

Research Article

Protective Effect of a Mixture of *Astragalus membranaceus* and *Lithospermum erythrorhizon* Extract against Hepatic Steatosis in High Fat Diet-Induced Nonalcoholic Fatty Liver Disease Mice

Doo Jin Choi ¹, Seong Cheol Kim,² Gi Eun Park,² Bo-Ram Choi,¹ Dae Young Lee,¹ Young-Seob Lee ¹, Sung-Bum Park,³ Yong Il Park ², and Geum Soog Kim ¹

¹Department of Herbal Crop Research, National Institute of Horticultural and Herbal Science, RDA, Eumseong 27709, Republic of Korea

²Department of Biotechnology, The Catholic University of Korea, Bucheon, Gyeonggi-do 14662, Republic of Korea

³Dong IL PharmTec Co. Ltd., Gangnam, Seoul 06296, Republic of Korea

Correspondence should be addressed to Yong Il Park; yongil382@catholic.ac.kr and Geum Soog Kim; kings0725@korea.kr

Received 10 October 2019; Revised 21 January 2020; Accepted 14 February 2020; Published 19 March 2020

Academic Editor: Sokcheon Pak

Copyright © 2020 Doo Jin Choi et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The present study aimed to evaluate the potential synergistic and protective effects of ALM16, a mixture of *Astragalus membranaceus* (AM) and *Lithospermum erythrorhizon* (LE) extract in a ratio of 7 : 3, against hepatic steatosis in high fat diet (HFD)-induced nonalcoholic fatty liver disease (NAFLD) mice. Forty-eight mice were randomly divided into eight groups and orally administered daily for 6 weeks with a normal diet (ND) or high fat diet alone (HFD), HFD with AM (HFD + 100 mg/kg AM extract), HFD with LE (HFD + 100 mg/kg LE extract), HFD with ALM16 (HFD + 50, 100, and 200 mg/kg ALM16), or HFD with MT (HFD + 100 mg/kg Milk thistle extract) as a positive control. ALM16 significantly decreased the body and liver weight, serum and hepatic lipid profiles, including triglyceride (TG), total cholesterol (TC), high-density lipoprotein-cholesterol (HDL), and low-density lipoprotein-cholesterol (LDL), and serum glucose levels, compared to the HFD group. Moreover, ALM16 significantly ameliorated the HFD-induced increased hepatic injury markers, including aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), lactate dehydrogenase (LDH), and gamma-glutamyltransferase (GGT)-1. Furthermore, as compared to the mice fed HFD alone, ALM16 increased the levels of phosphorylated AMP-activated protein kinase (p-AMPK) and acetyl-CoA carboxylase (p-ACC), thereby upregulating the expression of carnitine palmitoyltransferase (CPT)-1 and downregulating the expression of sterol regulatory element-binding protein (SREBP)-1c and fatty acid synthase (FAS). These results demonstrated that ALM16 markedly inhibited HFD-induced hepatic steatosis in NAFLD mice by modulating AMPK and ACC signaling pathways, and may be more effective than the single extracts of AM or LE.

1. Introduction

Nonalcoholic fatty liver disease (NAFLD) is one of the common chronic liver diseases in metabolic syndrome, resulting from the hepatic lipid accumulation of >5% of liver weight [1]. The liver is the major contributor to whole-body lipogenesis, gluconeogenesis, and cholesterol metabolism [2, 3]. The development of NAFLD is characterized by abnormal liver function and can progress to more severe liver diseases such as nonalcoholic steatohepatitis (NASH), fibrosis, and cirrhosis [4, 5]. NAFLD has become a critical

health concern worldwide owing to its increased morbidity and mortality. However, there are no approved agents currently available for the treatment of NAFLD [6]. NAFLD patients generally require long-term treatment, which can potentially induce serious or diverse side effects. Therefore, the identification of potential therapeutic compounds with better biological efficacy and safety, and fewer side effects is imperative [7].

Until now, the pathogenesis of NAFLD remained poorly understood; the ‘two-hit’ concept is generally recognized as the most important theory. The ‘first hit’ is the excessive

accumulation of triglycerides (TG) in hepatocytes, and ‘second hits’ are associated with increased inflammatory adipokines/cytokines, such as adiponectin, tumor necrosis factor (TNF)- α and interleukin (IL)-6, oxidative stress, and mitochondrial impairment [8, 9]. Accordingly, therapeutic strategies for NAFLD have focused on identifying potential agents that can exert effects to prevent hepatic steatosis and inflammation.

The AMP-activated protein kinase (AMPK) signaling pathway plays an important role in maintaining cellular energy metabolism [10]. AMPK, involved in mediating lipogenesis and fatty acid oxidation in the liver, has become a major target for the treatment and/or prevention of NAFLD [11]. Activation of AMPK mediates energy metabolism through the suppression of acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS) expression, via the decreased transcriptional activation of the sterol regulatory element-binding protein (SREBP)-1c [12]. Inactivation of ACC by activated AMPK leads to the reduced synthesis of malonyl-CoA, which decreases fatty acid synthesis and increases mitochondrial fatty acid oxidation via regulation of carnitine palmitoyltransferase (CPT)-1 [13].

Recently, many medicinal herbs have gained attention as potential therapeutic agents in the treatment of various human diseases. In traditional oriental medicine, major medicinal herbs are frequently used in combination with other herbs for the treatment of chronic or complex diseases, including liver disease [14]. Herb-herb combinations have long been used in Oriental medicine for thousands of years, yet scientific evidence of their therapeutic benefits was lacking. Accordingly, scientific interest to study the combination therapy for the treatment of diseases is increasing since scientific literature demonstrating evidence of enhanced efficacy, thus called ‘synergistic effects’, for certain diseases when treated in a combination of different herbs than treated individually, is now piling up [15–17].

Astragalus membranaceus (AM), called Hwanggi in Korea, is an edible herb widely consumed for various medical symptoms owing to its diverse pharmaceutical activities, including modulation of the immune system, and antitumor properties [18, 19]. AM is enriched with iso-flavonoids, triterpene saponins, and polysaccharides [20]. Among them, calycosin and calycosin-7-*O*- β -D-glucoside are used as chemical markers for the comprehensive quality evaluation of AM, due to the presence of various biological activities [21, 22]. Previously, calycosin was shown to exert hepatoprotective effects in acute liver injury and chronic liver diseases [23, 24]. The AM extract has been frequently consumed in combination with various other herbal mixtures in traditional folk medicines in Korea due to its many health-beneficial activities and synergistic effects under different pathological conditions, *in vitro* and *in vivo* [25, 26].

Lithospermum erythrorhizon (LE) has also been used in traditional medicine for its various pharmacological effects. LE contains naphthoquinones, including shikonin and its derivatives, identified as the major phytochemicals [27]. Recent studies have reported the various biological activities of LE, including anti-inflammatory, antiobesity, and

antiprogenic activities [28–30]. Especially, Gwon et al. reported that ethanol extract of LE, which contains acetylshikonin as the main compound, exerted antiobesity effect through inhibition of lipogenic and adipogenic gene expression [29]. They also showed that the same sample prevented the hepatic fat accumulation, although detailed underlying action mechanism on its inhibitory activity against hepatic fat accumulation was not extensively addressed [29]. Like AM, LE has also garnered much attention as a complementary herb that can act simultaneously on various targets in diseased states [31, 32]. A study by Roh [33] reported that Gangjihwan, a polyherbal drug composed of LE as the main ingredient, demonstrated a beneficial effect in NAFLD mice via the regulation of the genes involved in lipid metabolism and inflammation.

In our previous study, we selected the effective ratio (7:3, w/w) of a combination of AM and LE extracts to measure the activities of matrix metalloproteinases (MMP) –1, –3, and –13 [34]. Subsequently, we reported that a mixture of these two herbal extracts at the ratio of 7:3 (AM:LE, 7:3, w/w; referred to as ALM16), as a new herbal composition, has an antiosteoarthritis activity based on synergistic action [35]. However, despite the reported health benefits and potential synergistic actions associated with these two herbs, the hepatoprotective and synergistic effects of a mixture of AM and LE have not been explored yet. In this regard, to extend the possibility of the combined mixture of AM and LE as a new and/or improved therapeutic modality to other health problems beyond its antiosteoarthritis activity, thereby increasing the medicinal applicability of these two herbs, this study was aimed to evaluate a possible positive effect of ALM16 against NAFLD. For this purpose, this study was designed to investigate the protective effects of ALM16 on hepatic steatosis using the high fat diet (HFD)-induced NAFLD mouse model. The underlying mechanisms of the antihepatic steatosis activity in the NAFLD model are also discussed. In addition, we examined whether the combined treatment with AM and LE, rather than the use of individual herb alone, could possibly show an enhanced effect against NAFLD. For comparison, *Silybum marianum* (milk thistle) extract (MT), known for its antisteatosis activity, was used as a positive control in the *in vivo* experiments [36].

2. Materials and Methods

2.1. Ethics Statement. All mice were treated in accordance with the Guide for the Care and Use of Laboratory Animals, as approved by the Institutional Animal Care and Use Committee of the Catholic University of Korea (Proved No. 018–011), and all efforts have been made to minimize the number of animals used.

2.2. Preparation of Herbal Mixture (ALM16). AM and LE were collected from Jecheon, Chungcheongbuk-do, Korea. AM and LE, which were assigned voucher specimen number (MPS005087 and MPS004961, respectively), were authenticated by Dr. Jeong Hoon Lee, Ph.D., from the National Institute of Horticultural and Herbal Science, Rural

Development Administration, Korea. The mixture of AM and LE extracts (ALM16) was prepared according to the method described in our previous study [35]. Briefly, the dried roots of AM and LE were extracted with 50% and 70% aqueous fermented ethanol at 80°C for 4 h, respectively. Individual AM and LE dried extracts were mixed well in a ratio of 7 : 3 (w/w), to prepare the ALM16. The appropriate amounts of AM, LE, and ALM16 were dissolved in phosphate-buffered saline (PBS) for the experiments, and stored at -20°C.

2.3. Animal Experimental Design. Male C57BL/6 mice (5 weeks old), purchased from the ORIENT BIO INC (Gyeonggi-do, Korea), were housed in the specific pathogen-free (SPF) facility, and maintained under controlled temperatures (22–23°C), on a 12/12 h light-dark cycle in cages. After the acclimatization period, 48 male C57BL/6 mice (initially weighing approximately 19.3–20.3 g) were randomly divided into eight groups ($n=6$) as follows: (1) normal diet group (ND, 16% calories from fat), (2) high fat diet group (HFD, 60% calories from fat), (3) HFD + 100 mg/kg AM extract group (AM), (4) HFD + 100 mg/kg LE extract (LE), (5) HFD + 50 mg/kg ALM16 (ALM16-L), (6) HFD + 100 mg/kg ALM16 (ALM16-M), (7) HFD + 200 mg/kg ALM16 (ALM16-H), and (8) HFD + 100 mg/kg Milk thistle extract group (MT, as a positive control group). After 7 weeks of HFD-induced NAFLD, all samples were dissolved in PBS and orally administered daily for 6 weeks. The groups fed with ND or HFD only were administered PBS. Body weights and food intake were measured every week. At the end of the experimental period, all mice underwent fasting for 12 h and were then anesthetized using a mixture of Zoletil (50 mg/ml, Virbac, Carros, France)/Rompun (23.32 mg/ml, Bayer, Germany) (1 : 3). Blood samples were collected by cardiac puncture. The liver tissues were then collected and weighed.

2.4. Serum and Tissue Biochemical Analysis. Serum levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), triglyceride (TG), total cholesterol (TC), high-density lipoprotein-cholesterol (HDL), low-density lipoprotein-cholesterol (LDL), and glucose were measured using an AU680 Automatic Analyzer (Beckman coulter, CA, USA), and lactate dehydrogenase (LDH) was measured using a commercial LDH assay kit (Sigma, St. Louis, MO, USA). Lipid in the liver tissue was extracted using modified methods, as previously described [37]. Briefly, stored liver tissues were homogenized in chloroform-methanol-distilled water (2 : 1 : 1, v/v/v) using a homogenizer for 1 min, and then centrifuged at 13,000 rpm for 15 min at 4°C. The solvent (chloroform) phase obtained from the extraction was dried and dissolved in methanol, and then the hepatic TG levels were measured using a triglyceride assay kit (Cayman Chemical, Ann Arbor, MI, USA).

2.5. Western Blot Analysis. The liver tissues were homogenized using a tissue grinder (Nippon genetic, Germany), with 500 μ l of the lysis buffer for 30 s. The supernatant was

then obtained by repeated centrifugation for 15 min at 13,000 rpm, at 4°C. The protein concentrations of lysates were determined using the Bio-Rad protein assay reagent (Bio-Rad, Hercules, GA, USA). Equal amounts of protein were separated on 6–13% sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis (PAGE) and transferred onto a nitrocellulose membrane. Membranes were blocked using a 5% nonfat milk blocking solution for 2 h. The membranes were incubated with the primary antibodies against AMP-activated protein kinase (AMPK), phospho-AMPK, acetyl-CoA carboxylase (ACC), phospho-ACC, FAS (Cell Signaling Technology, Beverly, MA, USA), CPT-1, SREBP-1c (Santa Cruz Biotechnology, Dallas, TX, USA) and gamma-glutamyltransferase (GGT)-1 (Abcam, Cambridge, MA, UK) at 4°C overnight, washed 3 times with Tris-buffered saline containing Tween 20 (TBST) for 5 min and then incubated with HRP-conjugated secondary antibodies for 1 h. Immunoreactive bands were then developed using a Chemiluminescence detection reagent (Pierce Biotechnology, Rockford, IL) and exposed to X-ray film (Konica, Tokyo, Japan). β -Actin was used as a loading control. The bands were quantified by densitometric analysis with Image J software.

2.6. Histopathological Analysis. To investigate the effect of ALM16 on hepatic steatosis induced by HFD in mice, liver tissues were examined by histopathological analysis. The liver tissue was fixed in 10% buffered formalin for 48 h, embedded in paraffin, and sectioned into 3- μ m-thick sections. The liver sections were stained with hematoxylin and eosin (H&E) for histological analysis. Images were acquired using a light microscope (Olympus, Tokyo, Japan). Histopathological examinations were performed by a board certified toxicologic pathologist, blinded to the treatment strategy.

2.7. Statistical Analysis. All values are expressed as mean \pm standard error of mean (SEM). The results were evaluated using one-way ANOVA with Dunnett's multiple comparison test using Graph-Pad Prism® software version 5.01 (San Diego, CA, USA). The differences between the data from the HFD and sample-treated groups were considered statistically significant when $P < 0.05$.

3. Results

3.1. ALM16 Inhibits Hepatic Steatosis in the Liver of HFD-Fed Mice. The body and liver weights in mice are shown in Figures 1(a) and 1(b). Both body ($P < 0.001$) and liver weights ($P < 0.01$) significantly increased in the HFD group, compared to the ND group. However, the body weight ($P < 0.001$) and liver weight ($P < 0.001$) in ALM16-treated groups (50, 100 and 200 mg/kg) significantly decreased by 30.8%, 32.8%, and 24.8%, respectively, compared to the HFD group. During the experiment period, no significant differences in food intake were observed between the HFD group (2.8 g/day/mice) and all sample-treated groups (2.5–2.8 g/day/mice, Supplementary Figure 1). AM, LE, and

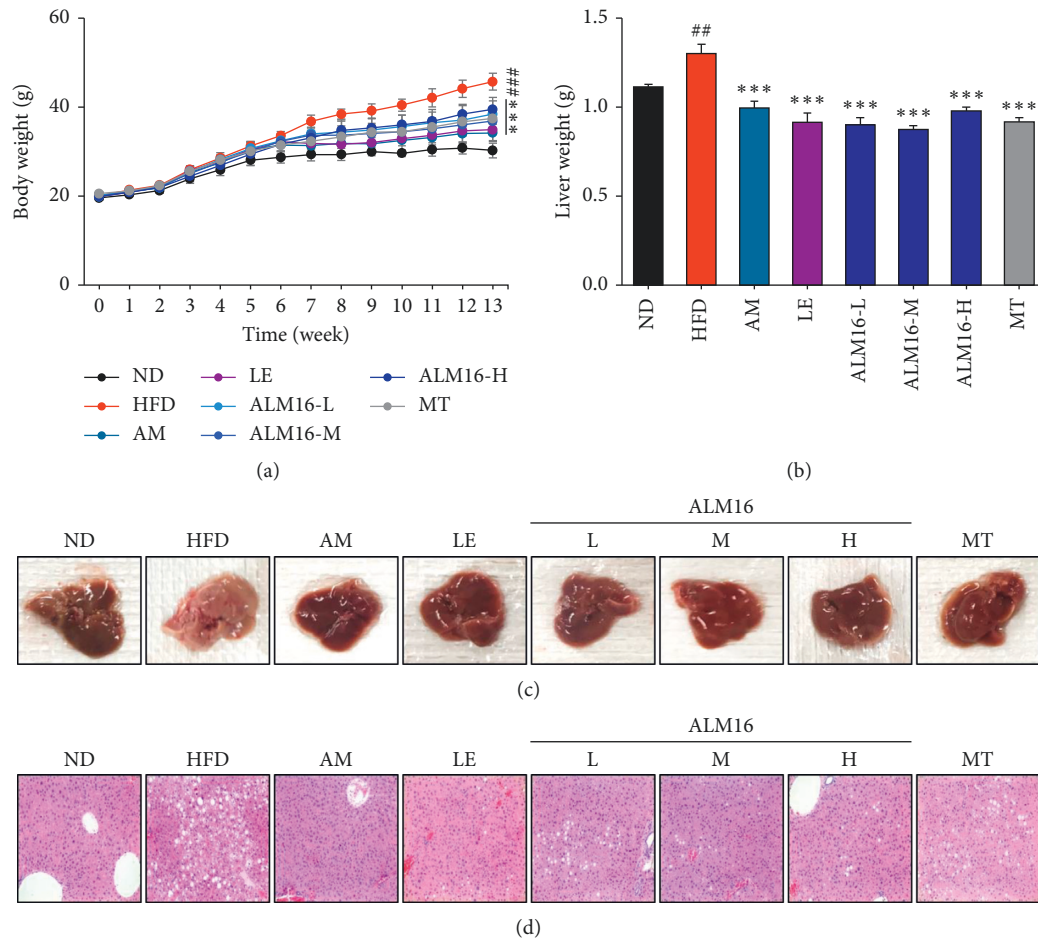


FIGURE 1: Effects of ALM16 and individual extracts (AM and LE) on hepatic steatosis in the liver of HFD-fed mice. (a) Body weight and (b) Liver weight ($n = 6$). (c) Representative macroscopic images of the liver. (d) Representative histopathological images of the liver tissue, stained with hematoxylin and eosin (H&E), of HFD-fed mice (magnification, 200x). Data are presented as mean \pm standard error of the mean (SEM). * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ vs ND group; * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ vs HFD group. ND: normal diet; HFD: high fat diet; AM: HFD + AM extract (100 mg/kg); LE: HFD + LE extract (100 mg/kg); ALM16-L: HFD + ALM16 (50 mg/kg); ALM16-M: HFD + ALM16 (100 mg/kg); ALM16-H: HFD + ALM16 (200 mg/kg); MT: HFD + milk thistle extract (100 mg/kg).

MT also significantly inhibited liver weight gain by 23.5% ($P < 0.001$), 29.7% ($P < 0.001$), and 29.4% ($P < 0.001$), respectively, compared with the HFD group. Although ALM16 demonstrated similar inhibitory effects on body weight gain, it inhibited liver weight gain more effectively compared with that in AM, LE, or MT-treated groups. In addition, the liver tissue of the HFD group was pale and yellow in appearance with severe accumulation of lipid droplets compared to that of the ND group which showed normal hepatic conditions (Figures 1(c) and 1(d)). However, a drastic decrease in hepatic lipid deposition was observed in all sample-treated groups (Figures 1(c) and 1(d)). The ALM16 (100 mg/kg) group demonstrated mild steatosis in the liver tissue, similar to the AM and LE-treated groups, but was more effective than MT (100 mg/kg) group (Figures 1(c) and 1(d)). In the ALM16 groups, although no dose-dependent differences were observed, the group administered 100 mg/kg showed greater effective amelioration of hepatic steatosis than those groups treated with 50 and 200 mg/kg ALM16. These results suggested that ALM16 (100 mg/kg) has a strong inhibitory effect against HFD-induced hepatic steatosis.

3.2. Effects of ALM16 on Liver Damage Markers in HFD-Fed Mice. The hepatic injury caused by HFD and the protective effects of ALM16 were assessed by measuring the levels of hepatic biochemical markers in the serum and liver. As shown in Figures 2(a)–2(d), the serum levels of AST ($P < 0.05$), ALT ($P < 0.001$), ALP, and LDH increased in the HFD group, compared to those in ND group, indicating that hepatic damage was induced by HFD administration. However, these HFD-induced elevated serum levels of AST, ALT, ALP, and LDH were markedly reduced following the administration of ALM16 (100 mg/kg) by 54.0% ($P < 0.05$), 59.9% ($P < 0.001$), 30.0% ($P < 0.01$), and 63.7% ($P < 0.05$), respectively, compared with the levels in the HFD group. Furthermore, serum levels of these markers were much lower in the ALM16-treated group, especially the ALM16-M (100 mg/kg)-treated group, compared to the groups treated with AM, LE, or MT alone. The results demonstrated that the HFD-induced hepatic damage is more attenuated by ALM16 than in individual extracts (AM or LE). Additionally, the hepatic GGT-1 level significantly increased in the HFD group compared with that in the ND group (Figure 2(e)),

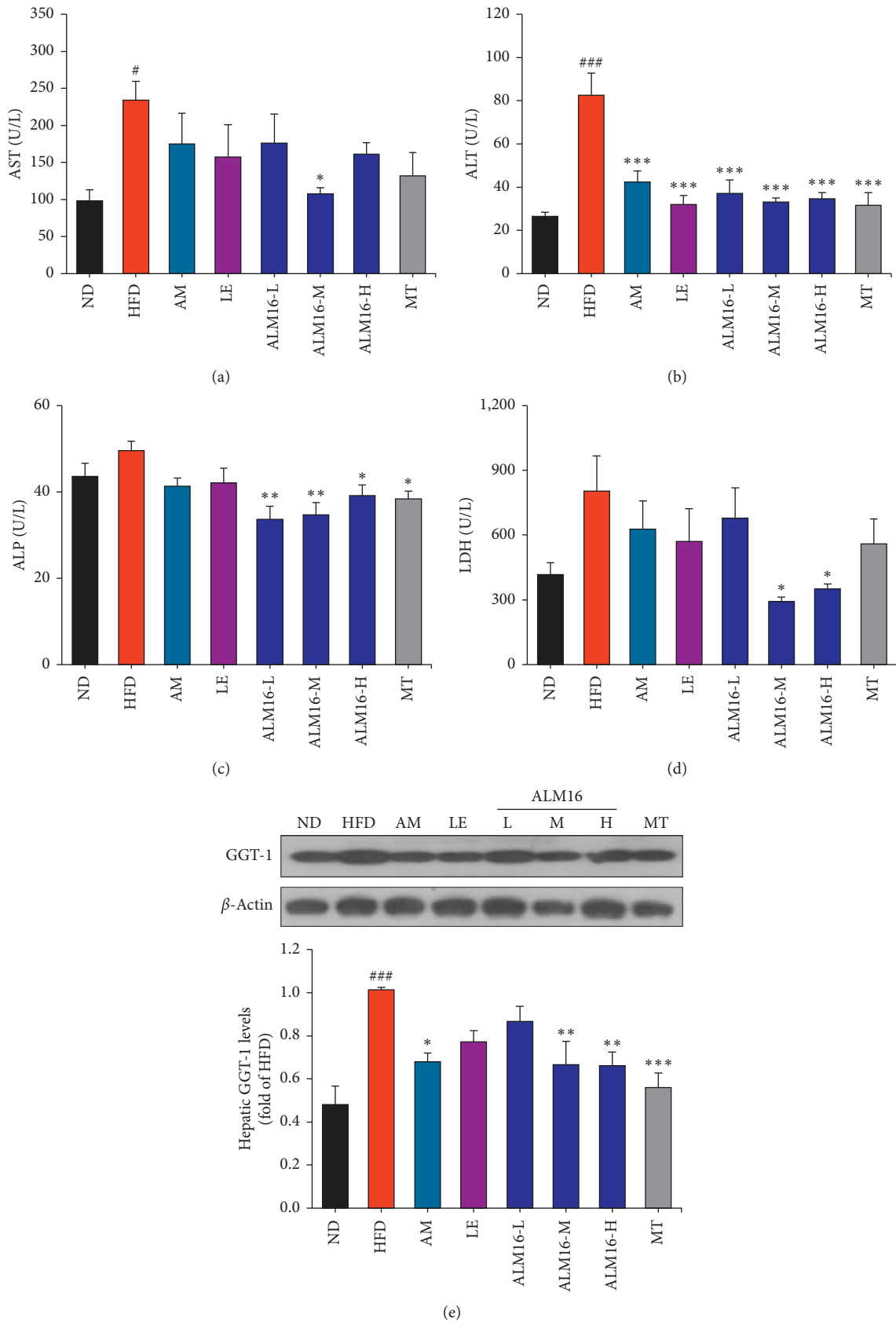


FIGURE 2: Effects of ALM16 and individual extracts (AM and LE) on biochemical parameters in the serum of NAFLD mice. Serum levels of (a) AST, (b) ALT, (c) ALP, and (d) LDH in HFD-fed mice ($n = 5$). (e) Protein expression levels of hepatic GGT-1 ($n = 4$). Data are expressed as mean \pm standard error of the mean (SEM). [#] $P < 0.05$, ^{**} $P < 0.01$ and ^{###} $P < 0.001$ vs ND group; ^{*} $P < 0.05$, ^{**} $P < 0.01$, and ^{***} $P < 0.001$ vs HFD group. ND: normal diet; HFD: high fat diet; AM: HFD + AM extract (100 mg/kg); LE: HFD + LE extract (100 mg/kg); ALM16-L: HFD + ALM16 (50 mg/kg); ALM16-M: HFD + ALM16 (100 mg/kg); ALM16-H: HFD + ALM16 (200 mg/kg); MT: HFD + milk thistle extract (100 mg/kg).

($P < 0.001$). In comparison with the hepatic GGT-1 level of HFD group, AM and LE decreased the hepatic GGT-1 level by 33.1% ($P < 0.05$) and 23.9%, respectively, and ALM16 (50, 100 and 200 mg/kg) treatment also significantly reduced the GGT-1 level by 14.6%, 34.4% ($P < 0.01$), and 34.8% ($P < 0.01$), respectively. These results clearly suggest that ALM16 ameliorates the HFD-induced liver damage and is more effective than AM or LE extract alone.

3.3. Effects of ALM16 on Serum and Liver Lipids in HFD-Fed Mice. As shown in Figure 3, ALM16 (100 mg/kg) significantly reduced the serum and hepatic TG (Figures 3(a) and 3(b)), serum TC (Figure 3(d)), HDL (Figure 3(e)), and LDL (Figure 3(f)) levels by 44.1%, 56.5%, 21.6%, 14.5%, and 43.7%, respectively, compared to the HFD group. Moreover, the elevated serum glucose level in the HFD group significantly decreased in the ALM16 (100 mg/kg)-treated group by 51.2% compared with the HFD group (Figure 3(c)). Although the serum TC, HDL, and LDL levels, in ALM16 (100 mg/kg) group, were similar to the other sample groups, the serum and hepatic TG and glucose levels were more significantly reduced by ALM16 compared to the other sample groups.

3.4. ALM16 Increases Phosphorylation of AMPK and ACC in the Liver of HFD-Fed Mice. As presented in Figure 4(a) and 4(b), phosphorylated AMPK (p-AMPK)/AMPK levels decreased in the HFD group compared with the ND group. The AM group demonstrated a 2.5-fold ($P < 0.05$) increase in the p-AMPK/AMPK levels compared with that of the HFD group (1.0-fold), but the LE-treated group showed no significant difference. ALM16 (50, 100 and 200 mg/kg) groups significantly increased p-AMPK/AMPK levels by 2.8-fold ($P < 0.01$), 3.4-fold ($P < 0.001$) and 2.5-fold ($P < 0.01$), respectively, compared to the HFD group. Moreover, compared to the HFD group, levels of phosphorylated ACC (p-ACC)/ACC increased in the AM and LE groups with no significant difference (Figure 4(c)). However, ALM16 (100 mg/kg) significantly increased the p-ACC/ACC levels by 4.2-fold ($P < 0.001$), compared to the HFD group. These results showed that ALM16 has a higher ability to activate the AMPK/ACC pathway than individual extract, possibly based on synergistic action. Thus, administration of the combined mixture of these two herbal extracts (AM:LE, 7:3, w/w; referred to as ALM16) was shown to be more effective against AMPK/ACC pathway-related NAFLD, compared with the results from individual herb (AM or LE) treatment.

3.5. Effects of ALM16 on the Protein Expression of CPT-1, SREBP-1c, and FAS. The protein expressions of CPT-1, SREBP-1c, and FAS in the liver of HFD-fed mice are shown in Figure 5(a). In the HFD group, the protein expression of CPT-1 decreased, and the protein levels of SREBP-1c ($P < 0.01$) and FAS ($P < 0.001$) significantly increased, compared to the ND group. The reduced CPT-1 level in the HFD-fed mice was increased in the ALM16 ($P < 0.05$) groups, but the AM and LE group showed no significant change (Figure 5(b)). Compared to the HFD group, LE and ALM16 (100 mg/kg) groups

significantly suppressed the protein expression of SREBP-1c by 42.5% ($P < 0.05$) and 54.7% ($P < 0.01$), respectively, but the AM group revealed no significant change (Figure 5(c)). Furthermore, as shown in Figure 5(d), all samples (AM, LE, and ALM16) at a dose of 100 mg/kg significantly decreased the FAS protein expression by 63.0% ($P < 0.001$), 46.8% ($P < 0.01$), and 74.7% ($P < 0.001$), respectively. MT (100 mg/kg) increased the protein expression of CPT-1 by 2.0-fold ($P < 0.01$) and suppressed the protein expression of SREBP-1c and FAS by 68.8% ($P < 0.001$) and 69.4% ($P < 0.001$), respectively, compared to the HFD group.

4. Discussion

Nonalcoholic fatty liver disease (NAFLD) is defined as impaired lipid and glucose metabolism caused by abnormal liver function, such as hepatic steatosis presented as excessive lipid accumulation in the liver [8]. However, no clinically effective therapeutic agents for the treatment of NAFLD are currently available [38]. Recent studies have reported that many herbal medicines for the treatment of NAFLD tend to use multiple herbs in complex formulations, owing to multitargeting and multifunctional properties, complementary interactions, and synergistic effects demonstrating better or enhanced efficacies over treatment with individual herbs [39–41].

In our previous study, we demonstrated that ALM16, mainly composed of calycosin, calycosin-7-*O*- β -D-glucoside, and lithospermic acid as index compounds, exerts a synergistic chondroprotective effect in a monosodium iodoacetate (MIA)-induced osteoarthritis rat model [35]. A previous study by Duan [24] reported that calycosin, a major component of *Astragalus radix*, ameliorates HFD-induced NAFLD by inhibiting gluconeogenesis through the modulation of Farnesoid X receptor (FXR) activation. In addition, shikonin, which is one of the naphthoquinones in LE, suppresses lipogenesis by increasing the phosphorylated AMPK in hepatocytes [30]. Based on these findings, we postulated that ALM16 would possess a certain level of cartilage protective effect and could exert multiple biological activities under various pathological conditions. To verify this hypothesis, the present study investigated the potential beneficial and synergistic effects of ALM16 on NAFLD using the HFD-fed mouse model. HFD feeding reportedly increases body and liver weights and elevates the levels of lipid parameters and hepatic injury markers in mice, resulting in the development of hepatic steatosis and injury [42]. Thus, rodent NAFLD models induced with dietary interventions (high fat, carbohydrate, and/or cholesterol diets) have generally been accepted as essential tools for research in NAFLD, as they mimic the pathophysiological and phenotypical symptoms in human NAFLD [43].

In this study, the NAFLD mouse model was established by feeding the HFD for 7 weeks. The resulting NAFLD mice reported increased body and liver weights and elevated levels of serum and hepatic lipid profiles. In HFD-fed mice, our results demonstrated that these increased body and liver weights and elevated levels of serum and hepatic lipid parameters were significantly decreased following ALM16 administration. In comparison to AM and LE, ALM16 (100 mg/kg)-treated group demonstrated relatively lower serum TG levels, whereas similar

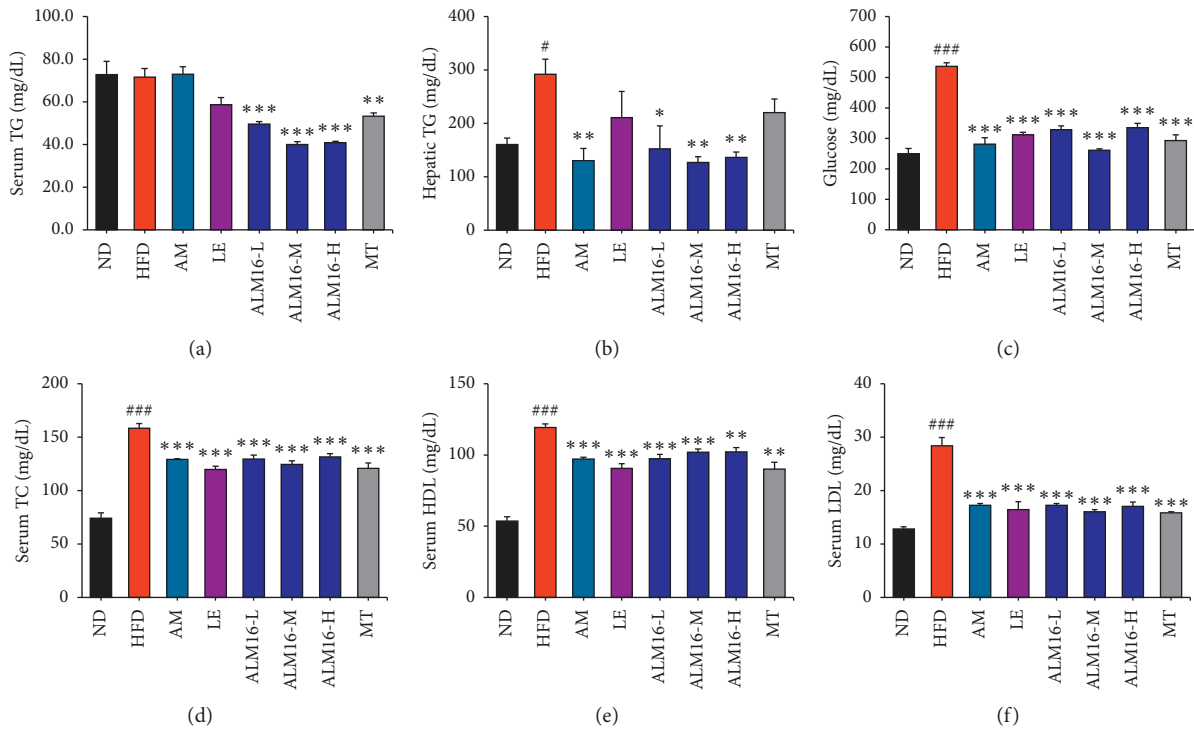


FIGURE 3: Effects of ALM16 and individual extracts (AM and LE) on serum lipids and glucose levels in HFD-fed mice. (a) Serum TG and (b) hepatic TG levels. (c) Serum glucose levels. Serum levels of (d) TC, (e) HDL, and (f) LDL cholesterol in HFD-fed mice ($n = 5$). Values are presented as mean \pm standard error of the mean (SEM). $^{\#}P < 0.05$, $^{\#\#}P < 0.01$, and $^{\#\#\#}P < 0.001$ vs ND group; $^*P < 0.05$, $^{**}P < 0.01$, and $^{***}P < 0.001$ vs HFD group. ND: normal diet; HFD: high fat diet; AM: HFD + AM extract (100 mg/kg); LE: HFD + LE extract (100 mg/kg); ALM16-L: HFD + ALM16 (50 mg/kg); ALM16-M: HFD + ALM16 (100 mg/kg); ALM16-H: HFD + ALM16 (200 mg/kg); MT: HFD + milk thistle extract (100 mg/kg).

effects with AM or LE, on the hepatic TG, serum TC, HDL, and LDL levels, were observed. HFD feeding is also known to be implicated in the impairment of hepatic glucose metabolism by reducing the ability of insulin to inhibit gluconeogenesis in the liver [44, 45]. Previous studies have reported that HFD feeding in rodents elevates serum glucose levels due to an increase in the hepatic insulin resistance [46, 47]. In this study, ALM16 (100 mg/kg) also effectively reversed the highly elevated serum glucose levels in HFD-fed mice. These results indicated that ALM16 could improve hepatic lipid profiles and glucose metabolism in HFD-fed mice.

It is well known that the serum levels of AST, ALT, ALP, and LDH, which are major enzymes presented in hepatocytes, increase following hepatocellular injury [48]. Among them, especially ALT, is a sensitive biomarker as an indicator associated with NAFLD [49]. In many clinical studies on NAFLD, an increased level of ALT has been considered as an independent predictor [50]. It is also generally accepted that GGT, which is an enzyme abundant in the liver, is elevated as a result of obesity and liver damage, and thus regarded as one of the predictors of liver mortality [51]. ALM16 administration significantly decreased the elevated serum AST, ALT, ALP, and LDH levels caused by HFD and reduced GGT-1 levels in the liver tissue of HFD-fed mice. In particular, ALM16 (100 mg/kg) was more effective on the serum levels of AST, ALP, and LDH than AM or LE, although the effects on the serum ALT and hepatic GGT-1 levels were similar to AM or LE groups. Moreover,

histopathological analysis of liver tissues with H&E showed that ALM16 administration markedly attenuated the excessive formation and accumulation of lipid droplets in hepatocytes. Hepatic steatosis, the first hallmark of NAFLD, is known to be characterized by excessive accumulation of lipid in hepatocytes [52]. Several previous studies have provided evidence regarding the hepatoprotective properties against HFD-induced NAFLD by reducing the liver damage markers and inhibiting the accumulation of lipid droplets in the liver tissues of HFD-fed rodents [53, 54]. In parallel, our results demonstrated that ALM16 can exert protective effects and synergistic action against hepatic steatosis and liver injury in HFD-fed mice.

We further investigated the molecular mechanism by which ALM16 provides a protective effect against hepatic steatosis by monitoring the lipid metabolism (lipogenesis and fatty acid oxidation) associated proteins and AMPK and ACC pathways. AMPK is considered as a major therapeutic target for the treatment of NAFLD as it plays an important role in the control of lipid metabolism by regulating lipogenesis and fatty acid oxidation in the liver [12]. The activation of AMPK induces the inactivation of ACC, which decreases the conversion of acetyl-CoA to malonyl-CoA, thereby increasing CPT-1 expression [55]. Increased CPT-1 expression stimulates fatty acid oxidation, and consequently reduces hepatic lipid accumulation [56]. Our study revealed that ALM16 markedly increased the phosphorylation of AMPK and ACC in the liver tissues of HFD-fed mice. Based

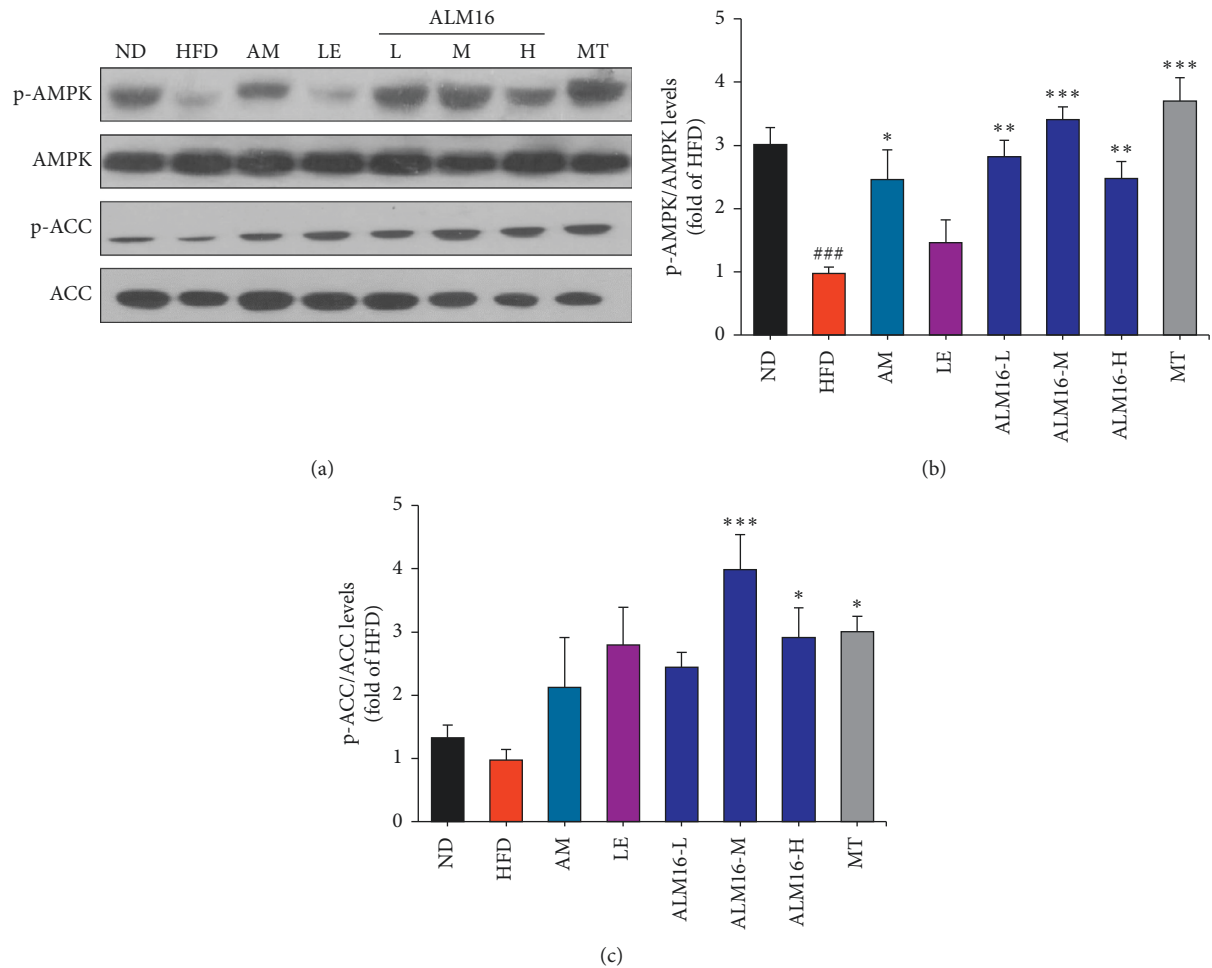


FIGURE 4: Effects of ALM16 and individual extracts (AM and LE) on hepatic AMPK and ACC phosphorylation in HFD-fed mice. (a) Western blot analysis of AMPK, p-AMPK, ACC, and p-ACC in the liver tissues of HFD-fed mice. Bands were quantified as the ratio of (b) p-AMPK/AMPK and (c) p-ACC/ACC, respectively, with values normalized to the HFD group ($n = 4$). Values are presented as mean \pm standard error of the mean (SEM). * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ vs ND group; * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$ vs HFD group. ND: normal diet; HFD: high fat diet; AM: HFD + AM extract (100 mg/kg); LE: HFD + LE extract (100 mg/kg); ALM16-L: HFD + ALM16 (50 mg/kg); ALM16-M: HFD + ALM16 (100 mg/kg); ALM16-H: HFD + ALM16 (200 mg/kg); MT: HFD + milk thistle extract (100 mg/kg).

on the increase in the phosphorylation of AMPK and ACC by ALM16, we further investigated whether ALM16 downregulated lipid metabolism related to lipogenesis and lipid oxidation in the liver of HFD-fed mice. The results demonstrated that ALM16 treatment negatively regulated the lipogenesis-related proteins, including SREBP-1c and FAS, in the liver tissue of HFD-fed mice. It is generally recognized that the expression of lipogenesis-related proteins, such as SREBP-1c and FAS, are regulated by AMPK activation [12]. In the liver, SREBP-1c is an important transcription factor regulating fatty acid, cholesterol, and TG synthesis, and FAS is involved in lipid accumulation [57]. Previous studies have reported that downregulation of SREBP-1c and FAS, by the increased activation of AMPK, decreases lipid accumulation, and deposition, thereby improving hepatic steatosis [58, 59]. Our results showed that ALM16 reversed the HFD-induced downregulation of CPT-1, involved in fatty acid oxidation, in the HFD-fed mice. Similarly, several studies have reported that the protective effects against hepatic steatosis are mediated by the

downregulation of genes involved in lipogenesis and upregulation genes associated with fatty acid oxidation via enhanced AMPK signaling [60, 61]. The results showed that AM treatment alone downregulated the FAS protein expression by activating AMPK phosphorylation, while it had no effect on the SREBP-1c and CPT-1 expressions. On the other hand, LE inhibited the expressions of SREBP-1c and FAS, while it had no effect on AMPK activation, suggesting that LE may act in a different way from AM, following other mechanisms where AMPK was not involved. Therefore, although the action mechanism between AM and LE underlying the observed positive effect of ALM16 on NAFLD is not fully understood at the moment, one of the possible explanations would be that it might be due to the cross-acting or complementary interaction between the constituent compounds in the two herb extracts, AM and LE, in ALM16, thereby resulting in a complementary multitarget effect on NAFLD. However, this preclinical study also has some limitations, including certain subjective biases due to the relatively small sample sizes, and some differences in

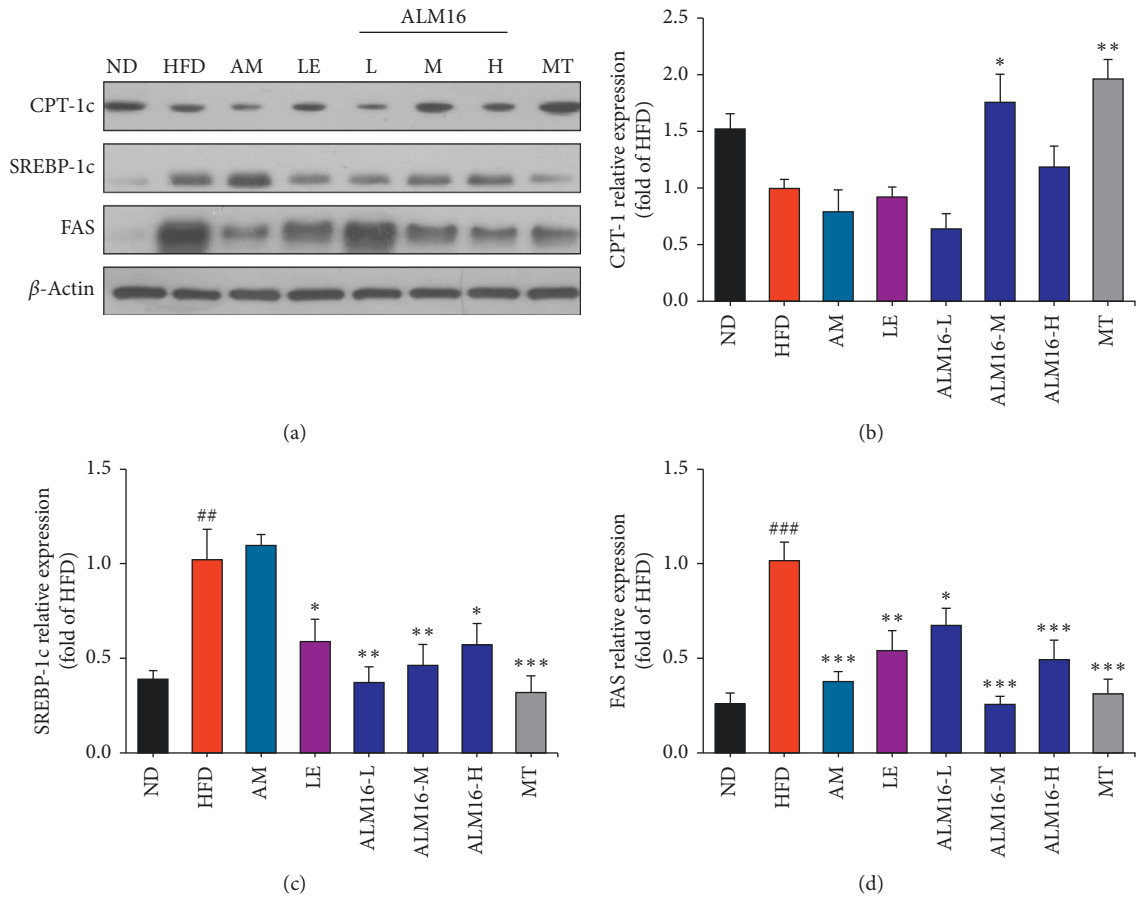


FIGURE 5: Effects of ALM16 and individual extracts (AM and LE) on the protein expression in hepatic lipid metabolism, CPT-1, SREBP-1c, and FAS, in HFD-fed mice. (a) Western blot analysis of CPT-1, SREBP-1c, and FAS in the liver tissues of HFD-fed mice. The protein expression levels of (b) CPT-1, (c) SREBP-1c, and (d) FAS normalized to β -actin relative to HFD-fed mice ($n = 4$). Values are presented as mean \pm standard error of the mean (SEM). $^*P < 0.05$, $^{**}P < 0.01$, and $^{***}P < 0.001$ vs. ND group; $^*P < 0.05$, $^{**}P < 0.01$, and $^{***}P < 0.001$ vs. HFD group. ND: normal diet; HFD: high fat diet; AM, HFD + AM extract (100 mg/kg); LE: HFD + LE extract (100 mg/kg); ALM16-L: HFD + ALM16 (50 mg/kg); ALM16-M: HFD + ALM16 (100 mg/kg); ALM16-H: HFD + ALM16 (200 mg/kg); MT: HFD + milk thistle extract (100 mg/kg).

features relevant to human NAFLD, which needs to be further clarified in the future.

Collectively, our results clearly demonstrated that ALM16 at a dose of 100 mg/kg could ameliorate NAFLD, through the downregulation of lipid metabolism-related proteins, especially modulating the AMPK and ACC signaling pathways. Although the detailed mechanism underlying the complementary interaction between the two herbs in ALM16 remains unknown, more potent effect on NAFLD was observed with ALM16 rather than individual extract (AM or LE) treatment.

5. Conclusions

The observed positive effects of ALM16 against hepatic steatosis and liver damage were, at least partly, shown to be mediated by the downregulation of SREBP-1c and FAS expression and upregulation of CPT-1 expression, through the modulation of AMPK and ACC signaling, in the established NAFLD *in vivo* model. Furthermore, this positive effect was more significant than the individual treatments with AM or LE. Although the clinical effectiveness of the observed hepatoprotective properties of ALM16 needs

further investigation, these findings suggest that ALM16 could be used as a functional food supplement for the prevention or treatment of NAFLD.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon reasonable request.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Acknowledgments

This work was carried out with support from the "Cooperative Research Program for Agriculture Science and Technology Development (project no. PJ01329101)," Rural Development Administration, Republic of Korea.

Supplementary Materials

Supplementary Figure 1: effects of ALM16 and individual extracts (AM and LE) on food intake ($n = 6$). (*Supplementary Materials*)

References

- [1] B. M. Herrera, S. Keildson, and C. M. Lindgren, "Genetics and epigenetics of obesity," *Maturitas*, vol. 69, no. 1, pp. 41–49, 2011.
- [2] B. Viollet, M. Foretz, B. Guigas et al., "Activation of AMP-activated protein kinase in the liver: a new strategy for the management of metabolic hepatic disorders," *The Journal of Physiology*, vol. 574, no. 1, pp. 41–53, 2006.
- [3] V. T. Samuel and G. I. Shulman, "Nonalcoholic fatty liver disease as a nexus of metabolic and hepatic diseases," *Cell Metabolism*, vol. 27, no. 1, pp. 22–41, 2018.
- [4] G. Marchesini, E. Bugianesi, G. Forlani et al., "Nonalcoholic fatty liver, steatohepatitis, and the metabolic syndrome," *Hepatology*, vol. 37, no. 4, pp. 917–923, 2003.
- [5] G. A. Michelotti, M. V. Machado, and A. M. Diehl, "NAFLD, NASH and liver cancer," *Nature Reviews Gastroenterology & Hepatology*, vol. 10, no. 11, pp. 656–665, 2013.
- [6] L. A. Adams, J. F. Lymp, J. St. Sauver et al., "The natural history of nonalcoholic fatty liver disease: a population-based cohort study," *Gastroenterology*, vol. 129, no. 1, pp. 113–121, 2005.
- [7] S. Li, X. Liao, F. Meng et al., "Therapeutic role of ursolic acid on ameliorating hepatic steatosis and improving metabolic disorders in high-fat diet-induced non-alcoholic fatty liver disease rats," *PLoS One*, vol. 9, no. 1, Article ID e86724, 2014.
- [8] M. Durazno, P. Belci, A. Collo, E. Grisoglio, and S. Bo, "Focus on therapeutic strategies of nonalcoholic fatty liver disease," *International Journal of Hepatology*, vol. 2012, Article ID 464706, 9 pages, 2012.
- [9] C. P. Day and O. F. W. James, "Steatohepatitis: a tale of two "hits"?" *Gastroenterology*, vol. 114, no. 4, pp. 842–845, 1998.
- [10] D. Carling, C. Thornton, A. Woods, and M. J. Sanders, "AMP-activated protein kinase: new regulation, new roles?" *Biochemical Journal*, vol. 445, no. 1, pp. 11–27, 2012.
- [11] B. Viollet, B. Guigas, J. Leclerc et al., "AMP-activated protein kinase in the regulation of hepatic energy metabolism: from physiology to therapeutic perspectives," *Acta Physiologica*, vol. 196, no. 1, pp. 81–98, 2009.
- [12] B. K. Smith, K. Marcinko, E. M. Desjardins, J. S. Lally, R. J. Ford, and G. R. Steinberg, "Treatment of nonalcoholic fatty liver disease: role of AMPK," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 311, no. 4, pp. E730–E740, 2016.
- [13] M. M. Assifi, G. Suchankova, S. Constant, M. Prentki, A. K. Saha, and N. B. Ruderman, "AMP-activated protein kinase and coordination of hepatic fatty acid metabolism of starved/carbohydrate-refed rats," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 289, no. 5, pp. E794–E800, 2005.
- [14] C.-q. Zhao, Y. Zhou, J. Ping, and L.-m. Xu, "Traditional Chinese medicine for treatment of liver diseases: progress, challenges and opportunities," *Journal of Integrative Medicine*, vol. 12, no. 5, pp. 401–408, 2014.
- [15] C.-T. Che, Z. Wang, M. Chow, and C. Lam, "Herb-herb combination for therapeutic enhancement and advancement: theory, practice and future perspectives," *Molecules*, vol. 18, no. 5, pp. 5125–5141, 2013.
- [16] E. Williamson, "Synergy and other interactions in phyto-medicines," *Phytomedicine*, vol. 8, no. 5, pp. 401–409, 2001.
- [17] D. Wheatley, "Stress-induced insomnia treated with kava and valerian: singly and in combination," *Human Psychopharmacology: Clinical and Experimental*, vol. 16, no. 4, pp. 353–356, 2001.
- [18] E. Bedir, N. Pugh, I. Calis, D. S. Pasco, and I. A. Khan, "Immunostimulatory effects of cycloartane-type triterpene glycosides from *astragalus* species," *Biological & Pharmaceutical Bulletin*, vol. 23, no. 7, pp. 834–837, 2000.
- [19] W. C. S. Cho and K. N. Leung, "In vitro and in vivo anti-tumor effects of *Astragalus membranaceus*," *Cancer Letters*, vol. 252, no. 1, pp. 43–54, 2007.
- [20] J. Fu, Z. Wang, L. Huang et al., "Review of the botanical characteristics, phytochemistry, and pharmacology of *Astragalus membranaceus* (huangqi)," *Phytotherapy Research*, vol. 28, no. 9, pp. 1275–1283, 2014.
- [21] X. Su, Q. Huang, J. Chen et al., "Calycosin suppresses expression of pro-inflammatory cytokines via the activation of p62/Nrf2-linked heme oxygenase 1 in rheumatoid arthritis synovial fibroblasts," *Pharmacological Research*, vol. 113, no. Pt A, pp. 695–704, 2016.
- [22] S. I. Choi, T. R. Heo, B.-H. Min, J. H. Cui, B. H. Choi, and S. R. Park, "Alleviation of osteoarthritis by calycosin-7-O- β -D-glucopyranoside (CG) isolated from *Astragali radix* (AR) in rabbit osteoarthritis (OA) model," *Osteoarthritis and Cartilage*, vol. 15, no. 9, pp. 1086–1092, 2007.
- [23] X. Chen, Q. Meng, C. Wang et al., "Protective effects of calycosin against CCl4-induced liver injury with activation of FXR and STAT3 in mice," *Pharmaceutical Research*, vol. 32, no. 2, pp. 538–548, 2015.
- [24] X. Duan, Q. Meng, C. Wang et al., "Effects of calycosin against high-fat diet-induced nonalcoholic fatty liver disease in mice," *Journal of Gastroenterology and Hepatology*, vol. 33, no. 2, pp. 533–542, 2018.
- [25] X. Xu, F. Li, X. Zhang et al., "In vitro synergistic antioxidant activity and identification of antioxidant components from *Astragalus membranaceus* and *Paeonia lactiflora*," *PLoS One*, vol. 9, no. 5, Article ID e96780, 2014.
- [26] K. Cui, S. Zhang, X. Jiang, and W. Xie, "Novel synergic antidiabetic effects of *Astragalus* polysaccharides combined with *Crataegus* flavonoids via improvement of islet function and liver metabolism," *Molecular Medicine Reports*, vol. 13, no. 6, pp. 4737–4744, 2016.
- [27] I. Andújar, J. Rios, R. Giner, and M. Recio, "Pharmacological properties of shikonin - a review of literature since 2002," *Planta Medica*, vol. 79, no. 18, pp. 1685–1697, 2013.
- [28] X. Chen, L. Yang, J. J. Oppenheim, and O. M. Z. Howard, "Cellular pharmacology studies of shikonin derivatives," *Phytotherapy Research*, vol. 16, no. 3, pp. 199–209, 2002.
- [29] S. Y. Gwon, J. Y. Ahn, C. H. Chung, B. Moon, and T. Y. Ha, "Lithospermum erythrorhizon suppresses high-fat diet-induced obesity, and acetylshikonin, a main compound of *Lithospermum erythrorhizon*, inhibits adipocyte differentiation," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 36, pp. 9089–9096, 2012.
- [30] R. A. Velliquette, A. Rajgopal, J. Rebhun, and K. Glynn, "Lithospermum erythrorhizon root and its naphthoquinones repress SREBP1c and activate PGC1 α through AMPK α ," *Obesity*, vol. 26, no. 1, pp. 126–134, 2018.
- [31] Y. H. Choi, G. S. Kim, J. H. Choi et al., "Ethanol extract of *Lithospermum erythrorhizon* Sieb. et Zucc. promotes osteoblastogenesis through the regulation of Runx2 and Osterix,"

- International Journal of Molecular Medicine*, vol. 38, no. 2, pp. 610–618, 2016.
- [32] W. Zheng, L. Cao, Z. Xu, Y. Ma, and X. Liang, “Anti-angiogenic alternative and complementary medicines for the treatment of endometriosis: a review of potential molecular mechanisms,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2018, Article ID 4128984, 28 pages, 2018.
- [33] J. S. Roh, H. Lee, J. Lim et al., “Effect of Gangjihwan on hepatic steatosis and inflammation in high fat diet-fed mice,” *Journal of Ethnopharmacology*, vol. 206, pp. 315–326, 2017.
- [34] D. J. Choi, B. R. Choi, D. Y. Lee, S. I. Choi, Y. S. Lee, and G. S. Kim, “Inhibitory effect of mixed extracts obtained from *Astragal radix* and *lithospermi radix* on matrix metalloproteinases in IL-1 β -induced SW1353 cells and quantitative analysis of active compounds,” *Korean Journal of Medicinal Crop Science*, vol. 27, no. 4, pp. 247–258, 2019.
- [35] D. J. Choi, S. I. Choi, B. R. Choi, Y. S. Lee, D. Y. Lee, and G. S. Kim, “Cartilage protective and anti-analgesic effects of ALM16 on monosodium iodoacetate induced osteoarthritis in rats,” *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, p. 325, 2019.
- [36] P. Pais and M. D’Amato, “In vivo efficacy study of milk thistle extract (ETHIS-094) in STAM model of nonalcoholic steatohepatitis,” *Drugs in R&D*, vol. 14, no. 4, pp. 291–299, 2014.
- [37] X. Ren, X. Zhao, F. Turcotte, J. S. Deschênes, R. Tremblay, and M. Jolicoeur, “Current lipid extraction methods are significantly enhanced adding a water treatment step in *Chlorella protothecoides*,” *Microbial Cell Factories*, vol. 16, no. 1, p. 26, 2017.
- [38] M. A. Konerman, J. C. Jones, and S. A. Harrison, “Pharmacotherapy for NASH: current and emerging,” *Journal of Hepatology*, vol. 68, no. 2, pp. 362–375, 2018.
- [39] Y. Y. Choi, M. H. Kim, J. Hong, K. Kim, and W. M. Yang, “Effect of Dangguibohtul-Tang, a mixed extract of *Astragalus membranaceus* and *Angelica sinensis*, on allergic and inflammatory skin reaction compared with single extracts of *Astragalus membranaceus* or *Angelica sinensis*,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2016, Article ID 5936354, 9 pages, 2016.
- [40] T. G. Ahn, J. Y. Lee, S. Y. Cheon, H. J. An, and Y. B. Kook, “Protective effect of Sam-Hwang-Sa-Sim-Tang against hepatic steatosis in mice fed a high-cholesterol diet,” *BMC Complementary and Alternative Medicine*, vol. 13, p. 366, 2013.
- [41] X. Zhou, S. W. Seto, D. Chang et al., “Synergistic effects of Chinese herbal medicine: a comprehensive review of methodology and current research,” *Frontiers in Pharmacology*, vol. 7, p. 201, 2016.
- [42] G. Kanuri and I. Bergheim, “In vitro and in vivo models of non-alcoholic fatty liver disease (NAFLD),” *International Journal of Molecular Sciences*, vol. 14, no. 6, pp. 11963–11980, 2013.
- [43] S. Zhang, L. Zheng, D. Dong et al., “Effects of flavonoids from *Rosa laevigata* Michx fruit against high-fat diet-induced non-alcoholic fatty liver disease in rats,” *Food Chemistry*, vol. 141, no. 3, pp. 2108–2116, 2013.
- [44] E. Fabbri and F. Magkos, “Hepatic steatosis as a marker of metabolic dysfunction,” *Nutrients*, vol. 7, no. 6, pp. 4995–5019, 2015.
- [45] J. Xu, Y. Li, W.-D. Chen et al., “Hepatic carboxylesterase 1 is essential for both normal and farnesoid X receptor-controlled lipid homeostasis,” *Hepatology*, vol. 59, no. 5, pp. 1761–1771, 2014.
- [46] A. Dittmann, N. J. Kennedy, N. L. Soltero et al., “High-fat diet in a mouse insulin-resistant model induces widespread rewiring of the phosphotyrosine signaling network,” *Molecular Systems Biology*, vol. 15, no. 8, Article ID e8849, 2019.
- [47] A. Ulla, M. A. Alam, B. Sikder et al., “Supplementation of *Syzygium cumini* seed powder prevented obesity, glucose intolerance, hyperlipidemia and oxidative stress in high carbohydrate high fat diet induced obese rats,” *BMC Complementary and Alternative Medicine*, vol. 17, no. 1, p. 289, 2017.
- [48] J. W. Kim, Y. S. Lee, D. J. Seol et al., “Anti-obesity and fatty liver-preventing activities of *Lonicera caerulea* in high-fat diet-fed mice,” *International Journal of Molecular Medicine*, vol. 42, no. 6, pp. 3047–3064, 2018.
- [49] F. Hadizadeh, E. Faghihmani, and P. Adibi, “Nonalcoholic fatty liver disease: diagnostic biomarkers,” *World Journal of Gastrointestinal Pathophysiology*, vol. 8, no. 2, pp. 11–26, 2017.
- [50] L. G. Draijer, S. Feddoui, A. E. Bohte et al., “Comparison of diagnostic accuracy of screening tests ALT and ultrasound for pediatric non-alcoholic fatty liver disease,” *European Journal of Pediatrics*, vol. 178, no. 6, pp. 863–870, 2019.
- [51] P. T. Donnan, D. McLernon, J. F. Dillon et al., “Development of a decision support tool for primary care management of patients with abnormal liver function tests without clinically apparent liver disease: a record-linkage population cohort study and decision analysis (ALFIE),” *Health Technology Assessment*, vol. 13, no. 25, pp. 1–134, 2009.
- [52] J. D. Browning and J. D. Horton, “Molecular mediators of hepatic steatosis and liver injury,” *Journal of Clinical Investigation*, vol. 114, no. 2, pp. 147–152, 2004.
- [53] D. K. Yang and D. G. Jo, “Mulberry fruit extract ameliorates nonalcoholic fatty liver disease (NAFLD) through inhibition of mitochondrial oxidative stress in rats,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2018, Article ID 8165716, 9 pages, 2018.
- [54] N. K. S. Tran, G. T. Kim, S. H. Park, D. Lee, S. M. Shim, and T. S. Park, “Fermented *Cordyceps militaris* extract prevents hepatosteatosis and adipocyte hypertrophy in high fat diet-fed mice,” *Nutrients*, vol. 11, no. 5, p. E1015, 2019.
- [55] S. Bijland, S. J. Mancini, and I. P. Salt, “Role of AMP-activated protein kinase in adipose tissue metabolism and inflammation,” *Clinical Science*, vol. 124, no. 8, pp. 491–507, 2013.
- [56] J.-F. Liu, Y. Ma, Y. Wang, Z.-Y. Du, J.-K. Shen, and H.-L. Peng, “Reduction of lipid accumulation in HepG2 cells by luteolin is associated with activation of AMPK and mitigation of oxidative stress,” *Phytotherapy Research*, vol. 25, no. 4, pp. 588–596, 2011.
- [57] A. Moslehi and Z. Hamidi-zad, “Role of SREBPs in liver diseases: a mini-review,” *Journal of Clinical and Translational Hepatology*, vol. 6, no. 3, pp. 332–338, 2018.
- [58] P. Yu, X. Xu, J. Zhang et al., “Liraglutide attenuates non-alcoholic fatty liver disease through adjusting lipid metabolism via SHP1/AMPK signaling pathway,” *International Journal of Endocrinology*, vol. 2019, Article ID 1567095, 11 pages, 2019.
- [59] Q. Wei, B. Zhou, G. Yang et al., “JAZF1 ameliorates age and diet-associated hepatic steatosis through SREBP-1c-dependent mechanism,” *Cell Death & Disease*, vol. 9, no. 9, p. 859, 2018.
- [60] E. Kim, Y. Choi, J. Jang, and T. Park, “Carvacrol protects against hepatic steatosis in mice fed a high-fat diet by enhancing SIRT1-AMPK signalling,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, Article ID 290104, 10 pages, 2013.
- [61] S. Cheng, S. Liang, Q. Liu et al., “Diosgenin prevents high-fat diet-induced rat non-alcoholic fatty liver disease through the AMPK and LXR signaling pathways,” *International Journal of Molecular Medicine*, vol. 41, no. 2, pp. 1089–1095, 2018.