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Advances in the *In vitro* and *In vivo* Pharmacology of Alpha4beta2 Nicotinic Receptor Positive Allosteric Modulators

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Introduction

Nicotinic receptors are ionotropic, ligand-gated ion channels that were originally defined by activation with nicotine, an alkaloid found in tobacco plants, and products made with tobacco. Nicotinic receptors are composed of five membrane spanning subunits (Cooper, 1991). As their full name implies, nAChR also are receptors for the endogenous neurotransmitter acetylcholine (ACh). Multiple neuronal nAChR subunits have been identified so far, named α 2 to α 10 and β 2 to β 4. Although many different possible combinations of subunits could come together to form an ion channel, certain α subunits are required for a functional binding site. In mammalian brain, homomeric receptors, which include five of the same α subunit, are thought to be limited to those containing five α 7 subunits. In contrast, multiple heteromeric nAChR subtypes containing both α and β subunits have been identified. Each heteromeric nAChR subtype has a distinct biophysical property, physiological role, and pharmacology (Gotti et al., 2009).

The most prevalent heteromeric nAChR subtype in the mammalian brain are those made up of α 4 and β 2 subunits assembled with other nAChR subunits (Flores et al., 1997; Whiting and Lindstrom, 1987; Zoli et al., 2002), and is generally denoted as α 4 β 2* nAChR with * added to denote the possible involvement of additional subunits. When assembled alone, the α 4 and β 2 subunits form two distinct functional isoforms: low-sensitivity (α 4)3(β 2)2 and high-sensitivity (α 4)2(β 2)3 nAChRs with the low ACh sensitivity (α 4)3(β 2)2 representing

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the majority of nAChRs in the cortex (DeDominicis et al., 2017). The designation of low- or high- affinity reflects the effect of subunit ratio on potency. The combination of (3) $\alpha 4$ plus (2) $\beta 2$ subunit ratios has been found to possess low sensitivity to ACh (i.e., EC₅₀ = 100 μ M), and is in contrast to the combination of (2) $\alpha 4$ plus (3) $\beta 2$ subunit ratios, which has been found to have high sensitivity to ACh (i.e., EC₅₀ = 1 μ M), (Moroni et al., 2006; Nelson et al., 2003). This effect is further reviewed in (Bertrand and Terry, 2018). The X-ray structure of human ($\alpha 4$)2($\beta 2$)3 nAChR has recently been published (Morales-Perez et al., 2016). The $\alpha 4$ and $\beta 2$ subunits are highly homologous and share similar secondary structure, which consists of a 10-strand β sandwich N-terminal extracellular domain and a 4-helix bundle (TM1-TM4) transmembrane domain; however, they differ remarkably regarding their respective intracellular domains.

There is abundant evidence that the $\alpha 4\beta 2^*$ subtype is of particular importance to the abuse potential of nicotine (Hurst et al., 2013; Besson et al., 2006; Picciotto et al., 1998). Anatomically, a4\beta2* nAChR are found on midbrain (i.e., ventral tegmental area and substantia nigra) dopaminergic neuronal projections, which are well-characterized to be involved in the development and maintenance of drug dependence (Klink et al., 2001). Chronic (i.e., 2 mg/kg twice daily for 10 days) nicotine treatment in rats has been documented to lead to substantial and selective upregulation of the α 4 and β 2 nAChR subtypes (Flores et al., 1992). Varenicline, a currently approved smoking cessation aid, is believed to exert at least some of its therapeutic effects via its actions as a $\alpha 4\beta 2$ nAChR partial agonist (Reus et al., 2007). In rats varenicline decreases the acquisition, expression and reinstatement of nicotine's effects in the place preference assay (Biala et al., 2010), and reduces the potency of nicotine in nicotine-induced intracranial self-stimulation (Vann et al., 2010). Additionally, in rats both varenicline and cytisine, another compound that has actions as a $\alpha 4\beta 2$ nAChR partial agonist, decrease nicotine withdrawal-induced elevations in nicotine-induced intracranial self-stimulation thresholds, suggesting a decrease in nicotine withdrawal-induced dysphoria (Igari et al., 2014). The importance of the β 2 subunit in the development and maintenance of nicotine dependence is supported by studies of $\beta 2$ knockout mice, which do not self-administer nicotine. However, when $\beta 2$ subunit functionality is restored, these mice self-administer nicotine (Picciotto et al., 1998). Additionally, when compared to wild-type mice, $\alpha 4$ knockout mice express fewer nicotine binding sites in the brain and have reduced neuronal activity and antinociceptive effects of nicotine (Marubio et al., 1999). Thus, pharmacotherapies that selectively target the $\alpha 4\beta 2^*$ subtype of nAChR may be useful as smoking cessation therapeutics (Mohamed et al., 2015).

Both acute nicotine exposure and nicotine withdrawal in dependent subjects are welldocumented to impact cognitive function (Loughead et al., 2010; Potter and Newhouse, 2008; Maskos et al., 2005). Likewise, nAChR, including the $\alpha 4\beta 2^*$ subtype, play a role in cognitive disorders, and nAChR agonists can produce improvement in neurological diseases that impact cognitive function (Potter and Newhouse, 2008; Hurst et al., 2013). Specifically, the $\alpha 4\beta 2$ partial agonist ispronicline (TC-1734, AZD3480) improves memory tasks in both rats and mice, as well as humans (Dunbar et al., 2011; Gatto et al., 2004). Meanwhile, the $\alpha 4$ knockout mouse exhibits a significant decrease of nicotine binding in the brain and an increase in anxiety-like behaviors, such as a decrease in time spent in the open arm of the elevated plus maze (Ross et al., 2000). Meanwhile, deletion of the $\beta 2$ nAChR subunit in

mice leads to improved responses in the passive avoidance task, which measures associative memory (Picciotto et al., 1995). However, these mice exhibit loss of sensitivity to nicotine's ability to increase performance in the same paradigm (Picciotto et al., 1995).

Alzheimer's disease is one neurological disease that may benefit from pharmacological interventions targeting the $\alpha 4\beta 2$ nAChR. One hallmark of Alzheimer's Disease is the deterioration of the cholinergic system, including nAChR function (Kasa et al., 1997). It is believed that basal cholinergic tone in the forebrain is critical for normal cognition, and deterioration of this tone may be responsible for some of the cognitive impairments associated with Alzheimer's Disease (Auld et al., 2002). Further, a study of human postmortem brain tissue found a significant decrease in choline acetyltransferase, the enzyme responsible for ACh synthesis, but not nAChR density or function (Flynn and Mash, 1986). However, in human post-mortem tissue, significant decreases in α 4 and α 7 subunits are correlated with increased hyper-phosporylated tau protein (Wevers et al., 1999). Additional evidence shows that the $\alpha 4\beta 2$ nAChR located in the temporal cortex are particularly susceptible in the pathology of Alzheimer's Disease (Warpman and Nordberg, 1995). In rats, the $\alpha 4\beta 2$ nAChR is expressed within the motor cortex (DeDominicis et al., 2017), and in humans, it is believed that the motor cortex is significantly involved in the pathology of impaired motor function towards the later stages of Alzheimer's Disease (Suva et al., 1999). In rat cell culture experiments nicotine protects neurons from the cytotoxic effects of beta amyloid peptide, well known to be associated with the pathobiology of Alzheimer's Disease (Kihara et al., 1998). Further, these protective effects are mediated by the $\alpha 4\beta 2$ nAChR, and cytisine, a $\alpha 4\beta 2$ nAChR selective agonist, also produces similar protective effects (Kihara et al., 1998). Taken together, pharmacotherapies that selectively target the $\alpha 4\beta 2^*$ subtype of nAChR may be useful as therapeutics to treat neurological diseases that impact cognitive function.

There is substantial evidence that nAChR, including the $\alpha 4\beta 2^*$ subtype, play a role in mediating pathological pain, and that nAChR agonists can produce analgesia (Hurst et al., 2013; Jain, 2004). Systemic delivery of nicotine both reverses and prevents mechanical allodynia in a mouse model of chemotherapy-induced peripheral neuropathy (Kyte et al., 2018). Anatomically, nAChR, including the $\alpha 4\beta 2^*$ subtype, are found on neurons within peripheral nociceptive nerve fibers, dorsal root ganglia, the dorsal horn of the spinal cord, rostral ventromedial medulla in the brainstem, and the periaqueductal grey – all sites that mediate pain and pain processing (Hone et al., 2011; Umana et al., 2013). Additionally, astrocytes within the spinal cord and pain-processing regions in the brain also express $\alpha 4\beta 2$ nAChR (Gotti and Clementi, 2004). Further, the $\alpha 4\beta 2$ nAChR is upregulated in the rat ventralposterolateral thalamic nucleus starting at two weeks after a nerve injury model of neuropathic pain (Ueda et al., 2010). In rats, administration of 5-iodo-3-(2(S)azetidinylmethoxy)pyridine (5-iodo-A-85380), a selective and potent a4\beta2 nAChR agonist, administered into the ventral posterolateral thalamic nucleus dose-dependently reverses mechanical allodynia (Ueda et al., 2010). Additionally, the selective $\alpha 4\beta 2$ nAChR agonist A-366833 dose-dependently reverses mechanical allodynia in a multitude of rat neuropathic pain models (i.e., chronic constriction injury of the sciatic nerve (CCI), partial sciatic nerve ligation, spinal nerve ligation, as well as diabetic- and chemotherapeutic-induced peripheral neuropathy), and in the Complete Freund's adjuvant-induced inflammatory pain model

(Nirogi et al., 2011). Recent evidence suggests that systemically administered $\alpha 4\beta 2$ nAChR agonist TC-2559 decreases peripheral macrophage and spinal cord microglial activation concomitant with reversal of mechanical allodynia in a mouse partial sciatic nerve ligation model of neuropathic pain (Kiguchi et al., 2018). Thus, $\alpha 4\beta 2$ nAChR orthosteric agonists may hold therapeutic promise for the treatment of pathological pain (Nirogi et al., 2013). In addition to nAChR that possess $\alpha 4\beta 2$ subunits, both the $\alpha 5$ and $\alpha 7$ nAChR subunits have also shown preclinical promise as targets for treating both inflammatory and neuropathic pathological pain (Freitas et al., 2013; Bagdas et al., 2015).

Positive allosteric modulators (PAMs) of α4β2 nAChRs

As previously mentioned, the $\alpha 4\beta 2$ nAChR is an attractive therapeutic target for nicotine dependence, neurological diseases that impair cognitive function, and pathological pain (Hurst et al., 2013). However, these therapeutic indications, especially with the use of orthosteric agonists of $\alpha 4\beta 2$ nAChR, come with the possibility of producing addiction and dependence with prolonged use. Positive allosteric modulators (PAMs) can increase the binding affinity and/or efficacy of an orthosteric agonist (Pandya and Yakel, 2013), and may be a more advantageous long-term therapeutic strategy over othosteric $\alpha 4\beta 2$ nAChR agonists (Wang and Lindstrom, 2018). Furthermore, numerous nAChR allosteric sites are thought to exist, with the exact number dependent on the subunit types that comprise the receptor (Wang and Lindstrom, 2018; Taly et al., 2009). Thus, PAMs represent a 'tunable' and highly selective method to influence nAChR pore activity. PAMs may produce increased receptor binding in several different ways. The prevailing consensus is that nAChRs are comprised of dynamic proteins, capable of multiple different states. Here we limit discussion to three possible states: closed, open and desensitized. At baseline equilibrium, when exogenous and endogenous signaling does not occur, nAChRs remain preferentially in the closed state. However, with an extremely low probability of shifting to the open state, the receptor may shift to the desensitized state. If a high concentration of agonist, sufficient to saturate all the possible binding sites is rapidly applied, $\alpha 4\beta 2$ nAChRs have an 80% probability of simultaneous shifting transiently into the open state before reaching a new equilibrium in the desensitized state (Li and Steinbach, 2010). Furthermore, it is known that once receptors are in the desensitized state, agonists bind with much greater affinity (Gielen and Corringer, 2018; Papke et al., 2011).

One way that PAMs can exert their effects is by increasing the agonist binding to the closed state of the receptor, which is experimentally represented by an increase in the potency of the agonist. This type of modulation might be most advantageous under a condition where there is a low concentration of agonist which would not otherwise produce a maximal response. This lower concentration of an agonist is thought to limit potential off-target, or unwanted side-effects. Meanwhile, by increasing the potency of the agonist, lower concentrations would be able to produce the maximum intended target therapeutic response of the agonist (Grupe et al., 2015). However, it would remain impossible to exceed the maximum response if the modulator only changes the potency of the orthosteric ligand.

PAMs are often observed to increase the efficacy of an agonist. One way a PAM might accomplish this is via shifting the equilibrium between the open and closed states, making it

easier to move from closed to open state. This would result in not only more receptors moving to the open state, but also a greater likelihood that they might move from closed to open more than once before shifting to the desensitized state. Thus, this would yield a concurrent decrease in rate of desensitization. Functionally, this could be observed as an overall increase in the time spent in the open state. Having this effect, a PAM can produce a transient increase in efficacy (Uteshev, 2014). Finally, a PAM might exert its effect by making the desensitized state less favorable. This would not be manifest as an increase in efficacy, or the maximum effect. Instead, it would be most apparent under conditions less affected by receptor desensitization.

The above is a simplified explanation of a theoretical model. Although there is experimental evidence that is consistent with these scenarios, it is not possible to prove or disprove them. These scenarios simply do not violate the current scientific knowledge. Furthermore, it has been reported that, at least under some conditions, only a small percentage of available nAChRs are capable of being activated at once (Li and Steinbach, 2010; McNerney et al., 2000). Thus, the ability of a modulator to change receptors from the inactive to the active state is yet another possible mechanism by which it might enhance agonist activity. Most commonly PAMs of nAChR are simplify classified into Type I or Type II PAMs based on above mentioned effects on nAChR gating kinetics and ACh-mediate responses. Type I generally refers to compound that enhances ACh sensitivity and peak ion current with no alteration of channel gating kinetics. Type II refers to compounds that have additional effects on channel gating kinetics (e.g. increase open channel duration, decrease desensitization) (Bertrand and Gopalakrishnan, 2007; Williams et al., 2011). Furthermore, a distinguishing feature of type II PAMs is that they can transiently reactivate desensitized nAChRs. Regardless of their exact mechanism, positive allosteric modulation of a4β2 nAChRs represents a potentially attractive therapeutic strategy which may circumvent the limitations inherent in targeting an orthosteric site. Importantly, PAMs possess the ability to amplify the amplitude of response patterns without necessarily altering the response pattern itself, such as what is observed with an agonist, which would result in sustained activation or enhanced desensitization pattern of responses. Examples of compounds that potentiate $\alpha 4\beta 2$ nAChRs include physostigmine, galantamine, desformylflustrabromine (dFBr; N-(2-[6-bromo-2(1,1dimethyl-2-propyl)-1H-indol-3-yl]ethyl-N-methylamine), NS9283 (3-[3-(pyridin-3yl)-1,2,4-oxadiazol-5-yl]benzonitrile, CMPI (3-(2-chlorophenyl)-5-(5-methyl-1-(piperidin-4-yl)-1H-pyrrazol-4-yl)isoxazole), and LY2087101 ([2-[(4-Fluorophenyl)amino]-4-methyl-5-thiazolyl]-3-thienylmethanone). Figure 1 shows the structures of these compounds.

Physostigmine and Galantamine

Physostigmine and galantamine are acetylcholinesterase (AChE) inhibitors and among the first nAChR allosteric potentiators to be identified (Maelicke et al., 2000). They are thought to potentiate muscle and neuronal type nAChRs but not muscarinic acetylcholine receptors (Samochocki et al., 2003; Farlow, 2003; Maelicke et al., 2001). Binding sites for galantamine and physostigmine on nAChRs have been identified using photoaffinity labeling, mutational analyses and computational docking (Luttmann et al., 2009; Hamouda et al., 2013). Physostigmine is thought to bind at both the agonist-binding and non-agonist

binding subunit extracellular interfaces with the latter being equivalent to the positive allosteric binding site of benzodiazepines at γ -aminobutyric acid (GABA) type A receptors. Additional mutational analysis revealed physostigmine selectivity to the low-sensitivity $(\alpha 4)3(\beta 2)2$ nAChRs and physostigmine's ability to potentiate $(\alpha 4)2\alpha 5(\beta 2)2$ nAChR (Jin et al., 2014). Galantamine is one of the most widely prescribed drugs for Alzheimer's disease, and is also under examination as a smoking cessation pharmacotherapy (Schilström et al., 2007). Galantamine has been found to enhance dopaminergic neurotransmission through potentiation of both α 7 and α 4 β 2 nAChRs (Schilstrom et al., 2007). Altered dopamine signaling in the brain reward pathway (i.e., ventral tegmental area, nucleus accumbens, prefrontal cortex) is one of the major mechanisms that is critical for the development and maintenance of nicotine addiction and dependence (Mansvelder et al., 2002). Galantamine has been shown to attenuate nicotine self-administration in rats (Hopkins et al., 2012; Liu, 2013), and to reduce cigarette smoking among alcohol-dependent patients (Diehl et al., 2006). However, recent studies suggest that galantamine does not functionally act at human α4β2 or α7 nAChRs as a PAM (Kowal et al., 2018). Further, if galantamine or physostigmine produce PAM-like effects via increasing acetylcholine tone, and thus sufficient to produce nicotine-like effects, then one might predict other acetylcholinesterase inhibitors may produce similar results when applied to smoking cessation. Thus, it is unclear which mechanism of galantamine, AChE inhibition or allosteric modulation of nAChRs, might be responsible for its capacity to promote smoking cessation. Regardless of the exact mechanism, the pharmacology of galantamine, and possibly physostigmine, represents a potential repurposed target that may have potential as an aid for smoking cessation.

dFBr (desformylflustrabromine)

Desformylflustrabromine (dFBr) is one of the most studied nAChR PAMs and was discovered by serendipity. dFBr was first isolated, along with many other alkaloid metabolites, from the marine bryozoan *Flustra foliacea* and identified as a novel bromotryptamine derivative with pharmacological effects on nAChR (Peter et al., 2002; Sala et al., 2005). Following this discovery, syntheses of dFBr (in base form, water-soluble salts, and radioactive [³H]dFBr form) a large library of dFBr analogues has been described and used to investigate the *in vitro* nAChR effects, identify binding sites, and determine the structure-activity relationship of this novel nAChR pharmacophore (Lindel et al., 2006; Kim et al., 2007; German et al., 2011; Weltzin and Schulte, 2010; Pandya and Yakel, 2011; Hamouda et al., 2015, 2016; Deba et al., 2018; Dukat et al., 2018).

The effects of dFBr at muscle and neuronal nAChRs heterologously expressed in *Xenpous laves* oocytes or mammalian cell lines have been characterized using two-electrode voltageclamp and patch-clamp electrophysiological recordings, respectively. dFBr by itself did not elicit nAChR-mediated current (i.e. did not activate nAChR). However, it enhanced peak ACh-induced current responses of α 4 β 2 nAChR and α 2 β 2 at submicromolar concentrations (Sala et al., 2005; Kim et al., 2007; Weltzin and Schulte, 2010; Pandya and Yakel, 2011). Further, it was found that dFBr potentiates both (α 4)2(β 2)3 and (α 4)3(β 2)2 nAChR isoforms with potentiation EC₅₀s of ~0.4 and ~1.6 µM and potentiation Imax, of ~400 and ~300%, respectively, suggesting a stronger potentiation of (α 4)3(β 2)2 than (α 4)2(β 2)3 nAChR (Hamouda et al. 2016). The effect of dFBr on the ACh concentration-response

curves of $(\alpha 4)3(\beta 2)2$ and $(\alpha 4)2(\beta 2)3$ nAChRs was characterized by ~4-fold increase in ACh efficacy with minimal effects on ACh potency (Hamouda et al., 2016). Single channel recordings of $\alpha 4\beta 2$ nAChR current in the presence of ACh in the absence or presence of dFBr suggested an increase in ion channel open probability, which could be a result of an increase in the opening rate constant and/or a decrease the closing rate constant (Sala et al., 2005). Therefore, dFBr is considered a Type II PAM of the $\alpha 4\beta 2$ nAChR as it enhances peak current of low and saturated concentrations of ACh and alters channel gating (Wang and Lindstrom, 2017).

Interestingly, dFBr did not potentiate ACh-induced responses of muscle-type, $\alpha 3\beta 2$, $\alpha 3\beta 4$, $\alpha 4\beta 4$, or $\alpha 7$ nAChRs. Instead, dFBr inhibited muscle-type and $\alpha 7$ nAChRs with an IC₅₀ of ~1 μ M and at higher concentration (>10 μ M). dFBr inhibited ACh responses of most neuronal nAChRs including the $\alpha 4\beta 2$ nAChRs (Sala et al., 2005; Kim et al., 2007; Weltzen and Schulte, 2010; Hamouda et al., 2015). Reversable binding analyses and [³H]dFBr photolabeling identified a higher affinity binding site for dFBr within the ion channel of muscle type nAChR (Hamouda et al., 2015), consistent with the notion that dFBr, at higher concentrations, inhibits nAChR responses by acting as an open channel blocker (Weltzen and Schulte, 2010; Hamouda et al., 2015). In addition, dFBr was found to bind at sites identified for physostigmine at the agonist-binding and non-agonist binding subunit extracellular interfaces in muscle-type nAChR (Hamouda et al., 2013; Hamouda et al., 2015).

There have been three recent publications using mutational analyses to identify dFBr binding sites in $\alpha 4\beta 2$ nAChRs (Weltzin and Schulte, 2015; Alcaino et al., 2017; Deba et al., 2018). The first study (Weltzin and Schulte, 2015) reported that amino acid mutations in the β 2 subunit significantly reduce dFBr potency in (α 4)3(β 2)2 and (α 4)2(β 2)3 nAChRs, suggesting an important role of the β 2 subunit extracellular domain in dFBr modulation, especially the amino acid chain projecting from the $\beta 2$ to the $\alpha 4$ subunit. However, a subsequent study (Alcaino et al., 2017) reported no effect of these mutations within the β 2 subunit on dFBr modulation and using alanine substitution and substituted cysteine accessibility assay predicted a dFBr binding site within the the a4 subunit. The latter results were supported by mutational and computational docking analyses (Deba et al., 2018) which identified two district dFBr binding sites within the transmembrane domain of $\alpha 4\beta 2$ nAChRs; an intrasubunit binding site within the a4 subunit helix bundle (equivalent to site identified in Alcaino et al., 2017) and an intersubunit site at the α 4: α 4 subunit interface. Deba et al. 2018 also modeled the binding mode for dFBr within these sites as well as possible interactions between dFBr and amino acids within these binding sites which were found consistent with experimental values obtained with a panel of dFBr analogues (Dukat et al., 2018).

So far, all published dFBr structure activity data come from the Richard Glennon's group (German et al., 2011; Dukat et al., 2018). While dozens of dFBr analogues have been reported in these systematic structure activity studies, the naturally occurring structure of dFBr was found nearly optimal and none of these analogues were found to be substantially superior to dFBr in both efficacy and potency at $\alpha 4\beta 2$ nAChR. Furthermore, when compared to dFBr, none of these dFBr analogues have exhibited significant improvement of

The first published *in vivo* use of dFBr was a study of rats trained to self-administer nicotine (0.03 mg/kg/infusion) (Liu, 2013). In the study, lower doses of dFBr (0.1 and 1 mg/kg) had no effect on nicotine self-administration, but larger doses (3 and 6 mg/kg) reduced the number of nicotine infusions earned. By itself, dFBr did not produce general depression of behavior. The elimination half-life of dFBr was estimated to be 8.6 hours and was present in the cerebrospinal fluid at about 30% of the concentration seen in plasma, indicating that dFBr crossed the blood-brain barrier. The observation that dFBr reduces nicotine self-administration has been attributed to the production of aversive behaviors resulting from increased potency of nicotine. Specifically, $\alpha 4\beta 2\alpha 5$ nAChR in the medial habenula have been reported contribute to aversive behaviors in response to high nicotine doses (Fowler and Kenny, 2013). Additionally, in mice, by itself dFBr produces dose-dependent hypothermia (Moerke et al., 2016) and when paired with nicotine dFBr produced synergistic effects to produce a nicotine discriminative stimulus (Moerke et al., 2016). Recent *in vivo* studies demonstrate that dFBr can reverse behavioral signs of nicotine withdrawal in nicotine-dependent mice (Hamouda et al., 2018).

Other *in vivo* studies have shown that dFBr decreases anxiety-like behavior in the mouse marble burying assay as well as in the open-field assay (Mitra et al., 2017). Further, dFBr has been found to block the inhibitory effect of β Amyloid (A β 1–42) peptide and to restore the function of both α 2 β 2 and α 4 β 2 nAChR expressed in *Xenopus* oocytes (Pandya and Yakel, 2011). However, more studies are needed to determine the *in vivo* therapeutic implications of dFBr for the treatment of neurological diseases that impact cognitive function.

The nAChR PAM dFBr may also hold promise as a treatment for pathological pain. Specifically, dFBr increases the anti-allodynic effects of nicotine in a mouse model of CCI (Bagdas et al., 2017). Meanwhile, administration of dFBr alone appears to produce analgesic effects in both early and late phases of the formalin assay, as well as in the acetic acid writhing assay (Weggel and Pandya, 2019). The effects of the nAChR PAM dFBr are summarized in Table 1.

Overall, several lines of research, both *in vitro* and *in vivo*, have established the preclinical efficacy and tolerability of dFBr as a potent nAChR PAM that preferentially potentiates the $\alpha 4\beta 2^*$ nAChRs. Nevertheless, major limitations still face the preclinical development of dFBr into a clinically useful drug including: 1) dFBr inhibits the $\alpha 7$ nAChRs and muscle-type nAChR at concentrations that are equivalent or only slightly higher than that required for $\alpha 4\beta 2$ nAChR potentiation.; 2) following subcutaneous administration, only one third of a dFBr dose crossed the blood-brain barrier; and 3) variability of dFBr effects, alone or co-administered with agonist, in different animal models of pain; and 4) the fact that $\alpha 4$ and $\beta 2$ nAChR subunits assemble in two stoichiometries (($\alpha 4$)3($\beta 2$)2 and ($\alpha 4$)3($\beta 2$)2 nAChRs) and can assemble with other nAChR subunits (e.g. $\alpha 6$, $\alpha 5$) forming a number of functionally and pharmacological unique nAChR subtypes that can be affected by dFBr. Therefore, the overall extent of dFBr's therapeutic effect is predicted to depend on the expression level of

each isoform in the affected brain area, the relative contributions of these subtypes to a specific disease state, and the extent to which dFBr can potentiate each $\alpha.4\beta2^*$ nAChR isoform. However, the extent to which these potential issues might be relevant to the clinical use of dFBr is currently unknown.

LY 2087101 ([2-[(4-Fluorophenyl)amino]-4-methyl-5-thiazolyl]-3-thienylmethanone)

Drug development and high-throughput screening of the chemical library at Eli Lilly and Company has identified three 2-amino-2-keto-thiazole derivatives (LY2087101, LY1078733, and LY2087133) with nAChR PAM activity (Broad et al., 2006). In vitro, LY2087101 potentiated current responses of the $\alpha 4\beta 2$, $\alpha 4\beta 4$, and $\alpha 7$ nAChR but not in $\alpha 3\beta 2$ or $\alpha 3\beta 4$ nAChRs establishing LY2087101 as a nAChR PAM that interacts mainly with the a4 and a7 subunits (Broad et al., 2006). LY2087101 is considered a Type I nAChR PAM because it produced an increase in both potency and magnitude of nicotine-induced currents (Broad et al., 2006) with little effect on the rate of receptor desensitization (Young et al., 2008). Specifically regarding a4β2 nAChRs, LY2087101 was an equipotent potentiator (potentiation EC₅₀ ~1 μ M) of both the low-sensitivity (a4)3(β2)2 nAChR and highsensitivity $(\alpha 4)2(\beta 2)3$ nAChR isoform but 2-fold more efficacious at the $(\alpha 4)3(\beta 2)2$ than (α4)2(β2)3 nAChR (Deba et al., 2018). LY2087101 does not appear to alter ACh potency and its effect on ACh efficacy was higher in the $(\alpha 4)3(\beta 2)2$ than $(\alpha 4)2(\beta 2)3$ nAChRs. The more profound effect of LY2087101 at the $(\alpha 4)3(\beta 2)2$ nAChR is thought to be consistent with the number of a4 subunits which confer LY2087101 binding to nAChRs. Subsequent mutational and computational analyses have identified two LY2087101 binding sites within the transmembrane domain of the $(\alpha 4)3(\beta 2)2$ nAChRs: one at the interface between two adjacent a4 subunits (intersubunit binding site) and the other within the a4 subunit transmembrane helix bundle (intrasubunit binding site), (Deba et al., 2018). The intrasubunit binding site within the a4 subunit was found to be equivalent to the binding site identified for LY2087101 in the helix bundle of a7 nAChR (Young et al., 2008). Molecular docking studies predicted that LY2087101 binding within the α 4 subunit helix bundle (intrasubunit site) is governed by hydrogen bond interactions with amino acids in the third transmembrane helix (TM3) whereas LY2087101 binding within the intersubunit site is governed by multiple non-bond hydrophobic interactions with amino acids at the interface of the adjacent a4 subunits (Deba et al., 2018).

In vivo, LY2087101 failed to produce substitution for 1.0 mg/kg nicotine at any dose tested in the mouse drug discrimination assay, even at doses that produce significant reduction of schedule-controlled responding (Moerke et al., 2016). Further, when LY 2087101 is paired with doses of nicotine that do not produce significant substitution for 1 mg/kg nicotine, no potentiation is observed (Moerke et al., 2016). Thus, there appears to be a disconnect between the *in vitro* and *in vivo* literature regarding whether LY 2087101 is a true functional nAChR PAM. The effects of LY 2087101 are summarized in Table 1.

CMPI (3-(2-chlorophenyl)-5-(5-methyl-1-(piperidin-4-yl)-1H-pyrrazol-4yl)isoxazole)

CMPI was developed at Amgen Inc. via chemical modification of substituted piperidine structure with nAChR PAM activity (Albrecht et al., 2008; Springer et al., 2008). At submicromolar concentrations, CPMI potentiated $\alpha 4\beta 2$ nAChR but not any other major nAChR subtype (Albrecht et al., 2008). Subsequent studies have established CMPI as a nAChR PAM that potentiates the low-sensitivity ($\alpha 4$)3($\beta 2$)2 nAChR but not the highsensitivity ($\alpha 4$)2($\beta 2$)3 nAChR (Hamouda et al., 2016) and identified the binding site for CMPI within the extracellular domain at the $\alpha 4$: $\alpha 4$ subunit interface which exists only in the ($\alpha 4$)3($\beta 2$)2 nAChR subtype (Wang et al., 2017). The effect of CMPI on ACh responses of ($\alpha 4$)3($\beta 2$)2 nAChR was characterized by enhancement of ACh potency by ~100 fold with no significant effect on the efficacy of ACh (Wang et al., 2017). While the selectively of CMPI to ($\alpha 4$)3($\beta 2$)2 nAChRs, the major subpopulation of nAChRs in the cortex (DeDominicis et al., 2017), is viewed as pharmacologically favorable, it has yet to be determined if CMPI has *in vivo* biological activity.

NS9283 (3-[3-(pyridin-3-yl)-1,2,4-oxadiazol-5-yl]benzonitrile

NS9283 is an a2- and a4-selective nAChR PAM that was discovered via research efforts at Neurosearch A/S (Timmerman et al., 2012). It was found that NS9283 increased the potency of currents evoked with ACh in HEK293 cells transfected with human a4β2 nAChR. Further, it was found that NS9283 did not alter the rate of desensitization of currents evoked with ACh (Grupe et al., 2013). It has been found that NS9283 selectively and preferentially acts on nAChR with the combination of (3) α 4 plus (2) β 2 subunit ratios (Grupe et al., 2013; Timmermann et al., 2012). Single channel recordings have shown that $(\alpha 4)3(\beta 2)2$ nAChRs show high single channel conductance, brief mean open lifetime, and high potentiation by NS9283, whereas $(\alpha 4)3(\beta 2)2$ nAChRs display low single channel conductance, long mean open lifetime, and are not potentiated by NS9283 (Mazzaferro et al., 2017). It has been thought that NS9283 is a PAM which is only able to potentiate $(\alpha 4)3(\beta 2)2$ nAChRs by binding to a site at the a4:a4 subunit extracellular interface (Timmerman et al., 2012; Mazzaferro et al., 2011; Harpose et al. 2011; Eaton et al., 2014). However, studies suggest that NS9283 not actually a PAM, but is rather an agonist selective for the a4:a4 ACh binding site in the $(\alpha 4)$ 3 stoichiometry (Wang et al., 2015). Further, in a recent single channel recording study, mutations of amino acid in the β 2 subunit that contribute to the β 2:a4 subunit extracellular interface decreased NS9283 effect, indicating that other subunit interfaces play essential role in NS9283 binding and/or potentiation of $(\alpha 4)3(\beta 2)2$ nAChR (Mazzaferro et al., 2019).

In the rat drug discrimination assay, NS9283 fails to produce substitution for 0.4 mg/kg nicotine at any dose tested (Mohler et al., 2014). When NS9283 is paired with doses of nicotine that do not produce significant substitution for 0.4 mg/kg nicotine, full substitution is observed. In the rat self-administration assay, NS9283 is not readily self-administered (Maskos et al., 2017). However, both acute and repeated administration of NS9283 dose-dependently reduces nicotine self-administration in rats (Maurer et al., 2017). The

observation that NS9283 reduces nicotine self-administration in rats suggest that it might contribute to enhanced desensitization of the $(\alpha 4)3$ stoichiometry or that the $(\alpha 4)3$ stoichiometry contributes to responses that trigger aversion, such those observed after high nicotine doses.

NS9283 alone did not alter pain threshold for mechanical allodynia in neuropathic pain model; However, it enhanced the analgesic efficacy of nicotinic agonist in same model suggested that NS9283 can be used to improve the therapeutic index of nicotinic agonist in pain treatment (Lee et al., 2011). NS9283 has shown promise in improving cognitive function in rodents. Specifically, in rats NS9283 improves social memory, task acquisition in the hippocampal-dependent spatial memory Morris Water Maze task, as well as attention performance in the five-choice serial reaction time task (Timmerman et al., 2012). However, more studies are needed to determine the *in vivo* therapeutic implications of NS9283 for the treatment of neurological diseases that impact cognitive function. The effects of the nAChR PAM NS9283 are summarized in Table 1.

Conclusions

A major challenge in the development of novel nAChR therapeutics lies in the apparent probability that several different nAChR subtypes play important roles in the development and maintenance of nicotine dependence and addiction, cognitive function, and pathological pain. Further, these receptors can be dynamically regulated in different manners, which adds to the complexity of designing therapeutic interventions with limited side-effects. However, rightly so, the $\alpha 4\beta 2$ nAChR remains a substantial focus of research for novel compounds to treat these diseases. The therapeutic utility of orthosteric $\alpha 4\beta 2$ nAChR agonists leaves room for the development of better therapeutics for smoking cessation aids, cognitive and neurological diseases as well as pathological pain. PAMs at $\alpha 4\beta 2$ nAChR are attractive alternatives, as lower concentration of an agonists is needed to produce maximal therapeutic effects, and this decrease in agonist dose is thought to limit potential off-target, or unwanted side-effects. Interestingly, it has recently been reported that the binding sites for dFBr, LY2087101, and Br-PBTC, a novel nAChR PAM at multiple subunits including $\alpha 4\beta 2$, have been reported to be very close to one another (Norleans et al., 2019). In this study, derivatives of Br-PBTC have been found that interact with a 5 subunits, which suggests that it might be possible to develop PAMs selective for $a4a.5\beta2$ nAChRs. Further, one limitation to the experimental compounds listed in this review is that when administered in vivo, most were administered via a systemic injection, leaving other routes of administration unexplored thus far. It may be that varying the route of administration may enhance the therapeutic window of these compounds. Regardless, $\alpha 4\beta 2$ nAChR PAMs may hold promise as smoking cessation aids, as well as treatments for neurological diseases and pathological pain.

Nonstandard Abbreviations:

ACh	acetylcholine
CCI	chronic constriction injury

nAChR nicotinic acetylcholine receptorPAM Positive Allosteric Modulator

REFERENCES:

- Albrecht BK, Berry V, Boezio AA, Cao L, Clarkin K, Guo W, Harmange JC, Hierl M, Huang L, Janosky B, Knop J, Malmberg A, McDermott JS, Nguyen HQ, Springer SK, Waldon D, Woodin K, and McDonough SI, 2008 Discovery and optimization of substituted piperidines as potent, selective, CNS-penetrant alpha4beta2 nicotinic acetylcholine receptor potentiators. Bioorg Med Chem Lett. 18, 5209–12. 10.1016/j.bmcl.2008.08.080. [PubMed: 18789861]
- Alcaino C, Musgaard M, Minguez T, Mazzaferro S, Faundez M, Iturriaga-Vasquez P, Biggin PC, Bermudez I, 2017 Role of the Cys Loop and Transmembrane Domain in the Allosteric Modulation of α4β2 Nicotinic Acetylcholine Receptors. J Biol Chem. 292, 551–562. 10.1074/ jbc.M116.751206. [PubMed: 27864368]
- Auld DS, Kornecook TJ, Bastianetto S, Quirion R, 2002 Alzheimer's disease and the basal forebrain cholinergic system: relations to beta-amyloid peptides, cognition, and treatment strategies. Prog Neurobiol. 68, 209–45. 10.1016/S0301-0082(02)00079-5. [PubMed: 12450488]
- Bagdas D, AlSharari SD, Freitas K, Tracy M, Damaj MI, 2015 The role of alpha5 nicotinic acetylcholine receptors in mouse models of chronic inflammatory and neuropathic pain. Biochem. Pharmacol 97, 590–600. 10.1016/j.bcp.2015.04.013. [PubMed: 25931144]
- Bagdas D, Ergun D, Jackson A, Toma W, Schulte MK, Damaj MI, 2017 Allosteric modulation of $\alpha 4\beta 2^*$ nicotinic acetylcholine receptors: Desformylflustrabromine potentiates antiallodynic response of nicotine in a mouse model of neuropathic pain. Eur J Pain. 1, 84–93. 10.1002/ejp.1092.
- Bertrand D, Terry AV Jr., 2018 The wonderland of neuronal nicotinic acetylcholine receptors. Biochem Pharmacol 151, 214–225 10.1016/j.bcp.2017.12.008. [PubMed: 29248596]
- Besson M, David V, Suarez S, Cormier A, Cazala P, Changeux JP, Granon S, 2006 Genetic dissociation of two behaviors associated with nicotine addiction: beta-2 containing nicotinic receptors are involved in nicotine reinforcement but not in withdrawal syndrome. Psychopharmacology (Berl) 187, 189–199. 10.1007/s00213-006-0418-z. [PubMed: 16752141]
- Biala G, Staniak N, Budzynska B, 2010 Effects of varenicline and mecamylamine on the acquisition, expression, and reinstatement of nicotine-conditioned place preference by drug priming in rats. Naunyn Schmiedebergs Arch Pharmacol 381, 361–370. 10.1007/s00210-010-0498-5. [PubMed: 20217050]
- Broad LM, Zwart R, Pearson KH, Lee M, Wallace L, McPhie GI, Emkey R, Hollinshead SP, Dell CP, Baker SR, Sher E, 2006 Identification and pharmacological profile of a new class of selective nicotinic acetylcholine receptor potentiators. J Pharmacol Exp Ther. 318, 1108–1117. 10.1124/ jpet.106.104505. [PubMed: 16738207]
- Cooper E, Couturier S, Ballivet M, 1991 Pentameric structure and subunit stoichiometry of a neuronal nicotinic acetylcholine receptor. Nature 350, 235–238. 10.1038/350235a0. [PubMed: 2005979]
- Deba F, Ali HI, Tairu A, Ramos K, Ali JH, Hamouda A, 2018 LY2087101 and dFBr share transmembrane binding sites in the (α4)3(β2)2 Nicotinic Acetylcholine Receptor. Scientific Reports 8, 1249 10.1038/s41598-018-19790-4. [PubMed: 29352227]
- DeDominicis KE, Sahibzada N, Olson TT, Xiao Y, Wolfe BB, Kellar KJ, Yasuda RP, 2017 The (α4)3(β2)2 Stoichiometry of the Nicotinic Acetylcholine Receptor Predominates in the Rat Motor Cortex. Mol Pharmacol. 92, 327–337. 10.1124/mol.116.106880. [PubMed: 28698187]
- Diehl A, Nakovics H, Croissant B, Smolka MN, Batra A, Mann K, 2006 Galantamine reduces smoking in alcohol-dependent patients: a randomized, placebo-controlled trial. Int J Clin Pharmacol Ther 44, 614–622. 10.5414/cpp44614. [PubMed: 17190371]
- Dukat M, Jain A, German N, Ferrara-Pontoriero R, Huang Y, Ma Y, Schulte MK, Glennon RA, 2018 des-Formylflustrabromine (dFBr): A Structure-Activity Study on Its Ability To Potentiate the Action of Acetylcholine at α4β2 Nicotinic Acetylcholine Receptors. ACS Chem Neurosci. 12, 2984–2996. 10.1021/acschemneuro.8b00156.

- Dunbar GC, Kuchibhatla RV, Lee G, 2011 A randomized double-blind study comparing 25 and 50 mg TC-1734 (AZD3480) with placebo, in older subjects with age-associated memory Impairment. J Psychopharmacol 25, 1020–1029. 10.1111/j.1527-3458.2004.tb00010.x [PubMed: 20542923]
- Farlow MR, 2003 Clinical pharmacokinetics of galantamine. Clin Pharmacokinet 42, 1383–1392. 10.2165/00003088-200342150-00005. [PubMed: 14674789]
- Flores CM, Rogers SW, Pabreza LA, Wolfe BB, Kellar KJ, 1992 A subtype of nicotinic cholinergic receptor in rat brain is composed of alpha 4 and beta 2 subunits and is up-regulated by chronic nicotine treatment. Mol Pharmacol 41, 31–7. [PubMed: 1732720]
- Flores CM, Davila-Garcia MI, Ulrich YM, Kellar KJ, 1997 Differential regulation of neuronal nicotinic receptor binding sites following chronic nicotine administration. J Neurochem 69, 2216– 2219. 10.1046/j.1471-4159.1997.69052216.x. [PubMed: 9349569]
- Flynn DD, Mash DC, 1986 Characterization of L-[3H] nicotine binding in human cerebral cortex: comparison between Alzheimer's disease and the normal. J Neurochem. 6, 1948–54. 10.1111/ j.1471-4159.1986.tb13113.x.
- Fowler CD, Kenny PJ, 2013 Nicotine aversion: Neurobiological mechanisms and relevance to tobacco dependence vulnerability. Neuropharmacology 76 Pt B, 533–544. DOI: 10.1016/ j.neuropharm.2013.09.008 [PubMed: 24055497]
- Freitas K, Carroll FI, Damaj MI, 2013 The antinociceptive effects of nicotinic receptors a7-positive allosteric modulators in murine acute and tonic pain models. J Pharmacol Exp Ther. 11, 264–75. https://doi.org/0.1124/jpet.112.197871.
- Gatto GJ, Bohme GA, Caldwell WS, Letchworth SR, Traina VM, Obinu MC, Laville M, Reibaud M, Pradier L, Dunbar G, Bencherif M, 2004 TC-1734: an orally active neuronal nicotinic acetylcholine receptor modulator with antidepressant, neuroprotective and long-lasting cognitive effects. CNS Drug Rev 10, 147–166. 10.1111/j.1527-3458.2004.tb00010.x [PubMed: 15179444]
- German N, Kim JS, Jain A, Dukat M, Pandya A, Ma Y, Weltzin M, Schulte MK, Glennon RA, 2011 Deconstruction of the α4β2 nicotinic acetylcholine receptor positive allosteric modulator desformylflustrabromine. J Med Chem. 54, 7259–67. 10.1021/jm200834x. [PubMed: 21905680]
- Gielen M, Corringer PJ, 2018 The dual-gate model for pentameric ligand-gated ion channels activation and desensitization. J Physiol 596, 1873–1902. 10.1113/JP275100 [PubMed: 29484660]
- Gotti C, Clementi F, 2004 Neuronal nicotinic receptors: from structure to pathology. Prog Neurobiol. 74, 363–96. 10.1016/j.pneurobio.2004.09.006. [PubMed: 15649582]
- Grupe M, Grunnet M, Bastlund JF, Jensen AA, 2015 Targeting α4β2 nicotinic acetylcholine receptors in central nervous system disorders: perspectives on positive allosteric modulation as a therapeutic approach. Basic Clin. Pharmacol. Toxicol 116, 187–200. 10.1111/bcpt.12361. [PubMed: 25441336]
- Grupe M, Jensen AA, Ahring PK, Christensen JK, Grunnet M, 2013 Unravelling the mechanism of action of NS9283, a positive allosteric modulator of (alpha4)3(beta2)2 nicotinic ACh receptors. Br J Pharmacol 168, 2000–2010. 10.1111/bph.12095. [PubMed: 23278456]
- Hamouda AK, Deba F, Wang ZJ, Cohen JB, 2016 Photolabeling a Nicotinic Acetylcholine Receptor (nAChR) with an (α4)3(β2)2 nAChR-Selective Positive Allosteric Modulator. Mol Pharmacol. 89, 575–84. 10.1124/mol.116.103341 [PubMed: 26976945]
- Hamouda AK, Jayakar SS, Chiara DC, Cohen JB, 2014 Photoaffinity labeling of nicotinic receptors: diversity of drug binding sites! J Mol Neurosci. 53, 480–6. 10.1007/s12031-013-0150-1. [PubMed: 24158732]
- Hamouda AK, Kimm T, Cohen JB, 2013 Physostigmine and galanthamine bind in the presence of agonist at the canonical and noncanonical subunit interfaces of a nicotinic acetylcholine receptor. J Neurosci 33, 485–494. 10.1523/JNEUROSCI.3483-12.2013 [PubMed: 23303929]
- Hamouda AK, Wang ZJ, Stewart DS, Jain AD, Glennon RA, Cohen JB, 2015 Desformylflustrabromine (dFBr) and [3H]dFBr-Labeled Binding Sites in a Nicotinic Acetylcholine Receptor. Mol Pharmacol. 88, 1–11. 10.1124/mol.115.098913. [PubMed: 25870334]
- Hamouda AK, Jackson A, Bagdas D, Damaj MI, 2018 Reversal of Nicotine Withdrawal Signs Through Positive Allosteric Modulation of α4β2 Nicotinic Acetylcholine Receptors in Male Mice. Nicotine Tob Res. 20, 903–907. 10.1093/ntr/ntx183 [PubMed: 29059422]

- Harpsøe K, Ahring PK, Christensen JK, Jensen ML, Peters D, Balle T, 2011 Unraveling the high- and low-sensitivity agonist responses of nicotinic acetylcholine receptors. J. Neurosci 31, 10759– 10766. 10.1523/JNEUROSCI.1509-11.2011. [PubMed: 21795528]
- Hone AJ, Meyer EL, McIntyre M, McIntosh JM, 2011 Nicotinic acetylcholine receptors in dorsal root ganglion neurons include the $\alpha 6\beta 4^*$ subtype. FASEB J. 26, 91–26. 10.1096/fj.11-195883.
- Hopkins TJ, Rupprecht LE, Hayes MR, Blendy JA, Schmidt HD, 2012 Galantamine, an acetylcholinesterase inhibitor and positive allosteric modulator of nicotinic acetylcholine receptors, attenuates nicotine taking and seeking in rats. Neuropsychopharmacology 37, 2310– 2321. 10.1038/npp.2012.83. [PubMed: 22669169]
- Hurst R, Rollema H, Bertrand D, 2013 Nicotinic acetylcholine receptors: From basic science to therapeutics. Pharmacol. Ther 137, 22–54. 10.1016/j.pharmthera.08.012. [PubMed: 22925690]
- Igari M, Alexander JC, Ji Y, Qi X, Papke RL, Bruijnzeel AW, 2014 Varenicline and cytisine diminish the dysphoric-like state associated with spontaneous nicotine withdrawal in rats. Neuropsychopharmacology 39, 455–465. 10.1038/npp.2013.216. [PubMed: 23966067]
- Jain KK, 2004 Modulators of nicotinic acetylcholine receptors as analgesics. Curr Opin Investig Drugs. 5, 76–81.
- Jensen AA, Frølund B, Liljefors T, Krogsgaard-Larsen P, 2005 Neuronal nicotinic acetylcholine receptors: structural revelations, target identifications, and therapeutic inspirations. J Med Chem 48, 4705–4745. 10.1021/jm040219e. [PubMed: 16033252]
- Jin X, Bermudez I, Steinbach JH, 2014 The Nicotinic α5 Subunit Can Replace Either an Acetylcholine-Binding or Nonbinding Subunit in the α4β2* Neuronal Nicotinic Receptor. Mol Pharmacol. 85, 11–17. 10.1124/mol.113.089979. [PubMed: 24184962]
- Kalamida D, Poulas K, Avramopoulou V, Fostieri E, Lagoumintzis G, Lazaridis K, Sideri A, Zouridakis M, Tzartos SJ, 2007 Muscle and neuronal nicotinic acetylcholine receptors Structure, function and pathogenicity. FEBS 274, 3799–3845. 10.1111/j.1742-4658.2007.05935.x.
- Kasa P, Rakonczay Z, Gulya K, 1997 The cholinergic system in Alzheimer's disease. Prog. Neurobiol 52, 511–535. 10.1016/S0301-0082(97)00028-2 [PubMed: 9316159]
- Kiguchi N, Kobayashi D, Saika F, Matsuzaki S, Kishioka S, 2018 Inhibition of peripheral macrophages by nicotinic acetylcholine receptor agonists suppresses spinal microglial activation and neuropathic pain in mice with peripheral nerve injury. J Neuroinflam 15, 96 10.1186/ s12974-018-1133-5.
- Kihara T, Shimohama S, Urushitani M, Sawada H, Kimura J, Kume T, Maeda T, Akaike A, 1998 Stimulation of alpha4beta2 nicotinic acetylcholine receptors inhibits beta-amyloid toxicity. Brain Research 792, 331–334. 10.1016/s0006-8993(98)00138-3 [PubMed: 9593977]
- Kim JS, Padnya A, Weltzin M, Edmonds BW, Schulte MK, Glennon RA, 2007 Synthesis of desformylflustrabromine and its evaluation as an alpha4beta2 and alpha7 nACh receptor modulator. Bioorg Med Chem Lett 17, 4855–4860. 10.1016/j.bmcl.2007.06.047 [PubMed: 17604168]
- Klink R, de Kerchove d'Exaerde A, Zoli M, Changeux JP, 2001 Molecular and physiological diversity of nicotinic acetylcholine receptors in the midbrain dopaminergic nuclei. J Neurosci 21,1452– 1463. 10.1523/JNEUROSCI.21-05-01452.2001. [PubMed: 11222635]
- Kowal NM, Ahring PK, Liao VWY, Indurti DC, Harvey BS, O'Connor SM, Chebib M, Olafsdottir ES, Balle T, 2018 Galantamine is not a positive allosteric modulator of human alpha4beta2 or alpha7 nicotinic acetylcholine receptors. Br J Pharmacol 175, 2911–2925. 10.1111/bph.14329. [PubMed: 29669164]
- Kyte SL, Toma W, Bagdas D, Meade JA, Schurman LD, Lichtman AH, Chen ZJ, Del Fabbro E, Fang X, Bigbee JW, Damaj MI, Gewirtz DA., 2018 Nicotine Prevents and Reverses Paclitaxel-Induced Mechanical Allodynia in a Mouse Model of CIPN. J Pharmacol Exp Ther. 364,110–119. 10.1124/ jpet.117.243972 [PubMed: 29042416]
- Lee CH, Zhu C, Malysz J, Campbell T, Shaughnessy T, Honore P, Polakowski J, Gopalakrishnan M, 2011 $\alpha 4\beta 2$ neuronal nicotinic receptor positive allosteric modulation. An approach for improving the therapeutic index of $\alpha 4\beta 2$ nAChR agonists in pain. Biochem. Pharmacol 82, 959–966. 10.1016/j.bcp.2011.06.044. [PubMed: 21763685]

- Li P, Steinbach JH, 2010 The neuronal nicotinic alpha4beta2 receptor has a high maximal probability of being open. Br J Pharmacol 160, 1906–1915. 10.1111/j.1476-5381.2010.00761.x. [PubMed: 20649589]
- Lindel T, Bräuchle L, Golz G, Böhrer P, 2007 Total synthesis of flustramine C via dimethylallyl rearrangement. Org Lett 9, 283–6. 10.1021/ol0627348. [PubMed: 17217285]
- Liu X, 2013 Positive allosteric modulation of alpha4beta2 nicotinic acetylcholine receptors as a new approach to smoking reduction: evidence from a rat model of nicotine self-administration. Psychopharmacology (Berl) 230, 203–213. 10.1007/s00213-013-3145-2. [PubMed: 23712602]
- Loughead J, Ray R, Wileyto EP, Ruparel K, Sanborn P, Siegel S, Gur RC, Lerman C, 2010 Effects of the alpha4beta2 partial agonist varenicline on brain activity and working memory in abstinent smokers. Biol Psychiatry. 8, 715–21. 10.1016/j.biopsych.2010.01.016
- Luttmann E, Ludwig J, Höffle-Maas A, Samochocki M, Maelicke A, Fels G, 2009 Structural model for the binding sites of allosterically potentiating ligands on nicotinic acetylcholine receptors. Chem Med Chem 4, 1874–1882. 10.1002/cmdc.200900320 [PubMed: 19739198]
- Maelicke A, Samochocki M, Jostock R, Fehrenbacher A, Ludwig J, Albuquerque EX, Zerlin M, 2001 Allosteric sensitization of nicotinic receptors by galantamine, a new treatment strategy for Alzheimer's disease. Biol Psychiatry 49, 279–288. 10.1016/s0006-3223(00)01109-4. [PubMed: 11230879]
- Mansvelder HD, Keath JR, McGehee DS, 2002 Synaptic mechanisms underlie nicotine-induced excitability of brain reward areas. Neuron 33, 905–919. 10.1016/s0896-6273(02)00625-6. [PubMed: 11906697]
- Marubio LM, del Mar Arroyo-Jimenez M, Cordero-Erausquin M, Léna C, Le Novère N, de Kerchove d'Exaerde A, Huchet M, Damaj MI, Changeux JP 1999 Reduced antinociception in mice lacking neuronal nicotinic receptor subunits. Nature 398, 805–810. 10.1038/19756. [PubMed: 10235262]
- Maskos U, Molles BE, Pons S, Besson M, Guiard BP, Guilloux JP, Evrard A, Cazala P, Cormier A, Mameli-Engvall M, Dufour N, Tayarani-Cloëz I, Bemelmans AP, Mallet J, Gardier AM, David V, Faure P, Garnon S, Changeux JP, 2005 Nicotine reinforcement and cognition restored by targeted expression of nicotinic receptors. Nature 436, 103–107. 10.1038/nature03694. [PubMed: 16001069]
- Maurer JJ, Sandager-Nielsen K, Schmidt HD, 2017 Attenuation of nicotine taking and seeking in rats by the stoichiometry-selective alpha4beta2 nicotinic acetylcholine receptor positive allosteric modulator NS9283. Psychopharmacology (Berl). 234, 475–484. 10.1007/s00213-016-4475-7 [PubMed: 27844094]
- Mazzaferro S, Bermudez I, Sine SM, 2017 A4β2 nicotinic acetylcholine receptors: relationships between subunit stoichiometry and function at the single channel level. J Biol Chem. 292, 2729– 2740. 10.1074/jbc.M116.764183. [PubMed: 28031459]
- Mazzaferro S, Bermudez I, Sine SM, 2019 Potentiation of a neuronal nicotinic receptor via pseudoagonist site. Cell Mol Life Sci. 76, 1151–1167. 10.1007/s00018-018-2993-7. [PubMed: 30600358]
- Mazzaferro S, Benallegue N, Carbone A, Gasparri F, Vijayan R, Biggin PC, Moroni M, Bermudez I, 2011 Additional acetylcholine (ACh) binding site at alpha4/alpha4 interface of (alpha4beta2)2alpha4 nicotinic receptor influences agonist sensitivity. J Biol Chem. 286, 31043–54. 10.1074/jbc.M111.262014. [PubMed: 21757735]
- McNerney ME, Pardi D, Pugh PC, Nai Q, Margiotta JF, 2000 Expression and channel properties of alpha-bungarotoxin-sensitive acetylcholine receptors on chick ciliary and choroid neurons. J Neurophysiol 84, 1314–1329. 10.1152/jn.2000.84.3.1314. [PubMed: 10980005]
- Mitra S, Mucha M, Khatri SN, Glenon R, Schulte MK, Bult-Ito A, 2017 Attenuation of Compulsive-Like Behavior Through Positive Allosteric Modulation of α4β2 Nicotinic Acetylcholine Receptors in Non-Induced Compulsive-Like Mice. Front Behav Neurosci. 10, 244 10.3389/ fnbeh.2016.00244. [PubMed: 28105008]
- Moerke MJ, de Moura FB, Koek W, McMahon LR, 2016 Effects of nicotine in combination with drugs described as positive allosteric nicotinic acetylcholine receptor modulators in vitro: discriminative stimulus and hypothermic effects in mice. Eur J Pharmacol. 786, 169–178. 10.1016/ j.ejphar.2016.05.032. [PubMed: 27238974]

- Mohler EG, Franklin SR, Rueter LE, 2014 Discriminative-stimulus effects of NS9283, a nicotinic $\alpha 4\beta 2^*$ positive allosteric modulator, in nicotine-discriminating rats. Psychopharmacology (Berl). 231, 67–74. 10.1007/s00213-013-3207-5. [PubMed: 23925734]
- Morales-Perez CL, Noviello CM, Hibbs RE, 2016 X-ray structure of the human alpha4beta2 nicotinic receptor. Nature 538, 411–415. 10.1038/nature19785. [PubMed: 27698419]
- Moroni M, Zwart R, Sher E, Cassels BK and Bermudez I, 2006 alpha4beta2 nicotinic receptors with high and low acetylcholine sensitivity: pharmacology, stoichiometry, and sensitivity to long-term exposure to nicotine. Mol Pharmacol 70, 755–768. 10.1124/mol.106.023044. [PubMed: 16720757]
- Nelson ME, Kuryatov A, Choi CH, Zhou Y, Lindstrom J, 2003 Alternate stoichiometries of alpha4beta2 nicotinic acetylcholine receptors. Mol Pharmacol 63, 332–341. 10.1124/mol.63.2.332. [PubMed: 12527804]
- Nirogi R, Goura V, Abraham R, Jayarajan P, 2013 α4β2* neuronal nicotinic receptor ligands (agonist, partial agonist and positive allosteric modulators) as therapeutic prospects for pain. Eur J Pharmacol. 712, 22–9. 10.1016/j.ejphar.2013.04.021. [PubMed: 23660369]
- Nirogi R, Jabaris SL, Jayarajan P, Abraham R, Shanmuganathan D, Rasheed MA, Royapalley PK, Goura V, 2011 Antinociceptive activity of α4β2* neuronal nicotinic receptor agonist A-366833 in experimental models of neuropathic and inflammatory pain. Eur J Pharmacol. 668, 155–62. 10.1016/j.ejphar.2011.06.032. [PubMed: 21756895]
- Norleans J, Wang J, Kuryatov A, Leffler A, Doebelin C, Kamenecka TM, Lindstrom J, 2019 Discovery of an intrasubunit nicotinic acetylcholine receptor-binding site for the positive allosteric modulator Br-PBTC. J Biol Chem 294, 12132–12145. doi: 10.1074/jbc.RA118.006253 [PubMed: 31221718]
- Olsen JA, Kastrup JS, Peters D, Gajhede M, Balle T, Ahring PK, 2013 Two distinct allosteric binding sites at α4β2 nicotinic acetylcholine receptors revealed by NS206 and NS9283 give unique insights to binding activity-associated linkage at Cys-loop receptors. J Biol Chem. 288, 35997– 6006. 10.1074/jbc.M113.498618. [PubMed: 24169695]
- Olsen JA, Ahring PK, Kastrup JS, Gajhede M, Balle T, 2014 Structural and Functional Studies of the Modulator NS9283 Reveal Agonist-like Mechanism of Action at α4β2 Nicotinic Acetylcholine Receptors. J Biol Chem. 289, 24911–21. 10.1074/jbc.M114.568097. [PubMed: 24982426]
- Pandya A, Yakel JL, 2011 Allosteric modulator Desformylflustrabromine relieves the inhibition of $\alpha 2\beta 2$ and $\alpha 4\beta 2$ nicotinic acetylcholine receptors by β -amyloid(1–42) peptide. J Mol Neurosci 45, 42–47. 10.1007/s12031-011-9509-3. [PubMed: 21424792]
- Pandya AA, Yakel JL, 2013 Effects of neuronal nicotinic acetylcholine receptor allosteric modulators in animal behavior studies. Biochem Pharmacol 86, 1054–1062. 10.1016/j.bcp.2013.05.018. [PubMed: 23732296]
- Papke D, Gonzalez-Gutierrez G, Grosman C, 2011 Desensitization of neurotransmitter-gated ion channels during high-frequency stimulation: a comparative study of Cys-loop, AMPA and purinergic receptors. J Physiol 589, 1571–1585. 10.1113/jphysiol.2010.203315 [PubMed: 21300749]
- Peters L, König GM, Terlau H, Wright AD, 2002 Four new bromotryptamine derivatives from the marine bryozoan Flustra foliacea. J Nat Prod 65, 1633–1637. 10.1021/np0105984. [PubMed: 12444689]
- Picciotto MR, Zoli M, Léna C, Bessis A, Lallemand Y, Le Novère N, Vincent P, Pich EM, Brûlet P, Changeux JP, 1995 Abnormal avoidance learning in mice lacking functional high-affinity nicotine receptor in the brain. Nature 374, 65–67. 10.1038/374065a0 [PubMed: 7870173]
- Picciotto MR, Zoli M, Rimondini R, Léna C, Marubio LM, Pich EM, Fuxe K, Changeux JP, 1998 Acetylcholine receptors containing the beta2 subunit are involved in the reinforcing properties of nicotine. Nature 391, 173–177. 10.1038/34413. [PubMed: 9428762]
- Potter AS, Newhouse PA, 2008 Acute nicotine improves cognitive deficits in young adults with attention-deficit/hyperactivity disorder. Pharmacol Biochem Behav 88, 407–417. 10.1016/ j.pbb.2007.09.014. [PubMed: 18022679]
- Reus VI, Obach RS, Coe JW, Faessel H, Rollema H, Watsky E, Reeves K, 2007 Varenicline: new treatment with efficacy in smoking cessation. Drugs Today (Barc). 43, 65–75. 10.1358/ dot.2007.43.2.1069956 [PubMed: 17353944]

- Ross SA, Wong JY, Clifford JJ, Kinsella A, Massalas JS, Horne MK, Scheffer IE, Kola I, Waddington JL, Berkovic SF, Drago J, 2000 Phenotypic characterization of an alpha 4 neuronal nicotinic acetylcholine receptor subunit knock-out mouse. J Neurosci 20, 6431–6441. 10.1523/ JNEUROSCI.20-17-06431.2000. [PubMed: 10964949]
- Sala F, Mulet J, Reddy KP, Bernal JA, Wikman P, Valor LM, Peters L, König GM, Criado M, Sala S, 2005 Potentiation of human alpha4beta2 neuronal nicotinic receptors by a Flustra foliacea metabolite. Neurosci Lett. 373, 144–9. 10.1016/j.neulet.2004.10.002. [PubMed: 15567570]
- Samochocki M, Höffle A, Fehrenbacher A, Jostock R, Ludwig J, Christner C, Radina M, Zerlin M, Ullmer C, Pereira EF, Lübbert H, Albuquerque EX, Maelicke A, 2003 Galantamine is an allosterically potentiating ligand of neuronal nicotinic but not of muscarinic acetylcholine receptors. J Pharmacol Exp Ther 305, 1024–36. 10.1124/jpet.102.045773. [PubMed: 12649296]
- Schilström B, Ivanov VB, Wiker C, Svensson TH, 2007 Galantamine enhances dopaminergic neurotransmission in vivo via allosteric potentiation of nicotinic acetylcholine receptors. Neuropsychopharmacology 32, 43–53. 10.1038/sj.npp.1301087. [PubMed: 16641937]
- Springer SK, Woodin KS, Berry V, Boezio AA, Cao L, Clarkin K, Harmange JC, Hierl M, Knop J, Malmberg AB, McDermott JS, Nguyen HQ, Waldon D, Albrecht BK, McDonough SI, 2008 Synthesis and activity of substituted carbamates as potentiators of the alpha4beta2 nicotinic acetylcholine receptor. Bioorg Med Chem Lett. 18, 5643–7. 10.1016/j.bmcl.2008.08.092. [PubMed: 18805006]
- Suva D, Favre I, Kraftsik R, Esteban M, Lobrinus A, Miklossy J, 1999 Primary motor cortex involvement in Alzheimer disease. J Neuropathol Exp Neurol 58, 1125–1134. 10.1097/00005072-199911000-00002. [PubMed: 10560655]
- Taly A, Corringer PJ, Guedin D, Lestage P, Changeux JP, 2009 Nicotinic receptors: allosteric transitions and therapeutic targets in the nervous system. Nat Rev Drug Disc 8, 733–750. 10.1038/ nrd2927.
- Timmermann DB, Sandager-Nielsen K, Dyhring T, Smith M, Jacobsen AM, Nielsen EØ, Grunnet M, Christensen JK, Peters D, Kohlhaas K, Olsen GM, Ahring PK, 2012 Augmentation of cognitive function by NS9283, a stoichiometry-dependent positive allosteric modulator of α2- and α4containing nicotinic acetylcholine receptors. Br J Pharmacol. 167, 164–82. 10.1111/ j.1476-5381.2012.01989.x. [PubMed: 22506660]
- Ueda M, Iida Y, Tominaga A, Yoneyama T, Ogawa M, Magata Y, Nishimura H, Kuge Y, Saji H, 2010 Nicotinic acetylcholine receptors expressed in the ventralposterolateral thalamic nucleus play an important role in anti-allodynic effects. Br J Pharmacol. 6, 1201–10. 10.1111/ j.1476-5381.2009.00613.x.
- Umana IC, Daniele CA, McGehee DS, 2013 Neuronal nicotinic receptors as analgesic targets: it's a winding road. Biochem Pharmacol. 8, 1208–14. 10.1016/j.bcp.2013.08.001.
- Uteshev VV, 2014 The therapeutic promise of positive allosteric modulation of nicotinic receptors. Eur. J. Pharmacol 727, 181–185. 10.1016/j.ejphar.2014.01.072. [PubMed: 24530419]
- Vann R, Tobey K, Lobe S, Kipps B, Kwilasz A, Aceto M, Harris L, 2011 Varenicline does not alter brain stimulation reward thresholds and reverses nicotine-facilitated thresholds in rats. Drug Dev. Res 72, 310–314. 10.1007/s00213-012-2703-3.
- Walsh RM, Roh SH, Gharpure A, Morales-Perez CL, Teng J, Hibbs RE, 2018 Structural principles of distinct assemblies of the human α4β2 nicotinic receptor. Nature 557, 261–265. 10.1038/ s41586-018-0081-7. [PubMed: 29720657]
- Wang J, Kuryatov A, Sriram A, Jin Z, Kamenecka TM, Kenny PJ, Lindstrom J, 2015 An Accessory Agonist Binding Site Promotes Activation of alpha4beta2* Nicotinic Acetylcholine Receptors. J Biol Chem 290, 13907–13918. doi: 10.1074/jbc.M115.646786 [PubMed: 25869137]
- Wang J, Lindstrom J, 2018 Orthosteric and allosteric potentiation of heteromeric neuronal nicotinic acetylcholine receptors. Br J Pharmacol. 11, 1805–1821. 10.1111/bph.13745.
- Wang ZJ, Deba F, Mohamed TS, Chiara DC, Ramos K, Hamouda AK, 2017 Unraveling amino acid residues critical for allosteric potentiation of (alpha4)3(beta2)2-type nicotinic acetylcholine receptor responses. J Biol Chem. 292, 9988–10001. 10.1074/jbc.M116.771246. [PubMed: 28446611]

- Warpman U, Nordberg A, 1995 Epibatidine and ABT 418 reveal selective losses of α4β2 nicotinic receptors in Alzheimer brains. Neuroreport. 6, 2419–2423. 10.1097/00001756-199511270-00033 [PubMed: 8747166]
- Weggel LA, Pandya AA, 2019 Acute Administration of Desformylflustrabromine Relieves Chemically Induced Pain in CD-1 Mice. Molecules. 24, 944 10.3390/molecules24050944.
- Weltzin MM, Schulte MK, 2015 Desformylflustrabromine Modulates α4β2 Neuronal Nicotinic Acetylcholine Receptor High- and Low-Sensitivity Isoforms at Allosteric Clefts Containing the β2 Subunit. J Pharmacol Exp Ther. 354, 184–194. https://doi.org/0.1124/jpet.115.223933. [PubMed: 26025967]
- Weltzin MM, Huang Y, Schulte MK, 2014 Allosteric modulation of alpha4beta2 nicotinic acetylcholine receptors by HEPES. Eur J Pharmacol 732, 159–168. 10.1016/ j.ejphar.2012.06.001. [PubMed: 22732654]
- Wevers A, Monteggia L, Nowacki S, Bloch W, Schütz U, Lindstrom J, Pereira EF, Eisenberg H, Giacobini E, de Vos RA, Steur EN, Maelicke A, Albuquerque EX, Schröder H, 1999 Expression of nicotinic acetylcholine receptor subunits in the cerebral cortex in Alzheimer's disease: histotopographical correlation with amyloid plaques and hyperphosphorylated-tau protein. Eur J Neurosci. 11, 2551–65. 10.4103/1673-5374.147943. [PubMed: 10383644]
- Whiting P, Lindstrom J, 1987 Affinity labelling of neuronal acetylcholine receptors localizes acetylcholine-binding sites to their beta-subunits. FEBS Lett 213, 55–60. 10.1016/0014-5793(87)81464-3 [PubMed: 2435576]
- Williams DK, Wang J, Papke RL, 2011 Positive allosteric modulators as an approach to nicotinic acetylcholine receptor-targeted therapeutics: Advantages and limitations. Biochem. Pharmacol 82, 915–930. 10.1016/j.bcp.2011.05.001. [PubMed: 21575610]
- Young GT, Zwart R, Walker AS, Sher E, Millar NS, 2008 Potentiation of a7 nicotinic acetylcholine receptors via an allosteric transmembrane site. Proc. Natl. Acad. Sci. U.S.A 105, 14686–14691. 10.1073/pnas.0804372105. [PubMed: 18791069]
- Zoli M, Moretti M, Zanardi A, McIntosh JM, Clementi F, Gotti C, 2002 Identification of the nicotinic receptor subtypes expressed on dopaminergic terminals in the rat striatum. J Neurosci 22, 8785– 8789. 10.1523/JNEUROSCI.22-20-08785.2002 [PubMed: 12388584]
- Zwart R, Vijverberg HP, 1998 Four pharmacologically distinct subtypes of alpha4beta2 nicotinic acetylcholine receptor expressed in Xenopus laevis oocytes. Mol Pharmacol 54, 1124–1131. 10.1124/mol.54.6.1124. [PubMed: 9855643]

Highlights

• Antimicrobial mechanism of carvacrol against uropathogenic E. coli

- Carvacrol demonstrated membrane depolarization and oxidative burst in *E. coli*
- Carvacrol induced the release of DNA, proteins and ions from *E. coli* cells
- Carvacrol reduced the levels of inflammatory proteins COX-2 and iNOS.
- Carvacrol inhibited cell mortality and β -lactamase enzyme activity



Physostigmine



dFBr



Galantamine



LY2087101







NS9283

Figure 1. Structures of positive allosteric modulators of $\alpha 4\beta 2$ nAChR.

Table 1.

Positive allosteric modulators of $\alpha 4\beta 2$ nAChR and the work done thus far investigating them as novel therapeutics.

Compound	nAChR subtype activity in vitro	In vivo findings
Physostigmine	Full activity at $a4\beta2$, $a7$	Clinically used acetylcholinesterase inhibitor and antidot for anticholinergic toxicities.
Galantamine	Full activity at $\alpha 4\beta 2$, $\alpha 7$	-used clinically for the treatment of Alzheimer's Disease
		- Attenuates nicotine self-administration in rats
		- reduces cigarette smoking among alcohol-dependent patients
(dFBr)	Full activity at α4β2, partial activity at α7	-reduces nicotine self-administration in rats
		-blocks behavioral signs of nicotine withdrawal in mice
		- decreases anxiety-like behavior in the mouse marble burying assay, open-field assay
		- increases the anti-allodynic effects of nicotine in mouse CCI.
		- analgesic effects in formalin assay, and in the acetic acid writhing assay
NS9283	Selective activity at α4β2	-potentiates the effect of nicotine in the rat drug discrimination assay
		-acute and repeated dosing reduces nicotine self-administration in rats
		 improves social memory, task acquisition in the hippocampal-dependent spatial memory Morris Water Maze task, as well as attention performance in the five-choice serial reaction time task
		- Enhances analgesic efficacy of nAChR agonist in neuropathic pain model.
CMPI	Selective activity at $\alpha 4\beta 2$	N.A.
LY 2087101	Full activity at a4β2, a4β4, a7 subtypes	- does not potentiate the effect of nicotine in the mouse drug discrimination assay

* See article text under the corresponding compound for supporting references.