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## **Advances in the In vitro and In vivo Pharmacology of Alpha4beta2 Nicotinic Receptor Positive Allosteric Modulators**

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### **Introduction**

Nicotinic receptors are ionotropic, ligand-gated ion channels that were originally defined by activation with nicotine, an alkaloid found in tobacco plants, and products made with tobacco. Nicotinic receptors are composed of five membrane spanning subunits (Cooper, 1991). As their full name implies, nAChR also are receptors for the endogenous neurotransmitter acetylcholine (ACh). Multiple neuronal nAChR subunits have been identified so far, named  $\alpha$ 2 to  $\alpha$ 10 and  $\beta$ 2 to  $\beta$ 4. Although many different possible combinations of subunits could come together to form an ion channel, certain α subunits are required for a functional binding site. In mammalian brain, homomeric receptors, which include five of the same  $\alpha$  subunit, are thought to be limited to those containing five  $\alpha$ 7 subunits. In contrast, multiple heteromeric nAChR subtypes containing both  $\alpha$  and  $\beta$ subunits have been identified. Each heteromeric nAChR subtype has a distinct biophysical property, physiological role, and pharmacology (Gotti et al., 2009).

The most prevalent heteromeric nAChR subtype in the mammalian brain are those made up of α4 and β2 subunits assembled with other nAChR subunits (Flores et al., 1997; Whiting and Lindstrom, 1987; Zoli et al., 2002), and is generally denoted as  $\alpha$ 4 $\beta$ 2\* nAChR with \* added to denote the possible involvement of additional subunits. When assembled alone, the α4 and β2 subunits form two distinct functional isoforms: low-sensitivity (α4)3(β2)2 and high-sensitivity  $(\alpha 4)2(\beta 2)3$  nAChRs with the low ACh sensitivity  $(\alpha 4)3(\beta 2)2$  representing

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the majority of nAChRs in the cortex (DeDominicis et al., 2017). The designation of low- or high- affinity reflects the effect of subunit ratio on potency. The combination of (3) α4 plus (2) β2 subunit ratios has been found to possess low sensitivity to ACh (i.e.,  $EC_{50} = 100 \mu M$ ), and is in contrast to the combination of (2)  $\alpha$ 4 plus (3)  $\beta$ 2 subunit ratios, which has been found to have high sensitivity to ACh (i.e.,  $EC_{50} = 1 \mu M$ ), (Moroni et al., 2006; Nelson et al., 2003). This effect is further reviewed in (Bertrand and Terry, 2018). The X-ray structure of human (α4)2(β2)3 nAChR has recently been published (Morales-Perez et al., 2016). The α4 and β2 subunits are highly homologous and share similar secondary structure, which consists of a 10-strand β sandwich N-terminal extracellular domain and a 4-helix bundle (TM1-TM4) transmembrane domain; however, they differ remarkably regarding their respective intracellular domains.

There is abundant evidence that the  $\alpha$ 4 $\beta$ 2\* subtype is of particular importance to the abuse potential of nicotine (Hurst et al., 2013; Besson et al., 2006; Picciotto et al., 1998). Anatomically, α4β2\* nAChR are found on midbrain (i.e., ventral tegmental area and substantia nigra) dopaminergic neuronal projections, which are well-characterized to be involved in the development and maintenance of drug dependence (Klink et al., 2001). Chronic (i.e., 2 mg/kg twice daily for 10 days) nicotine treatment in rats has been documented to lead to substantial and selective upregulation of the α4 and β2 nAChR subtypes (Flores et al., 1992). Varenicline, a currently approved smoking cessation aid, is believed to exert at least some of its therapeutic effects via its actions as a α4β2 nAChR partial agonist (Reus et al., 2007). In rats varenicline decreases the acquisition, expression and reinstatement of nicotine's effects in the place preference assay (Biala et al., 2010), and reduces the potency of nicotine in nicotine-induced intracranial self-stimulation (Vann et al., 2010). Additionally, in rats both varenicline and cytisine, another compound that has actions as a α4β2 nAChR partial agonist, decrease nicotine withdrawal-induced elevations in nicotine-induced intracranial self-stimulation thresholds, suggesting a decrease in nicotine withdrawal-induced dysphoria (Igari et al., 2014). The importance of the β2 subunit in the development and maintenance of nicotine dependence is supported by studies of β2 knockout mice, which do not self-administer nicotine. However, when β2 subunit functionality is restored, these mice self-administer nicotine (Picciotto et al., 1998). Additionally, when compared to wild-type mice,  $\alpha$ 4 knockout mice express fewer nicotine binding sites in the brain and have reduced neuronal activity and antinociceptive effects of nicotine (Marubio et al., 1999). Thus, pharmacotherapies that selectively target the  $\alpha$ 4 $\beta$ 2<sup>\*</sup> subtype of nAChR may be useful as smoking cessation therapeutics (Mohamed et al., 2015).

Both acute nicotine exposure and nicotine withdrawal in dependent subjects are welldocumented to impact cognitive function (Loughead et al., 2010; Potter and Newhouse, 2008; Maskos et al., 2005). Likewise, nAChR, including the  $\alpha$ 4 $\beta$ 2<sup>\*</sup> subtype, play a role in cognitive disorders, and nAChR agonists can produce improvement in neurological diseases that impact cognitive function (Potter and Newhouse, 2008; Hurst et al., 2013). Specifically, the α4β2 partial agonist ispronicline (TC-1734, AZD3480) improves memory tasks in both rats and mice, as well as humans (Dunbar et al., 2011; Gatto et al., 2004). Meanwhile, the α4 knockout mouse exhibits a significant decrease of nicotine binding in the brain and an increase in anxiety-like behaviors, such as a decrease in time spent in the open arm of the elevated plus maze (Ross et al., 2000). Meanwhile, deletion of the β2 nAChR subunit in

mice leads to improved responses in the passive avoidance task, which measures associative memory (Picciotto et al., 1995). However, these mice exhibit loss of sensitivity to nicotine's ability to increase performance in the same paradigm (Picciotto et al., 1995).

Alzheimer's disease is one neurological disease that may benefit from pharmacological interventions targeting the α4β2 nAChR. One hallmark of Alzheimer's Disease is the deterioration of the cholinergic system, including nAChR function (Kasa et al., 1997). It is believed that basal cholinergic tone in the forebrain is critical for normal cognition, and deterioration of this tone may be responsible for some of the cognitive impairments associated with Alzheimer's Disease (Auld et al., 2002). Further, a study of human postmortem brain tissue found a significant decrease in choline acetyltransferase, the enzyme responsible for ACh synthesis, but not nAChR density or function (Flynn and Mash, 1986). However, in human post-mortem tissue, significant decreases in  $\alpha$ 4 and  $\alpha$ 7 subunits are correlated with increased hyper-phosporylated tau protein (Wevers et al., 1999). Additional evidence shows that the α4β2 nAChR located in the temporal cortex are particularly susceptible in the pathology of Alzheimer's Disease (Warpman and Nordberg, 1995). In rats, the α4β2 nAChR is expressed within the motor cortex (DeDominicis et al., 2017), and in humans, it is believed that the motor cortex is significantly involved in the pathology of impaired motor function towards the later stages of Alzheimer's Disease (Suva et al., 1999). In rat cell culture experiments nicotine protects neurons from the cytotoxic effects of beta amyloid peptide, well known to be associated with the pathobiology of Alzheimer's Disease (Kihara et al., 1998). Further, these protective effects are mediated by the α4β2 nAChR, and cytisine, a α4β2 nAChR selective agonist, also produces similar protective effects (Kihara et al., 1998). Taken together, pharmacotherapies that selectively target the α4β2\* subtype of nAChR may be useful as therapeutics to treat neurological diseases that impact cognitive function.

There is substantial evidence that nAChR, including the  $\alpha$ 4 $\beta$ 2\* subtype, play a role in mediating pathological pain, and that nAChR agonists can produce analgesia (Hurst et al., 2013; Jain, 2004). Systemic delivery of nicotine both reverses and prevents mechanical allodynia in a mouse model of chemotherapy-induced peripheral neuropathy (Kyte et al., 2018). Anatomically, nAChR, including the  $\alpha$ 4 $\beta$ 2\* subtype, are found on neurons within peripheral nociceptive nerve fibers, dorsal root ganglia, the dorsal horn of the spinal cord, rostral ventromedial medulla in the brainstem, and the periaqueductal grey – all sites that mediate pain and pain processing (Hone et al., 2011; Umana et al., 2013). Additionally, astrocytes within the spinal cord and pain-processing regions in the brain also express α4β2 nAChR (Gotti and Clementi, 2004). Further, the α4β2 nAChR is upregulated in the rat ventralposterolateral thalamic nucleus starting at two weeks after a nerve injury model of neuropathic pain (Ueda et al., 2010). In rats, administration of 5-iodo-3-(2(S) azetidinylmethoxy)pyridine (5-iodo-A-85380), a selective and potent α4β2 nAChR agonist, administered into the ventralposterolateral thalamic nucleus dose-dependently reverses mechanical allodynia (Ueda et al., 2010). Additionally, the selective α4β2 nAChR agonist A-366833 dose-dependently reverses mechanical allodynia in a multitude of rat neuropathic pain models (i.e., chronic constriction injury of the sciatic nerve (CCI), partial sciatic nerve ligation, spinal nerve ligation, as well as diabetic- and chemotherapeutic-induced peripheral neuropathy), and in the Complete Freund's adjuvant-induced inflammatory pain model

(Nirogi et al., 2011). Recent evidence suggests that systemically administered α4β2 nAChR agonist TC-2559 decreases peripheral macrophage and spinal cord microglial activation concomitant with reversal of mechanical allodynia in a mouse partial sciatic nerve ligation model of neuropathic pain (Kiguchi et al., 2018). Thus, α4β2 nAChR orthosteric agonists may hold therapeutic promise for the treatment of pathological pain (Nirogi et al., 2013). In addition to nAChR that possess  $\alpha$ 4 $\beta$ 2 subunits, both the  $\alpha$ 5 and  $\alpha$ 7 nAChR subunits have also shown preclinical promise as targets for treating both inflammatory and neuropathic pathological pain (Freitas et al., 2013; Bagdas et al., 2015).

### **Positive allosteric modulators (PAMs) of** α**4**β**2 nAChRs**

As previously mentioned, the α4β2 nAChR is an attractive therapeutic target for nicotine dependence, neurological diseases that impair cognitive function, and pathological pain (Hurst et al., 2013). However, these therapeutic indications, especially with the use of orthosteric agonists of α4β2 nAChR, come with the possibility of producing addiction and dependence with prolonged use. Positive allosteric modulators (PAMs) can increase the binding affinity and/or efficacy of an orthosteric agonist (Pandya and Yakel, 2013), and may be a more advantageous long-term therapeutic strategy over othosteric α4β2 nAChR agonists (Wang and Lindstrom, 2018). Furthermore, numerous nAChR allosteric sites are thought to exist, with the exact number dependent on the subunit types that comprise the receptor (Wang and Lindstrom, 2018; Taly et al., 2009). Thus, PAMs represent a 'tunable' and highly selective method to influence nAChR pore activity. PAMs may produce increased receptor binding in several different ways. The prevailing consensus is that nAChRs are comprised of dynamic proteins, capable of multiple different states. Here we limit discussion to three possible states: closed, open and desensitized. At baseline equilibrium, when exogenous and endogenous signaling does not occur, nAChRs remain preferentially in the closed state. However, with an extremely low probability of shifting to the open state, the receptor may shift to the desensitized state. If a high concentration of agonist, sufficient to saturate all the possible binding sites is rapidly applied, α4β2 nAChRs have an 80% probability of simultaneous shifting transiently into the open state before reaching a new equilibrium in the desensitized state (Li and Steinbach, 2010). Furthermore, it is known that once receptors are in the desensitized state, agonists bind with much greater affinity (Gielen and Corringer, 2018; Papke et al., 2011).

One way that PAMs can exert their effects is by increasing the agonist binding to the closed state of the receptor, which is experimentally represented by an increase in the potency of the agonist. This type of modulation might be most advantageous under a condition where there is a low concentration of agonist which would not otherwise produce a maximal response. This lower concentration of an agonist is thought to limit potential off-target, or unwanted side-effects. Meanwhile, by increasing the potency of the agonist, lower concentrations would be able to produce the maximum intended target therapeutic response of the agonist (Grupe et al., 2015). However, it would remain impossible to exceed the maximum response if the modulator only changes the potency of the orthosteric ligand.

PAMs are often observed to increase the efficacy of an agonist. One way a PAM might accomplish this is via shifting the equilibrium between the open and closed states, making it

easier to move from closed to open state. This would result in not only more receptors moving to the open state, but also a greater likelihood that they might move from closed to open more than once before shifting to the desensitized state. Thus, this would yield a concurrent decrease in rate of desensitization. Functionally, this could be observed as an overall increase in the time spent in the open state. Having this effect, a PAM can produce a

transient increase in efficacy (Uteshev, 2014). Finally, a PAM might exert its effect by making the desensitized state less favorable. This would not be manifest as an increase in efficacy, or the maximum effect. Instead, it would be most apparent under conditions less affected by receptor desensitization.

The above is a simplified explanation of a theoretical model. Although there is experimental evidence that is consistent with these scenarios, it is not possible to prove or disprove them. These scenarios simply do not violate the current scientific knowledge. Furthermore, it has been reported that, at least under some conditions, only a small percentage of available nAChRs are capable of being activated at once (Li and Steinbach, 2010; McNerney et al., 2000). Thus, the ability of a modulator to change receptors from the inactive to the active state is yet another possible mechanism by which it might enhance agonist activity. Most commonly PAMs of nAChR are simplify classified into Type I or Type II PAMs based on above mentioned effects on nAChR gating kinetics and ACh-mediate responses. Type I generally refers to compound that enhances ACh sensitivity and peak ion current with no alteration of channel gating kinetics. Type II refers to compounds that have additional effects on channel gating kinetics (e.g. increase open channel duration, decrease desensitization) (Bertrand and Gopalakrishnan, 2007; Williams et al., 2011). Furthermore, a distinguishing feature of type II PAMs is that they can transiently reactivate desensitized nAChRs. Regardless of their exact mechanism, positive allosteric modulation of α4β2 nAChRs represents a potentially attractive therapeutic strategy which may circumvent the limitations inherent in targeting an orthosteric site. Importantly, PAMs possess the ability to amplify the amplitude of response patterns without necessarily altering the response pattern itself, such as what is observed with an agonist, which would result in sustained activation or enhanced desensitization pattern of responses. Examples of compounds that potentiate α4β2 nAChRs include physostigmine, galantamine, desformylflustrabromine (dFBr; N-(2-[6-bromo-2(1,1 dimethyl-2-propyl)-1H-indol-3-yl]ethyl-N-methylamine), NS9283 (3-[3-(pyridin-3 yl)-1,2,4-oxadiazol-5-yl]benzonitrile, CMPI (3-(2-chlorophenyl)-5-(5-methyl-1- (piperidin-4-yl)-1H-pyrrazol-4-yl)isoxazole), and LY2087101 ([2-[(4- Fluorophenyl)amino]-4-methyl-5-thiazolyl]-3-thienylmethanone). Figure 1 shows the structures of these compounds.

### **Physostigmine and Galantamine**

Physostigmine and galantamine are acetylcholinesterase (AChE) inhibitors and among the first nAChR allosteric potentiators to be identified (Maelicke et al., 2000). They are thought to potentiate muscle and neuronal type nAChRs but not muscarinic acetylcholine receptors (Samochocki et al., 2003; Farlow, 2003; Maelicke et al., 2001). Binding sites for galantamine and physostigmine on nAChRs have been identified using photoaffinity labeling, mutational analyses and computational docking (Luttmann et al., 2009; Hamouda et al., 2013). Physostigmine is thought to bind at both the agonist-binding and non-agonist

binding subunit extracellular interfaces with the latter being equivalent to the positive allosteric binding site of benzodiazepines at  $\gamma$ -aminobutyric acid (GABA) type A receptors. Additional mutational analysis revealed physostigmine selectivity to the low-sensitivity (α4)3(β2)2 nAChRs and physostigmine's ability to potentiate (α4)2α5(β2)2 nAChR (Jin et al., 2014). Galantamine is one of the most widely prescribed drugs for Alzheimer's disease, and is also under examination as a smoking cessation pharmacotherapy (Schilström et al., 2007). Galantamine has been found to enhance dopaminergic neurotransmission through potentiation of both α7 and α4β2 nAChRs (Schilstrom et al., 2007). Altered dopamine signaling in the brain reward pathway (i.e., ventral tegmental area, nucleus accumbens, prefrontal cortex) is one of the major mechanisms that is critical for the development and maintenance of nicotine addiction and dependence (Mansvelder et al., 2002). Galantamine has been shown to attenuate nicotine self-administration in rats (Hopkins et al., 2012; Liu, 2013), and to reduce cigarette smoking among alcohol-dependent patients (Diehl et al., 2006). However, recent studies suggest that galantamine does not functionally act at human α4β2 or α7 nAChRs as a PAM (Kowal et al., 2018). Further, if galantamine or physostigmine produce PAM-like effects via increasing acetylcholine tone, and thus sufficient to produce nicotine-like effects, then one might predict other acetylcholinesterase inhibitors may produce similar results when applied to smoking cessation. Thus, it is unclear which mechanism of galantamine, AChE inhibition or allosteric modulation of nAChRs, might be responsible for its capacity to promote smoking cessation. Regardless of the exact mechanism, the pharmacology of galantamine, and possibly physostigmine, represents a potential repurposed target that may have potential as an aid for smoking cessation.

### **dFBr (desformylflustrabromine)**

Desformylflustrabromine (dFBr) is one of the most studied nAChR PAMs and was discovered by serendipity. dFBr was first isolated, along with many other alkaloid metabolites, from the marine bryozoan Flustra foliacea and identified as a novel bromotryptamine derivative with pharmacological effects on nAChR (Peter et al., 2002; Sala et al., 2005). Following this discovery, syntheses of dFBr (in base form, water-soluble salts, and radioactive  $[3H]$ dFBr form) a large library of dFBr analogues has been described and used to investigate the *in vitro* nAChR effects, identify binding sites, and determine the structure-activity relationship of this novel nAChR pharmacophore (Lindel et al., 2006; Kim et al., 2007; German et al., 2011; Weltzin and Schulte, 2010; Pandya and Yakel, 2011; Hamouda et al., 2015, 2016; Deba et al., 2018; Dukat et al., 2018).

The effects of dFBr at muscle and neuronal nAChRs heterologously expressed in Xenpous laves oocytes or mammalian cell lines have been characterized using two-electrode voltageclamp and patch-clamp electrophysiological recordings, respectively. dFBr by itself did not elicit nAChR-mediated current (i.e. did not activate nAChR). However, it enhanced peak ACh-induced current responses of α4β2 nAChR and α2β2 at submicromolar concentrations (Sala et al., 2005; Kim et al., 2007; Weltzin and Schulte, 2010; Pandya and Yakel, 2011). Further, it was found that dFBr potentiates both (α4)2(β2)3 and (α4)3(β2)2 nAChR isoforms with potentiation  $EC_{50}$ s of ~0.4 and ~1.6 μM and potentiation Imax, of ~400 and ~300%, respectively, suggesting a stronger potentiation of  $(\alpha 4)3(\beta 2)2$  than  $(\alpha 4)2(\beta 2)3$ nAChR (Hamouda et al. 2016). The effect of dFBr on the ACh concentration-response

curves of  $(α4)3(β2)2$  and  $(α4)2(β2)3$  nAChRs was characterized by ~4-fold increase in ACh efficacy with minimal effects on ACh potency (Hamouda et al., 2016). Single channel recordings of α4β2 nAChR current in the presence of ACh in the absence or presence of dFBr suggested an increase in ion channel open probability, which could be a result of an increase in the opening rate constant and/or a decrease the closing rate constant (Sala et al., 2005). Therefore, dFBr is considered a Type II PAM of the α4β2 nAChR as it enhances peak current of low and saturated concentrations of ACh and alters channel gating (Wang and Lindstrom, 2017).

Interestingly, dFBr did not potentiate ACh-induced responses of muscle-type, α3β2, α3β4, α4β4, or α7 nAChRs. Instead, dFBr inhibited muscle-type and α7 nAChRs with an IC<sub>50</sub> of  $\sim$ 1μM and at higher concentration (>10 μM). dFBr inhibited ACh responses of most neuronal nAChRs including the α4β2 nAChRs (Sala et al., 2005; Kim et al., 2007; Weltzen and Schulte, 2010; Hamouda et al., 2015). Reversable binding analyses and  $[3H]$ dFBr photolabeling identified a higher affinity binding site for dFBr within the ion channel of muscle type nAChR (Hamouda et al., 2015), consistent with the notion that dFBr, at higher concentrations, inhibits nAChR responses by acting as an open channel blocker (Weltzen and Schulte, 2010; Hamouda et al., 2015). In addition, dFBr was found to bind at sites identified for physostigmine at the agonist-binding and non-agonist binding subunit extracellular interfaces in muscle-type nAChR (Hamouda et al., 2013; Hamouda et al., 2015).

There have been three recent publications using mutational analyses to identify dFBr binding sites in α4β2 nAChRs (Weltzin and Schulte, 2015; Alcaino et al., 2017; Deba et al., 2018). The first study (Weltzin and Schulte, 2015) reported that amino acid mutations in the β2 subunit significantly reduce dFBr potency in  $(α4)3(β2)2$  and  $(α4)2(β2)3$  nAChRs, suggesting an important role of the β2 subunit extracellular domain in dFBr modulation, especially the amino acid chain projecting from the β2 to the α4 subunit. However, a subsequent study (Alcaino et al., 2017) reported no effect of these mutations within the β2 subunit on dFBr modulation and using alanine substitution and substituted cysteine accessibility assay predicted a dFBr binding site within the the α4 subunit. The latter results were supported by mutational and computational docking analyses (Deba et al., 2018) which identified two district dFBr binding sites within the transmembrane domain of α4β2 nAChRs; an intrasubunit binding site within the α4 subunit helix bundle (equivalent to site identified in Alcaino et al., 2017) and an intersubunit site at the  $\alpha$ 4: $\alpha$ 4 subunit interface. Deba et al. 2018 also modeled the binding mode for dFBr within these sites as well as possible interactions between dFBr and amino acids within these binding sites which were found consistent with experimental values obtained with a panel of dFBr analogues (Dukat et al., 2018).

So far, all published dFBr structure activity data come from the Richard Glennon's group (German et al., 2011; Dukat et al., 2018). While dozens of dFBr analogues have been reported in these systematic structure activity studies, the naturally occurring structure of dFBr was found nearly optimal and none of these analogues were found to be substantially superior to dFBr in both efficacy and potency at α4β2 nAChR. Furthermore, when compared to dFBr, none of these dFBr analogues have exhibited significant improvement of

The first published *in vivo* use of dFBr was a study of rats trained to self-administer nicotine (0.03 mg/kg/infusion) (Liu, 2013). In the study, lower doses of dFBr (0.1 and 1 mg/kg) had no effect on nicotine self-administration, but larger doses (3 and 6 mg/kg) reduced the number of nicotine infusions earned. By itself, dFBr did not produce general depression of behavior. The elimination half-life of dFBr was estimated to be 8.6 hours and was present in the cerebrospinal fluid at about 30% of the concentration seen in plasma, indicating that dFBr crossed the blood-brain barrier. The observation that dFBr reduces nicotine selfadministration has been attributed to the production of aversive behaviors resulting from increased potency of nicotine. Specifically,  $\alpha$ 4β2 $\alpha$ 5 nAChR in the medial habenula have been reported contribute to aversive behaviors in response to high nicotine doses (Fowler and Kenny, 2013). Additionally, in mice, by itself dFBr produces dose-dependent hypothermia (Moerke et al., 2016) and when paired with nicotine dFBr produced synergistic effects to produce a nicotine discriminative stimulus (Moerke et al., 2016). Recent in vivo studies demonstrate that dFBr can reverse behavioral signs of nicotine withdrawal in nicotine-dependent mice (Hamouda et al., 2018).

Other in vivo studies have shown that dFBr decreases anxiety-like behavior in the mouse marble burying assay as well as in the open-field assay (Mitra et al., 2017). Further, dFBr has been found to block the inhibitory effect of  $\beta$  Amyloid (A $\beta$ 1–42) peptide and to restore the function of both α2β2 and α4β2 nAChR expressed in Xenopus oocytes (Pandya and Yakel, 2011). However, more studies are needed to determine the *in vivo* therapeutic implications of dFBr for the treatment of neurological diseases that impact cognitive function.

The nAChR PAM dFBr may also hold promise as a treatment for pathological pain. Specifically, dFBr increases the anti-allodynic effects of nicotine in a mouse model of CCI (Bagdas et al., 2017). Meanwhile, administration of dFBr alone appears to produce analgesic effects in both early and late phases of the formalin assay, as well as in the acetic acid writhing assay (Weggel and Pandya, 2019). The effects of the nAChR PAM dFBr are summarized in Table 1.

Overall, several lines of research, both in vitro and in vivo, have established the preclinical efficacy and tolerability of dFBr as a potent nAChR PAM that preferentially potentiates the α4β2\* nAChRs. Nevertheless, major limitations still face the preclinical development of dFBr into a clinically useful drug including: 1) dFBr inhibits the α7 nAChRs and muscletype nAChR at concentrations that are equivalent or only slightly higher than that required for α4β2 nAChR potentiation.; 2) following subcutaneous administration, only one third of a dFBr dose crossed the blood-brain barrier; and 3) variability of dFBr effects, alone or coadministered with agonist, in different animal models of pain; and 4) the fact that α4 and β2 nAChR subunits assemble in two stoichiometries  $((α4)3(β2)2$  and  $(α4)3(β2)2$  nAChRs) and can assemble with other nAChR subunits (e.g.  $\alpha$ 6,  $\alpha$ 5) forming a number of functionally and pharmacological unique nAChR subtypes that can be affected by dFBr. Therefore, the overall extent of dFBr's therapeutic effect is predicted to depend on the expression level of

each isoform in the affected brain area, the relative contributions of these subtypes to a specific disease state, and the extent to which dFBr can potentiate each α4β2\* nAChR isoform. However, the extent to which these potential issues might be relevant to the clinical use of dFBr is currently unknown.

### **LY 2087101 ([2-[(4-Fluorophenyl)amino]-4-methyl-5-thiazolyl]-3 thienylmethanone)**

Drug development and high-throughput screening of the chemical library at Eli Lilly and Company has identified three 2-amino-2-keto-thiazole derivatives (LY2087101, LY1078733, and LY2087133) with nAChR PAM activity (Broad et al., 2006). In vitro, LY2087101 potentiated current responses of the α4β2, α4β4, and α7 nAChR but not in α3β2 or α3β4 nAChRs establishing LY2087101 as a nAChR PAM that interacts mainly with the α4 and α7 subunits (Broad et al., 2006). LY2087101 is considered a Type I nAChR PAM because it produced an increase in both potency and magnitude of nicotine-induced currents (Broad et al., 2006) with little effect on the rate of receptor desensitization (Young et al., 2008). Specifically regarding α4β2 nAChRs, LY2087101 was an equipotent potentiator (potentiation  $EC_{50} \sim 1 \mu M$ ) of both the low-sensitivity ( $\alpha$ 4)3( $\beta$ 2)2 nAChR and highsensitivity (α4)2(β2)3 nAChR isoform but 2-fold more efficacious at the (α4)3(β2)2 than (α4)2(β2)3 nAChR (Deba et al., 2018). LY2087101 does not appear to alter ACh potency and its effect on ACh efficacy was higher in the  $(\alpha 4)3(\beta 2)2$  than  $(\alpha 4)2(\beta 2)3$  nAChRs. The more profound effect of LY2087101 at the  $\alpha$ 4)3(β2)2 nAChR is thought to be consistent with the number of α4 subunits which confer LY2087101 binding to nAChRs. Subsequent mutational and computational analyses have identified two LY2087101 binding sites within the transmembrane domain of the  $(a4)3(B2)2$  nAChRs: one at the interface between two adjacent α4 subunits (intersubunit binding site) and the other within the α4 subunit transmembrane helix bundle (intrasubunit binding site), (Deba et al., 2018). The intrasubunit binding site within the α4 subunit was found to be equivalent to the binding site identified for LY2087101 in the helix bundle of α7 nAChR (Young et al., 2008). Molecular docking studies predicted that LY2087101 binding within the α4 subunit helix bundle (intrasubunit site) is governed by hydrogen bond interactions with amino acids in the third transmembrane helix (TM3) whereas LY2087101 binding within the intersubunit site is governed by multiple non-bond hydrophobic interactions with amino acids at the interface of the adjacent α4 subunits (Deba et al., 2018).

In vivo, LY2087101 failed to produce substitution for 1.0 mg/kg nicotine at any dose tested in the mouse drug discrimination assay, even at doses that produce significant reduction of schedule-controlled responding (Moerke et al., 2016). Further, when LY 2087101 is paired with doses of nicotine that do not produce significant substitution for 1 mg/kg nicotine, no potentiation is observed (Moerke et al., 2016). Thus, there appears to be a disconnect between the *in vitro* and *in vivo* literature regarding whether LY 2087101 is a true functional nAChR PAM. The effects of LY 2087101 are summarized in Table 1.

### **CMPI (3-(2-chlorophenyl)-5-(5-methyl-1-(piperidin-4-yl)-1H-pyrrazol-4 yl)isoxazole)**

CMPI was developed at Amgen Inc. via chemical modification of substituted piperidine structure with nAChR PAM activity (Albrecht et al., 2008; Springer et al., 2008). At submicromolar concentrations, CPMI potentiated α4β2 nAChR but not any other major nAChR subtype (Albrecht et al., 2008). Subsequent studies have established CMPI as a nAChR PAM that potentiates the low-sensitivity  $(α4)3(β2)2$  nAChR but not the highsensitivity  $(\alpha 4)2(\beta 2)3$  nAChR (Hamouda et al., 2016) and identified the binding site for CMPI within the extracellular domain at the  $\alpha$ 4: $\alpha$ 4 subunit interface which exists only in the (α4)3(β2)2 nAChR subtype (Wang et al., 2017). The effect of CMPI on ACh responses of (α4)3(β2)2 nAChR was characterized by enhancement of ACh potency by ~100 fold with no significant effect on the efficacy of ACh (Wang et al., 2017). While the selectively of CMPI to  $(\alpha 4)3(\beta 2)2$  nAChRs, the major subpopulation of nAChRs in the cortex (DeDominicis et al., 2017), is viewed as pharmacologically favorable, it has yet to be determined if CMPI has in vivo biological activity.

### **NS9283 (3-[3-(pyridin-3-yl)-1,2,4-oxadiazol-5-yl]benzonitrile**

NS9283 is an α2- and α4-selective nAChR PAM that was discovered via research efforts at Neurosearch A/S (Timmerman et al., 2012). It was found that NS9283 increased the potency of currents evoked with ACh in HEK293 cells transfected with human α4β2 nAChR. Further, it was found that NS9283 did not alter the rate of desensitization of currents evoked with ACh (Grupe et al., 2013). It has been found that NS9283 selectively and preferentially acts on nAChR with the combination of (3)  $\alpha$ 4 plus (2)  $\beta$ 2 subunit ratios (Grupe et al., 2013; Timmermann et al., 2012). Single channel recordings have shown that (α4)3(β2)2 nAChRs show high single channel conductance, brief mean open lifetime, and high potentiation by NS9283, whereas  $\alpha$ 4)3( $\beta$ 2)2 nAChRs display low single channel conductance, long mean open lifetime, and are not potentiated by NS9283 (Mazzaferro et al., 2017). It has been thought that NS9283 is a PAM which is only able to potentiate  $(\alpha 4)3(\beta 2)2$  nAChRs by binding to a site at the  $\alpha$ 4: $\alpha$ 4 subunit extracellular interface (Timmerman et al., 2012; Mazzaferro et al., 2011; Harpose et al. 2011; Eaton et al., 2014). However, studies suggest that NS9283 not actually a PAM, but is rather an agonist selective for the α4:α4 ACh binding site in the  $(\alpha 4)$ 3 stoichiometry (Wang et al., 2015). Further, in a recent single channel recording study, mutations of amino acid in the β2 subunit that contribute to the β2:α4 subunit extracellular interface decreased NS9283 effect, indicating that other subunit interfaces play essential role in NS9283 binding and/or potentiation of (α4)3(β2)2 nAChR (Mazzaferro et al., 2019).

In the rat drug discrimination assay, NS9283 fails to produce substitution for 0.4 mg/kg nicotine at any dose tested (Mohler et al., 2014). When NS9283 is paired with doses of nicotine that do not produce significant substitution for 0.4 mg/kg nicotine, full substitution is observed. In the rat self-administration assay, NS9283 is not readily self-administered (Maskos et al., 2017). However, both acute and repeated administration of NS9283 dosedependently reduces nicotine self-administration in rats (Maurer et al., 2017). The

observation that NS9283 reduces nicotine self-administration in rats suggest that it might contribute to enhanced desensitization of the  $(\alpha 4)$ 3 stoichiometry or that the  $(\alpha 4)$ 3 stoichiometry contributes to responses that trigger aversion, such those observed after high nicotine doses.

NS9283 alone did not alter pain threshold for mechanical allodynia in neuropathic pain model; However, it enhanced the analgesic efficacy of nicotinic agonist in same model suggested that NS9283 can be used to improve the therapeutic index of nicotinic agonist in pain treatment (Lee et al., 2011). NS9283 has shown promise in improving cognitive function in rodents. Specifically, in rats NS9283 improves social memory, task acquisition in the hippocampal-dependent spatial memory Morris Water Maze task, as well as attention performance in the five-choice serial reaction time task (Timmerman et al., 2012). However, more studies are needed to determine the *in vivo* therapeutic implications of NS9283 for the treatment of neurological diseases that impact cognitive function. The effects of the nAChR PAM NS9283 are summarized in Table 1.

### **Conclusions**

A major challenge in the development of novel nAChR therapeutics lies in the apparent probability that several different nAChR subtypes play important roles in the development and maintenance of nicotine dependence and addiction, cognitive function, and pathological pain. Further, these receptors can be dynamically regulated in different manners, which adds to the complexity of designing therapeutic interventions with limited side-effects. However, rightly so, the α4β2 nAChR remains a substantial focus of research for novel compounds to treat these diseases. The therapeutic utility of orthosteric α4β2 nAChR agonists leaves room for the development of better therapeutics for smoking cessation aids, cognitive and neurological diseases as well as pathological pain. PAMs at α4β2 nAChR are attractive alternatives, as lower concentration of an agonists is needed to produce maximal therapeutic effects, and this decrease in agonist dose is thought to limit potential off-target, or unwanted side-effects. Interestingly, it has recently been reported that the binding sites for dFBr, LY2087101, and Br-PBTC, a novel nAChR PAM at multiple subunits including  $\alpha$ 4 $\beta$ 2, have been reported to be very close to one another (Norleans et al., 2019). In this study, derivatives of Br-PBTC have been found that interact with α5 subunits, which suggests that it might be possible to develop PAMs selective for α4α5β2 nAChRs. Further, one limitation to the experimental compounds listed in this review is that when administered in vivo, most were administered via a systemic injection, leaving other routes of administration unexplored thus far. It may be that varying the route of administration may enhance the therapeutic window of these compounds. Regardless, α4β2 nAChR PAMs may hold promise as smoking cessation aids, as well as treatments for neurological diseases and pathological pain.

### **Nonstandard Abbreviations:**



**nAChR** nicotinic acetylcholine receptor **PAM** Positive Allosteric Modulator

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### **Highlights**

**•** Antimicrobial mechanism of carvacrol against uropathogenic *E. coli* 

- **•** Carvacrol demonstrated membrane depolarization and oxidative burst in E. coli
- Carvacrol induced the release of DNA, proteins and ions from *E. coli* cells
- **•** Carvacrol reduced the levels of inflammatory proteins COX-2 and iNOS.
- **•** Carvacrol inhibited cell mortality and β-lactamase enzyme activity



Physostigmine



dFBr



Galantamine



LY2087101







**NS9283** 

**Figure 1.**  Structures of positive allosteric modulators of α 4 β2 nAChR.

#### **Table 1.**

Positive allosteric modulators of α4β2 nAChR and the work done thus far investigating them as novel therapeutics.



\* See article text under the corresponding compound for supporting references.