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Short communication

Frequency of detection of respiratory viruses in the lower respiratory tract of hospitalized adults

C. Minosse^a, M. Selleri^a, M.S. Zaniratti^a, G. Cappiello^b, R. Longo^b, E. Schifano^c, A. Spanò^b, N. Petrosillo^d, F.N. Lauria^d, V. Puro^c, M.R. Capobianchi^{a,*}

^a Laboratory of Virology, National Institute for Infectious Diseases "L. Spallanzani", Via Portuense, 292, 00149 Rome, Italy

^b Hospital "S. Pertini", Via dei Monti Tiburtini, 385, 00157 Rome, Italy

^c Department of Epidemiology, National Institute for Infectious Diseases "L. Spallanzani", Via Portuense, 292, 00149 Rome, Italy ^d Clinical Department, National Institute for Infectious Diseases "L. Spallanzani", Via Portuense, 292, 00149 Rome, Italy

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Abstract

Background: Respiratory infections are the most common infections in humans. The prevalence of respiratory viruses in adults is largely underestimated, and relevant data mostly concern infants and children.

Objectives: To evaluate the prevalence of respiratory viruses in adults hospitalized in Italy.

Study design: During April 2004–May 2005, 510 consecutive lower respiratory tract samples were prospectively collected. These were evaluated with a molecular panel that detected 12 respiratory viruses.

Results: Two hundred and fifteen samples were positive for at least one viral pathogen, with an overall sample prevalence of 42.2%. Human rhinoviruses (HRVs) were the most commonly detected viruses (32.9%), followed by influenza virus (FLU)-A (9.0%); the other viruses were 2% or less. Multiple agents were detected in 30 samples from 29 patients, resulting in a co-infection rate of 6.7%.

Conclusions: This study shows a high prevalence of viruses in the lower respiratory tract samples of hospitalized adults, mostly HRV and FLU-A. It is not possible to establish the role of viruses detected at low frequency, but our findings suggest the necessity to consider them as potential causes or precursors of lower respiratory tract infections (LRTIs).

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Keywords: Lower respiratory tract infection; Respiratory viruses; Adults; Hospitalized

1. Introduction

The etiology of lower respiratory tract infections (LRTIs) in adults, which are an important cause of morbidity, remains undetermined in more than 50% of cases (Fine et al., 1999; Garbino et al., 2002; File, 2003), mostly due to limited diagnostic procedures. The prevalence of respiratory viruses

in adults is largely unexplored, as most relevant data concern infants and children (Choi et al., 2006; Costa et al., 2006; Kusel et al., 2006; Ma et al., 2006; Ordas et al., 2006; Pierangeli et al., 2007). Respiratory syncytial virus (RSV), parainfluenzaviruses (PIVs), influenzaviruses (FLUs) and adenoviruses (AdVs) are considered the leading causes of acute viral LRTIs (Henderson et al., 1979; Hall et al., 1990; Yun et al., 1995; Hong et al., 2001). However, many other respiratory viruses have been recently identified as possible causative agents of LRTIs. Human rhinoviruses (HRVs), which previously were believed to cause only mild upper respiratory illnesses, have been found in association with acute and chronic LRTIs, including asthma exacerbations and chronic obstructive pulmonary disease (COPD), although their role as causative agents has only been established for asthma (Singh and Busse, 2007; Papadopoulos et

Abbreviations: HRV, human rhinovirus; FLU, influenzavirus; hCoV, human coronavirus; AdV, adenovirus; PIV, parainfluenzavirus; RSV, respiratory syncytial virus; hMPV, human metapneumovirus; LRTIs, lower respiratory tract infections; SP, sputum; EA, endotracheal aspirate; BAL, bronchoalveolar lavage; COPD, chronic obstructive pulmonary disease; ARI, acute respiratory illness; hBoV, human bocavirus.

Corresponding author. Tel.: +39 06 55170434; fax: +39 06 5582346. *E-mail address:* capobianchi@inmi.it (M.R. Capobianchi).

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al., 2002; Friedlander and Busse, 2005; Xatzipsalti et al., 2005). Newly discovered viruses, such as human metapneumovirus (hMPV) and human coronaviruses (hCoVs) have been associated with LRTIs in children (van den Hoogen et al., 2001; Fouchier et al., 2004; van der Hoek et al., 2004; Choi et al., 2006).

The aim of this study was to assess the point prevalence of a full spectrum of respiratory viruses in the lower respiratory tract specimens of hospitalized adults by using a molecular diagnostic panel.

2. Methods

A prospective collection of 510 lower respiratory samples from 433 adult patients with pulmonary diseases, admitted to three Italian hospitals was performed. Clinicalepidemiological data were collected through a standardized questionnaire. Patients with suspect Mycobacterium tuberculosis infection were not included in the analysis. The study was approved by the Institutional Ethics Committees, and informed consent signed by participants. The selection criterion for patients' enrolment was to have any one of the following samples sent to the laboratory for routine microbiological evaluation: sputum (SP), bronchoalveolar lavages (BALs), or endotracheal aspirate (EA). To minimize the possibility that positive results reflected contamination from the upper respiratory tract, only samples containing more than 25 polymorphonuclear leucocytes per field $(40 \times)$ were processed (Murray and Washington, 1975; Geckler et al., 1977). Less than 4% of the samples did not meet this criterion and were discarded. Half of each sample was submitted to standard microbiological analysis (common microbiological culture and, in case of positive result, identification and antibody susceptibility test), while the remainder was stored at -80 °C for the molecular analysis. During April 2004–May 2005, a total of 510 consecutive lower tract respiratory samples (329 SP, 165 EA and 16 BAL), which were considered appropriate on the basis of microscope inspection, were analyzed.

The samples were thawed and liquefied by using 1:1 Sputasol (Oxoid Ltd., Basingstoke, England), and then underwent DNA/RNA extraction (Boom et al., 1990). RNA was reverse transcribed with random primers with a Reverse Transcriptase Kit (Invitrogen, Milan, Italy). Control DNA ((globin DNA) and RNA (MS2 phage genome) preparations were spiked into the samples before the extraction, to control their suitability for PCR/RT-PCR.

The molecular diagnostic panel included: FLU-A and -B, hMPV, AdV, PIV-1, -2 and -3, RSV, HRV, hCoV-OC43, -229E and -NL63. Specific primers and PCR or RT-PCR conditions were as previously described (Allard et al., 1994; Pitkaranta et al., 1997; Echevarria et al., 1998; Gröndahl et al., 1999; Allard et al., 2001; Steininger et al., 2001; Maggi et al., 2003; Fouchier et al., 2004; Bastien et al., 2005; Esper et al., 2005; Woo et al., 2005; Minosse et al., 2007). The sensitivity of the assays ranged between 1 and 10 copies for FLU-A, AdV, PIV-1, -2 and -3, RSV, HRV and hCoV-NL63; between 10 and 50 copies for FLU-B and hMPV A; and about 300 copies for hCoV-OC43. These sensitivity values, established by probit analysis on plasmids containing the target region as the insert, were confirmed by participation in the Quality Control for Molecular Diagnostics External Quality Assurance (QCMD EQA) programmes (Glasgow, UK). From the hMPV QCMD EQA programme MPV.RSV06, we determined that the sensitivity for hMPV B of our test was much lower (>1000 times) than for other respiratory viruses.

The amplified products were separated by agarose gel electrophoresis and UV-visualized after ethidium bromide staining.

3. Results

A molecular diagnostic panel for detection of 12 viruses was applied to 510 lower respiratory samples from 433 adult patients hospitalized with pulmonary diseases.

The most common respiratory illnesses in the patients were: COPD, 45.3%, pneumonia (30.5%) and acute respiratory illness (ARI, 17.1%); 19.6% of patients were in the intensive care unit, and 17.5% were receiving assisted ventilation (Table 1). Two hundred and fifteen specimens (42.2%) were positive for a respiratory virus (Table 2). The most common viruses were HRV (32.9%), FLU-A (9.0%) and hCoV-NL63 (2.0%). The frequency of the other viruses ranged from 0.2 to 1.4%. In 30 out of 510 samples (5.9%), corresponding to 29 out of 433 patients (6.7%), more than one virus was detected. Multiple detection included HRV + FLU-A (2.9% of total samples), HRV + FLU-B and HRV + hCoV-NL63 (0.6% each combina-

Table 1

Demographic and clinical characteristics of 433 adult hospitalized patients

Age				
Mean (\pm S.D.)	56.3 (18.2)			
Median (IQR)	57.0 (41–72)			
Sex				
Male (%)	291 (67.2)			
Female (%)	136(31.4)			
Not known (%)	6(1.4)			
Smoking status				
Smokers (%)	168 (38.8)			
Non-smokers (%)	235 (54.3)			
Not known (%)	30(6.9)			
Clinical findings				
COPD (%)	196 (45.3)			
Pneumonia (%)	132 (30.5)			
Acute respiratory failure (%)	76(17.5)			
Chronic respiratory failure (%)	15 (3.5)			
Assisted ventilation (%)	76(17.5)			
Asthma (%)	9(2.1)			

COPD, chronic obstructive pulmonary disease.

Table 2						
Detection	of respiratory	agents in 510	samples from	433 adult	hospitalized	patients

Respiratory agent	Distribution of positive specimens, n (%)					
	Overall positivity	SP	EA	BAL		
FLU-A [*]	46 (9.0)	24(7.3)	20(12.1)	2(12.5)		
FLU-B	5(1.0)	5(1.5)	0	0		
PIV-1	6(1.2)	6(1.8)	0	0		
PIV-2	0	0	0	0		
PIV-3	1 (0.2)	0	1 (0.6)	0		
RSV	1 (0.2)	1(0.3)	0	0		
hMPV	0	0	0	0		
HRV ^{**}	168 (32.9)	124 (37.7)	39 (23.6)	5(31.2)		
hCoV-OC43	7(1.4)	4(1.2)	3(1.8)	0		
hCoV-229E	0	0	0	0		
hCoV-NL63	10(2.0)	7(2.1)	3(1.8)	0		
AdV	6(1.2)	6(1.8)	0	0		
Samples positive to at least 1 agent***	215 (42.2)	156 (47.4)	58 (35.1)	6(37.5)		
Samples positive to >1 agent	30(5.9)	22(6.7)	8(4.8)	0		
Total analyzed samples	510(100)	329 (100)	165(100)	16(100)		

SP, sputum; EA, endotracheal aspirate; BAL, bronchoalveolar lavage; FLU, influenzavirus; PIV, parainfluenzavirus; RSV, respiratory syncytial virus; hMPV, human metapneumovirus; HRV, human rhinovirus; hCoV, human coronavirus; AdV, adenovirus.

p = 0.186.

** p = 0.007.

*** p = 0.031 for frequency in SP, EA and BAL in χ^2 test.

tion), HRV + hCoV-OC43 (0.4%), HRV + AdV, HRV + PIV-1, HRV + RSV, FLU-A + hCoV-OC43, AdV + hCoV-NL63, HRV + FLU-A + PIV-1 and HRV + FLU-A + AdV (0.2% each combination). The association with the sample type (either EA, or SP, or BAL) could be evaluated only for the most frequent viruses, i.e. FLU-A and HRV, showing that HRV, and not FLU-A, was unevenly distributed (more frequent in SP and BAL than in EA, Table 2). Data on the smoking status was available for 403/433 patients: the frequency of patients positive to any virus was similar in smokers and non-smokers (49.4% vs. 50.6%, p = 0.742), and the frequency of individual viruses did not significantly differ according to the smoker status (for instance, HRV positive patients were 42.2% in smokers and 34.5% in non-smokers, p = 0.137).

In addition, similar frequencies of virus-positive samples were obtained when comparing patients with COPD vs. patients without COPD (57.1% vs. 56.7%); patients with pneumonia vs. subjects with no record of pneumonia (54.4% in both cases); or patients with ARI vs. subjects without ARI (44.7% vs. 51.5%, p = 0.342). In addition, the frequency of virus-positive samples did not significantly differ among patients with pneumonia, COPD and ARI (p = 0.207).

The monthly distribution of respiratory agents over the study period (Fig. 1) highlights a peak in January–February for HRV and FLU-A.

4. Discussion

Many reports have highlighted the possible role of viruses as causative agents, or as possible co-factors, in LRTIs. We applied a large molecular diagnostic panel which included 12 respiratory viruses to evaluate the prevalence of respiratory pathogens in adults hospitalized in Italy, during April 2004-May 2005. Although we would be unable to detect some viral respiratory pathogens, such as bocavirus, coronavirus HKU1 and parainfluenzavirus 4, this is the first extensive investigation of lower respiratory tract secretions from adults that was capable of detecting such a wide selection of respiratory pathogens. We have made the assumption that the samples represent the lower respiratory tract, as a recognized microscope inspection criterion was used to validate their quality. However, as expectorated sputum is necessarily contaminated with oral contents, it is not possible to rule out that the virus detected in the samples (mainly SP) could actually derive from contamination by upper respiratory tract secretions. In fact, the rate of positivity in SP is higher than in EA and BAL (47.4% vs. 35.1% and 37.5%, respectively); however, this is not the case for all viruses (i.e. FLU-A). In addition, the presence of several viruses (FLU-A, PIV-3, HRV, hCoV-OC43 and hCoV-NL63) on both EA and BAL, suggests that contamination with oral secretions is not a major source of the virus present in the analyzed samples.

Our results confirm previous data that HRV are the most frequent viruses in lower respiratory tract samples (Garbino et al., 2004; Puro et al., 2005), and strongly support the association of HRV with LRTIs, already hypothesized in previous studies based on upper respiratory samples (Tsolia et al., 2004). However, based on the present data it is not possible to establish the precise role of HRV as a leading cause, as well as its possible involvement as co-factor in LRTIs.

Among the other viruses, accounting for 7.2% of positive samples, hCoV-NL63 was detected in 2.0% of samples. The involvement of hCoV (namely NL63) in lower respiratory diseases has been largely hypothesized, but available



Fig. 1. Monthly distribution of lower respiratory tract samples positive for respiratory viruses, during April 2004–May 2005. The line represents the total collected samples, while the bars represent the samples positive for the indicated viruses. First panel: absolute numbers; lower panels: frequency (%) of samples positive for the indicated viruses.

data are mostly based on upper respiratory samples, mostly on the basis of studies in pediatric patients (Arden et al., 2005; Bastien et al., 2005; Boivin et al., 2005; Chiu et al., 2005; Ebihara et al., 2005; Moes et al., 2005; Pierangeli et al., 2007). The circulation of hCoV-NL63 in Italy is probably underreported. There is only one other report concerning this virus and lower respiratory samples from adults (Gerna et al., 2006).

We did not detect hMPV in our patients, although this is a relatively common respiratory pathogen (van den Hoogen et al., 2001; Choi et al., 2006). The absence of hMPV is probably due to the low sensitivity of the RT-PCR assay used in this study (particularly for hMPV B). Alternatively, it is possible that sputasol, used to liquefy the samples, could have affected the sensitivity of the RT-PCR used. This issue could also apply to other viruses investigated in this study, such as RSV, and needs further investigation.

In conclusion, our study shows a high prevalence of viruses, especially HRV and influenza virus, in the lower respiratory tract samples of adult hospitalized patients with pulmonary diseases, without any significant difference in the frequency of virus-positivity among patients with pneumonia, COPD and ARI. Additional viruses were detected at low frequency. It is not possible to establish the role of viruses detected at low frequency as causative agents or as co-factors in LRTIs, but these results highlight the necessity to consider them as potential causes of LRTIs.

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References

- Allard A, Kajon A, Wadell G. Simple procedure for discrimination and typing of enteric adenoviruses after detection by polymerase chain reaction. J Med Virol 1994;44(3):250–7.
- Allard A, Albinsson B, Wadell G. Rapid typing of human adenoviruses by a general PCR combined with restriction endonuclease analysis. J Clin Microbiol 2001;39(2):498–505.
- Arden KE, Nissen MD, Sloots TP, Mackay IM. New human coronavirus, HCoV-NL63, associated with severe lower respiratory tract disease in Australia. J Med Virol 2005;75(3):455–62.
- Bastien N, Anderson K, Hart L, Van Caeseele P, Brandt K, Milley D, et al. Human coronavirus NL63 infection in Canada. J Infect Dis 2005;191(4):503–6.
- Boivin G, Baz M, Cote S, Gilca R, Deffrasnes C, Leblanc E, et al. Infections by human coronavirus-NL in hospitalized children. Pediatr Infect Dis J 2005;24(12):1045–8.
- Boom R, Sol CJ, Salimans MM, Jansen CL, Wertheim-van Dillen PM, van der Noordaa J. Rapid and simple method for purification of nucleic acids. J Clin Microbiol 1990;28(3):495–503.
- Chiu SS, Chan KH, Chu KW, Kwan SW, Guan Y, Poon LL, et al. Human coronavirus NL63 infection and other coronavirus infections in children hospitalized with acute respiratory disease in Hong Kong, China. Clin Infect Dis 2005;40(12):1721–9.
- Choi EH, Lee HJ, Kim SJ, Eun BW, Kim NH, Lee JA, et al. The association of newly identified respiratory viruses with lower respiratory tract infections in Korean children 2000–2005. Clin Infect Dis 2006;43(5): 585–92.
- Costa LF, Yokosawa J, Mantese OC, Oliveira TF, Silveira HL, Nepomuceno LL, et al. Respiratory viruses in children younger than five years old with acute respiratory disease from 2001 to 2004 in Uberlandia, MG, Brazil. Mem Inst Oswaldo Cruz 2006;101(3):301–6.
- Ebihara T, Endo R, Ma X, Ishiguro N, Kikuta H. Detection of human coronavirus NL63 in young children with bronchiolitis. J Med Virol 2005;75(3):463–5.
- Echevarria JE, Erdman DD, Swierkosz EM, Holloway BP, Anderson LJ. Simultaneous detection and identification of human parainfluenza viruses 1, 2 and 3 from clinical samples by multiplex PCR. J Clin Microbiol 1998;36(5):1388–91.

- Esper F, Weibel C, Ferguson D, Landry ML, Kahn JS. Evidence of a novel human coronavirus that is associated with respiratory tract disease in infants and young children. J Infect Dis 2005;191(4):492–8.
- File TM. Community-acquired pneumonia. Lancet 2003;362(9400):1991–2001.
- Fine MJ, Stone RA, Singer DE, Coley CM, Marrie TJ, Lave JR, et al. Processes and outcomes of care for patients with community-acquired pneumonia: results from the Pneumonia Patient Outcomes Research Team (PORT) cohort study. Arch Intern Med 1999;159(9):970–80.
- Fouchier RA, Hartwig NG, Bestebroer TM, Niemeyer B, de Jong JC, Simon JH, et al. A previously undescribed coronavirus associated with respiratory disease in humans. Proc Natl Acad Sci USA 2004;101(16):6212–6.
- Friedlander SL, Busse WW. The role of rhinovirus in asthma exacerbations. J Allergy Clin Immunol 2005;116(2):267–73.
- Garbino J, Sommer R, Gerber A, Regamey C, Vernazza P, Genne D, et al. Prospective epidemiologic survey of patients with community-acquired pneumonia requiring hospitalization in Switzerland. Int J Infect Dis 2002;6(4):288–93.
- Garbino J, Gerbase MW, Wunderli W, Deffernez C, Thomas Y, Rochat T, et al. Lower respiratory viral illnesses: improved diagnosis by molecular methods and clinical impact. Am J Respir Crit Care Med 2004;170(11):1197–203.
- Geckler RW, Gremillion DH, McAllister CK, Ellenbogen C. Microscopic and bacteriological comparison of paired sputa and transtracheal aspirates. J Clin Microbiol 1977;6:396–9.
- Gerna G, Campanini G, Rovida F, Percivalle E, Sarasini A, Marchi A, et al. Genetic variability of human coronavirus OC43-, 229E-, and NL63like strains and their association with lower respiratory tract infections of hospitalized infants and immunocompromised patients. J Med Virol 2006;78(7):938–49.
- Gröndahl B, Puppe W, Hoppe A, Kuhne I, Weigl JA, Schmitt HJ. Rapid identification of nine microorganisms causing acute respiratory tract infections by single-tube multiplex reverse transcription-PCR: feasibility study. J Clin Microbiol 1999;37(1):1–7.
- Hall CB, Walsh EE, Schnabel KC, Long CE, McConnochie KM, Hildreth SW, et al. Occurrence of groups A and B of respiratory syncytial virus over 15 years: associated epidemiologic and clinical characteristics in hospitalized and ambulatory children. J Infect Dis 1990;162(6):1283– 90.
- Henderson FW, Clyde Jr WA, Collier AM, Denny FW, Senior RJ, Sheaffer CI, et al. The etiologic and epidemiologic spectrum of bronchiolitis in pediatric practice. J Pediatr 1979;95(2):183–90.
- Hong JY, Lee HJ, Piedra PA, Choi EH, Park KH, Koh YY, et al. Lower respiratory tract infections due to adenovirus in hospitalized Korean children: epidemiology, clinical features, and prognosis. Clin Infect Dis 2001;32(10):1423–9.
- Kusel MM, de Klerk NH, Holt PG, Kebadze T, Johnston SL, Sly PD. Role of respiratory viruses in acute upper and lower respiratory tract illness in the first year of life: a birth cohort study. Pediatr Infect Dis J 2006;25(8):680–6.
- Ma X, Endo R, Ishiguro N, Ebihara T, Ishiko H, Ariga T, et al. Detection of human bocavirus in Japanese children with lower respiratory tract infections. J Clin Microbiol 2006;44(3):1132–4.
- Maggi F, Pifferi M, Vatteroni M, Fornai C, Tempestini E, Anzilotti S, et al. Human metapneumovirus associated with respiratory tract infections in a 3-year study of nasal swabs from infants in Italy. J Clin Microbiol 2003;41(7):2987–91.
- Minosse C, Selleri M, Zaniratti MS, Lauria FN, Puro V, Carletti F, et al. Improved detection of human influenza A and B viruses in respiratory tract specimens by hemi-nested PCR. J Virol Methods 2007;141(2):225–8.
- Moes E, Vijgen L, Keyaerts E, Zlateva K, Li S, Maes P, et al. A novel pancoronavirus RT-PCR assay: frequent detection of human coronavirus NL63 in children hospitalized with respiratory tract infections in Belgium. BMC Infect Dis 2005;5(1):6.
- Murray PR, Washington II JA. Microscopic and bacteriologic analysis od expectored sputum. Mayo Clin Proc 1975;50:339–44.

- Ordas J, Boga JA, Alvarez-Arguelles M, Villa L, Rodriguez-Dehli C, de Ona M, et al. Role of metapneumovirus in viral respiratory infections in young children. J Clin Microbiol 2006;44(8):2739–42.
- Papadopoulos NG, Moustaki M, Tsolia M, Bossios A, Astra E, Prezerakou A, et al. Association of rhinovirus infection with increased disease severity in acute bronchiolitis. Am J Respir Crit Care Med 2002;165(9): 1285–9.
- Pierangeli A, Gentile M, Di Marco P, Pagnotti P, Scagnolari C, Trombetti S, et al. Detection and typing by molecular techniques of respiratory viruses in children hospitalized for acute respiratory infection in Rome, Italy. J Med Virol 2007;79(4):463–8.
- Pitkaranta A, Arruda E, Malmberg H, Hayden FG. Detection of rhinovirus in sinus brushings of patients with acute community-acquired sinusitis by reverse transcription-PCR. J Clin Microbiol 1997;35(7):1791–3.
- Puro V, Minosse C, Cappiello G, Lauria FN, Capobianchi MR. Rhinovirus and lower respiratory tract infection in adults. Clin Infect Dis 2005;40(7):1068–9.
- Singh AM, Busse WW. Human rhinovirus models in asthma. Contrib Microbiol 2007;14:12–20.
- Steininger C, Aberle SW, Popow-Kraupp T. Early detection of acute rhinovirus infections by a rapid reverse transcription-PCR assay. J Clin Microbiol 2001;39(1):129–33.

- Tsolia MN, Psarras S, Bossios A, Audi H, Paldanius M, Gourgiotis D, et al. Etiology of community-acquired pneumonia in hospitalized school-age children: evidence for high prevalence of viral infections. Clin Infect Dis 2004;39(5):681–6.
- van den Hoogen BG, de Jong JC, Groen J, Kuiken T, de Groot R, Fouchier RA, et al. A newly discovered human pneumovirus isolated from young children with respiratory tract disease. Nat Med 2001;7(6):719– 24.
- van der Hoek L, Pyrc K, Jebbink MF, Vermeulen-Oost W, Berkhout RJ, Wolthers KC, et al. Identification of a new human coronavirus. Nat Med 2004;10(4):368–73.
- Woo PC, Lau SK, Chu CM, Chan KH, Tsoi HW, Huang Y, et al. Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. J Virol 2005;79(2):884– 95.
- Xatzipsalti M, Kyrana S, Tsolia M, Psarras S, Bossios A, Laza-Stanca V, et al. Rhinovirus viremia in children with respiratory infections. Am J Respir Crit Care Med 2005;172(8):1037–40.
- Yun BY, Kim MR, Park JY, Choi EH, Lee HJ, Yun CK. Viral etiology and epidemiology of acute lower respiratory tract infections in Korean children. Pediatr Infect Dis J 1995;14(12):1054–9.