




CORRESPONDENCE

Anti-IgE IgG autoantibodies isolated from therapeutic normal IgG intravenous immunoglobulin induce basophil activation

Caroline Galeotti^{1,2}, Anupama Karnam¹, Jordan D. Dimitrov^{1,3}, Alain Chevailler⁴, Srini V. Kaveri^{1,3} and Jagadeesh Bayry^{1,3} 

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Basophils are rare granulocytes. Despite representing only ~0.5% of all leukocytes, basophils have several important physiological functions.^{1,2} Although basophils lack the classic features of professional antigen-presenting cells,^{3–7} through the secretion of cytokines, they orient the immune response by polarizing Th2 differentiation and supporting B-cell differentiation and class switching. Basophils are also critical for mediating protection against helminth infection.^{1,2,8,9}

Basophils receive activation signals from diverse sources. It is well recognized that cytokines such as IL-3, granulocyte–macrophage colony-stimulating factor (GM-CSF), thymic stromal lymphopoietin (TSLP) and IL-33; various toll-like receptor ligands; allergen-bound IgE provide activation signals to basophils and induce the release of inflammatory mediators.^{10–15} In addition, several reports have also demonstrated the existence of anti-IgE autoantibodies that possess the capacity to induce basophil activation in patients with chronic spontaneous urticaria (CSU), atopic or non-atopic asthma or autoimmune disease.^{16–21} However, isolation and functional exploration of such anti-IgE IgG autoantibodies from either healthy donors or patients have not been attempted yet. By using a pooled normal IgG preparation from healthy donors, specifically intravenous immunoglobulin G (IVIg)²² that represents the complete IgG repertoire of a normal individual, we attempted to address this outstanding question in the field.

Recently, we reported that at a concentration (25 mg/0.5 million cells/mL) corresponding to the level of IgG reached in patients immediately following high-dose IVIg therapy, both IVIg and its F(ab')₂ fragments induce basophil activation.²³ Mechanistically, IVIg induces basophil activation by signaling through basophil surface-bound IgE in an IL-3- and Syk-dependent mechanism.²³ We first performed dose–response experiments with various concentrations of IVIg to determine whether a lower concentration of IVIg also induces basophil activation.

As a source of IVIg, Sandoglobulin® (CSL Behring, Switzerland) was used in our experiments. It was dialyzed several times against phosphate-buffered saline followed by final dialysis in RPMI-1640 medium at 4°C to remove stabilizing agents. Peripheral blood basophils were isolated from the buffy bags of healthy donors obtained from the Centre Necker-Cabanel, EFS, Paris (INSERM-EFS ethical permission N°18/EFS/033). The cellular fractions that contained peripheral blood mononuclear cells and basophils were collected by Ficoll density gradient centrifugation. From these

fractions, basophils were isolated by using a basophil isolation kit II (Miltenyi Biotec) and autoMACS⁺ (Miltenyi Biotec). The purity of the isolated basophils was 96–97%.

Basophils (0.1 × 10⁶ cells/well/200 µL) were seeded in 96-well U-bottomed plates. For dose–response experiments with IVIg, basophils were cultured either alone in serum-free X-VIVO 15 medium or with IL-3 (100 ng/mL, ImmunoTools) or IL-3 plus three different doses of IVIg (25, 15, or 10 mg/mL) for 24 h. IVIg was added to the basophils following 2 h of stimulation with IL-3. The basophils were analyzed for the expression of CD69 by flow cytometry (LSR II, BD Biosciences) using a CD69-APC/Cy7 MAb from BD Biosciences. Flow cytometry data were analyzed by BD FACS DIVA (BD Biosciences) and FlowJo (FlowJo LLC). Cell-free culture supernatants were used for cytokine analyses by ELISA (ELISA Ready-SET-Go, eBioscience Affymetrix).

As IVIg did not modify the activation status of either resting basophils or basophils primed with the cytokines IL-33, TSLP, IL-25, or GM-CSF,²³ we used IL-3 for priming throughout the experiments. The dose–response experiments revealed that even at a concentration of 10 mg/mL, which corresponds to circulating IgG levels in healthy donors, IVIg could induce basophil activation, as shown by the enhanced expression of CD69 and secretion of IL-4 (Fig. 1a, b). These results suggest that the occurrence of anti-IgE IgG autoantibodies is a common feature in the healthy population.

We then investigated whether the induction of basophil activation by IVIg is due to nonspecific cell activation caused by cell death. Therefore, we measured the viability of cells by Annexin V staining. Basophils were cultured in medium alone or with IL-3, IL-3 plus IVIg (25 mg/mL), or IL-3 plus human serum albumin (HSA, 10 mg/mL, Laboratoire Français du Fractionnement et des Biotechnologies, France) for 24 h. HSA was used as an irrelevant protein control for IVIg. The cells were stained with Annexin V (APC-Annexin V, BD Biosciences) and propidium iodide (PI; Invitrogen™). We indeed confirmed that basophil activation induced by IVIg was not because of nonspecific cell activation caused by cell death. In fact, cell viability as analyzed by Annexin V–PI staining was similar between the IL-3-treated, IVIg-treated, and HSA-treated cells (Fig. 1c, d).

These data prompted us to investigate the anti-IgE IgG autoantibodies in IVIg. An ELISA performed with recombinant human IgE-kappa (AbD Serotec) revealed that both IVIg and its F(ab')₂ fragments recognize IgE in a dose-dependent manner

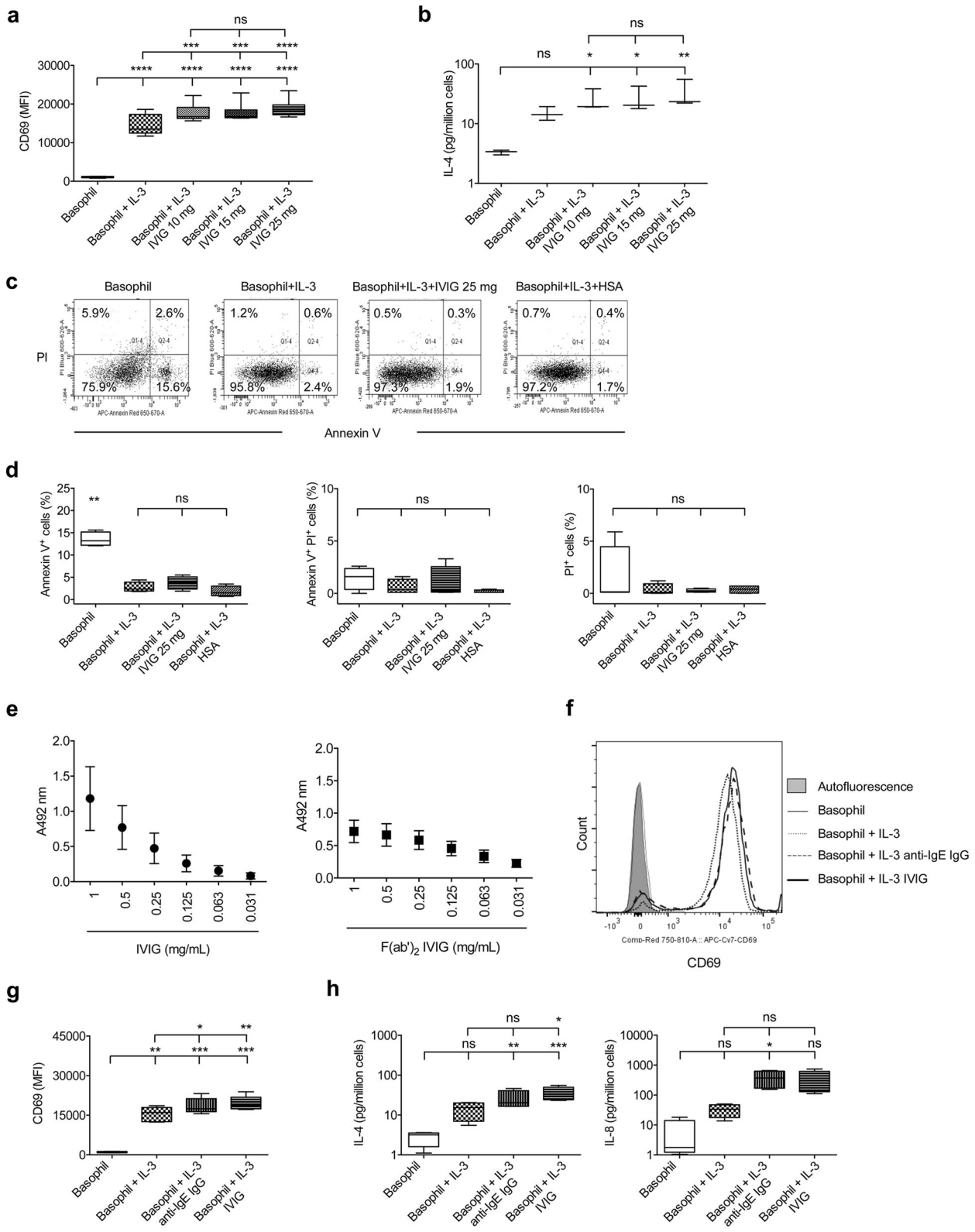
¹Institut National de la Santé et de la Recherche Médicale; Centre de Recherche des Cordeliers, Equipe-Immunopathologie et Immunointervention Thérapeutique, Sorbonne Université, Paris F-75006, France; ²Service de Rhumatologie Pédiatrique, Centre de Référence des Maladies Auto-Inflammatoires Rares et des Amyloses, CHU de Bicêtre, le Kremlin Bicêtre F-94270, France; ³Université Paris Descartes, Sorbonne Paris Cité, Paris F-75006, France and ⁴Laboratoire d'Immunologie et d'Allergologie; CHU d'Angers, Université d'Angers; INSERM Unité 1232; LabEx IGO "Immuno-Graft-Onco", Angers F-49933, France

Correspondence: Jagadeesh Bayry (jagadeesh.bayry@crc.jussieu.fr)

These authors contributed equally: Caroline Galeotti, Anupama Karnam

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(Fig. 1e). Therefore, we next aimed to isolate anti-IgE IgG autoantibodies from IVIG by affinity chromatography. The IgE myeloma protein was coupled to cyanogen bromide-activated sepharose 4B (Sigma-Aldrich). IVIG at a concentration of 60 mg/mL was added to the column and incubated at ambient temperature for 4 h on a rotator. After draining the unbound

IgG, the column was washed several times with phosphate-buffered saline. The anti-IgE IgG autoantibodies were eluted by using 3 M potassium thiocyanate, dialyzed against phosphate-buffered saline and concentrated. The IgG concentration was measured by using a spectrophotometer (NanoDrop Technologies).

Fig. 1 Anti-IgE IgG autoantibodies isolated from IVIG induce basophil activation. **a, b** Basophils isolated from healthy donors were cultured either alone (0.1×10^6 cells/well/200 μ L) or with IL-3 (100 ng/mL). Three different concentrations of IVIG (25, 15, or 10 mg/ml) were added to the cells following 2 h of stimulation with IL-3. After 24 h, the expression of CD69 (MFI, median fluorescence intensity; $n = 7$ donors) and the concentration of IL-4 ($n = 3$ donors) in the culture supernatants were analyzed. **c, d** The viability of basophils was analyzed by Annexin V and PI staining. Representative dot plots (**c**) and data from four experiments (**d**) using different donors are presented. human serum albumin (HSA) used at 10 mg/mL. **e** The recognition of IgE by IVIG (left panel) or (Fab')₂ fragments (right panel) was evaluated by ELISA. Data (mean \pm SEM) are from two experiments. **f, g** Basophils were cultured either alone or with IL-3 (100 ng/mL). IVIG (25 mg/mL) or anti-IgE IgG (1.5 mg/mL) was added to the cells following 2 h of stimulation with IL-3. After 24 h, the expression (MFI, median fluorescence intensity) of CD69 was analyzed. Representative histogram overlays (**f**) and data from five donors (**g**) are presented. **h** The amounts (pg) of IL-4 and IL-8 in culture supernatants from the above experiments were measured by ELISA. Data from four donors are presented. All data (except in panel **e**) are plotted using a box and whisker plot, wherein the whiskers denote the minimum and maximum values, and the dividing line in the box symbolizes the median. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; **** $P < 0.0001$; ns, not significant; one-way ANOVA with Tukey's multiple comparison test.

Based on the amount of eluted IgG, we estimated that anti-IgE IgG represent nearly 0.3–0.5% of total IgG (IVIG). We then investigated whether this isolated anti-IgE IgG is functional. IL-3-primed basophils from healthy individuals were stimulated with either IVIG (25 mg/0.5 $\times 10^6$ cells/mL) or isolated anti-IgE IgG (1.5 mg/0.5 $\times 10^6$ cells/mL) for 24 h. This concentration of anti-IgE IgG was selected according to the level of IgG and the number of basophils in the circulation of healthy individuals. We found that isolated anti-IgE IgG was able to induce basophil activation similar to that of IVIG even at a 16-fold lower concentration: both induced similar levels of CD69 expression (Fig. 1f, g) and IL-4 and IL-8 secretion (Fig. 1h).

The site of recognition of anti-IgE IgG autoantibodies from IVIG is not known. We used the IgE myeloma protein for the isolation of anti-IgE IgG, and yet these isolated IgG autoantibodies induced the activation of basophils from various donors carrying distinct IgE molecules, implying that anti-IgE IgG does not signal basophils by interacting with the Fab region of basophil-bound IgE. It is likely that these antibodies may recognize either the C ϵ 2 or C ϵ 4 domain of IgE but not the C ϵ 3 domain (which is the binding site of Fc ϵ RI and hence inaccessible for recognition by anti-IgE IgG), as indicated by a previous report;²⁴ however, we cannot rule out the possibility of recognition of the C ϵ 1 domain.

The precise function of anti-IgE IgG autoantibodies in healthy individuals is not clear at this stage. The isolation and characterization of such anti-IgE IgG monoclonal antibodies by using techniques, such as phage-display technology, should provide further insight into these antibodies.²⁵ Since anti-IgE autoantibodies induce IL-4 secretion in basophils, this function might support Th2 and B-cell differentiation. In the context of IVIG immunotherapy, however, we propose that IL-4 induction in basophils may help in the suppression of effector Th1 and Th17 cells²² and in the upregulation of inhibitory Fc γ RIIB expression on phagocytic cells to reduce the inflammatory response to IgG–autoantigen immune complexes.²⁶ Thus, distinct natural autoantibodies diversify the effects of IVIG on granulocytes.²⁷

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AUTHOR CONTRIBUTIONS

J.B. designed the study. C.G. and A.K. performed the experiments. C.G., A.K., J.D., S.V. K., and J.B. analyzed the data. A.C. provided essential tools. J.B. drafted the paper, and all the authors edited and approved the final version of the paper.

ADDITIONAL INFORMATION

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