. Multiple sclerosis—the plaque and its pathogenesis. *N Engl J Med* 2006;354:942–55.
- [121] Giovannoni G, Cutter GR, Lunemann J, Martin R, Munz C, Sriram S, et al. Infectious causes of multiple sclerosis. *Lancet Neurol* 2006;5:887–94.
- [122] Giovannoni G, Ebers G. Multiple sclerosis: the environment and causation. *Curr Opin Neurol* 2007;20:261–8.
- [123] Ebers GC, Sadovnick AD, Risch NJ. A genetic basis for familial aggregation in multiple sclerosis. Canadian Collaborative Study Group. *Nature* 1995;377:150–1.
- [124] Sadovnick AD, Ebers GC, Dyment DA, Risch NJ. Evidence for genetic basis of multiple sclerosis. The Canadian Collaborative Study Group. *Lancet* 1996;347:1728–30.
- [125] Barcellos LF, Sawcer S, Ramsay PP, Baranzini SE, Thomson G, Briggs F, et al. Heterogeneity at the HLA-DRB1 locus and risk for multiple sclerosis. *Hum Mol Genet* 2006;15:2813–24.
- [126] Hafler DA, Compston A, Sawcer S, Lander ES, Daly MJ, De Jager PL, et al. Risk alleles for multiple sclerosis identified by a genomewide study. *N Engl J Med* 2007;357:851–62.
- [127] Qu HQ, Bradfield JP, Belisle A, Grant SF, Hakonarson H, Polychronakos C. The type 1 diabetes association of the IL2RA locus. *Genes Immun* 2009;10(Suppl. 1):S42–8.
- [128] Arnson Y, Amital H, Shoenfeld Y. Vitamin D and autoimmunity: new aetiological and therapeutic considerations. *Ann Rheum Dis* 2007;66:1137–42.
- [129] Carvalho JF, Blank M, Kiss E, Tarr T, Amital H, Shoenfeld Y. Anti-vitamin D, vitamin D in SLE: preliminary results. *Ann N Y Acad Sci* 2007;1109:550–7.
- [130] Shoenfeld N, Amital H, Shoenfeld Y. The effect of melanin and vitamin D synthesis on the incidence of autoimmune disease. *Nat Clin Pract Rheumatol* 2009;5:99–105.
- [131] Shapira Y, Agmon-Levin N, Shoenfeld Y. Mycobacterium tuberculosis, autoimmunity, and vitamin D. *Clin Rev Allergy Immunol* 2010;38:169–77.
- [132] Amital H, Szekanez Z, Szucs G, Danko K, Nagy E, Csepány T, et al. Serum concentrations of 25-OH vitamin D in patients with systemic lupus erythematosus (SLE) are inversely related to disease activity: is it time to routinely supplement patients with SLE with vitamin D? *Ann Rheum Dis* 2010;69:1155–7.
- [133] Toubi E, Shoenfeld Y. The role of vitamin D in regulating immune responses. *Isr Med Assoc J* 2010;12:174–5.
- [134] Souberbielle JC, Body JJ, Lappe JM, Plebani M, Shoenfeld Y, Wang TJ, et al. Vitamin D and musculoskeletal health, cardiovascular disease, autoimmunity and cancer: recommendations for clinical practice. *Autoimmun Rev* 2010;9:709–15.
- [135] Agmon-Levin N, Blank M, Zandman-Goddard G, Orbach H, Meroni PL, Tincani A, et al. Vitamin D: an instrumental factor in the anti-phospholipid syndrome by inhibition of tissue factor expression. *Ann Rheum Dis* 2011;70:145–50.
- [136] Oren Y, Shapira Y, Agmon-Levin N, Kivity S, Zafrir Y, Altman A, et al. Vitamin D insufficiency in a sunny environment: a demographic and seasonal analysis. *Isr Med Assoc J* 2010;12:751–6.
- [137] Kivity S, Agmon-Levin N, Zisappi M, Shapira Y, Nagy EV, Danko K, et al. Vitamin D and autoimmune thyroid diseases. *Cell Mol Immunol* 2011;8:243–7.
- [138] Lerner A, Shapira Y, Agmon-Levin N, Pacht A, Ben-Ami Shor D, Lopez HM, et al. The clinical significance of 25OH-Vitamin D status in celiac disease. *Clin Rev Allergy Immunol* 2012;42:322–30.
- [139] Hajas A, Sandor J, Csathy L, Csipo I, Barath S, Paragh G, et al. Vitamin D insufficiency in a large MCTD population. *Autoimmun Rev* 2011;10:317–24.
- [140] Cutolo M, Pizzorni C, Sulli A. Vitamin D endocrine system involvement in autoimmune rheumatic diseases. *Autoimmun Rev* 2011;11:84–7.
- [141] Lucas RM, Ponsoby AL, Dear K, Valery PC, Pender MP, Taylor BV, et al. Sun exposure and vitamin D are independent risk factors for CNS demyelination. *Neurology* 2011;76:540–8.
- [142] Munger KL, Levin LI, Hollis BW, Howard NS, Ascherio A. Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. *JAMA* 2006;296:2832–8.
- [143] Handal AE, Williamson AJ, Disanto G, Dobson R, Giovannoni G, Ramagopalan SV. Smoking and multiple sclerosis: an updated meta-analysis. *PLoS One* 2011;6:e16149.
- [144] Hedstrom AK, Sundqvist E, Baarnhielm M, Nordin N, Hillert J, Kockum I, et al. Smoking and two human leukocyte antigen genes interact to increase the risk for multiple sclerosis. *Brain* 2011;134:653–64.
- [145] Stejskal V, Hudecek R, Stejskal J, Sterzl I. Diagnosis and treatment of metal-induced side-effects. *Neuro Endocrinol Lett* 2006;27(Suppl. 1):7–16.
- [146] Fleming JO. Helminths and multiple sclerosis: will old friends give us new treatments for MS? *J Neuroimmunol* 2011;233:3–5.
- [147] Gaisford W, Cooke A. Can infections protect against autoimmunity? *Curr Opin Rheumatol* 2009;21:391–6.
- [148] Albright AV, Lavi E, Black JB, Goldberg S, O'Connor MJ, Gonzalez-Scarano F. The effect of human herpesvirus-6 (HHV-6) on cultured human neural cells: oligodendrocytes and microglia. *J Neurovirol* 1998;4:486–94.
- [149] Chan PK, Ng HK, Cheng AF. Detection of human herpesviruses 6 and 7 genomic sequences in brain tumours. *J Clin Pathol* 1999;52:620–3.
- [150] Kim JS, Lee KS, Park JH, Kim MY, Shin WS. Detection of human herpesvirus 6 variant A in peripheral blood mononuclear cells from multiple sclerosis patients. *Eur Neurol* 2000;43:170–3.
- [151] Opsahl ML, Kennedy PG. Early and late HHV-6 gene transcripts in multiple sclerosis lesions and normal appearing white matter. *Brain* 2005;128:516–27.
- [152] Mirandola P, Stefan A, Brambilla E, Campadelli-Fiume G, Grimaldi LM. Absence of human herpesvirus 6 and 7 from spinal fluid and serum of multiple sclerosis patients. *Neurology* 1999;53:1367–8.
- [153] Sospedra M, Zhao Y, zur Hausen H, Muraro PA, Hamashin C, de Villiers EM, et al. Recognition of conserved amino acid motifs of common viruses and its role in autoimmunity. *PLoS Pathog* 2005;1:e41.
- [154] Bogdanos DP, Gershwin ME. What is new in primary biliary cirrhosis? *Dig Dis* 2012;30(Suppl. 1):20–31.
- [155] Corpechot C, Chretien Y, Chazouilleres O, Poupon R. Demographic, lifestyle, medical and familial factors associated with primary biliary cirrhosis. *J Hepatol* 2010;53:162–9.
- [156] Gershwin ME, Selmi C, Worman HJ, Gold EB, Watnik M, Utts J, et al. Risk factors and comorbidities in primary biliary cirrhosis: a controlled interview-based study of 1032 patients. *Hepatology* 2005;42:1194–202.
- [157] Parikh-Patel A, Gold EB, Worman H, Krivy KE, Gershwin ME. Risk factors for primary biliary cirrhosis in a cohort of patients from the United States. *Hepatology* 2001;33:16–21.
- [158] Prince MI, Ducker SJ, James OF. Case-control studies of risk factors for primary biliary cirrhosis in two United Kingdom populations. *Gut* 2010;59:508–12.
- [159] Invernizzi P, Selmi C, Gershwin ME. Update on primary biliary cirrhosis. *Dig Liver Dis* 2010;42:401–8.
- [160] Selmi C, Invernizzi P, Zuin M, Podda M, Gershwin ME. Genetics and geoepidemiology of primary biliary cirrhosis: following the footprints to disease etiology. *Semin Liver Dis* 2005;25:265–80.
- [161] Selmi C, Invernizzi P, Zuin M, Podda M, Seldin MF, Gershwin ME. Genes and (auto)immunity in primary biliary cirrhosis. *Genes Immun* 2005;6:543–56.
- [162] Smyk D, Cholongitas E, Kriese S, Rigopoulou EI, Bogdanos DP. Primary biliary cirrhosis: family stories. *Autoimmune Dis* 2011;2011:189585.
- [163] Bogdanos DP, Smyk DS, Rigopoulou EI, Mytilinaou MG, Heneghan MA, Selmi C, et al. Twin studies in autoimmune disease: genetics, gender and environment. *J Autoimmun* 2012;38:156–69.
- [164] Gershwin ME, Mackay IR. The causes of primary biliary cirrhosis: convenient and inconvenient truths. *Hepatology* 2008;47:737–45.
- [165] Konikoff F, Pecht M, Theodor E, Shoenfeld Y. Primary biliary cirrhosis: lymphocyte subsets and function in a time frame. *Hepatology* 1989;10:525–6.
- [166] Schlesinger M, Benbassat C, Shoenfeld Y. Complement profile in primary biliary cirrhosis. *Immunol Res* 1992;11:98–103.
- [167] Shoenfeld Y. Primary biliary cirrhosis and autoimmune rheumatic diseases: prediction and prevention. *Isr J Med Sci* 1992;28:113–6.
- [168] Weiss P, Shoenfeld Y. Primary biliary cirrhosis is a multisystem disorder. *Ann Med Interne (Paris)* 1991;142:283–7.
- [169] Weiss P, Shoenfeld Y. Primary biliary cirrhosis: increasing problem, approaching solution. *Isr J Med Sci* 1992;28:726–8.
- [170] Benbassat C, Schlesinger M, Shoenfeld Y. The complement system and primary biliary cirrhosis. *J Clin Lab Immunol* 1992;38:51–61.
- [171] Rotman P, Levy Y, Shoenfeld Y. Primary biliary cirrhosis—association or overlap with other autoimmune diseases. *Isr J Med Sci* 1997;33:823–5.
- [172] Shapira S, Bar-Dayan Y, Gershwin ME, Shoenfeld Y. Is it possible to predict and prevent primary biliary cirrhosis? *Harefuah* 1999;137:39–41.
- [173] Aoki CA, Roifman CM, Lian ZX, Bowls CL, Norman GL, Shoenfeld Y, et al. IL-2 receptor alpha deficiency and features of primary biliary cirrhosis. *J Autoimmun* 2006;27:50–3.
- [174] Barak V, Selmi C, Schlesinger M, Blank M, Agmon-Levin N, Kalickman I, et al. Serum inflammatory cytokines, complement components, and soluble interleukin 2 receptor in primary biliary cirrhosis. *J Autoimmun* 2009;33:178–82.
- [175] Bogdanos DP, Baum H, Vergani D. Antimitochondrial and other autoantibodies. *Clin Liver Dis* 2003;7:759–77 [vi].
- [176] Bogdanos DP, Baum H, Vergani D, Burroughs AK. The role of *E. coli* infection in the pathogenesis of primary biliary cirrhosis. *Dis Markers* 2010;29:301–11.
- [177] Bogdanos DP, Vergani D. Origin of cross-reactive autoimmunity in primary biliary cirrhosis. *Liver Int* 2006;26:633–5.

- [178] Bogdanos DP, Vergani D. Bacteria and primary biliary cirrhosis. *Clin Rev Allergy Immunol* 2009;36:30–9.
- [179] Gershwin ME, Mackay IR. Primary biliary cirrhosis: paradigm or paradox for autoimmunity. *Gastroenterology* 1991;100:822–33.
- [180] Mackay IR, Whittingham S, Fida S, Myers M, Ikuno N, Gershwin ME, et al. The peculiar autoimmunity of primary biliary cirrhosis. *Immunol Rev* 2000;174:226–37.
- [181] Shimoda S, Nakamura M, Ishibashi H, Hayashida K, Niho Y. HLA DRB4 0101-restricted immunodominant T cell autoepitope of pyruvate dehydrogenase complex in primary biliary cirrhosis: evidence of molecular mimicry in human autoimmune diseases. *J Exp Med* 1995;181:1835–45.
- [182] Shimoda S, Nakamura M, Shigematsu H, Tanimoto H, Gushima T, Gershwin ME, et al. Mimicry peptides of human PDC-E2 163–176 peptide, the immunodominant T-cell epitope of primary biliary cirrhosis. *Hepatology* 2000;31:1212–6.
- [183] Shimoda S, Van de Water J, Ansari A, Nakamura M, Ishibashi H, Coppel RL, et al. Identification and precursor frequency analysis of a common T cell epitope motif in mitochondrial autoantigens in primary biliary cirrhosis. *J Clin Invest* 1998;102:1831–40.
- [184] Vergani D, Bogdanos DP. Positive markers in AMA-negative PBC. *Am J Gastroenterol* 2003;98:241–3.
- [185] Hohenester S, Oude-Elferink RP, Beuers U. Primary biliary cirrhosis. *Semin Immunopathol* 2009;31:283–307.
- [186] Neuberger J. Primary biliary cirrhosis. *Lancet* 1997;350:875–9.
- [187] Bogdanos DP, Invernizzi P, Mackay IR, Vergani D. Autoimmune liver serology: current diagnostic and clinical challenges. *World J Gastroenterol* 2008;14:3374–87.
- [188] Bogdanos DP, Komorowski L. Disease-specific autoantibodies in primary biliary cirrhosis. *Clin Chim Acta* 2011;412:502–12.
- [189] Dahnrich C, Pares A, Caballeria L, Rosemann A, Schlumberger W, Probst C, et al. New ELISA for detecting primary biliary cirrhosis-specific antimitochondrial antibodies. *Clin Chem* 2009;55:978–85.
- [190] Liu H, Norman GL, Shums Z, Worman HJ, Krawitt EL, Bizzaro N, et al. PBC screen: an IgG/IgA dual isotype ELISA detecting multiple mitochondrial and nuclear autoantibodies specific for primary biliary cirrhosis. *J Autoimmun* 2010;35:436–42.
- [191] Ma Y, Thomas MG, Okamoto M, Bogdanos DP, Nagl S, Kerker N, et al. Key residues of a major cytochrome P450D6 epitope are located on the surface of the molecule. *J Immunol* 2002;169:277–85.
- [192] Rigopoulou EI, Davies ET, Bogdanos DP, Liaskos C, Mytilinaiou M, Koukoulis GK, et al. Antimitochondrial antibodies of immunoglobulin G3 subclass are associated with a more severe disease course in primary biliary cirrhosis. *Liver Int* 2007;27:1226–31.
- [193] Wen L, Ma Y, Bogdanos DP, Wong FS, Demaine A, Mieli-Vergani G, et al. Pediatric autoimmune liver diseases: the molecular basis of humoral and cellular immunity. *Curr Mol Med* 2001;1:379–89.
- [194] Invernizzi P, Lleo A, Podda M. Interpreting serological tests in diagnosing autoimmune liver diseases. *Semin Liver Dis* 2007;27:161–72.
- [195] Rigopoulou EI, Bogdanos DP, Liaskos C, Koutsoumpas A, Baum H, Vergani D, et al. Anti-mitochondrial antibody immunofluorescent titres correlate with the number and intensity of immunoblot-detected mitochondrial bands in patients with primary biliary cirrhosis. *Clin Chim Acta* 2007;380:118–21.
- [196] Gilburd B, Ziporen L, Zharhary D, Blank M, Zurgil N, Scheinberg MA, et al. Antimitochondrial (pyruvate dehydrogenase) antibodies in leprosy. *J Clin Immunol* 1994;14:14–9.
- [197] Konikoff F, Isenberg DA, Barrison I, Theodor E, Shoenfeld Y. Antinuclear autoantibodies in chronic liver diseases. *HepatoGastroenterology* 1989;36:341–5.
- [198] Konikoff F, Isenberg DA, Kooperman O, Kennedy RC, Rauch J, Theodor E, et al. Common lupus anti-DNA antibody idiotypes in chronic liver diseases. *Clin Immunol Immunopathol* 1987;43:265–72.
- [199] Konikoff F, Shoenfeld Y, Isenberg DA, Barrison I, Sobe T, Theodor E, et al. Anti-Rnp antibodies in chronic liver diseases. *Clin Exp Rheumatol* 1987;5:359–61.
- [200] Krause I, Hacham S, Gilburd B, Damianovitch M, Blank M, Shoenfeld Y. Absence of anti-idiotypic antibodies in IVIG preparations to autoantibodies of rare autoimmune diseases. *Clin Immunol Immunopathol* 1995;77:229–35.
- [201] Lorber M, Kra-Oz Z, Guilbrud B, Shoenfeld Y. Natural (antiphospholipid-PDH₄-DNA) autoantibodies and their physiologic serum inhibitors. *Isr J Med Sci* 1995;31:31–5.
- [202] Maran R, Dueymes M, Adler Y, Shoenfeld Y, Youinou P. Isotypic distribution of anti-pyruvate dehydrogenase antibodies in patients with primary biliary cirrhosis and their family members. *J Clin Immunol* 1994;14:323–6.
- [203] Shoenfeld Y, Beresovski A, Zharhary D, Tomer Y, Swissa M, Sela E, et al. Natural autoantibodies in sera of patients with Gaucher's disease. *J Clin Immunol* 1995;15:363–72.
- [204] Zurgil N, Bakimer R, Kaplan M, Youinou P, Shoenfeld Y. Anti-pyruvate dehydrogenase autoantibodies in primary biliary cirrhosis. *J Clin Immunol* 1991;11:239–45.
- [205] Zurgil N, Bakimer R, Moutsopoulos HM, Tzioufas AG, Youinou P, Isenberg DA, et al. Antimitochondrial (pyruvate dehydrogenase) autoantibodies in autoimmune rheumatic diseases. *J Clin Immunol* 1992;12:201–9.
- [206] Zurgil N, Bakimer R, Slor H, Kaplan M, Moutsopoulos H, Shoenfeld Y. Pyruvate dehydrogenase as an antigen to detect antimitochondrial antibodies. *Isr J Med Sci* 1990;26:682–5.
- [207] Zurgil N, Konikoff F, Bakimer R, Slor H, Shoenfeld Y. Detection of antimitochondrial antibodies: characterization by enzyme immunoassay and immunoblotting. *Autoimmunity* 1989;4:289–97.
- [208] Tishler M, Alosachie I, Barka N, Lin HC, Gershwin ME, Peter JB, et al. Primary Sjogren's syndrome and primary biliary cirrhosis: differences and similarities in the autoantibody profile. *Clin Exp Rheumatol* 1995;13:497–500.
- [209] Bean P, Sutphin MS, Liu Y, Anton R, Reynolds TB, Shoenfeld Y, et al. Carbohydrate-deficient transferrin and false-positive results for alcohol abuse in primary biliary cirrhosis: differential diagnosis by detection of mitochondrial autoantibodies. *Clin Chem* 1995;41:858–61.
- [210] Agmon-Levin N, Shapira Y, Selmi C, Barzilai O, Ram M, Szyper-Kravitz M, et al. A comprehensive evaluation of serum autoantibodies in primary biliary cirrhosis. *J Autoimmun* 2010;34:55–8.
- [211] Metcalf JV, Mitchison HC, Palmer JM, Jones DE, Bassendine MF, James OF. Natural history of early primary biliary cirrhosis. *Lancet* 1996;348:1399–402.
- [212] Dubel L, Tanaka A, Leung PS, Van de Water J, Coppel R, Roche T, et al. Autoepitope mapping and reactivity of autoantibodies to the dihydroliipoamide dehydrogenase-binding protein (E3BP) and the glycine cleavage proteins in primary biliary cirrhosis. *Hepatology* 1999;29:1013–8.
- [213] Leung PS, Coppel RL, Ansari A, Munoz S, Gershwin ME. Antimitochondrial antibodies in primary biliary cirrhosis. *Semin Liver Dis* 1997;17:61–9.
- [214] Palmer JM, Jones DE, Quinn J, McHugh A, Yeaman SJ. Characterization of the autoantibody responses to recombinant E3 binding protein (protein X) of pyruvate dehydrogenase in primary biliary cirrhosis. *Hepatology* 1999;30:21–6.
- [215] Van de Water J, Fregeau D, Davis P, Ansari A, Danner D, Leung P, et al. Autoantibodies of primary biliary cirrhosis recognize dihydroliipoamide acetyltransferase and inhibit enzyme function. *J Immunol* 1988;141:2321–4.
- [216] Bogdanos DP, Liaskos C, Pares A, Norman G, Rigopoulou EI, Caballeria L, et al. Anti-gp210 antibody mirrors disease severity in primary biliary cirrhosis. *Hepatology* 2007;45:1583 [author reply 4].
- [217] Invernizzi P, Podda M, Battezzati PM, Crosignani A, Zuin M, Hitchman E, et al. Autoantibodies against nuclear pore complexes are associated with more active and severe liver disease in primary biliary cirrhosis. *J Hepatol* 2001;34:366–72.
- [218] Miyachi K, Hankins RW, Matsushima H, Kikuchi F, Inomata T, Horigome T, et al. Profile and clinical significance of anti-nuclear envelope antibodies found in patients with primary biliary cirrhosis: a multicenter study. *J Autoimmun* 2003;20:247–54.
- [219] Nakamura M, Kondo H, Mori T, Komori A, Matsuyama M, Ito M, et al. Anti-gp210 and anti-centromere antibodies are different risk factors for the progression of primary biliary cirrhosis. *Hepatology* 2007;45:118–27.
- [220] Rigopoulou EI, Davies ET, Pares A, Zachou K, Liaskos C, Bogdanos DP, et al. Prevalence and clinical significance of isotype specific antinuclear antibodies in primary biliary cirrhosis. *Gut* 2005;54:528–32.
- [221] Itoh S, Ichida T, Yoshida T, Hayakawa A, Uchida M, Tashiro-Itoh T, et al. Autoantibodies against a 210 kDa glycoprotein of the nuclear pore complex as a prognostic marker in patients with primary biliary cirrhosis. *J Gastroenterol Hepatol* 1998;13:257–65.
- [222] Lassoued K, Guilly MN, Andre C, Paintrand M, Dhumeaux D, Danon F, et al. Autoantibodies to 200 kD polypeptide(s) of the nuclear envelope: a new serologic marker of primary biliary cirrhosis. *Clin Exp Immunol* 1988;74:283–8.
- [223] Muratori P, Muratori L, Ferrari R, Cassani F, Bianchi G, Lenzi M, et al. Characterization and clinical impact of antinuclear antibodies in primary biliary cirrhosis. *Am J Gastroenterol* 2003;98:431–7.
- [224] Wesierska-Gadek J, Penner E, Battezzati PM, Selmi C, Zuin M, Hitchman E, et al. Correlation of initial autoantibody profile and clinical outcome in primary biliary cirrhosis. *Hepatology* 2006;43:1135–44.
- [225] Yang WH, Yu JH, Nakajima A, Neuberger D, Lindor K, Bloch DB. Do antinuclear antibodies in primary biliary cirrhosis patients identify increased risk for liver failure? *Clin Gastroenterol Hepatol* 2004;2:1116–22.
- [226] Duarte-Rey C, Bogdanos D, Yang CY, Roberts K, Leung PS, Anaya JM, et al. Primary biliary cirrhosis and the nuclear pore complex. *Autoimmun Rev* 2012;11:898–902.
- [227] Duarte-Rey C, Bogdanos DP, Leung PS, Anaya JM, Gershwin ME. IgM predominance in autoimmune disease: genetics and gender. *Autoimmun Rev* 2012;11:A404–12.
- [228] Bach N, Schaffner F. Familial primary biliary cirrhosis. *J Hepatol* 1994;20:698–701.
- [229] Floreani A, Naccarato R, Chiaromonte M. Prevalence of familial disease in primary biliary cirrhosis in Italy. *J Hepatol* 1997;26:737–8.
- [230] Tsuji K, Watanabe Y, Van De Water J, Nakanishi T, Kajiyama G, Parikh-Patel A, et al. Familial primary biliary cirrhosis in Hiroshima. *J Autoimmun* 1999;13:171–8.
- [231] Lazaridis KN, Juran BD, Boe GM, Slusser JP, de Andrade M, Homburger HA, et al. Increased prevalence of antimitochondrial antibodies in first-degree relatives of patients with primary biliary cirrhosis. *Hepatology* 2007;46:785–92.
- [232] Douglas JG, Finlayson ND. Are increased individual susceptibility and environmental factors both necessary for the development of primary biliary cirrhosis? *BMJ* 1979;2:419–20.
- [233] Fagan E, Williams R, Cox S. Primary biliary cirrhosis in mother and daughter. *BMJ* 1977;2:1195.
- [234] Tong MJ, Nies KM, Reynolds TB, Quismorio FP. Immunological studies in familial primary biliary cirrhosis. *Gastroenterology* 1976;71:305–7.
- [235] Poupon R. Primary biliary cirrhosis: a 2010 update. *J Hepatol* 2010;52:745–58.
- [236] Corpechot C, Carrat F, Poupon R, Poupon RE. Primary biliary cirrhosis: incidence and predictive factors of cirrhosis development in ursodiol-treated patients. *Gastroenterology* 2002;122:652–8.
- [237] Smyk DS, Rigopoulou EI, Bogdanos DP. Potential roles for infectious agents in the pathophysiology of primary biliary cirrhosis: what's new? *Curr Infect Dis Rep* Nov 29 2012 [Electronic publication ahead of print].

- [238] Chuang YH, Ridgway WM, Ueno Y, Gershwin ME. Animal models of primary biliary cirrhosis. *Clin Liver Dis* 2008;12:333–47 [ix].
- [239] Hirschfield GM, Invernizzi P. Progress in the genetics of primary biliary cirrhosis. *Semin Liver Dis* 2011;31:147–56.
- [240] Lleo A, Invernizzi P, Mackay IR, Prince H, Zhong RQ, Gershwin ME. Etiopathogenesis of primary biliary cirrhosis. *World J Gastroenterol* 2008;14:3328–37.
- [241] Selmi C, Zuin M, Gershwin ME. The unfinished business of primary biliary cirrhosis. *J Hepatol* 2008;49:451–60.
- [242] Wu SJ, Yang YH, Tsuneyama K, Leung PS, Illarionov P, Gershwin ME, et al. Innate immunity and primary biliary cirrhosis: activated invariant natural killer T cells exacerbate murine autoimmune cholangitis and fibrosis. *Hepatology* 2011;53:915–25.
- [243] Hirschfield GM, Gershwin ME. Primary biliary cirrhosis: one disease with many faces. *Isr Med Assoc J* 2011;13:55–9.
- [244] Invernizzi P. Human leukocyte antigen in primary biliary cirrhosis: an old story now reviving. *Hepatology* 2011;2:714–23 [PMID: 21563204].
- [245] Hemminki K, Li X, Sundquist K, Sundquist J. Shared familial aggregation of susceptibility to autoimmune diseases. *Arthritis Rheum* 2009;60:2845–7.
- [246] Hirschfield GM, Liu X, Han Y, Gorlov IP, Lu Y, Xu C, et al. Variants at IRF5-TNPO3, 17q12–21 and MMEL1 are associated with primary biliary cirrhosis. *Nat Genet* 2010;42:655–7.
- [247] Hirschfield GM, Liu X, Xu C, Lu Y, Xie G, Gu X, et al. Primary biliary cirrhosis associated with HLA, IL12A, and IL12RB2 variants. *N Engl J Med* 2009;360:2544–55.
- [248] Liu X, Invernizzi P, Lu Y, Kosoy R, Bianchi I, Podda M, et al. Genome-wide meta-analyses identify three loci associated with primary biliary cirrhosis. *Nat Genet* 2010;42:658–60.
- [249] Mellis GF, Floyd JA, Morley KI, Cordell HJ, Franklin CS, Shin SY, et al. Genome-wide association study identifies 12 new susceptibility loci for primary biliary cirrhosis. *Nat Genet* 2011;43:329–32.
- [250] Tanaka A, Invernizzi P, Ohira H, Kikuchi K, Nezu S, Kosoy R, et al. Replicated association of 17q12–21 with susceptibility of primary biliary cirrhosis in a Japanese cohort. *Tissue Antigens* 2011;78:65–8.
- [251] Tanaka A, Ohira H, Kikuchi K, Nezu S, Shibuya A, Bianchi I, et al. Genetic association of Fc receptor-like 3 polymorphisms with susceptibility to primary biliary cirrhosis: ethnic comparative study in Japanese and Italian patients. *Tissue Antigens* 2011;77:239–43.
- [252] Invernizzi P, Miozzo M, Battezzati PM, Bianchi I, Grati FR, Simoni G, et al. Frequency of monosomy X in women with primary biliary cirrhosis. *Lancet* 2004;363:533–5.
- [253] Invernizzi P, Pasini S, Selmi C, Gershwin ME, Podda M. Female predominance and X chromosome defects in autoimmune diseases. *J Autoimmun* 2009;33:12–6.
- [254] Miozzo M, Selmi C, Gentilin B, Grati FR, Sirchia S, Oertelt S, et al. Preferential X chromosome loss but random inactivation characterize primary biliary cirrhosis. *Hepatology* 2007;46:456–62.
- [255] Invernizzi P, Selmi C, Poli F, Frison S, Floreani A, Alvaro D, et al. Human leukocyte antigen polymorphisms in Italian primary biliary cirrhosis: a multicenter study of 664 patients and 1992 healthy controls. *Hepatology* 2008;48:1906–12.
- [256] Bogdanos DP, Baum H, Butler P, Rigopoulou EI, Davies ET, Ma Y, et al. Association between the primary biliary cirrhosis specific anti-sp100 antibodies and recurrent urinary tract infection. *Dig Liver Dis* 2003;35:801–5.
- [257] Bogdanos DP, Baum H, Grasso A, Okamoto M, Butler P, Ma Y, et al. Microbial mimics are major targets of crossreactivity with human pyruvate dehydrogenase in primary biliary cirrhosis. *J Hepatol* 2004;40:31–9.
- [258] Bogdanos DP, Pares A, Rodes J, Vergani D. Primary biliary cirrhosis specific anti-nuclear antibodies in patients from Spain. *Am J Gastroenterol* 2004;99:763–4 [author reply 5].
- [259] Bogdanos DP, Baum H, Okamoto M, Montalto P, Sharma UC, Rigopoulou EI, et al. Primary biliary cirrhosis is characterized by IgG3 antibodies cross-reactive with the major mitochondrial autoepitope and its *Lactobacillus* mimic. *Hepatology* 2005;42:458–65.
- [260] Bogdanos DP, Baum H, Sharma UC, Grasso A, Ma Y, Burroughs AK, et al. Antibodies against homologous microbial caseinolytic proteases P characterise primary biliary cirrhosis. *J Hepatol* 2002;36:14–21.
- [261] Fussey SP, Lindsay JG, Fuller C, Perham RN, Dale S, James OF, et al. Autoantibodies in primary biliary cirrhosis: analysis of reactivity against eukaryotic and prokaryotic 2-oxo acid dehydrogenase complexes. *Hepatology* 1991;13:467–74.
- [262] Koutsoumpas A, Mytilinaou D, Polymeros D, Dalekos GN, Bogdanos DP. Anti-*Helicobacter pylori* antibody responses specific for VacA do not trigger primary biliary cirrhosis-specific antimicrobial antibodies. *Eur J Gastroenterol Hepatol* 2009;21:1220.
- [263] Abdulkarim AS, Petrovic LM, Kim WR, Angulo P, Lloyd RV, Lindor KD. Primary biliary cirrhosis: an infectious disease caused by *Chlamydia pneumoniae*? *J Hepatol* 2004;40:380–4.
- [264] Burroughs AK, Butler P, Sternberg MJ, Baum H. Molecular mimicry in liver disease. *Nature* 1992;358:377–8.
- [265] Burroughs AK, Rosenstein IJ, Epstein O, Hamilton-Miller JM, Brumfitt W, Sherlock S. Bacteriuria and primary biliary cirrhosis. *Gut* 1984;25:133–7.
- [266] Butler P, Hamilton-Miller J, Baum H, Burroughs AK. Detection of M2 antibodies in patients with recurrent urinary tract infection using an ELISA and purified PBC specific antigens. Evidence for a molecular mimicry mechanism in the pathogenesis of primary biliary cirrhosis? *Biochem Mol Biol Int* 1995;35:473–85.
- [267] Butler P, Hamilton-Miller JM, McIntyre N, Burroughs AK. Natural history of bacteriuria in women with primary biliary cirrhosis and the effect of antimicrobial therapy in symptomatic and asymptomatic groups. *Gut* 1995;36:931–4.
- [268] Butler P, Valle F, Hamilton-Miller JM, Brumfitt W, Baum H, Burroughs AK. M2 mitochondrial antibodies and urinary rough mutant bacteria in patients with primary biliary cirrhosis and in patients with recurrent bacteriuria. *J Hepatol* 1993;17:408–14.
- [269] Donaldson PT. Genetics of liver disease: immunogenetics and disease pathogenesis. *Gut* 2004;53:599–608.
- [270] Floreani A, Bassendine MF, Mitchison H, Freeman R, James OF. No specific association between primary biliary cirrhosis and bacteriuria? *J Hepatol* 1989;8:201–7.
- [271] Leung PS, Park O, Matsumura S, Ansari AA, Coppel RL, Gershwin ME. Is there a relation between *Chlamydia* infection and primary biliary cirrhosis? *Clin Dev Immunol* 2003;10:227–33.
- [272] Mason A, Xu L, Shen Z, Fodera B, Joplin R, Neuberger J, et al. Patients with primary biliary cirrhosis make anti-viral and anti-mitochondrial antibodies to mouse mammary tumor virus. *Gastroenterology* 2004;127:1863–4 [author reply 4–5].
- [273] McNally RJ, Ducker S, James OF. Are transient environmental agents involved in the cause of primary biliary cirrhosis? Evidence from space-time clustering analysis. *Hepatology* 2009;50:1169–74.
- [274] Selmi C, Balkwill DL, Invernizzi P, Ansari AA, Coppel RL, Podda M, et al. Patients with primary biliary cirrhosis react against a ubiquitous xenobiotic-metabolizing bacterium. *Hepatology* 2003;38:1250–7.
- [275] Smyk D, Mytilinaou MG, Rigopoulou EI, Bogdanos DP. PBC triggers in water reservoirs, coal mining areas and waste disposal sites: from Newcastle to New York. *Dis Markers* 2010;29:337–44.
- [276] Xu L, Sakalian M, Shen Z, Loss G, Neuberger J, Mason A. Cloning the human betaretrovirus proviral genome from patients with primary biliary cirrhosis. *Hepatology* 2004;39:151–6.
- [277] Xu L, Shen Z, Guo L, Fodera B, Keogh A, Joplin R, et al. Does a betaretrovirus infection trigger primary biliary cirrhosis? *Proc Natl Acad Sci U S A* 2003;100:8454–9.
- [278] Selmi C, De Santis M, Cavaciocchi F, Gershwin ME. Infectious agents and xenobiotics in the etiology of primary biliary cirrhosis. *Dis Markers* 2010;29:287–99.
- [279] Bogdanos DP, Pustl T, Rust C, Vergani D, Beuers U. Primary biliary cirrhosis following *Lactobacillus* vaccination for recurrent vaginitis. *J Hepatol* 2008;49:466–73.
- [280] Oldstone MB. Molecular mimicry and autoimmune disease. *Cell* 1987;50:819–20.
- [281] Bogdanos DP, Choudhuri K, Vergani D. Molecular mimicry and autoimmune liver disease: virtuous intentions, malign consequences. *Liver* 2001;21:225–32.
- [282] Vergani D, Bogdanos DP, Baum H. Unusual suspects in primary biliary cirrhosis. *Hepatology* 2004;39:38–41.
- [283] Rigopoulou EI, Smyk DS, Matthews CE, Billinis C, Burroughs AK, Lenzi M, et al. Epstein-Barr virus as a trigger of autoimmune liver diseases. *Adv Virol* 2012;2012:987471.
- [284] Vergani D, Choudhuri K, Bogdanos DP, Mieli-Vergani G. Pathogenesis of autoimmune hepatitis. *Clin Liver Dis* 2002;6:727–37.
- [285] Vergani D, Longhi MS, Bogdanos DP, Ma Y, Mieli-Vergani G. Autoimmune hepatitis. *Semin Immunopathol* 2009;31:421–35.
- [286] Ahn J, Yang L, Paster BJ, Ganly I, Morris L, Pei Z, et al. Oral microbiome profiles: 16S rRNA pyrosequencing and microarray assay comparison. *PLoS One* 2011;6:e22788.
- [287] Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, et al. Enterotypes of the human gut microbiome. *Nature* 2011;473:174–80.
- [288] Gill SR, Pop M, Deboy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, et al. Metagenomic analysis of the human distal gut microbiome. *Science* 2006;312:1355–9.
- [289] Kurokawa K, Itoh T, Kuwahara T, Oshima K, Toh H, Toyoda A, et al. Comparative metagenomics revealed commonly enriched gene sets in human gut microbiomes. *DNA Res* 2007;14:169–81.
- [290] Marchesi JR. Human distal gut microbiome. *Environ Microbiol* 2011.
- [291] Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 2010;464:59–65.
- [292] Saulnier DM, Riehle K, Mistretta TA, Diaz MA, Mandal D, Raza S, et al. Gastrointestinal microbiome signatures of pediatric patients with irritable bowel syndrome. *Gastroenterology* 2011.
- [293] Siezen RJ, Kleerebezem M. The human gut microbiome: are we our enterotypes? *Microb Biotechnol* 2011;4:550–3.
- [294] Turnbaugh PJ, Hamady M, Yatsunenkov T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. *Nature* 2009;457:480–4.
- [295] Nasidze I, Li J, Schroeder R, Creasey JL, Li M, Stoneking M. High diversity of the saliva microbiome in Batwa pygmies. *PLoS One* 2011;6:e23352.
- [296] Smyk DS, Bogdanos DP, Kriese S, Billinis C, Burroughs AK, Rigopoulou EI. Urinary tract infection as a risk factor for autoimmune liver disease: from bench to bedside. *Clin Res Hepatol Gastroenterol* 2011.
- [297] Varyani FK, West J, Card TR. An increased risk of urinary tract infection precedes development of primary biliary cirrhosis. *BMC Gastroenterol* 2011;11:95.
- [298] Smyk D, Rigopoulou EI, Baum H, Burroughs AK, Vergani D, Bogdanos DP. Autoimmunity and environment: am I at risk? *Clin Rev Allergy Immunol* 2011.
- [299] Natesan M, Ulrich RG. Protein microarrays and biomarkers of infectious disease. *Int J Mol Sci* 2010;11:5165–83.
- [300] Quintana FJ, Hagedorn PH, Elizur G, Merbl Y, Domany E, Cohen IR. Functional immunomics: microarray analysis of IgG autoantibody repertoires predicts the future response of mice to induced diabetes. *Proc Natl Acad Sci U S A* 2004;101(Suppl. 2):14615–21.
- [301] Larman HB, Zhao Z, Laserson U, Li MZ, Ciccio A, Gakidis MA, et al. Autoantigen discovery with a synthetic human peptidome. *Nat Biotechnol* 2011;29:535–41.

- [302] Drebber U, Kasper HU, Ratering J, Wedemeyer I, Schirmacher P, Dienes HP, et al. Hepatic granulomas: histological and molecular pathological approach to differential diagnosis—a study of 442 cases. *Liver Int* 2008;28:828–34.
- [303] O'Donoghue J, Fidler H, Garcia-Barcelo M, Nouri-Aria K, Williams R, McFadden J. Mycobacterial DNA not detected in liver sections from patients with primary biliary cirrhosis. *J Hepatol* 1998;28:433–8.
- [304] Tanaka A, Prindiville TP, Gish R, Solnick JV, Coppel RL, Keeffe EB, et al. Are infectious agents involved in primary biliary cirrhosis? A PCR approach. *J Hepatol* 1999;31:664–71.
- [305] Vilagut L, Pares A, Rodes J, Vila J, Vinas O, Gines A, et al. Mycobacteria—related to the aetiopathogenesis of primary biliary cirrhosis? *J Hepatol* 1996;24:125.
- [306] Chen EC, Miller SA, DeRisi JL, Chiu CY. Using a pan-viral microarray assay (Virochip) to screen clinical samples for viral pathogens. *J Vis Exp* 2011.
- [307] Kistler A, Avila PC, Rouskin S, Wang D, Ward T, Yagi S, et al. Pan-viral screening of respiratory tract infections in adults with and without asthma reveals unexpected human coronavirus and human rhinovirus diversity. *J Infect Dis* 2007;196:817–25.
- [308] Nakamura S, Nakaya T, Iida T. Metagenomic analysis of bacterial infections by means of high-throughput DNA sequencing. *Exp Biol Med (Maywood)* 2011;236:968–71.
- [309] Tang P, Chiu C. Metagenomics for the discovery of novel human viruses. *Future Microbiol* 2010;5:177–89.
- [310] Moore RA, Warren RL, Freeman JD, Gustavsen JA, Chenard C, Friedman JM, et al. The sensitivity of massively parallel sequencing for detecting candidate infectious agents associated with human tissue. *PLoS One* 2011;6:e19838.
- [311] Broome U, Scheynius A, Hultcrantz R. Induced expression of heat-shock protein on biliary epithelium in patients with primary sclerosing cholangitis and primary biliary cirrhosis. *Hepatology* 1993;18:298–303.
- [312] Singhal S, Dian D, Keshavarzian A, Fogg L, Fields JZ, Farhadi A. The role of oral hygiene in inflammatory bowel disease. *Dig Dis Sci* 2011;56:170–5.
- [313] Ochoa-Reparaz J, Mielcarz DW, Ditrilo LE, Burroughs AR, Foureau DM, Haque-Begum S, et al. Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. *J Immunol* 2009;183:6041–50.
- [314] Helenius LM, Meurman JH, Helenius I, Kari K, Hietanen J, Suuronen R, et al. Oral and salivary parameters in patients with rheumatic diseases. *Acta Odontol Scand* 2005;63:284–93.
- [315] Mason AL, Xu L, Guo L, Munoz S, Jaspan JB, Bryer-Ash M, et al. Detection of retroviral antibodies in primary biliary cirrhosis and other idiopathic biliary disorders. *Lancet* 1998;351:1620–4.
- [316] Alvaro D, Alpini G, Onori P, Franchitto A, Glaser SS, Le Sage G, et al. Alfa and beta estrogen receptors and the biliary tree. *Mol Cell Endocrinol* 2002;193:105–8.
- [317] Cunningham M, Gilkeson G. Estrogen receptors in immunity and autoimmunity. *Clin Rev Allergy Immunol* 2011;40:66–73.
- [318] Duvic M, Steinberg AD, Klassen LW. Effect of the anti-estrogen, Nafoxidine, on NZB/W autoimmune disease. *Arthritis Rheum* 1978;21:414–7.
- [319] Holmdahl R. Estrogen exaggerates lupus but suppresses T-cell-dependent autoimmune disease. *J Autoimmun* 1989;2:651–6.
- [320] Walker SE. Estrogen and autoimmune disease. *Clin Rev Allergy Immunol* 2011;40:60–5.
- [321] Mizutani T, Shinoda M, Tanaka Y, Kuno T, Hattori A, Usui T, et al. Autoantibodies against CYP2D6 and other drug-metabolizing enzymes in autoimmune hepatitis type 2. *Drug Metab Rev* 2005;37:235–52.
- [322] Obermayer-Straub P, Strassburg CP, Manns MP. Autoimmune hepatitis. *J Hepatol* 2000;32:181–97.
- [323] Garratty G. Drug-induced immune hemolytic anemia. *Hematology Am Soc Hematol Educ Program* 2009;73–9.
- [324] Khokhar O, Gange C, Clement S, Lewis J. Autoimmune hepatitis and thyroiditis associated with rifampin and pyrazinamide prophylaxis: an unusual reaction. *Dig Dis Sci* 2005;50:207–11.
- [325] Takasu N, Takara M, Komiya I. Rifampin-induced hypothyroidism in patients with Hashimoto's thyroiditis. *N Engl J Med* 2005;352:518–9.
- [326] Ahrens N, Genth R, Salama A. Belated diagnosis in three patients with rifampicin-induced immune haemolytic anaemia. *Br J Haematol* 2002;117:441–3.
- [327] Heurgue-Berlot A, Bernard-Chabert B, Diebold MD, Thieffin G. Drug-induced autoimmune-like hepatitis: a case of chronic course after drug withdrawal. *Dig Dis Sci* 2011;56:2504–5 [author reply 5].
- [328] Bjornsson E, Talwalkar J, Treeprasertsuk S, Kamath PS, Takahashi N, Sanderson S, et al. Drug-induced autoimmune hepatitis: clinical characteristics and prognosis. *Hepatology* 2010;51:2040–8.
- [329] Geddes MR, Sinnreich M, Chalk C. Minocycline-induced dermatomyositis. *Muscle Nerve* 2010;41:547–9.
- [330] Angulo JM, Sigal LH, Espinoza LR. Minocycline induced lupus and autoimmune hepatitis. *J Rheumatol* 1999;26:1420–1.
- [331] Bachmeyer C, Cadranet JF. Minocycline-induced lupus and autoimmune hepatitis: family autoimmune disorders as possible risk factors. *Dermatology* 2002;205:185–6.
- [332] Bhat G, Jordan Jr J, Sokalski S, Bajaj V, Marshall R, Berkelhammer C. Minocycline-induced hepatitis with autoimmune features and neutropenia. *J Clin Gastroenterol* 1998;27:74–5.
- [333] Chamberlain MC, Schwarzenberg SJ, Akin EU, Kurth MH. Minocycline-induced autoimmune hepatitis with subsequent cirrhosis. *J Pediatr Gastroenterol Nutr* 2006;42:232–5.
- [334] Colmegna I, Perandones CE, Chaves JG. Minocycline induced lupus and autoimmune hepatitis. *J Rheumatol* 2000;27:1567–8.
- [335] Gough A, Chapman S, Wagstaff K, Emery P, Elias E. Minocycline induced autoimmune hepatitis and systemic lupus erythematosus-like syndrome. *BMJ* 1996;312:169–72.
- [336] Healy J, Alexander B, Eapen C, Roberts-Thomson IC. Minocycline-induced autoimmune hepatitis. *Intern Med J* 2009;39:487–8.
- [337] Della Corte C, Carlucci A, Francalanci P, Alisi A, Nobili V. Autoimmune hepatitis type 2 following anti-papillomavirus vaccination in a 11-year-old girl. *Vaccine* 2011;29:4654–6.
- [338] Karali Z, Basaranoglu ST, Karali Y, Oral B, Kilic SS. Autoimmunity and hepatitis A vaccine in children. *J Investig Allergol Clin Immunol* 2011;21:389–93.
- [339] Stubgen JP. Neuromuscular disorders associated with hepatitis B vaccination. *J Neurol Sci* 2010;292:1–4.
- [340] Cacoub P, Terrier B. Hepatitis B-related autoimmune manifestations. *Rheum Dis Clin North Am* 2009;35:125–37.
- [341] Aron-Maor A, Shoenfeld Y. Vaccination and systemic lupus erythematosus: the bidirectional dilemmas. *Lupus* 2001;10:237–40.
- [342] Borchers AT, Keen CL, Shoenfeld Y, Silva Jr J, Gershwin ME. Vaccines, viruses, and voodoo. *J Investig Allergol Clin Immunol* 2002;12:155–68.
- [343] Chen RT, Pless R, Destefano F. Epidemiology of autoimmune reactions induced by vaccination. *J Autoimmun* 2001;16:309–18.
- [344] Cohen AD, Shoenfeld Y. Vaccine-induced autoimmunity. *J Autoimmun* 1996;9:699–703.
- [345] Nadler JP. Multiple sclerosis and hepatitis B vaccination. *Clin Infect Dis* 1993;17:928–9.
- [346] Ravel G, Christ M, Horand F, Descotes J. Autoimmunity, environmental exposure and vaccination: is there a link? *Toxicology* 2004;196:211–6.
- [347] Shoenfeld Y, Aharon-Maor A, Sherer Y. Vaccination as an additional player in the mosaic of autoimmunity. *Clin Exp Rheumatol* 2000;18:181–4.
- [348] Shoenfeld Y, Aron-Maor A. Vaccination and autoimmunity-'vaccinosis': a dangerous liaison? *J Autoimmun* 2000;14:1–10.
- [349] Shoenfeld Y, Aron-Maor A, Tanai A, Ehrenfeld M. Bcg and autoimmunity: another two-edged sword. *J Autoimmun* 2001;16:235–40.
- [350] Costenbader KH, Gay S, Riquelme ME, Iaccarino L, Doria A. Genes, epigenetic regulation and environmental factors which is the most relevant in developing autoimmune diseases? *Autoimmun Rev* 2011.
- [351] Arnon Y, Shoenfeld Y, Amital H. Effects of tobacco smoke on immunity, inflammation and autoimmunity. *J Autoimmun* 2010;34:258–65.
- [352] Brusselle GG, Joos GF, Bracke KR. New insights into the immunology of chronic obstructive pulmonary disease. *Lancet* 2011;378:1015–26.
- [353] Simard JF, Costenbader KH. What can epidemiology tell us about systemic lupus erythematosus? *Int J Clin Pract* 2007;61:1170–80.
- [354] Costenbader KH, Karlson EW. Cigarette smoking and autoimmune disease: what can we learn from epidemiology? *Lupus* 2006;15:737–45.
- [355] Rubin RL, Hermanson TM, Bedrick EJ, McDonald JD, Burchiel SW, Reed MD, et al. Effect of cigarette smoke on autoimmunity in murine and human systemic lupus erythematosus. *Toxicol Sci* 2005;87:86–96.
- [356] Asherson RA, Shoenfeld Y, Jacobs P, Bosman C. An unusually complicated case of primary Sjogren's syndrome: development of transient "lupus-type" autoantibodies following silicone implant rejection. *J Rheumatol* 2004;31:196–7.
- [357] Bar-Meir E, Ehrenfeld M, Shoenfeld Y. Silicone gel breast implants and connective tissue disease—a comprehensive review. *Autoimmunity* 2003;36:193–7.
- [358] Bar-Meir E, Teuber SS, Lin HC, Alasocie I, Goddard G, Terybery J, et al. Multiple autoantibodies in patients with silicone breast implants. *J Autoimmun* 1995;8:267–77.
- [359] Vasey FB, Zarabadi SA, Seleznick M, Ricca L. Where there's smoke there's fire: the silicone breast implant controversy continues to flicker: a new disease that needs to be defined. *J Rheumatol* 2003;30:2092–4.
- [360] Vermeulen RC, Scholte HR. Rupture of silicone gel breast implants and symptoms of pain and fatigue. *J Rheumatol* 2003;30:2263–7.
- [361] Zandman-Goddard G, Blank M, Ehrenfeld M, Gilburd B, Peter J, Shoenfeld Y. A comparison of autoantibody production in asymptomatic and symptomatic women with silicone breast implants. *J Rheumatol* 1999;26:73–7.
- [362] Hajdu SD, Agmon-Levin N, Shoenfeld Y. Silicone and autoimmunity. *Eur J Clin Invest* 2011;41:203–11.
- [363] Alotaibi S, Kennedy J, Tellier R, Stephens D, Banwell B. Epstein-Barr virus in pediatric multiple sclerosis. *JAMA* 2004;291:1875–9.
- [364] DeLorenzo GN, Munger KL, Lennette ET, Orentreich N, Vogelmann JH, Ascherio A. Epstein-Barr virus and multiple sclerosis: evidence of association from a prospective study with long-term follow-up. *Arch Neurol* 2006;63:839–44.
- [365] Goodin DS. The causal cascade to multiple sclerosis: a model for MS pathogenesis. *PLoS One* 2009;4:e4565.
- [366] Levin LI, Munger KL, O'Reilly EJ, Falk KI, Ascherio A. Primary infection with the Epstein-Barr virus and risk of multiple sclerosis. *Ann Neurol* 2010;67:824–30.
- [367] Levin LI, Munger KL, Rubertone MV, Peck CA, Lennette ET, Spiegelman D, et al. Temporal relationship between elevation of Epstein-Barr virus antibody titers and initial onset of neurological symptoms in multiple sclerosis. *JAMA* 2005;293:2496–500.
- [368] Lindberg C, Andersen O, Vahlne A, Dalton M, Runmarker B. Epidemiological investigation of the association between infectious mononucleosis and multiple sclerosis. *Neuroepidemiology* 1991;10:62–5.
- [369] Lunemann JD, Edwards N, Muraro PA, Hayashi S, Cohen JI, Munz C, et al. Increased frequency and broadened specificity of latent EBV nuclear antigen-1-specific T cells in multiple sclerosis. *Brain* 2006;129:1493–506.
- [370] Lunemann JD, Huppke P, Roberts S, Bruck W, Gartner J, Munz C. Broadened and elevated humoral immune response to EBNA1 in pediatric multiple sclerosis. *Neurology* 2008;71:1033–5.
- [371] Lunemann JD, Jelcic I, Roberts S, Lutterotti A, Tackenberg B, Martin R, et al. EBNA1-specific T cells from patients with multiple sclerosis cross react with myelin antigens and co-produce IFN-gamma and IL-2. *J Exp Med* 2008;205:1763–73.

- [372] Operskalski EA, Visscher BR, Malmgren RM, Detels R. A case-control study of multiple sclerosis. *Neurology* 1989;39:825–9.
- [373] Peferoen LA, Lamers F, Lodder LN, Gerritsen WH, Huitinga I, Melief J, et al. Epstein Barr virus is not a characteristic feature in the central nervous system in established multiple sclerosis. *Brain* 2010;133:e137.
- [374] Sargsyan SA, Shearer AJ, Ritchie AM, Burgoon MP, Anderson S, Hemmer B, et al. Absence of Epstein-Barr virus in the brain and CSF of patients with multiple sclerosis. *Neurology* 2010;74:1127–35.
- [375] Serafini B, Rosicarelli B, Franciotta D, Magliozzi R, Reynolds R, Cinque P, et al. Dysregulated Epstein-Barr virus infection in the multiple sclerosis brain. *J Exp Med* 2007;204:2899–912.
- [376] Thorley-Lawson DA. Epstein-Barr virus: exploiting the immune system. *Nat Rev Immunol* 2001;1:75–82.
- [377] Warner HB, Carp RI. Multiple sclerosis and Epstein-Barr virus. *Lancet* 1981;2:1290.
- [378] Willis SN, Stadelmann C, Rodig SJ, Caron T, Gattenloehner S, Mallozzi SS, et al. Epstein-Barr virus infection is not a characteristic feature of multiple sclerosis brain. *Brain* 2009;132:3318–28.
- [379] Sriram S, Stratton CW, Yao S, Tharp A, Ding L, Bannan JD, et al. *Chlamydia pneumoniae* infection of the central nervous system in multiple sclerosis. *Ann Neurol* 1999;46:6–14.
- [380] Challoner PB, Smith KT, Parker JD, MacLeod DL, Coulter SN, Rose TM, et al. Plaque-associated expression of human herpesvirus 6 in multiple sclerosis. *Proc Natl Acad Sci U S A* 1995;92:7440–4.
- [381] Dockrell DH, Smith TF, Paya CV. Human herpesvirus 6. *Mayo Clin Proc* 1999;74:163–70.
- [382] Chmielewska-Badora J, Cisak E, Dutkiewicz J. Lyme borreliosis and multiple sclerosis: any connection? A seroepidemiological study. *Ann Agric Environ Med* 2000;7:141–3.
- [383] Mancuso R, Delbue S, Borghi E, Pagani E, Calvo MG, Caputo D, et al. Increased prevalence of varicella zoster virus DNA in cerebrospinal fluid from patients with multiple sclerosis. *J Med Virol* 2007;79:192–9.
- [384] Ordonez G, Pineda B, Garcia-Navarrete R, Sotelo J. Brief presence of varicella-zoster viral DNA in mononuclear cells during relapses of multiple sclerosis. *Arch Neurol* 2004;61:529–32.
- [385] Sotelo J, Martinez-Palomo A, Ordonez G, Pineda B. Varicella-zoster virus in cerebrospinal fluid at relapses of multiple sclerosis. *Ann Neurol* 2008;63:303–11.
- [386] Birnbaum G, Kotilinek L, Albrecht L. Spinal fluid lymphocytes from a subgroup of multiple sclerosis patients respond to mycobacterial antigens. *Ann Neurol* 1993;34:18–24.
- [387] Wroblewska Z, Gilden D, Devlin M, Huang ES, Rorke LB, Hamada T, et al. Cytomegalovirus isolation from a chimpanzee with acute demyelinating disease after inoculation of multiple sclerosis brain cells. *Infect Immun* 1979;25:1008–15.
- [388] Perron H, Garson JA, Bedin F, Beseme F, Paranhos-Baccala G, Komurian-Pradel F, et al. Molecular identification of a novel retrovirus repeatedly isolated from patients with multiple sclerosis. The Collaborative Research Group on Multiple Sclerosis. *Proc Natl Acad Sci U S A* 1997;94:7583–8.
- [389] Rasmussen HB, Geny C, Deforges L, Perron H, Tourtelotte W, Heltberg A, et al. Expression of endogenous retroviruses in blood mononuclear cells and brain tissue from multiple sclerosis patients. *Acta Neurol Scand Suppl* 1997;169:38–44.
- [390] Murray RS, Brown B, Brian D, Cabirac GF. Detection of coronavirus RNA and antigen in multiple sclerosis brain. *Ann Neurol* 1992;31:525–33.
- [391] Stewart JN, Mounir S, Talbot PJ. Human coronavirus gene expression in the brains of multiple sclerosis patients. *Virology* 1992;191:502–5.
- [392] Ferrante P, Omodeo-Zorini E, Caldarelli-Stefano R, Mediati M, Fainardi E, Granieri E, et al. Detection of JC virus DNA in cerebrospinal fluid from multiple sclerosis patients. *Mult Scler* 1998;4:49–54.
- [393] Stoner GL, Agostini HT, Ryschkeiwitsch CF, Baumhufner RW, Tourtelotte WW. Characterization of JC virus DNA amplified from urine of chronic progressive multiple sclerosis patients. *Mult Scler* 1996;1:193–9.
- [394] Forghani B, Cremer NE, Johnson KP, Ginsberg AH, Likosky WH. Viral antibodies in cerebrospinal fluid of multiple sclerosis and control patients: comparison between radioimmunoassay and conventional techniques. *J Clin Microbiol* 1978;7:63–9.
- [395] ter Meulen V, Koprowski H, Iwasaki Y, Kackell YM, Muller D. Fusion of cultured multiple-sclerosis brain cells with indicator cells: presence of nucleocapsids and virions and isolation of parainfluenza-type virus. *Lancet* 1972;2:1–5.
- [396] Haase AT, Ventura P, Gibbs Jr CJ, Tourtelotte WW. Measles virus nucleotide sequences: detection by hybridization in situ. *Science* 1981;212:672–5.
- [397] Jacobson S, Flerlage ML, McFarland HF. Impaired measles virus-specific cytotoxic T cell responses in multiple sclerosis. *J Exp Med* 1985;162:839–50.
- [398] Alperovitch A, Berr C, Cambon-Thomsen A, Puel J, Dugoujon JM, Ruidavets JB, et al. Viral antibody titers, immunogenetic markers, and their interrelations in multiple sclerosis patients and controls. *Hum Immunol* 1991;31:94–9.
- [399] Ortega-Hernandez OD, Levin NA, Altman A, Shoenfeld Y. Infectious agents in the pathogenesis of primary biliary cirrhosis. *Dis Markers* 2010;29:277–86.
- [400] Shigematsu H, Shimoda S, Nakamura M, Matsushita S, Nishimura Y, Sakamoto N, et al. Fine specificity of T cells reactive to human PDC-E2 163-176 peptide, the immunodominant autoantigen in primary biliary cirrhosis: implications for molecular mimicry and cross-recognition among mitochondrial autoantigens. *Hepatology* 2000;32:901–9.
- [401] Liu HY, Deng AM, Zhang J, Zhou Y, Yao DK, Tu XQ, et al. Correlation of *Chlamydia pneumoniae* infection with primary biliary cirrhosis. *World J Gastroenterol* Jul 14, 2005;11(26):4108–10.
- [402] Bogdanos DP, Pares A, Baum H, Caballeria L, Rigopoulou EI, Ma Y, et al. Disease-specific cross-reactivity between mimicking peptides of heat shock protein of *Mycobacterium gordonae* and dominant epitope of E2 subunit of pyruvate dehydrogenase is common in Spanish but not British patients with primary biliary cirrhosis. *J Autoimmun* 2004;22:353–62.
- [403] Vilagut L, Pares A, Vinas O, Vila J, Jimenez de Anta MT, Rodes J. Antibodies to mycobacterial 65-kD heat shock protein cross-react with the main mitochondrial antigens in patients with primary biliary cirrhosis. *Eur J Clin Invest* 1997;27:667–72.
- [404] Vilagut L, Vila J, Vinas O, Pares A, Gines A, Jimenez de Anta MT, et al. Cross-reactivity of anti-*Mycobacterium gordonae* antibodies with the major mitochondrial autoantigens in primary biliary cirrhosis. *J Hepatol* 1994;21:673–7.
- [405] Kita H, Matsumura S, He XS, Ansari AA, Lian ZX, Van de Water J, et al. Analysis of TCR antagonism and molecular mimicry of an HLA-A0201-restricted CTL epitope in primary biliary cirrhosis. *Hepatology* 2002;36:918–26.
- [406] Yamaguchi H, Miura H, Ohsumi K, Ishimi N, Taguchi H, Ishiyama N, et al. Detection and characterization of antibodies to bacterial heat-shock protein 60 in sera of patients with primary biliary cirrhosis. *Microbiol Immunol* 1994;38:483–7.
- [407] Nickowitz RE, Worman HJ. Autoantibodies from patients with primary biliary cirrhosis recognize a restricted region within the cytoplasmic tail of nuclear pore membrane glycoprotein Gp210. *J Exp Med* 1993;178:2237–42.
- [408] Stemerowicz R, Hopf U, Moller B, Wittenbrink C, Rodloff A, Reinhardt R, et al. Are antimicrobial antibodies in primary biliary cirrhosis induced by R(rough)-mutants of Enterobacteriaceae? *Lancet* 1988;2:1166–70.
- [409] Haruta I, Kikuchi K, Hashimoto E, Kato H, Hirota K, Kobayashi M, et al. A possible role of histone-like DNA-binding protein of *Streptococcus intermedius* in the pathogenesis of bile duct damage in primary biliary cirrhosis. *Clin Immunol* 2008;127:245–51.
- [410] Sayers TJ, Baum H. Possible cross-reactivity of human anti-mitochondrial antibodies with membrane vesicles of *Paracoccus denitrificans*. *Biochem Soc Trans* 1976;4:138–9.
- [411] Bogdanos DP, Koutsoumpas A, Baum H, Vergani D. *Borrelia burgdorferi*: a new self-mimicking trigger in primary biliary cirrhosis. *Dig Liver Dis* 2006;38:781–2 [author reply 2–3].
- [412] Harada K, Tsuneyama K, Sudo Y, Masuda S, Nakanuma Y. Molecular identification of bacterial 16S ribosomal RNA gene in liver tissue of primary biliary cirrhosis: is *Propionibacterium acnes* involved in granuloma formation? *Hepatology* 2001;33:530–6.
- [413] Berg CP, Kannan TR, Klein R, Gregor M, Baseman JB, Wesselborg S, et al. *Mycoplasma* antigens as a possible trigger for the induction of antimicrobial antibodies in primary biliary cirrhosis. *Liver Int* 2009;29:797–809.
- [414] Jan G, Le Henaff M, Fontenelle C, Wroblewski H. Biochemical and antigenic characterisation of *Mycoplasma gallisepticum* membrane proteins P52 and P67 (pMGA). *Arch Microbiol* 2001;177:81–90.
- [415] Ninomiya M, Ueno Y, Shimosegawa T. PBC: animal models of cholangiopathies and possible endogenous viral infections. *Int J Hepatol* 2012;2012 (649290). <http://dx.doi.org/10.1155/2012/649290> (Electronic publication ahead of print 2011 Aug 8).
- [416] Zhang G, Chen M, Graham D, Subsin B, McDougall C, Gilady S, et al. Mouse mammary tumor virus in anti-mitochondrial antibody producing mouse models. *J Hepatol* 2011;55:876–84.
- [417] Morshed SA, Nishioka M, Saito I, Komiyama K, Moro I. Increased expression of Epstein-Barr virus in primary biliary cirrhosis patients. *Gastroenterol Jpn* 1992;27:751–8.
- [418] Uzoegwu P, Baum H, Williamson J. Correlation of oligomycin-sensitive ATPase activity in trypanosomes with their content of an antigen to primary biliary cirrhosis. *Cell Biol Int Rep* 1984;8:981–6.
- [419] Uzoegwu PN, Baum H, Williamson J. The occurrence and localization in trypanosomes and other endo-parasites of an antigen cross-reacting with mitochondrial antibodies of primary biliary cirrhosis. *Comp Biochem Physiol B* 1987;88:1181–9.
- [420] Sakly W, Jeddi M, Ghedira I. Anti-*Saccharomyces cerevisiae* antibodies in primary biliary cirrhosis. *Dig Dis Sci* 2008;53:1983–7.